

# Pharmacokinetics of Dexamethasone in Tuberculous Meningitis

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**Background.** Dexamethasone is recommended as adjunctive therapy for tuberculous meningitis (TBM). Co-administration with rifampicin is expected to reduce dexamethasone exposure in TBM, an effect that may be more pronounced with the higher rifampicin doses currently being evaluated in clinical trials.

**Methods.** This pharmacokinetic study was nested in a randomized controlled trial comparing the safety of high-dose rifampicin (oral, 35 mg/kg; intravenous, 20 mg/kg) plus linezolid, with or without aspirin, versus standard-dose rifampicin (10 mg/kg) for adults with HIV-associated TBM. All participants received adjunctive oral dexamethasone every 12 hours starting at a dose of 0.4 mg/kg/day. Dexamethasone concentrations were measured on intensively sampled plasma on day 3 after study enrollment and analysed using nonlinear mixed-effects modeling.

**Results.** In total, 261 dexamethasone concentrations from 43 participants were available for model development. Eight (18%) participants were on efavirenz-based ART and five (11%) were on a lopinavir/ritonavir-based regimen. The median duration of rifampicin therapy at the time of pharmacokinetic sampling was 4 days (range: 0–7). Dexamethasone pharmacokinetics was best described by a one-compartment disposition model with first-order absorption and elimination. Typical oral clearance (CL/F) was 131 L/h, reduced to 11.5 L/h with concomitant lopinavir/ritonavir. High-dose rifampicin had no significant additional effect on dexamethasone pharmacokinetic parameters compared with the standard-dose.

**Conclusions.** In adults with HIV-associated TBM, there was high dexamethasone clearance, likely related to a drug-drug interaction with rifampicin. High-dose rifampicin had no additional effect on dexamethasone exposure.

**Keywords.** tuberculous meningitis; dexamethasone; pharmacokinetics; drug-drug interaction; rifampicin.

Tuberculous meningitis (TBM) is associated with high mortality, particularly among people with HIV, and survivors are often left with chronic neurological disability [1]. Disease severity in TBM is driven by intracerebral inflammation, caused by a dysregulated immune response to *Mycobacterium tuberculosis*. Host inflammation is modulated by dexamethasone, which confers modest survival benefit when provided with antituberculosis drugs in randomized controlled trials [2–4]. Adjunctive dexamethasone is therefore

recommended by international treatment guidelines for all adults with TBM.

Dexamethasone is a substrate of cytochrome P450 3A4 (CYP3A4) and P-glycoprotein (P-gp), making it susceptible to drug-drug interactions (DDI). Multiple doses of dexamethasone can also increase the transcription of both proteins [5]. Co-administration with rifampicin, the cornerstone of TBM treatment and a potent inducer of CYP3A4 and P-gp transporter activity, may reduce dexamethasone exposure [6, 7], potentially affecting anti-inflammatory efficacy. This DDI may be more pronounced with higher rifampicin doses ( $\geq 20$  mg/kg) currently being evaluated in TBM trials [8]. Additionally, anti-retroviral drugs may be CYP3A4 inhibitors or inducers, further increasing the risk for dexamethasone DDI in HIV-associated TBM.

We aimed to characterize the pharmacokinetics of dexamethasone co-administered with standard (10 mg/kg) or high-dose rifampicin (35 mg/kg) among adults with HIV-associated TBM, specifically to evaluate whether use of high-dose rifampicin reduced dexamethasone exposure in this population.

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## METHODS

### Study Design

This pharmacokinetic study was nested in the LASER-TBM trial (NCT03927313), which evaluated safety of intensified antituberculosis therapy among South African adults with HIV-associated TBM [9]. Participants were randomized within five days of starting TBM treatment to receive either the standard antituberculosis regimen containing rifampicin 10 mg/kg together with isoniazid, pyrazinamide, and ethambutol (R<sub>10</sub>HZE) or an experimental regimen containing high-dose rifampicin (oral 35 mg/kg or intravenous 20 mg/kg, determined by a second randomization) (R<sub>35</sub>HZE) and adding linezolid, with or without daily aspirin 1000 mg. Oral dexamethasone was administered to all participants every 12 hours for 4 weeks, starting at a dose of 0.4 mg/kg/day and tapering by 0.1 mg/kg per week, as recommended by national treatment guidelines. Experimental therapy was provided for 56 days, after which participants continued standard treatment.

### Pharmacokinetic Sampling

Plasma samples were collected on day 3 ( $\pm 2$  days) after study enrollment pre-dose, at 0.5, 1, 2, 3, 6, and 8–10 hours following the morning dose, and at 12 hours post-evening dose. Immediately following collection, samples were processed on-site and stored at  $-80^{\circ}\text{C}$ . Dexamethasone concentrations were quantified using a validated liquid chromatography-tandem mass spectrometry assay (lower limit of quantification [LLOQ]: 0.938 ng/mL) performed at the Division of Clinical Pharmacology, University of Cape Town (Supplementary material 1).

### Pharmacokinetic Modeling

Dexamethasone concentrations were described using nonlinear mixed-effects modeling in NONMEM v7.5.1 [10]. One- and two-compartment disposition models were tested with first-order absorption (with or without lag time or chain of transit compartments) and first-order elimination. Allometric scaling was applied for disposition parameters, testing body weight or fat-free mass (FFM) [11] as body size descriptors, with the exponents for clearance and volume fixed to 0.75 and 1, respectively [12]. Other covariates, including creatinine clearance (calculated using the Cockcroft-Gault formula [13]), age, treatment arm, time on rifampicin, and concomitant antiretroviral therapy (ART), were also assessed on pharmacokinetic parameters. Model development was guided by improvements in the objective function value ( $\Delta\text{OFV}$ ), goodness-of-fit plots, physiological plausibility and clinical relevance.

Random effects were included on the pharmacokinetic parameters if statistically significant, using a log-normal distribution [10]. Between-subject variability (BSV) was explored for disposition parameters, and between-occasion variability

(BOV) was explored for absorption parameters and bioavailability, with an occasion defined as a dosing event and its subsequent observations.

Residual unexplained variability was described using an error model with both additive and proportional components, with the additive component constrained to be at least 20% of the assay's LLOQ. Concentrations below the limit of quantification (BLQ) were handled using an adaptation of the M6 method proposed by Wijk et al. [14]. Details of population pharmacokinetic modeling and handling of missing covariate data are presented in Supplementary material 2.

The final population pharmacokinetics model was used to estimate dexamethasone 12-hour area under the curve ( $\text{AUC}_{0-12\text{h}}$ ) and maximum concentration ( $C_{\text{max}}$ ). Geometric mean ratios (GMR) for secondary pharmacokinetic parameters were computed for high-dose versus standard-dose rifampicin.

A post hoc power calculation showed that our study had  $>80\%$  power at an alpha of 0.05 to detect a 40% reduction in dexamethasone exposure due to high-dose rifampicin.

## RESULTS

### Study Data

Dexamethasone concentrations from 43 individuals were available, consisting of 261 observations after excluding samples taken 12 hours post-evening dose which could not be used because the exact dose timing was unknown (Supplementary Figure 1). 35/261 (13%) of the samples were below the assay's lower limit of quantification, most of which were pre-dose observations. The median duration of rifampicin therapy at the time of pharmacokinetic sampling was 4 days (range: 0–7). All participants were HIV-positive, 8 (18%) of whom were on efavirenz-based ART and 5 (11%) on lopinavir/ritonavir-based ART, provided at double the standard-dose (Table 1). Participants receiving high-dose rifampicin had lower body weight (57 kg; range 30–96) and thus received a lower total dexamethasone dose (9 mg; range 6–16) compared with those on standard-dose rifampicin (64 kg; range 42–107 and 12 mg; range 8–20, respectively) (Supplementary Figure 2). Other baseline characteristics were similar between the groups (Supplementary Table 1).

### Pharmacokinetic Modeling

Dexamethasone pharmacokinetics was best described by a one-compartment disposition model with first-order absorption and elimination. The effect of body size was best characterized using allometric scaling based on FFM ( $\Delta\text{OFV} = -17.7$ ), compared with total body weight ( $\Delta\text{OFV} = -12.3$ ). The typical subject (FFM: 45 kg) was estimated to have a volume of distribution (V/F) of 26.6 L and an oral clearance (CL/F) of 131 L/h, which was reduced to 11.5 L/h with concomitant lopinavir/ritonavir ( $\Delta\text{OFV} = -34.6$ , 1 degree of freedom,  $P < .001$ ) (Table 2). High-dose rifampicin had no significant effect on

**Table 1. Demographic and Clinical Characteristics**

	(n = 43)
Males	22 (52)
Female	21 (48)
Weight (kg)	60 (30–107)
Height (cm) <sup>a,c</sup>	160 (148–180) [15]
Fat-free mass (kg) <sup>b,c</sup>	45 (30–59)
Age (y)	39 (25–78)
Creatinine clearance (mL/min)	94 (53–148)
Days on rifampicin <sup>d</sup>	4 (0–7)
Antiretroviral therapy (ART)	
On ART	13 (30)
Efavirenz-based regimen	8 (18)
Lopinavir/ritonavir-based regimen	5 (11)
ART Naïve	19 (43)
Previous ART	11 (27)

Data is presented as median (range: min-max) or n (%). Numbers within square brackets indicate the count of IDs with missing values.

<sup>a</sup>Heights were missing in 26 participants. The missing heights were estimated using the provided details in the supplement S2.

<sup>b</sup>Fat-free mass was calculated by applying the formula from Janmahasatian et al [11].

<sup>c</sup>The reported median values along with the range (min—max) pertain exclusively to the non-missing data; the imputed values were not included in these calculations.

<sup>d</sup>The total number of days corresponds to the duration since the initiation of treatment, which commenced approximately 1–3 days before the start date for this study. Participants were assumed to be on a standard-dose (10 mg/kg) of rifampicin at the commencement of treatment and prior to study enrollment.

dexamethasone pharmacokinetic parameters compared with the standard-dose (CL/F:  $\Delta$ OFV =  $-1.54$ ,  $P > .05$ ; bioavailability:  $\Delta$ OFV =  $-0.09$ ,  $P > .05$ ) (Figure 1). No other covariates, including efavirenz and aspirin, significantly influenced dexamethasone pharmacokinetics. Goodness-of-fit plots for the final model are provided in Supplementary Figure 3.

The median dexamethasone AUC<sub>0–12h</sub> among participants not receiving lopinavir/ritonavir-based ART was 0.0864 mg·h/L (range: 0.0333–0.201 mg·h/L), following a median dose of 10.5 mg (range: 6–20 mg). Among those co-treated with lopinavir/ritonavir-based ART, the median AUC<sub>0–12h</sub> was approximately 12 times higher, at 1.06 mg·h/L (range: 0.591–2.51 mg·h/L), after a median dose of 10 mg (range: 8–16 mg) (Figure 2). Dexamethasone median AUC<sub>0–12h</sub> was 0.0841 mg·h/L (range: 0.0333–1.05) in the high-dose rifampicin group and 0.111 mg·h/L (range: 0.0364–2.51) in the standard-dose group, corresponding to a GMR of 0.80 (95% CI: 0.43–1.50). Median C<sub>max</sub> was 0.0275 mg/L (range: 0.0113–0.128) and 0.0347 mg/L (range: 0.00998–0.314) in the high-dose and standard-dose groups, respectively (GMR: 0.77; 95% CI: 0.47–1.25) (Figure 3). These differences are in line with the lower median total dexamethasone dose in the high-dose rifampicin group compared with the standard-dose group (GMR: 0.83; 95% CI: 0.72–1.00).

## DISCUSSION

We characterized the pharmacokinetics of oral dexamethasone among adults with HIV-associated TBM receiving rifampicin-

**Table 2. Final Pharmacokinetic Parameter Estimates for Dexamethasone**

Parameter (units)	Typical Values (95% CI) <sup>a</sup>
Clearance (L/h) <sup>b</sup>	131 (108–148)
Clearance with lopinavir/ritonavir-based ART (L/h)	11.5 (6.15–20.2)
Volume of distribution (L) <sup>b</sup>	26.6 (17.9–41.0)
Bioavailability (.)	1 Fixed
First-order absorption rate constant (h <sup>-1</sup> )	0.397 (0.359–0.437)
Between-subject variability in clearance (%)	61.8 (45.9–81.6)
Between-occasion variability in bioavailability	43.2 (25.7–55.4)
Between-occasion variability in first-order absorption rate constant (%)	27.0 (17.9–35.1)
Proportional error (%)	33.4 (31.0–40.5)
Additive error (x10 <sup>-3</sup> mg/L) <sup>c</sup>	0.180 Fixed

<sup>a</sup>Values in parentheses represent the 95% confidence interval, computed using sampling importance resampling (SIR) on the final model.

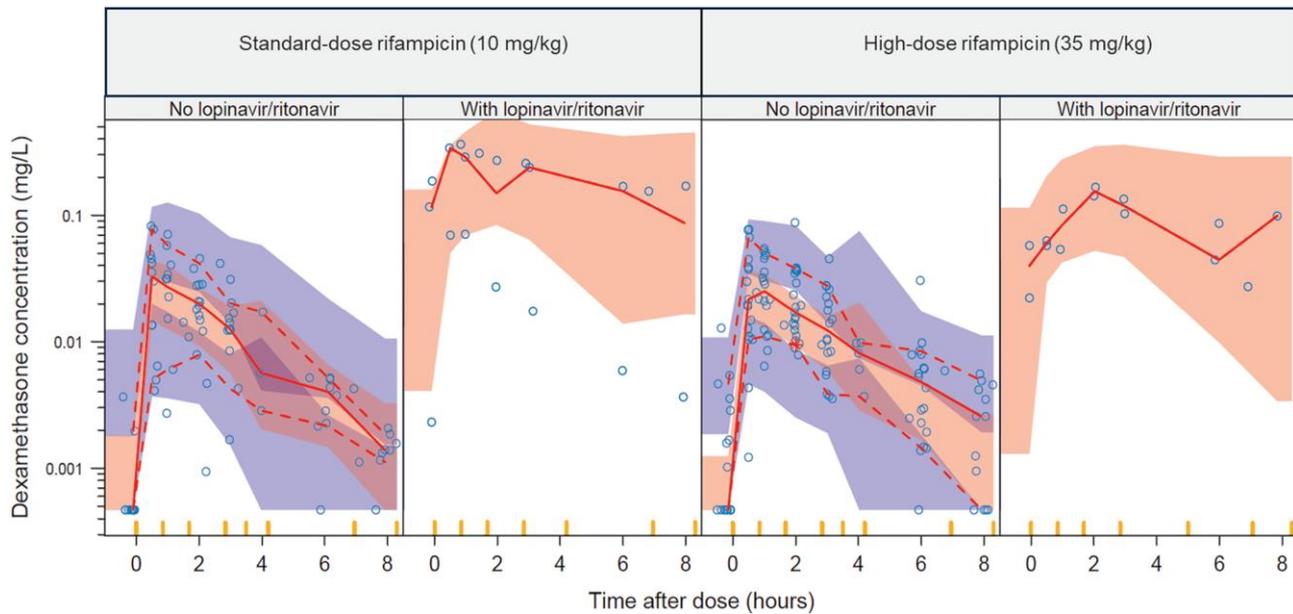
<sup>b</sup>All the disposition parameters were allometrically scaled using fat-free mass (FFM). The reported typical values refer to the typical individual in the cohort with FFM of 45 kg.

<sup>c</sup>The estimate of the additive component of the residual unexplained variability did not significantly differ from its lower boundary of 20% of LLOQ, it was consequently fixed to this value.

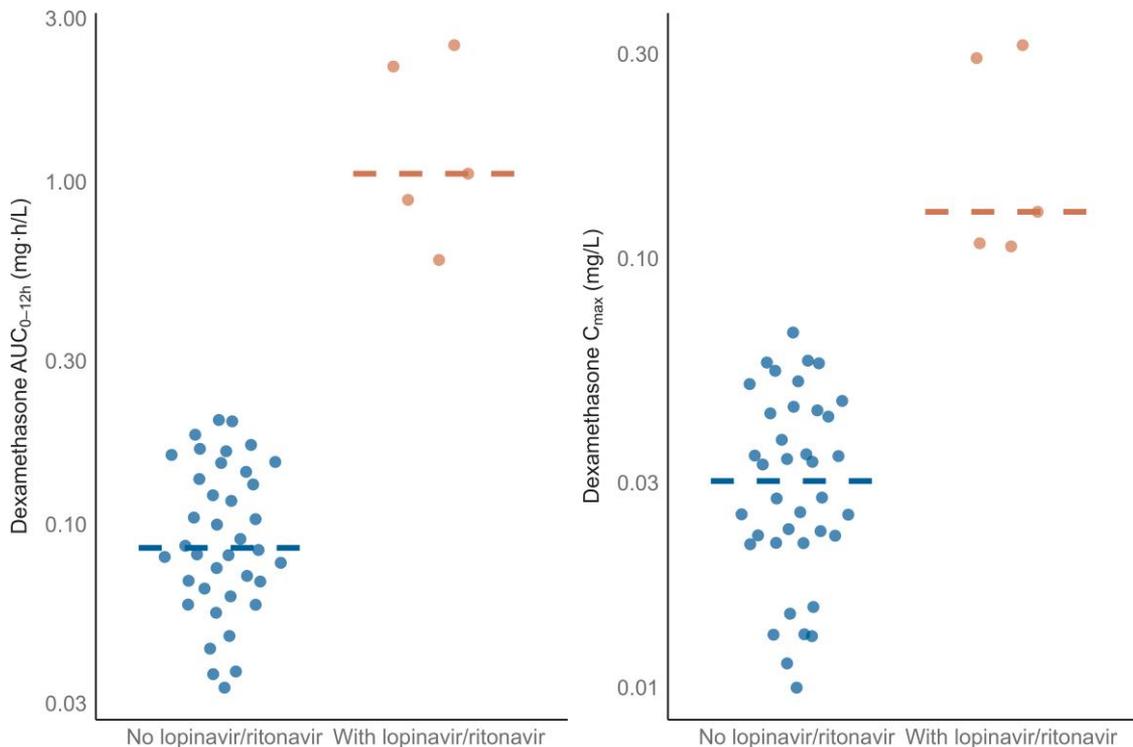
based treatment. In this clinical trial population, receipt of higher rifampicin doses had no additional effect on dexamethasone pharmacokinetics, and dexamethasone exposure was similar among participants receiving high-dose rifampicin (35 mg/kg) compared with standard dose rifampicin (10 mg/kg). By contrast, there was a strong inhibitory effect from lopinavir/ritonavir-based ART on dexamethasone clearance.

Limited data exist regarding dexamethasone pharmacokinetics in TBM patients, as well as its interactions with antituberculosis and antiretroviral drugs. Pharmacokinetic studies in healthy individuals receiving dexamethasone as monotherapy have reported CL/F values of 15.6 L/h following oral administration of 1.5 mg [5], and 18.1 L/h following an average intravenous dose of 5.7 mg [16]. We observed substantially higher dexamethasone CL/F of 131 L/h in our cohort of South African patients with HIV-associated TBM. This value exceeds hepatic blood flow (~90 L/h) [17], indicating low bioavailability from significant first-pass metabolism [18].

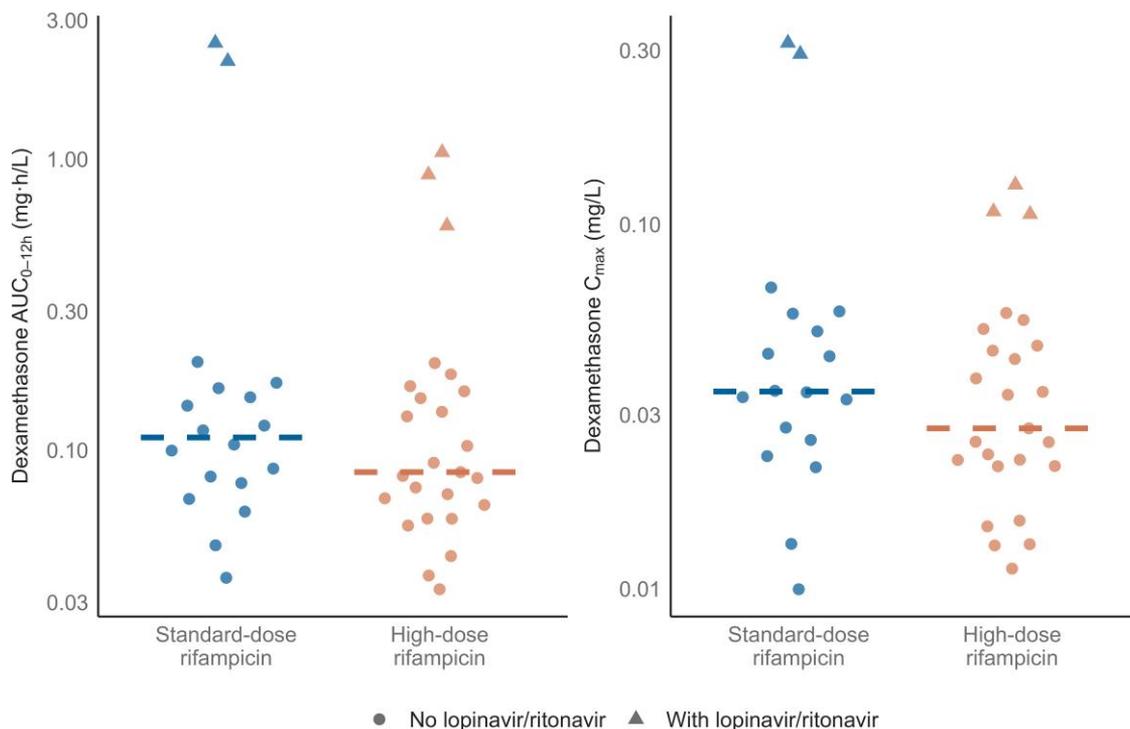
A plausible explanation for the large dexamethasone CL/F observed in our study is a DDI with rifampicin via induction of CYP3A4 and increased expression of P-gp, enhancing dexamethasone clearance and reducing its bioavailability [19]. Previous reports support this hypothesis. A Japanese cohort (n = 27) involving TB patients and healthy volunteers receiving intravenous dexamethasone reported a 5-fold increase in clearance among individuals co-administered rifampicin compared with those receiving dexamethasone alone [7]. Similarly, another study involving healthy volunteers (n = 16) found that, following oral administration, dexamethasone concentrations measured at 8-hours post-dose were 5- to 10-fold lower in participants co-treated with rifampicin compared with those not receiving rifampicin [6].



**Figure 1.** Visual predictive check of dexamethasone concentrations versus time after dose, stratified by co-administered rifampicin dose level and further stratified by lopinavir/ritonavir co-administration. Circles represent observed data. Solid and dashed lines indicate the 50th, 10th, and 90th percentiles of the observed data, while the shaded areas represent the 95% model-predicted confidence intervals for the same percentiles.



**Figure 2.** Area under the concentration–time curve from 0 to 12 h post-dose (AUC<sub>0–12h</sub>) and maximum concentration (C<sub>max</sub>), stratified by lopinavir/ritonavir co-administration. Dots represent individual values; lines indicate the median.



**Figure 3.** Area under the concentration–time curve from 0 to 12 h post-dose (AUC<sub>0-12h</sub>) and maximum concentration (C<sub>max</sub>), stratified by rifampicin dose level. Dots represent individual values; lines indicate the median.

Indirect confirmation of the DDI with rifampicin is the much higher dexamethasone exposure observed among participants co-treated with lopinavir/ritonavir-based ART, likely due to a DDI with ritonavir, a potent CYP3A4 inhibitor [20]. Dexamethasone CL/F in this subgroup (11.5 L/h) is consistent with previous reports of dexamethasone administered alone [5], suggesting that the potent inhibitory effect of ritonavir counteracts rifampicin induction.

There is an established relationship between rifampicin concentration and CYP3A4 induction. This has been demonstrated in vitro, where a concentration-dependent effect from rifampicin on CYP3A4 induction was shown in liver cell lines [21], and among pulmonary TB patients ( $n = 24$ ), where higher doses of rifampicin (40 mg/kg) reduced exposure of the CYP3A4 probe drug midazolam by 38% compared with standard rifampicin doses (10 mg/kg) [8]. Our study had >80% power to detect such a drop in dexamethasone exposure due to high-dose rifampicin, but no trend towards faster CL/F in this group was visible in our data.

An explanation for lack of observed effect in our study is that differences in probe substrate exposure between high-dose and standard-dose rifampicin are minor compared with the large overall effect of rifampicin itself (vs no rifampicin) [22]. Illustrating this, the mean oral midazolam exposure after a single 15 mg dose without rifampicin is 170  $\mu\text{g}\cdot\text{h/L}$  [22] and, in the study of pulmonary TB patients, was reduced by 95.8% (to

7.10  $\mu\text{g}\cdot\text{h/L}$ ) with 10 mg/kg rifampicin and by 97.4% (to 4.4  $\mu\text{g}\cdot\text{h/L}$ ) with 40 mg/kg rifampicin [8]. This may indicate that the small additional effect of higher rifampicin doses on CYP3A4 induction does not necessarily translate into clinically significant lower dexamethasone exposures, as demonstrated in our study. The slightly lower exposures we observed in the high-dose group were attributable to receipt of lower total dexamethasone doses rather than a DDI effect from high-dose rifampicin.

Our analysis, and the probe substrate data, suggests that CYP3A4 induction from rifampicin significantly reduces dexamethasone exposure, which could potentially influence its efficacy. The recommended dexamethasone dose was established from a randomized controlled trial ( $n = 545$ ) conducted in Vietnam, which demonstrated a modest survival benefit among predominantly HIV-negative patients receiving rifampicin-based TBM treatment [2]. Dexamethasone was administered intravenously in that trial, at the same dose used for oral administration in our study. Oral dexamethasone has a bioavailability of approximately 70%–80% [23] resulting in lower systemic concentrations compared with intravenous dosing; this reduced bioavailability could be exacerbated because rifampicin induction is likely to have a more pronounced effect on oral dexamethasone due to extensive CYP3A4-mediated first-pass metabolism [24]. Therefore, providing oral dexamethasone alongside rifampicin to TBM patients may achieve

substantially lower dexamethasone exposures than those in the Vietnam trial, with potentially reduced clinical benefit.

A more recent trial found a smaller, non-significant effect from adjunctive intravenous dexamethasone on survival among patients with HIV-associated TBM in Vietnam and Indonesia ( $n = 520$ ) [3]. The reason for the reduced efficacy in this population is uncertain. It may be related to secular trends in standard of care which had better outcomes compared with the original trial, reducing the relative effect from dexamethasone on survival. Another potential explanation is a DDI with efavirenz, a CYP3A4 inducer [25], resulting in increased dexamethasone clearance, as around half of participants in the dexamethasone arm (104/263) were on efavirenz-based ART [3]. In our study no differences in dexamethasone pharmacokinetics were observed among the eight participants on efavirenz-based ART, although this small sample size may have limited power to detect an effect. Additionally, since the enzyme induction pathway of efavirenz largely overlaps with that of rifampicin via activation of the pregnane X receptor [25], it is possible that no discernible effect of efavirenz was observed because participants had already been exposed to rifampicin for a median of 4 days by the time of the pharmacokinetic visit.

Our study has some limitations. First, we were unable to directly evaluate the DDI between rifampicin and dexamethasone in this population because there was no comparator group without rifampicin. However, the inhibitory effect of lopinavir/ritonavir—resulting in dexamethasone clearance values comparable to those reported historically for dexamethasone administered alone—provides indirect evidence of a DDI with rifampicin. Second, measurement of dexamethasone concentrations occurred after a median of 4 days of rifampicin exposure, prior to the full induction effect on CYP3A4, expected after about 1 week [26]. This may lead to underestimation of a DDI effect from high-dose rifampicin, which may be more pronounced after more prolonged coadministration. However, later impact on dexamethasone exposure is difficult to predict because of interindividual variability in CYP3A4 activity and dynamic rifampicin exposure from autoinduction [15]. Finally, since all patients in our study received dexamethasone orally, we were unable to estimate dexamethasone bioavailability and separate the potentially different effects of rifampicin induction on first-pass metabolism and systemic clearance.

In summary, our analysis suggests that CYP3A4 induction from rifampicin significantly reduces oral dexamethasone exposure, which could potentially influence its efficacy. Although the clinical implications are unclear—given that the target exposure for dexamethasone in TBM is unknown—prior knowledge of CYP3A4 substrate metabolism suggests that the reduction in dexamethasone exposure with oral dosing is much more pronounced than with intravenous dosing. The

similar dexamethasone exposures observed between standard- and high-dose rifampicin in our study reduce the likelihood of potential harm from high-dose rifampicin in TBM due to reduced dexamethasone exposure and, consequently, a reduced anti-inflammatory effect.

### Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

### Notes

**Author Contributions.** S. W. conceptualized the study with methodology input from P. D., A. D. and C. S. collected the data, provided by S. W., G. M. and R. J. W., L. W. did the drug concentration assays. J. M. C., J. E. R.-G., and N. A. did the analysis and visualization, supervised by P. D., J. M. C. and S. W. wrote the original draft; all co-authors reviewed and edited the manuscript. All authors approved the final draft of the manuscript. The corresponding author had full access to all the data in the study and had final responsibility for the decision to submit for publication.

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**Potential conflicts of interest.** All authors declare no conflict of interest. The authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest.

**Data Availability.** The data and model codes supporting the findings of this study are available from the corresponding author, S. W., upon reasonable request.

### References

- Dodd PJ, Osman M, Cresswell FV, et al. The global burden of tuberculous meningitis in adults: a modelling study. *PLOS Global Public Health* 2021; 1:e0000069.
- Thwaites GE, Bang ND, Dung NH, et al. Dexamethasone for the treatment of tuberculous meningitis in adolescents and adults. *N Engl J Med* 2004; 351:1741–51.

3. Donovan J, Bang ND, Imran D, et al. Adjunctive dexamethasone for tuberculous meningitis in HIV-positive adults. *N Engl J Med* **2023**; 389:1357–67.
4. Prasad K, Singh MB, Ryan H. Corticosteroids for managing tuberculous meningitis. *Cochrane Database Syst Rev* **2016**; 4:CD002244.
5. Loew D, Schuster O, Graul EH. Dose-dependent pharmacokinetics of dexamethasone. *Eur J Clin Pharmacol* **1986**; 30:225–30.
6. Kyriazopoulou V, Vagenakis AG. Abnormal overnight dexamethasone suppression test in subjects receiving rifampicin therapy. *J Clin Endocrinol Metab* **1992**; 75:315–7.
7. Kawai S. A comparative study of the accelerated metabolism of cortisol, prednisolone and dexamethasone in patients under rifampicin therapy. *Folia Endocrinol Japonica* **1985**; 61:145–61.
8. Stemkens R, de Jager V, Dawson R, et al. Drug interaction potential of high-dose rifampicin in patients with pulmonary tuberculosis. *Antimicrob Agents Chemother* **2023**; 67:e0068323.
9. Davis AG, Wasserman S, Stek C, et al. A phase 2A trial of the safety and tolerability of increased dose rifampicin and adjunctive linezolid, with or without aspirin, for human immunodeficiency virus–associated tuberculous meningitis: the LASER-TBM trial. *Clin Infect Dis* **2023**; 76:1412–22.
10. Mould DR, Upton RN. Basic concepts in population modeling, simulation, and model-based drug development—part 2: introduction to pharmacokinetic modeling methods. *CPT Pharmacomet Syst Pharmacol* **2013**; 2:38.
11. Janmahasatian S, Duffull SB, Ash S, Ward LC, Byrne NM, Green B. Quantification of lean bodyweight. *Clin Pharmacokinet* **2005**; 44:1051–65.
12. Anderson BJ, Holford NHG. Mechanism-based concepts of size and maturity in pharmacokinetics. *Annu Rev Pharmacol Toxicol* **2008**; 48:303–32.
13. Cockcroft DW, Gault H. Prediction of creatinine clearance from Serum creatinine. *Nephron* **1976**; 16:31–41.
14. Wijk M, Wasmann RE, Jacobson KR, Svensson EM, Denti P. A pragmatic approach to handling censored data below the lower limit of quantification in pharmacokinetic modeling. *CPT Pharmacomet Syst Pharmacol* **2025**; 14:1042–9.
15. Svensson RJ, Aarnoutse RE, Diacon AH, et al. A population pharmacokinetic model incorporating saturable pharmacokinetics and autoinduction for high rifampicin doses. *Clin Pharmacol Ther* **2018**; 103:674–83.
16. Hong Y, Mager DE, Blum RA, Jusko WJ. Population pharmacokinetic/pharmacodynamic modeling of systemic corticosteroid inhibition of whole blood lymphocytes: modeling interoccasion pharmacodynamic variability. *Pharm Res* **2007**; 24:1088–97.
17. Pang KS, Rowland M. Hepatic clearance of drugs. I. Theoretical considerations of a “well-stirred” model and a “parallel tube” model. Influence of hepatic blood flow, plasma and blood cell binding, and the hepatocellular enzymatic activity on hepatic drug clearance. *J Pharmacokinet Biopharm* **1977**; 5:625–53.
18. Thummel KE, O’Shea D, et al. Oral first-pass elimination of midazolam involves both gastrointestinal and hepatic CYP3A-mediated metabolism\*. *Clin Pharmacol Ther* **1996**; 59:491–502.
19. Jacobs TG, Marzolini C, Back DJ, Burger DM. Dexamethasone is a dose-dependent perpetrator of drug–drug interactions: implications for use in people living with HIV. *J Antimicrob Chemother* **2022**; 77:568–73.
20. Katzenmaier S, Markert C, Riedel K-D, Burhenne J, Haefeli WE, Mikus G. Determining the time course of CYP3A inhibition by potent reversible and irreversible CYP3A inhibitors using a limited sampling strategy. *Clin Pharmacol Ther* **2011**; 90:666–73.
21. Williamson B, Dooley KE, Zhang Y, Back DJ, Owen A. Induction of influx and efflux transporters and cytochrome P450 3A4 in primary human hepatocytes by rifampin, rifabutin, and rifapentine. *Antimicrob Agents Chemother* **2013**; 57:6366–9.
22. Backman JT, Olkkola KT, Neuvonen PJ. Rifampin drastically reduces plasma concentrations and effects of oral midazolam. *Clin Pharmacol Ther* **1996**; 59:7–13.
23. Duggan DE, Yeh KC, Matalia N, Ditzler CA, McMahon FG. Bioavailability of oral dexamethasone. *Clin Pharmacol Ther* **1975**; 18:205–9.
24. Thummel KE. Gut instincts: CYP3A4 and intestinal drug metabolism. *J Clin Invest* **2007**; 117:3173–6.
25. Hariparsad N, Nallani SC, Sane RS, Buckley DJ, Buckley AR, Desai PB. Induction of CYP3A4 by efavirenz in primary human hepatocytes: comparison with rifampin and phenobarbital. *J Clin Pharmacol* **2004**; 44:1273–81.
26. Niemi M, Backman JT, Fromm MF, Neuvonen PJ, Kivist KT. Pharmacokinetic interactions with rifampicin. *Clin Pharmacokinet* **2003**; 42:819–50.