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# Simultaneous carriage of multiple serotypes of Group B *Streptococcus*: Systematic review and meta-analysis



Vaccine



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#### ABSTRACT

*Background:* Epidemiological studies evaluating the distribution of Group B *Streptococcus* (GBS) serotypes are crucial for serotype-specific vaccine development and post-licensure surveillance. However, there is a paucity of data about the prevalence of simultaneous carriage of multiple serotypes.

*Methods:* We conducted a systematic review of three databases (Medline, Embase, PubMed) to identify studies reporting GBS serotype co-carriage at the same anatomical site (multiple serotypes in one sample) or different anatomical sites (paired samples from one individual with different serotypes). We conducted a random-effects meta-analysis to evaluate the prevalence of co-carriage.

*Results:* 18 articles met the inclusion criteria, representing at least 12,968 samples from 14 countries. In a random-effects meta-analysis, we identified that 10 % (95 % CI: 4–19) of the positive samples taken from one anatomical site have more than one serotype, and 11 % (95 % CI: 5–20) of positive participants with samples taken from two anatomical sites carried different serotypes. When reported, the number of serotypes simultaneously carried ranged from 1 to 4. The serotypes most often associated with co-carriage are III (20.3 %), V (20.3 %) and Ia (19.5 %).

*Conclusion:* This systematic review demonstrates that co-carriage is a minor but definite phenomenon, but the data are too limited to give a precise picture of the current epidemiology. Co-colonisation detection needs to be taken into consideration in the design and methods of future GBS carriage surveillance studies to estimate and evaluate the potential for serotype replacement once vaccines are introduced. © 2022 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/bu/4.0/)

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Review

#### 1. Introduction

Streptococcus agalactiae - also known as Group B Streptococcus (GBS) - is a commensal bacterium of the intestinal and genital flora, occasionally found in the throat and urethra. GBS is a leading cause of mortality and morbidity among neonates and young infants [1]. Thanks to the introduction of intrapartum antibiotic prophylaxis (IAP), the incidence of GBS disease has been much reduced. However, IAP coverage is not optimal, even in good screening settings, and it has no impact on preterm, stillbirths and acquisition of GBS after the first few days of life [2]. The bacterium may also cause invasive disease in pregnant women, the elderly, immunocompromised individuals, and adults with underlying health conditions [2]. Together with the current efforts to control the emergence of antimicrobial resistance, these are strong motivators to find another solution to combat GBS disease that does not include antibiotics [2]. One such solution is immunisation of pregnant women. Several maternal vaccines are currently under development. The most advanced candidate targets the capsular polysaccharide antigen of six of the ten known serotypes [3].

Epidemiological studies evaluating the worldwide distribution of GBS serotypes are crucial for serotype-specific vaccine development and post-licensure surveillance [2]. However, most studies serotype a single colony per clinical sample, which may introduce a bias towards the predominant and easiest-to-pick isolate rather than giving details about all the potential carried isolates [4,5]. There is a paucity of data about the prevalence of the carriage of multiple serotypes simultaneously. Knowledge about the possibility and frequency of co-carriage is needed to predict the risk of serotype replacement and potential horizontal gene transfer. This is specifically important for the genes leading to capsular switching, as a capsular polysaccharide vaccine might put a selective pressure on virulent strains to evade vaccine coverage [6,7]. Both phenomena have been observed after the introduction of the pneumococcal capsular polysaccharide vaccine [8].

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In this regard, we undertook a systematic review of human GBS co-carriage, defined as the simultaneous carriage of multiple sero-types of GBS at one or multiple anatomical sites of a human individual, in the published literature up to November 2021.



Fig. 1. Flow diagram of the data search and included studies.

#### Table 1

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Abstracted data from the included studies. *Total samples or paired samples* represents the number of samples (or participants, if the co-carriage has only been reported per participant) or pairs of samples taken from the population and tested for GBS carriage. In the case of same sample analysis, anatomical sites separated by commas indicate that all these sites were serotyped for multiple carriage, while anatomical sites separated by OR indicate that co-carriage was reported per participant without disclosing which sample was concerned. *Total positive samples or paired samples* represents either the number of positive samples/participants (with the possibility of including multiple samples from the same individual at the same or different visits) or the number of pairs of samples from the same individual, with at least one positive sample and all of the positive samples serotyped (with the possibility of including the same sample in multiple pairs within the same individual if more than two anatomical sites are investigated). NR: Non-reported; yr: years.

1	Study	Country of collection	Year of collection	Type of co- carriage	Population (age)	Anatomical site(s)	Total samples or paired samples	Total positive samples or paired samples	Co- carriage events	Bias assessment score
2	Anthony et al., 1978 [13]	The USA	1973– 1976	paired samples	Pregnant women (13-44 yr)	cervix, urethra	1488	NR	5	5.5/9
3	Anthony et al., 1981 [14]	The USA	1979– 1980	paired samples	Pregnant women (NR)	genitals, rectum	295	64	1	5.5/9
4	Anthony et al., 1981 [14]	The USA	1979– 1980	paired samples	Pregnant women (NR)	rectum, stool	135	33	1	5.5/9
5	Anthony et al., 1981[14]	The USA	1979– 1980	paired samples	Pregnant women (NR)	genitals, stool	135	37	1	5.5/9
6	Ferrieri et al., 2004 [15]	The USA	1998– 2000	paired samples	Non-pregnant women (18– 30 yr)	vagina, rectum	NR	102	18	4/6
7	Whitney et al., 2004 [16]	Thailand, The Philippines, Zimbabwe, Myanmar, Ireland, the USA	1999– 2001	paired samples	Pregnant women (23–31 yr)	cervix, vagina, urine	1308	128	1	6.5/9
8	Taylor et al., 2007 [17]	Australia	2003– 2005	paired samples	Pregnant and non-pregnant women (18–50 yr)	vagina, anus	374	70	12	5.5/9
9	El Aila et al., 2009 [18]	Belgium	2007	paired samples	Pregnant women (NR)	vagina, rectum	150	36	4	3.5/6
10	Palmeiro et al., 2010 [19]	Brazil	2006– 2008	paired samples	Pregnant women and healthy patients (0- >64 yr)	rectum, urethra	NR	NR	1	3/6
11	Slotved et al., 2017 [20]	Ghana	2012– 2013	paired samples	Pregnant women (<20–>30 yr)	vagina, rectum	400	107	1	8/9
12	To et al., 2021 [5]	The Gambia	2014	paired samples	Women post-delivery (>18 yr) and infants (0- 89 days)	rectovaginal, breastmilk, nasopharyngeal, rectal	NR	NR	12	4/6
13	Furfaro et al., 2019 [21]	Australia	2015– 2017	paired samples	Pregnant women (16–50 yr)	vagina, rectum	1381	337	35	8/9
14	Jisuvei et al., 2020 [22]	Kenya	2017	paired samples	Pregnant women (<25->36)	vagina, rectum	288	53	30	7.5/9
15	Maurer et al., 1979[23]	The USA	NR	paired samples	Children (0–14 yr)	throat, anus, vagina	415	47	2	6.5/9
16	Hoogkamp-Korstanje et al., 1982 [24]	The Netherlands	NR	paired samples	Pregnant women (NR)	vagina, cervix, rectum	762	106	24	3/9
17	Anthony et al., 1978 [13]	The USA	1973– 1976	same sample	Pregnant women (13–44 yr)	cervix OR urethra	1488	NR	4	5.5/9
18	Anthony et al., 1981 [14]	The USA	1979– 1980	same sample	Pregnant women (NR)	stool, rectal, genitals	743	134	2	5.5/9
19	Ferrieri et al., 2004 [15]	The USA	1998– 2000	same sample	Non-pregnant women (18– 30 yr)	vagina OR rectum	NR	102	4	4/6
20	Taylor et al., 2007 [17]	Australia	2003– 2005	same sample	Pregnant and non-pregnant women (18–50 yr)	vagina, anus	374	92	15	5.5/9
21	El Aila et al., 2009 [18]	Belgium	2007	same sample	Pregnant women (NR)	vagina OR rectum	150	36	11	3.5/6
22	Khatami et al., 2019 [4]	The USA	2010– 2012	same sample	Non-pregnant women (18- 55 yr)	vagina	433	91	6	4/6
23	Foster-Nyarko et al., 2016 [25]	The Gambia	2011– 2012	same sample	Infants (2 months)	nasopharynx	1170	NR	2	6.5/9
24	Jisuvei et al., 2020 [22]	Kenya	2017	same sample	Pregnant women (<25->36)	vagina OR rectum	292	53	7	7.5/9
25	Baker et al., 1976 [26]	The USA	NR	same sample	Non-pregnant women (NR)	vagina	210	79	4	3.5/6
26	Pérez-Ruiz et al., 2004 [27]	Spain	2001– 2002	same (including rectovaginal)	Pregnant women (NR)	vagina-rectum	NR	30	1	3/6
27	To et al., 2021 [5]	The Gambia	2014	same (including rectovaginal)	Women post-delivery (>18 yr) and infants (0– 89 days)	rectovaginal, breastmilk, nasopharyngeal, rectal	NR	96	31	4/6
28	Foxman et al., 2006 [1]	The USA	2001	unclear	Young adults (17–28 yr)	urine, rectum, vagina	977	NR	1	5.5/9

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#### 2. Methods

#### 2.1. Definitions

Studies reporting at least one case of serotype co-carriage, defined as the simultaneous carriage of multiple serotypes of GBS at one or multiple anatomical sites of a human individual within a population of asymptomatic GBS-positive individuals were included, irrespectively of the sample type, culture, serotyping techniques, population type and size.

#### 2.2. Search strategy

The published literature dated from 1946 up to the 2nd of November 2021 was searched using the Medline (1946–2021), Embase (1974–2021), and PubMed (1976–2021) databases with detailed search terms (Supplementary Material S1). The relevant articles were searched using snowballing techniques to identify additional related references. Abstracts were screened using the Rayyan software [9] by answering sequentially the questions (1) and (2), full-text articles were then screened to answer (3) and (4).

- (1) Is the study about GBS?
- (2) Does the study investigate multiple clinical samples from a population carrying GBS asymptomatically?
- (3) Are co-carriage of multiple strains or multiple colony-picks mentioned in the study?
- (4) Are the strains serotyped?

Data from the published studies and correspondence with their authors were abstracted into an Excel sheet by two independent reviewers (CB and MS), disagreements were resolved through discussion and with other reviewers (KLD and SL). Data are reported using the PRISMA guidelines [10].

#### 2.3. Quality assessment

Each study was scored independently by two reviewers (CB and MS) according to questions adapted from the Joanna Briggs Institute (JBI) Critical Appraisal Checklist for Prevalence Studies [11]. Questions 1, 2 and 9 were not relevant for studies whose primary aim was not prevalence; thus, these studies were scored out of six while prevalence studies were scored out of nine. Each positive answer scores one point. Unclear or negative answers score zero points.

## 2.4. Data analysis

The data for the prevalence of co-carriage with two, three or four serotypes were collected from studies designed to identify more than two serotypes. Studies with more than two colony picks or reporting more than two serotypes carriage were included. The data for the serotype distribution in co-carriage events were collected from the studies giving a detailed composition of each combination. The data for the meta-analyses of co-carriage prevalence were collected from studies whose design could have had identified co-carriage at the same or different anatomical site. The data were analysed in RStudio 1.4.1106 with the *meta* 4.19–1 and *metafor* 3.0–2 packages. After double-arcsine transformation, a random-effects model with Der Simonian and Laired method was conducted to weigh the proportions, as described elsewhere [12]. Results are reported as means with 95 % confidence intervals.

#### 3. Results

#### 3.1. General characteristics

Out of the 79 identified studies, 18 met the inclusion criteria (Fig. 1), representing more than 12,968 samples from various populations, including pregnant and non-pregnant women, neonates, children, female and male adults and from 14 different countries, screened between 1973 and 2017 (Table 1). Each study was assessed for bias. The scores are reported in Table 1, and the details of each score are reported in Supplementary Material S2. The scores rank from 3 to 4 out of 6 for studies whose primary endpoint was not prevalence and from 5.5 to 8 out of 9 for prevalence studies (Table 1). The main weaknesses were low sample size and suboptimal serotyping methods, as not all ten serotypes were tested (Supplementary Material S2).

#### 3.2. Number of co-carried isolates

The number of co-carried serotypes observed goes from two to four. However, some studies only refer to "more than one" or "different" serotypes in the same sample or individual. In the studies designed to identify more than two serotypes, co-carriage of two serotypes is more common than three or four (Table 2). One study found that three colony picks enable the accurate identification of all serotypes present for 91.1 % of the screened samples [15].

# 3.3. Prevalence of co-carriage at the same and different anatomical sites

In a random-effects meta-analysis of the studies reporting the incidence of co-carriage, 10 % (95 % CI: 4–19) of the positive samples/participants had more than one serotype at the same anatomical site. 11 % (95 % CI: 5–20) of participants with samples taken from two anatomical sites and at least one of these samples being positive carried different serotypes (Fig. 2A). The meta-analyses of same site co-carriage in pregnant women versus non-pregnant

#### Table 2

Prevalence of co-carriage with two, three or four serotypes in studies designed to identify more than two serotypes. Studies with more than two colony picks or reporting more than two serotypes carriage were included. NR: Non-reported.

1	Study	Colony picks (average)	Samples with 2 serotypes (%)	Samples with 3 serotypes (%)	Samples with 4 serotypes (%)	Total positive samples analysed
2	Baker et al., 1976	5	4 (5)	0 (0)	0 (0)	79
3	Ferrieri et al., 2004	10	20 (20)	2 (2)	0 (0)	102
4	Khatami et al., 2019	NR	5 (5)	1(1)	0 (0)	91
5	Pérez-Ruiz et al.,	15	1 (3)	0 (0)	0 (0)	30
	2004					
6	Taylor, 2006	NR	13 (14)	2 (2)	0 (0)	92
7	To et al., 2021	10	18 (19)	4 (4)	1 (1)	96

95%-CI Weight

#### А Studies with different sites co-carriage N



#### Studies with same site co-carriage N

Anthony et al. 1981 Perez-Ruiz et al., 2004 Ferrieri et al., 2004 Baker et al., 1976 Khatami et al., 2019 Jisuvei et al., 2020 Taylor, 2007 El Aila et al., 2009 To et al., 2021

Random effects model

Heterogeneity:  $I^2 = 89\%$ , p < 0.01



0.01	[0.00; 0.05]	11.9%
0.03	[0.00; 0.17]	9.7%
0.04	[0.01; 0.10]	11.6%
0.05	[0.01; 0.12]	11.3%
0.07	[0.02; 0.14]	11.5%
0.13	[0.05; 0.25]	10.8%
0.16	[0.09; 0.25]	11.5%
0.31	[0.16; 0.48]	10.1%
0.32	[0.23; 0.43]	11.6%

0.10 [0.04; 0.19] 100.0%

Proportion

#### В Studies with same site co-carriage in non-pregnant women N Proportion 95%-CI Weight Ferrieri et al., 2004 102 0.04 [0.01; 0.10] 37.5% 0.05 [0.01; 0.12] Baker et al., 1976 79 29.1% Khatami et al., 2019 91 0.07 [0.02; 0.14] 33.5% Random effects model 0.05 [0.03; 0.08] 100.0% Heterogeneity: $I^2 = 0\%$ , p = 0.720.02 0.04 0.06 0.08 0.1 0.12 Studies with same site co-carriage in pregnant women Ν Proportion 95%-CI Weight 0.01 [0.00; 0.05] Anthony et al. 1981 134 27.2% Perez-Ruiz et al., 2004 30 0.03 [0.00: 0.17] 23.4% Jisuvei et al., 2020 53 0.13 [0.05; 0.25] 25.3% El Aila et al., 2009 36 0.31 [0.16; 0.48] 24.1% Random effects model 0.09 [0.01; 0.25] 100.0% Heterogeneity: $I^2 = 89\%$ , p < 0.010.2 0.1 0.3 0.4

Fig. 2. A. Meta-analyses of the proportion of same and different site(s) co-carriage. Same site co-carriage proportion is defined as the number of samples/participants from which more than one serotype was recovered at the same anatomical site, among all positive samples/participants (N) identified during the study. Different sites co-carriage proportion is defined as the number of pairs of clinical samples taken simultaneously from the same individual at different anatomical sites and that retrieve discordant serotypes, among the total number of individuals who have given multiple samples with at least one of them being positive (N). B. Meta-analyses of the proportion of same site co-carriage defined as the number of samples/participants from which more than one serotype was recovered at the same anatomical site, among all positive samples/participants (N) in non-pregnant versus pregnant women. Random-effect models were used to weigh the studies.

women were unable to show a significant difference in prevalence (Fig. 2B). Moreover, the data we have do not allow to conclude on differences between male and female.

It is to be noted that Pérez-Ruiz and colleagues [27] and To and colleagues [5] found one and three rectovaginal swabs, respectively, each with two serotypes, which were counted as same site carriage in order not to bias the proportions. Considering that the measure of heterogeneity I<sup>2</sup> is high, a bias score was determined for each study (Table 1), but no study was excluded if the incidence data were available.

A													В							
Combinat	tions of												C	omb	oina	ations	s of	Total	per	
2 serotyp												>	>2 serotypes at sero					ре		
the same												tł	ne sa	ame	e site		la	7		
la	la												la	a			IV	lb	3	
lb	2	lb												IT					6	
II	2												la	a	lb	V			5	
	6	2	3	III									H	o I	II			IV	3	
IV	4	1		2	IV								la	a	lb	III		V	3	
V	4		6	6	4	V							la	a		III		VI	0	
VI							V	/1			la	a II		V		VII	0			
VII						1			VII					a		IV		VIII	0	
VIII				1		1				VI	II		la	a	IV	V		IX	0	
IX											IX							NT	1	
NT		1		1	2	1							NT	T						
Total	18	6	11	21	13	23	(	)	1	2	0	)	5	10	)0					
C																				
0			la	lb				IV	Т	V	VI	<b>_</b>	VII	VII	I	IX	N	Г То	tal	
Same	Numb	ber	25	9	17	26	;	16	Т	26	0	Γ	1	2	Τ	0	6	12	28	
site	Percen	tage	19.5	7.0	13.3	3 20.	3	12.5	5 2	20.3	0.0	(	).8	1.6	;	0.0	4.	7 100	0.0	
All	Numb	ber	57	20	48	57	7 26		T	59	6		1	3	╈	1	18	3 29	6	
studies	Percen	tage	19.3	6.8	16.2	2 19.	3	8.8	ŀ	19.9	2.0	(	).3	1.0	)	0.3	6.1	1 100	0.0	

Fig. 3. Serotype distribution in co-carriage events. A. Combinations of two serotypes carriage and total occurrence per serotype. B. Combinations of more than two serotypes carriage and total occurrence per serotype. C. Summary of the serotype prevalence in co-carriage events, for same site co-carriage and all studies (same site, different sites and unclear). NT: non-typeable.

#### 3.4. Prevalence of serotypes in co-carriage events

A review of the 59 cases of same site co-carriage with identified serotypes demonstrated that serotypes III and V are the most often co-carried (20.3 % each). This is followed by Ia (19.5 %), II (13.3 %), IV (12.5 %), and Ib (7.0 %). The most frequent combinations of two serotypes are Ia/III, III/V and II/V. Serotype Ia is the serotype most often associated with co-carriage of more than two serotypes (Fig. 3).

#### 4. Discussion

Our systematic review shows that more than one serotype carriage is a minor but definite phenomenon. According to our data, this would be the case in 10 % (95 % CI: 4-19) in case of same site co-carriage and 11 % (95 % CI: 5-20) in case of different sites cocarriage. Given the limitations of the available data in terms of reported numbers and serotypes, we believe this to be a minimum estimate and we advocate for improved surveillance to better understand this phenomenon.

With a commercialised serotype-specific vaccine covering only a subset of the more than 90 known serotypes, *Streptococcus pneumonia* is of interest [4,6,28]. Co-carriage with multiple pneumococcus serotypes is reported as common in children in low and middle-income countries [29]. In a longitudinal study conducted in Indonesian infants, 34.9 % of the positive infants in a total sample size of 198 participants showed multiple serotypes carriage [29]. Given that we also report multiple serotype carriage with GBS, the possibility of serotype replacement and serotype switching after vaccine introduction is to be considered. Indeed, the introduction of the PCV7 vaccine was followed by the replacement of vaccine strains by non-vaccine strains both among carriers and in disease [28]. Capsular switching is a regular occurrence among pneumococcus strains [8] and drives evasion in the context of vaccine-induced pressure. Capsular switching among GBS strains has also been documented [6].

Some of the studies of our review suggest that co-carriage may only be a transient phenomenon. Anthony and colleagues found that carriage of multiple GBS serotypes is never associated with chronicity [13]. Furfaro and colleagues observed that co-carriage occurs « significantly less than what would happen by chance », meaning that the presence of one GBS serotype decreases the chance of acquiring a second one [21]. Murad and colleagues were able to differentiate pneumococcus serotype replacement, stable co-colonisation, and short-term colonisation, which represent, respectively, 12.6 %, 4.8 %, and 4.8 % among all colonisation events, making stable co-colonisation a rare event [29]. Longitudinal studies evaluating GBS colonisation are lacking to confirm this trend. The capacity of certain colonising strains to compete with others is important due to its implication for vaccine-induced pressure on the ecology of GBS colonisation. In the present review, cocarriages with serotypes III, V and, Ia were the most common. They are also the most prevalent maternal colonising serotypes in most regions [30]. Further research is needed to evaluate if the cocarriage bias toward these three serotypes comes from their global prevalence or if, inversely, their prevalence is due to a capacity to out-compete other serotypes and thereby replace them.

Nonetheless, the incidence of co-carriage could be underestimated: anatomical sites, culture techniques, and protocol design may all impact upon the detection of GBS, and thus the detection of the serotypes. The results of our meta-analyses must be taken cautiously because different methods and protocols were used, which makes it difficult to aggregate the data, as demonstrated by our bias assessment score. Indeed, the optimal number of colony picks to identify all carried serotypes has not been universally defined. In addition, studies assessing different site co-carriage may underestimate the prevalence because only one isolate per swab is serotyped. Our knowledge and capacity to identify all ten serotypes have improved with time, therefore older studies might also underestimate the co-carriage phenomenon. Refined techniques such as sweep-agglutination, microarray, and multiplex PCR benefitted the detection of pneumococcal serotype cocarriage [31]. Recently, a Random Amplified Polymorphic DNA (RAPD) PCR protocol has been developed by To and colleagues to quickly screen the presence of multiple strains in clinical samples [5].

Serotype distribution varies geographically and historically, and carriage may have different implications in different populations, for example, pregnant women versus non-pregnant adults [30,32]. Our analysis was not able to find a significant difference in the prevalence of co-carriage between pregnant and non-pregnant women. Disaggregated analysis by age, population and geography, restricted to recent sample collections would be relevant to evaluate the serotype co-carriage combinations in different contexts. Non-epidemiological studies that would focus on a few participants and multiple colonies serotyping within host would be informative but feasibility would limit its applicability to inform worldwide serotype carriage and thus vaccine development.

Our data demonstrate that multiple GBS serotypes are present in a small number of carriage samples. This should encourage the design of improved epidemiological studies, able to detect multiple serotypes per participant sample, to monitor serotype distribution and replacement in preparation for the introduction of a capsular polysaccharide-based specific vaccine.

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### Data availability

Data will be made available on request.

#### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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#### Appendix A. Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.vaccine.2022.11.024.

### References

- [1] Foxman B, Gillespie B, Manning SD, Howard LJ, Tallman P, Zhang L, et al. Incidence and Duration of Group B Streptococcus by Serotype among Male and Female College Students Living in a Single Dormitory. Am J Epidemiol 2006;163(6):544–51. <u>https://doi.org/10.1093/aje/kwj075</u>.
- [2] Carreras-Abad C, Ramkhelawon L, Heath PT, Le Doare K. A Vaccine Against Group B Streptococcus: Recent Advances. Infect Drug Resist 2020;13:1263–72. <u>https://doi.org/10.2147/IDR.S203454</u>.

- [3] Absalon J, Segall N, Block SL, Center KJ, Scully IL, Giardina PC, et al. Safety and immunogenicity of a novel hexavalent group B streptococcus conjugate vaccine in healthy, non-pregnant adults: a phase 1/2, randomised, placebocontrolled, observer-blinded, dose-escalation trial. Lancet Infect Dis 2021;21 (2):263–74. <u>https://doi.org/10.1016/51473-3099(20)30478-3</u>.
- [4] Khatami A, Randis TM, Tavares L, Gegick M, Suzman E, Ratner AJ. Vaginal cocolonization with multiple Group B Streptococcus serotypes. Vaccine 2019;37:409–11. <u>https://doi.org/10.1016/j.vaccine.2018.12.001</u>.
- [5] To K-N, Powell O, Jamrozy D, Kopunova R, Anastasiadou K, Faal A, et al. RAPD PCR detects co-colonisation of multiple group B streptococcus genotypes: A practical molecular technique for screening multiple colonies. J Microbiol Methods 2021;190:106322. <u>https://doi.org/10.1016/i.mimet.2021.106322</u>.
- [6] Bellais S, Six A, Fouet A, Longo M, Dmytruk N, Glaser P, et al. Capsular Switching in Group B Streptococcus CC17 Hypervirulent Clone: A Future Challenge for Polysaccharide Vaccine Development. J Infect Dis 2012;206 (11):1745–52. <u>https://doi.org/10.1093/infdis/jiis605</u>.
- [7] Bröker G, Spellerberg B. Surface proteins of Streptococcus agalactiae and horizontal gene transfer. Int J Med Microbiol 2004;294:169–75. <u>https://doi.org/10.1016/i.iimm.2004.06.018</u>.
- [8] Wyres KL, Lambertsen LM, Croucher NJ, McGee L, von Gottberg A, Linares J, et al. Pneumococcal Capsular Switching: A Historical Perspective. J Infect Dis 2013;207(3):439–49. <u>https://doi.org/10.1093/infdis/iis703</u>.
- [9] Ouzzani M, Hammady H, Fedorowicz Z, Elmagarmid A. Rayyan-a web and mobile app for systematic reviews. Syst Rev 2016;5:210. <u>https://doi.org/ 10.1186/s13643-016-0384-4</u>.
- [10] Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ 2021;372:n71. https://doi.org/10.1136/bmj.n71.
- [11] Munn Z, Moola S, Lisy K, Riitano D, Tufanaru C. Methodological guidance for systematic reviews of observational epidemiological studies reporting prevalence and cumulative incidence data. Int J Evid Based Healthc 2015;13:147–53. <u>https://doi.org/10.1097/XEB.000000000000054</u>.
- [12] Wang N. How to Conduct a Meta-Analysis of Proportions in R: A Comprehensive Tutorial. 2018. https://doi.org/10.13140/RG.2.2.27199.00161.
- [13] Anthony BF, Okada DM, Hobel CJ. Epidemiology of group B Streptococcus: longitudinal observations during pregnancy. J Infect Dis 1978;137:524–30. https://doi.org/10.1093/infdis/137.5.524.
- [14] Anthony BF, Eisenstadt R, Carter J, Kim KS, Hobel CJ. Genital and Intestinal Carriage of Group B Streptococci During Pregnancy. J Infect Dis 1981;143:761–6. <u>https://doi.org/10.1093/infdis/143.6.761</u>.
- [15] Ferrieri P, Hillier SL, Krohn MA, Moore D, Paoletti LC, Flores AE. Characterization of vaginal & rectal colonization with multiple serotypes of group B streptococci using multiple colony picks. Indian J Med Res 2004;119 (Suppl):208–12.
- [16] Whitney CG, Daly S, Limpongsanurak S, Festin MR, Thinn KK, Chipato T, et al. The international infections in pregnancy study: group B streptococcal colonization in pregnant women. J Matern Fetal Neonatal Med 2004;15 (4):267–74. <u>https://doi.org/10.1080/14767050410001668617</u>.
- [17] Taylor KL. A study of Group B Streptococcus in Brisbane: the epidemiology, detection by PCR assay & serovar prevalence 2006:151.
- [18] El Aila NA, Tency I, Claeys G, Saerens B, De Backer E, Temmerman M, et al. Genotyping of Streptococcus agalactiae(group B streptococci) isolated from vaginal and rectal swabs of women at 35–37 weeks of pregnancy. BMC Infect Dis 2009;9(1). <u>https://doi.org/10.1186/1471-2334-9-153</u>.
- [19] Palmeiro JK, Dalla-Costa LM, Fracalanzza SEL, Botelho ACN, da Silva Nogueira K, Scheffer MC, et al. Phenotypic and Genotypic Characterization of Group B Streptococcal Isolates in Southern Brazil. J Clin Microbiol 2010;48 (12):4397–403. <u>https://doi.org/10.1128/ICM.00419-10</u>.
- [20] Slotved H-C, Dayie NTKD, Banini JAN, Frimodt-Møller N. Carriage and serotype distribution of Streptococcus agalactiae in third trimester pregnancy in southern Ghana. BMC Pregnancy Childbirth 2017;17:238. <u>https://doi.org/ 10.1186/s12884-017-1419-0</u>.
- [21] Furfaro LL, Nathan EA, Chang BJ, Payne MS. Group B streptococcus prevalence, serotype distribution and colonization dynamics in Western Australian pregnant women. J Med Microbiol 2019;68:728–40. <u>https://doi.org/10.1099/ imm.0.000980</u>.
- [22] Jisuvei SC, Osoti A, Njeri MA. Prevalence, antimicrobial susceptibility patterns, serotypes and risk factors for group B streptococcus rectovaginal isolates among pregnant women at Kenyatta National Hospital, Kenya; a crosssectional study. BMC Infect Dis 2020:20. <u>https://doi.org/10.1186/s12879-020-05035-1</u>.
- [23] Maurer M, Thirumoorthi MC, Dajani AS. Group B streptococcal colonization in prepubertal children. Pediatrics 1979;64:65–7.
- [24] Hoogkamp-Korstanje JAA, Gerards LJ, Cats BP. Maternal Carriage and Neonatal Acquisition of Group B Streptococci. J Infect Dis 1982;145:800–3. <u>https://doi. org/10.1093/infdis/145.6.800</u>.
- [25] Foster-Nyarko E, Kwambana B, Aderonke O, Ceesay F, Jarju S, Bojang A, et al. Associations between nasopharyngeal carriage of Group B Streptococcus and other respiratory pathogens during early infancy. BMC Microbiol 2016;16(1). https://doi.org/10.1186/s12866-016-0714-7.
- [26] Baker CJ, Goroff DK, Alpert SL, Hayes C, McCormack WM. Comparison of bacteriological methods for the isolation of group of B Streptococcus from vaginal cultures. J Clin Microbiol 1976;4(1):46–8.
- [27] Pérez-Ruiz M, Rodríguez-Granger JM, Bautista-Marín MF, Romero-Noguera J, Rosa-Fraile M. Genetic diversity of Streptococcus agalactiae strains colonizing the same pregnant woman. Epidemiol Infect 2004;132(2):375–8.

- [28] Weinberger DM, Malley R, Lipsitch M. Serotype replacement in disease following pneumococcal vaccination: A discussion of the evidence. Lancet 2011;378:1962-73. https://doi.org/10.1016/S0140-6736(10)@
- [29] Murad C, Dunne EM, Sudigdoadi S, Fadlyana E, Tarigan R, Pell CL, et al. Pneumococcal carriage, density, and co-colonization dynamics: A longitudinal study in Indonesian infants. Int J Infect Dis 2019;86:73-81. https://doi.org/ 10.1016/j.ijid.2019.06.024.
- [30] Bianchi-Jassir F, Paul P, To K-N, Carreras-Abad C, Seale AC, Jauneikaite E, et al. Systematic review of Group B Streptococcal capsular types, sequence types

- and surface proteins as potential vaccine candidates. Vaccine 2020;38 (43):6682–94. <u>https://doi.org/10.1016/j.vaccine.2020.08.052</u>.
  [31] Shak JR, Vidal JE, Klugman KP. Influence of bacterial interactions on pneumococcal colonization of the nasopharynx. Trends Microbiol 2013;21:129–35. <u>https://doi.org/10.1016/j.tim.2012.11.005</u>.
  [32] Le Doare K, Heath PT. An overview of global GBS epidemiology. Vaccine 2013;31:D7–D12. <u>https://doi.org/10.1016/j.vaccine.2013.01.009</u>.