Bayesian Inference Associates Rare *KDR* Variants with Specific Phenotypes in Pulmonary Arterial Hypertension

Running title: Swietlik et al.; A genotype-phenotype association study in PAH

Emilia M. Swietlik, MD¹; Daniel Greene, PhD^{2,3}; Na Zhu, PhD^{4,5}, Karyn Megy, PhD^{2,3}; Marcella Cogliano, MSc⁶; Smitha Rajaram, MD⁷; Divya Pandya, MSc¹; Tobias Tilly, MSc¹; Katie A. Lutz, BS⁸; Carrie C.L. Welch, PhD⁴; Michael W. Pauciulo, BS, MBA^{8,9}; Laura Southgate, PhD¹⁰; Jennifer M. Martin, MSt³; Carmen M. Treacy, BSc¹; Christopher J. Penkett, PhD^{2,3}; Jonathan C. Stephens, BSc^{2,3}; Harm J. Bogaard, MD, PhD¹¹; Colin Church, PhD¹²; Gerry Coghlan, MD¹³; Anna W. Coleman, MS⁸; Robin Condliffe, MD¹⁴; Christina A. Eichstaedt, PhD¹⁵⁻¹⁷; Mélanie Eyries, PhD¹⁸; Henning Gall, MD, PhD¹⁹; Stefano Ghio, MD²⁰; Barbara Girerd, PhD²¹; Ekkehard Grünig, MD^{16,17}; Simon Holden, PhD²²; Luke Howard, MD, PhD²³; Marc Humbert, MD, PhD²¹; David G. Kiely, MD¹⁴; Gabor Kovacs, MD^{24,25}; Jim Lordan, PhD²⁶; Rajiv D. Machado, PhD¹⁰; Robert V. MacKenzie Ross, MB, BChir²⁷; Colm McCabe, PhD^{23,28}; Shahin Moledina, MBChB²⁹; David Montani, MD, PhD²¹; Horst Olschewski, MD^{24,25}; Joanna Pepke-Zaba, PhD³⁰; Laura Price, PhD^{23,28}; Christopher J. Rhodes, PhD²³; Anton Vonk Noordegraaf, MD¹¹; John Wharton, PhD²³; James M. Wild, PhD⁶; Stephen John Wort, PhD^{23,28}; NIHR BioResource for Translational Research - Rare Diseases³¹; National Cohort Study of Idiopathic and Heritable PAH³²; PAH Biobank Enrolling Centers' Investigators³³; Allan Lawrie, PhD⁶; Martin R. Wilkins, MD²³; Richard C. Trembath, FRCP³⁴; Yufeng Shen, PhD^{5,35}; Wendy K. Chung, MD³⁶; Andrew J. Swift, PhD⁶; William C. Nichols, PhD^{8,9}; Nicholas W. Morrell, MD^{1,3,22,30}; Stefan Gräf, PhD¹⁻³

¹Dept of Medicine, ²Dept of Haematology, Univ of Cambridge; ³NIHR BioResource for Translational Research, Cambridge, UK; ⁴Dept of Pediatrics, ⁵Dept of Systems Biology, ³³Dept of Biomedical Informatics, Columbia Univ, New York, NY; ⁶Dept of Infection, Immunity & Cardiovascular Disease, Univ of Sheffield; ⁷⁵Neffield Teaching Hospitals NHS Foundation Trust, Sheffield, UK; ⁸Division of Human Genetics, Cincinnati Children's Hospital Medical Ctr; ⁹Dept of Pediatrics, Univ of Cincinnati College of Medicine, Cincinnati, OH; ¹⁰Molecular & Clinical Sciences Rsrch Inst, St George's, Univ of London, London, UK; ¹¹Dept of Clinical Genetics, Amsterdam UMC, Vrije Universiteit Amsterdam, Amsterdam, the Netherlands; ¹²Golden Jubilee National Hospital, Glaggow; ¹³Royal Free Hospital, London; ¹⁴Sheffield Pulmonary Vascular Disease Unit, Royal Hallamshire Hospital, Sheffield, UK; ¹⁵Laboratory for Molecular Genetic Diagnostics, Inst of Human Genetics, Heidelberg Univ; ¹⁶Ctr for Pulmonary Hypertension, Thoraxklinik gGmBH Heidelberg at Heidelberg Univ Hospital; ¹⁷Translational Lung Rsearch (DZL), Heidelberg, Germany; ¹⁶Département de génétique, hôpital Pitié-Salpëtrière, Assistance Publique-Hôpitau & Paris & UMR, S 1166-ICAN, INSERM UMR, UPMC Sorbonne Universités, Paris, France; ¹⁹Univ of Giessen & Marburg Lung Center (UGMLC), member of the German Center for Lung Research (DZL) and of the Excellence Cluster Cardio-Pulmonary Inst (CPI), Giessen, Germany; ²⁰Fondaziona IRCCS Policlinico San Matteo, Pavia, Italy; ²¹Université Paris-Sud, Faculté de Médecine, Université Paris-Saclay; AP-HP, Service de Pneumologie, Centre de référence de l'hypertension pulmonarie; INSERM UMR, S 999, Hôpital Bicétre, Les Kremlin-Bicètre, Paris, ¹²Fondarion IRCCS Policlinico San Matteo, Pavia, Italy; ²¹Université Paris-Sud, Faculté de Médecine, Université Paris-Sud, Faculté de Médecine, Université Paris-Saclay; AP-HP, Service de Pneumologie, Centre de référence de l'hypertension pulmonarie; INSERM UMR, S 999,

Correspondence: Dr Stefan Gräf, PhD Department of Medicine University of Cambridge, Level 5 Cambridge University Hospitals, Box 157 Cambridge Biomedical Campus Cambridge, CB2 0QQ, UK Tel: (+44) 1223 588036 Email: sg550@cam.ac.uk

Prof. Nicholas W. Morrell, MD Department of Medicine University of Cambridge, Level 5 Cambridge University Hospitals, Box 157 Cambridge Biomedical Campus Cambridge, CB2 0QQ, UK Tel:(+44) 1223 331666 Email: <u>nwm23@cam.ac.uk</u>

Journal Subject Terms: Genetic, Association Studies; Pulmonary Hypertension

Abstract

Background - Approximately 25% of patients with pulmonary arterial hypertension (PAH) have been found to harbor rare mutations in disease-causing genes. To identify missing heritability in PAH we integrated deep phenotyping with whole-genome sequencing data using Bayesian statistics.

Methods - We analyzed 13,037 participants enrolled in the NIHR BioResource - Rare Diseases (NBR) study, of which 1,148 were recruited to the PAH domain. To test for genetic associations between genes and selected phenotypes of pulmonary hypertension (PH), we used the Bayesian rare-variant association method BeviMed.

Results - Heterozygous, high impact, likely loss-of-function variants in the Kinase Insert Domain Receptor (*KDR*) gene were strongly associated with significantly reduced transfer coefficient for carbon monoxide (KCO, posterior probability (PP)=0.989) and older age at diagnosis (PP=0.912). We also provide evidence for familial segregation of a rare nonsense *KDR* variant with these phenotypes. On computed tomographic imaging of the lungs, a range of parenchymal abnormalities were observed in the five patients harboring these predicted deleterious variants in *KDR*. Four additional PAH cases with rare likely loss-of-function variants in *KDR* were independently identified in the US PAH Biobank cohort with similar phenotypic characteristics.

Conclusions - The Bayesian inference approach allowed us to independently validate *KDR*, which encodes for the Vascular Endothelial Growth Factor Receptor 2 (VEGFR2), as a novel PAH candidate gene. Furthermore, this approach specifically associated high impact likely loss-of-function variants in the genetically constrained gene with distinct phenotypes. These findings provide evidence for *KDR* being a clinically actionable PAH gene and further support the central role of the vascular endothelium in the pathobiology of PAH.

Key words: pulmonary hypertension; genetics, association studies; vascular endothelium; vascular endothelial growth factor receptor; family history; computed tomography

Nonstandard Abbreviations and Acronyms

11011516	inuaru Abbi eviations anu Aci onyms
6MWD	Six minute walking distance
ACVRL1	Activin-Like Kinase 1
APAH	Associated Pulmonary Arterial Hypertension
APAH:CHI	
APAH:CTE	
APAH:HIV	-PAH PAH associated with HIV
APAH:PPH	
AQP1	Aquaporin 1
ASD	Atrial Septal Defect
ATP13A3	ATPase 13A3
BA	Bronchial artery
BeviMed	Bayesian Evaluation of Variant Involvement in Mendelian Disease
BF	Bayes factor
BHF	British Heart Foundation
BMI	Body Mass Index
BMPR2	Bone Morphogenetic Protein Receptor Type 2
BNP	B-type Natriuretic Peptide
CAD	Coronary artery disease
CADD	Combined Annotation Dependent Depletion
CAVI	Caveolin-1
CHMC	Cincinnati Children's Hospital Medical Center
CI	Cardiac index
CKD	Chronic kidney disease
CO	Cardiac output
COPD	Chronic obstructive pulmonary disease
CRP	C reactive protein
CT	Computerized Tomography
CTEPH	Chronic thromboembolic pulmonary hypertension
CTPA	Computerised Tomography Pulmonary Angiogram
CUMC	Columbia University Medical Center
CVA	Celebro-vascular accident
DM	Diabetes mellitus
eCRF	Electronic Case Report Form
EIF2AK4	Eukaryotic Translation Initiation Factor 2 Alpha Kinase 4
ENG	Endoglin
FEV ₁	Forced expiratory volume in one second
FHx	Family history
FPAH	Familial PAH
FVC	
	Forced Vital Capacity
GDF2	Growth Differentiation Factor 2
GerpN	Conservation score of each nucleotide in multi-species alignment
GGO	Ground-glass opacities
gnomAD	Genome Aggregation Database
GRCh37	Genome Reference Consortium human genome build 37
	Haemoglobin
HGVS	Human Genome Variation Society
HGVSc	HGVS notation of coding sequence
HGVSp	HGVS notation of protein sequence
HRCT	High-Resolution Computerised Tomography

HTN	Systemic hypertension
I/HPAH	Idiopathic/Hereditary Pulmonary Arterial Hypertension
ICC	Intraclass Correlation Coefficient
ILD	Interstitial lung disease
KCNK3	Potassium Two Pore Domain Channel Subfamily K Member 3
KCO	Transfer coefficient for carbon monoxide
KDR	Kinase Insert Domain Receptor
MAF	Minor Allele Frequency
mPAP mRAP	Mean pulmonary artery pressure Mean right atrial pressure
NBR	NIHR BioResource - Rare Diseases
NIHR	National Institute for Health Research
NO	Nitric oxide
	N-terminal pro B-type Natriuretic Peptide
OSA	Obstructive sleep apnoea
PAD	Peripheral artery disease
PAH	Pulmonary Arterial Hypertension
PAHBB	US PAH Biobank
PAWP	Pulmonary Artery Wedge Pressure
PH	Pulmonary Hypertension
PH-LD	Pulmonary hypertension associated with lung disease
PH-LHD	Pulmonary hypertension associated with left heart disease
PH-multifac	torial Multifactorial pulmonary hypertension
pLI	Probability of being loss-of-function intolerant
PMAF	Probability that the minor allele count is at least the observed
	minor allele count
PolyPhen-2	Polymorphism Phenotyping v2 score
PP	Posterior probability
PVOD/PCH	Pulmonary veno-occlusive disease/ Pulmonary capillary
	hemangiomatosis
PVR	Pulmonary vascular resistance
RDW	Red cell distribution width
RHC	Right heart catheterisation Association.
Shared	Indicates in which cohort(s) the given variant was identified; i.e.
Sharea	$PAH(_2)$, $BPD(_1)$ means $_2$ PAH cases and $_1$ BPD case harbour
	this variant
SIFT	Sorting Intolerant From Tolerant prediction score
SMAD ₁	SMAD family member 1
SMAD ₄	SMAD family member 4
SMAD ₄ SMAD ₉	SMAD family member 9
SNIAD ₉	
	Single Nucleotide Variants SRY-box 17
SOX17	
SpO ₂ post	Arterial oxygen saturation post exercise
SpO ₂ pre	Arterial oxygen saturation pre exercise
U5416	Sugen
SvO ₂	Mixed venous oxygen saturation
TBX4	T-Box Transcription Factor 4
TLC	Total Lung Capacity
TOPMed	NHLBI Trans-Omics for Precision Medicine Hb
TSH	Thyroid Stimulating Hormone
VEGFR2	Vascular Endothelial Growth Factor Receptor 2
WBC	White Blood Cell Count
WHO FC	World Health Organisation functional class

Introduction

Pulmonary arterial hypertension (PAH) is characterized by pulmonary vascular constriction and

obliteration, causing elevation of pulmonary vascular resistance and ultimately, right ventricular

failure. Molecular mechanisms such as aberrant angiogenesis¹, metabolic reprogramming and

resistance to apoptosis² have been proposed to explain pulmonary vessel remodeling. A

breakthrough in our understanding of the pathobiology underlying PAH was the discovery of heterozygous germline mutations in the gene encoding the bone morphogenetic protein receptor type 2 (BMPR2)³, responsible for over 70% of familial PAH (FPAH) cases and 15-20% of idiopathic PAH (IPAH) cases. A smaller proportion (up to 10%) of PAH cases are caused by mutations in activin-like kinase 1 (ACVRL1)⁴, endoglin (ENG)⁵, SMAD family member 9 (SMAD9)⁶, caveolin-1 (CAV1), involved in colocalization of BMP receptors⁷, and the potassium channel KCNK3, responsible for membrane potential and vascular tone⁸. We recently identified rare pathogenic variants in growth differentiation factor 2 (GDF2), which encodes BMP9, a major ligand of the BMPR2/ALK1 receptor complex, as well as rare variants in ATPase 13A3 (ATP13A3), aquaporin 1 (AQP1) and SRY-box 17 (SOX17), and reported a list of additional putative genes potentially contributing to the pathobiology of PAH⁹. Together, the established genes explain approximately 25% of cases with IPAH, allowing their reclassification as heritable PAH (HPAH) cases. To identify additional genes harboring potentially causal rare variants in IPAH cases, we increased the cohort size¹⁰ and deployed a recently developed Bayesian methodology (BeviMed)¹¹ that incorporates phenotypic data to increase the power to detect rare risk variants.

Methods

Figure 1A provides an overview of the analysis strategy. The method details are described in the Supplemental Material. The data of the NBR study have been deposited in the European Genome-Phenome Archive¹⁰. The data from the US PAH Biobank (PAHBB) and the Columbia University Medical Center (CUMC) are available via an application¹².

Patients recruited to the NBR study provided informed consent for genetic analysis and clinical data capture (REC REF: 13/EE/0325); patients recruited by European collaborators

consented to genetic testing and clinical data collection locally. Institutional review boards at Cincinnati Children's Hospital Medical Center (CHMC) and CUMC, and the PAHBB Centers approved the validation cohort studies and written informed consent was obtained at enrollment.

Results

Characterization of study cohorts and tag definition

Whole-genome sequencing was performed in 13,037 participants of the NBR study, of which 1,148 were recruited to the PAH domain¹⁰. The PAH domain included 23 unaffected parents and three cases with an unknown phenotype, which were removed from the analysis (Figure 1B). Of the remaining 1,122 participants, 972 (86.6%) had a clinical diagnosis of IPAH, 73 (6.5%) of HPAH, and 20 (1.8%) were diagnosed with PVOD/PCH. Diagnosis verification revealed that 57 participants (5%) had a diagnosis other than IPAH, HPAH or PVOD/PCH. These cases were subsequently relabelled and moved to the respective tag group for analysis (Table 1). The comprehensive clinical characterisation of the study cohort is shown in Supplemental Table 1. In summary, the median age at diagnosis was 49[35;63] years with a female predominance of 68%. Europeans constituted 84% of the study cohort. Overall survival in the studied population was 97% at one year, 91% at three years and 84% at five years. As expected, there was a significant difference in survival between prevalent and incident cases. In prevalent cases, survival at one, three, and five years was 98%, 93%, and 87%, whereas in incident cases it was 97%, 84%, and 72%, respectively. Median transfer coefficient for carbon monoxide (KCO) in the entire studied population was 71[52;86]% predicted. Cases in the lower tertile or below the KCO threshold of 50% predicted were more commonly male, older at diagnosis, had a current or past history of cigarette smoking and an increased number of cardiorespiratory comorbidities (Supplemental Tables 2, 3, and 4). Survival in these groups was significantly worse than in those with preserved

or mildly reduced KCO (Supplemental Figure 1A-D). After adjusting for confounding factors (age, sex, comorbidities, smoking status and whether the case was prevalent or incident), KCO remained an independent predictor of survival (Supplemental Table 5).

Age at diagnosis was calculated as age at the time of diagnostic right heart catheterisation and was available in all but 10 cases. Patients in the higher age tertile showed more functional impairment despite milder hemodynamics, lower FEV1/FVC ratio and KCO [% predicted], as well as mild emphysematous and fibrotic changes on CT scans (Supplemental Figure 1E and F and Supplemental Table 6).

Rare variants in previously established genes

We identified variants in previously established genes (namely, *BMPR2*, *ACVRL1*, *ENG*, *SMAD1*, *SMAD4*, *SMAD9*, *KCNK3*, *TBX4*, *EIF2AK4*, *AQP1*, *ATP13A3*, *GDF2*, *SOX17*) in 271 (24.2%) of the 1,122 cases and interpreted them based on the ACMG standards and guidelines¹³. The majority of these variants have already been described in Gräf *et al.*⁹ (see Supplemental Material).

Rare variant association testing

We used BeviMed to consolidate previously reported PAH genes and to discover novel genotype-phenotype associations. Of note, cases explained by rare deleterious variants in previously established genes were only included for the association testing with the respective disease gene (see Methods). This analysis identified 40 significant gene-tag associations with posterior probability (PP) above 0.75 (Table 2 and Figure 2A). *BMPR2*, *TBX4*, *EIF2AK4*, *ACVRL1* and *AQP1* showed the highest association (PP \geq 0.99) but we also confirmed significant associations in the majority of other previously identified genes. Individuals with rare variants in *BMPR2*, *TBX4* (high impact), *EIF2AK4* (biallelic) and *SOX17* had a significantly younger age of

disease onset (tag: young age). We also confirmed the association of rare variants in *AQP1* with FPAH (log(BF)=10.023, PP=0.958). The refined phenotype approach corroborated the association between high impact variants in *BMPR2* and preserved KCO (KCO higher tertile, log(BF)=99.923, PP=1) together with an association of biallelic *EIF2AK4* mutations with significantly reduced KCO (KCO <50% predicted, log(BF)= 29.741, PP=1).

Under an autosomal dominant mode of inheritance, high impact variants in the Kinase Insert Domain Receptor (*KDR*) were associated with a significantly reduced KCO (KCO lower tertile, log(BF)=11.362, PP=0.989) and older age at diagnosis (tag: old age, log(BF)=9.249, PP=0.912).

Rare high impact variants in the new PAH candidate gene *KDR*

We identified five ultra-rare high impact variants in *KDR* in the study cohort. Ultra-rare variants exist in the general population only at a frequency of less than 1 in 10,000 (0.01%). Four of these were in PAH cases: one frameshift variant in exon 3 of 30 (c.183del, p.Tryp61CysfsTer16), two nonsense variants, one in exon 3 (c.183G>A, p.Trp61Ter) and one in exon 22 (c.3064C>T, p.Arg1022Ter) and one splice acceptor variant in intron 4 of 29 (c.490-1G>A). In addition, one nonsense variant was identified in exon 27 (p.Glu1206Ter) in a non-PAH control subject (Table 3). This latter nonsense variant appears late in the amino acid sequence, in exon 27 of 30, and hence is likely to escape nonsense-mediated decay, but this remains to be studied functionally. All loss-of-function variants were confirmed by Sanger sequencing (Figure 3 and Supplemental Figure 2). Furthermore, 13 PAH cases (1%) and 108 non-PAH controls (0.9%) harbored rare, predicted-deleterious *KDR* missense variants of moderate impact (Figure 3). The missense variant carriers, however, did not exhibit a reduced KCO or older age at diagnosis. Instead, these patients show the opposite trend in KCO (Figure 2B and C). Importantly, seven of the 13 *KDR*

missense variants seen in PAH cases were also detected in several non-PAH controls and thus, are of unknown significance. Furthermore, three of these missense variants co-occurred with a predicted-deleterious variant in an established PAH risk gene (two patients carried also a variant in *BMPR2* and one a variant in *AQP1*).

Clinical characterization of KDR mutation carriers

Patients with high impact variants in KDR were older and exhibited significantly reduced KCO similar to biallelic *EIF2AK4* mutation carriers and in contrast to *KDR* missense variant and BMPR2 mutation carriers (Figure 2B and C). Three of the four cases did not have a history of smoking. CT scans for all four patients showed a range of mild lung parenchymal changes (Figure 4). W000229 had evidence of mild mainly subpleural interstitial lung disease (ILD), mild emphysema, and air trapping. W000274 had signs of ILD with traction bronchiectasis in the lower zones, mild air trapping, and mild diffuse ground-glass opacities (GGO) and neovascularity. E001392 showed mild centrilobular GGO in addition to moderate pleural effusion and a trace of air trapping, but no ILD. In these cases, it seemed likely that the observed parenchymal changes contributed to the low KCO. In contrast, E003448 had a low KCO despite only a trace of central nonspecific GGO on the CT images. Comparisons of CT findings between patients harboring deleterious mutations in BMPR2, EIF2AK4, KDR, other PAH risk genes and patients without mutations are presented in Supplemental Table 7. There were no differences in the frequency of comorbidities between patients harboring missense and loss-of-function variants in KDR although the frequency of systemic hypertension was high (44%) (Supplemental Table 8). Survival analysis could not be conducted due to the small number of mutation carriers, as well as only two events occurring in this group. Following the death of W000229, his daughter, aged 53, was diagnosed with PAH and had a reduced KCO at 40% predicted. On the CT scan,

mild interstitial fibrosis was observed (Supplemental Figure 3). Sanger sequencing confirmed that father and daughter carried the same deleterious *KDR* nonsense variant p.Trp61Ter (Figure 3B).

Additional KDR cases in US PAH cohorts

To seek further evidence for *KDR* as a new candidate gene for PAH, we analyzed subjects recruited to the PAHBB¹² and the CUMC¹⁴ to identify additional patients carrying predicted pathogenic rare variants. Four additional individuals harboring rare high impact *KDR* variants were identified. These comprised, two nonsense variants, one in exon 3 (c.303C>A, p.Tyr101Ter) and one in exon 22 (c.3064C>T, p.Arg1022Ter) and two splice donor variants, one in intron 2 of 29 (c.161+1G>T) and one in intron 5 (c.658+1G>A). Interestingly, the nonsense variant p.Arg1022Ter appeared in both cohorts (Figure 3). Patient-level data for these individuals are summarized in Table 3. Three of the four patients were diagnosed with idiopathic PAH at 72, 65 and 42 years respectively, whereas one patient was diagnosed at age four with PAH associated with double outlet right ventricle. The diffusing capacity of carbon monoxide was available for one patient and was decreased at 35% predicted, with minor pleural scarring in the left upper lobe found on CT imaging. Two out of four patients (50%) harboring a high impact variant in *KDR* had been diagnosed with systemic hypertension.

Discussion

One of the critical steps in identifying novel, causative genes in rare disorders is the discovery of genotype-phenotype associations to inform patient care and outcomes. A pragmatic focus on deeply phenotyped individuals and "smart" experimental design provides additional leverage to identify novel risk variants¹⁵. To deploy this approach in PAH we brought together phenotypic and genetic data using BeviMed¹¹. This Bayesian framework allows the inclusion of prior

information regarding the hypothesis being tested in a flexible manner and compares a range of possible genetic models in a single analysis. To generate case-control labels, we tagged PAH cases with diagnostic labels and stratified them by age at diagnosis and KCO. Analyses were then performed to identify associations between tags and ultra-rare gene variants under dominant and recessive modes of inheritance and different variant impact categories.

Our BeviMed analysis provided strong statistical evidence of an association between ultra-rare, high impact variants in *KDR* and PAH with significantly reduced KCO and older age at diagnosis under a dominant mode of inheritance. Strikingly, likely loss-of-function variants in *KDR* exist in the general population with a frequency of only 4-7 per 100,000 (see Table 4). In contrast, we identified four PAH cases in the NBR cohort which equates to almost 2 in 1,000. Additionally, the statistical constraint metrics provided by gnomAD¹⁶ strongly suggest that loss-of-function variants in *KDR* are not tolerated (pLI = 1; o/e = 0.15 (0.09 - 0.25)). Besides the statistical evidence, we also identified one additional case with a family history, which together with a recently published case report of two families in which loss-of-function variants in *KDR* segregated with PAH and significantly reduced KCO¹⁷, amounts to three reported familial cases with a distinct phenotype.

VEGFR2, which is encoded by *KDR*¹⁸, binds VEGFA, a critical growth factor for physiological and pathological angiogenesis in vascular endothelial cells. In mice, even though VegfA haploinsufficiency is embryonically lethal¹⁹, heterozygosity of its receptor, Vegfr2, is compatible with life and unperturbed vascular development²⁰. The role of VEGF signaling in the pathogenesis of PAH has been an area of intense interest since increased expression of VEGF, VEGFR1 and VEGFR2 were reported in rat lung tissue in response to acute and chronic hypoxia²¹. An increase in lung VEGF has also been reported in rats with PH following

monocrotaline exposure²². In humans, VEGF-A is highly expressed in plexiform lesions in patients with IPAH²³. In addition, inhibition of VEGF signaling by SU5416 (sugen) combined with chronic hypoxia triggers severe angioproliferative PH²⁴. SU5416, a small-molecule inhibitor of the tyrosine kinase segment of VEGF receptors, inhibits VEGFR1²⁵ and VEGFR2²⁶ causing endothelial cell apoptosis, loss of lung capillaries and emphysema²⁷. Further evidence supporting the role of VEGF inhibition in the pathobiology of PAH comes from reports of PH in patients treated with bevacizumab²⁸ and the multi-tyrosine kinase inhibitors^{29,30}. Mutations in *KDR* have also been linked to congenital heart diseases. Bleyl *et al.* reported that *KDR* might be a candidate for familial total anomalous pulmonary venous return³¹. In addition, haploinsufficiency at the *KDR* locus has also been associated with tetralogy of Fallot³². We identified one patient in the CUMC cohort with PAH associated with congenital heart disease harboring a *KDR* likely protein-truncating splice donor variant (c.161+1G>T).

In the present study, we highlight that deep clinical phenotyping, in combination with genotype data, can improve the identification of novel disease risk genes and disease subtypes. *KDR* was already identified as a possible candidate gene, which did not achieve genome-wide significance, in our previous rare variant association study⁹. In combination with deep phenotyping data, *KDR* reached in the present study a significance level comparable to the most commonly affected genes in PAH. Reduced KCO, which reflects impairment of alveolar-capillary membrane function, has been noted in the analysis of early PAH registry data³³ to be an independent predictor of survival. Decreased KCO was also found in patients with PVOD/PCH with or without biallelic *EIF2AK4* mutations³⁴. Although some reduction in KCO is one of the typical features of pulmonary vascular disease, PVOD patients show the lowest KCO values when compared to IPAH or CTEPH. In contrast, KCO is relatively preserved in *BMPR2*

This article is published in its accepted form; it has not been copyedited and has not appeared in an issue of the journal. Preparation for inclusion in an issue of *Circulation: Arrhythmia and Electrophysiology* involves copyediting, typesetting, proofreading, and author review, which may lead to differences between this accepted version of the manuscript and the final published version.

11

mutation carriers³⁵. Strong association with survival and a link with other causative mutations makes the KCO phenotype particularly attractive for stratification in genetic studies.

As lung disease should always be taken under consideration as a cause of low KCO, we applied the World Symposium on PH criteria³⁶ to exclude lung disease as a cause of PH: TLC \geq 70% pred., FVC \geq 70% pred., FEV1 \geq 60% pred., and no severe fibrosis and/or emphysema on chest CT. None of the cases carrying a high impact variant in KDR met these criteria, although two of the four patients did show evidence of early ILD. Another potential reason for low KCO in the PAH population is the diagnosis of PVOD/PCH³⁷. Careful analysis of CT scans and clinical data did not reveal convincing evidence for this diagnosis in KDR mutation carriers. Cigarette smoking is a well-known factor leading to the decrease of KCO. Only one of the four KDR high impact variant carriers had a significant 15 pack-years smoking history, but with no signs of emphysema on CT. These findings suggest that loss-of-function variants in KDR are associated with a form of PAH characterized by a range of lung parenchymal abnormalities, including small airways disease, emphysema and ILD, as two of the four patients harboring a high impact variant in *KDR* had mild fibrotic lung changes. Notably, patients with mutations in other PAH risk genes, or those without the identified genetic mutation, showed less than 10% incidence of fibrotic changes on CT imaging. Further larger studies are needed to determine the full range of lung parenchymal abnormalities in PAH cases with deleterious variants in KDR.

In this study, we have assumed that PAH is a monogenic condition, which is caused by either deleterious heterozygous or biallelic variants in a single gene. This assumption, although widely supported by the literature, may not be entirely accurate. Alternatively, some cases of PAH might represent an oligogenic inheritance involving two or more genes. Although not statistically explored in the current analysis we found a total of 22 PAH cases carrying

deleterious variants in more than one PAH gene. These variants could contribute as genetic modifiers, impacting penetrance and/or expressivity. In this analysis, we have explored only a limited number of clinical phenotypes. Further studies with larger numbers of phenotypic tags derived from clinical and molecular data will increase the power to detect new associations. Finally, KCO measurements were missing for a proportion of patients which could introduce a selection bias, although all the deleterious variants in *KDR* had phenotypic data available in the UK cohort.

In summary, this study shows that deep phenotyping enables patient stratification into

subgroups with shared pathobiology and with increased power to detect new genotype-

phenotype associations. We provide statistical evidence for an association between high impact,

likely loss-of-function variants in KDR and significantly decreased KCO and later disease onset,

further supported by familial segregation.

Appendix:

NIHR BioResource for Translational Research - Rare Diseases / National Cohort Study of Idiopathic and Heritable PAH collabor: Stephen Abbs, Lara Abulhoul, Julian Adlard, Munaza Ahmed, Timothy J Aitman, Hana Alachkar, David J Allsup, Philip Ancliff, Richard Antrobus, Ruth Armstrong, Gavin Arno, Sofie Ashford, William J Astle, Anthony Attwood, Paul Aurora, Christian Babbs, Chiara Bacchelli, Tamam Bakchoul, Siddharth Banka, Tadbir Bariana, Julian Barwell, Joana Batista, Helen E Baxendale, Phil L Beales, David L Bennett, Agnieszka Bierzynska, Tina Biss, Maria A K Bitner-Glindzicz, Graeme C Black, Marta Bleda, Iulia Blesneac, Detlef Bockenhauer, Harm Bogaard, Sara Boyce, John R Bradley, Gerome Breen, Paul Brennan, Carole Brewer, Matthew Brown, Andrew C Browning, Michael J Browning, Rachel J Buchan, Matthew S Buckland, Teofila Bueser, Carmen Bugarin Diz, John Burn, Siobhan O Burns, Oliver S Burren, Nigel Burrows, Carolyn Campbell, Gerald Carr-White, Keren Carss, Ruth Casey, Mark J Caulfield, Jenny Chambers, John Chambers, Melanie M Y Chan, Floria Cheng, Patrick F Chinnery, Manali Chitre, Martin T Christian, Colin Church, Jill Clayton-Smith, Maureen Cleary, Naomi Clements Brod, Gerry Coghlan, Elizabeth Colby, Trevor R P Cole, Janine Collins, Peter W Collins, Cecilia J Compton, Robin Condliffe, H Terence Cook, Stuart Cook, Nichola Cooper, Paul A Corris, Nicola S Curry, Matthew J Daniels, Mehul Dattani, Louise C Daugherty, John Davis, Anthony De Soyza, Sri V V Deevi, Timothy Dent, Charu Deshpande, Eleanor F Dewhurst, Peter H Dixon, Sofia Douzgou, Kate Downes, Anna M Drazyk, Elizabeth Drewe, Daniel Duarte, Tina Dutt, J David M Edgar, Karen Edwards, William Egner, Melanie N Ekani, Perry Elliott, Wendy N Erber, Marie Erwood, Maria C Estiu, Dafydd Gareth

Evans, Gillian Evans, Tamara Everington, Mèlanie Eyries, Hiva Fassihi, Remi Favier, Debra Fletcher, Frances A Flinter, R Andres Floto, Tom Fowler, James Fox, Amy J Frary, Courtney E French, Kathleen Freson, Mattia Frontini, Abigail Furnell, Daniel P Gale, Henning Gall, Vijeva Ganesan, Michael Gattens, Stefano Ghio, Hossein-Ardeschir Ghofrani, J Simon R Gibbs, Kate Gibson, Kimberly C Gilmour, Barbara Girerd, Nicholas S Gleadall, Sarah Goddard, Keith Gomez, Pavels Gordins, David Gosal, Stefan Gräf, Jodie Graham, Luigi Grassi, Daniel Greene, Lynn Greenhalgh, Andreas Greinacher, Paolo Gresele, Philip Griffiths, Sofia Grigoriadou, Detelina Grozeva, Mark Gurnell, Scott Hackett, Charaka Hadinnapola, Rosie Hague, William M Hague, Matthias Haimel, Matthew Hall, Helen L Hanson, Eshika Hague, Kirsty Harkness, Andrew R Harper, Claire L Harris, Daniel Hart, Ahamad Hassan, Grant Hayman, Alex Henderson, Archana Herwadkar, Jonathan Hoffman, Simon Holden, Rita Horvath, Henry Houlden, Arjan C Houweling, Luke S Howard, Fengyuan Hu, Gavin Hudson, Aarnoud P Huissoon, Marc Humbert, Matthew Hurles, Melita Irving, Louise Izatt, Roger James, Sally A Johnson, Stephen Jolles, Jennifer Jolley, Dragana Josifova, Neringa Jurkute, Mary A Kasanicki, Hanadi Kazkaz, Rashid Kazmi, Peter Kelleher, Anne M Kelly, Wilf Kelsall, Carly Kempster, David G Kiely, Nathalie Kingston, Nils Koelling, Myrto Kostadima, Gabor Kovacs, Ania Koziell, Roman Kreuzhuber, Taco W Kuijpers, Ajith Kumar, Dinakantha Kumararatne, Manju A Kurian, Michael A Laffan, Fiona Lalloo, Michele Lambert, Hana Lango Allen, Allan Lawrie, D Mark Layton, Claire Lentaigne, Tracy Lester, Adam P Levine, Rachel Linger, Hilary Longhurst, Lorena E Lorenzo, Eleni Louka, Paul A Lyons, Rajiv D Machado, Robert V MacKenzie Ross, Bella Madan, Eamonn R Maher, Jesmeen Maimaris, Samantha Malka, Sarah Mangles, Rutendo Mapeta, Kevin J Marchbank, Stephen Marks, Hugh S Markus, Hanns-Ulrich Marschall, Andrew Marshall, Jennifer Martin, Mary Mathias, Emma Matthews, Heather Maxwell, Paul McAlinden, Mark I McCarthy, Harriet McKinney, Stuart Meacham, Adam J Mead, Karyn Megy, Sarju G Mehta, Michel Michaelides, Carolyn Millar, Shehla N Mohammed, Shahin Moledina, David Montani, Anthony T Moore, Nicholas W Morrell, Monika Mozere, Keith W Muir, Andrew D Mumford, Andrea H Nemeth, William G Newman, Michael Newnham, Sadia Noorani, Paquita Nurden, Jennifer O'Sullivan, Samva Obaji, Chris Odhams, Steven Okoli, Andrea Olschewski, Horst Olschewski, Kai Ren Ong, S Helen Oram, Elizabeth Ormondroyd, Willem H Ouwehand, Claire Palles, Sofia Papadia, Soo-Mi Park, David Parry, Smita Patel, Joan Paterson, Andrew Peacock, Simon H Pearce, Kathelijne Peerlinck, Christopher J Penkett, Joanna Pepke-Zaba, Romina Petersen, Clarissa Pilkington, Kenneth E S Poole, Bethan Psaila, Angela Pyle, Richard Quinton, Shamima Rahman, Anupama Rao, F Lucy Raymond, Paula J Rayner-Matthews, Augusto Rendon, Tara Renton, Christopher J Rhodes, Andrew S C Rice, Alex Richter, Leema Robert, Irene Roberts, Sarah J Rose, Robert Ross-Russell, Catherine Roughley, Noemi B A Roy, Deborah M Ruddy, Omid Sadeghi-Alavijeh, Moin A Saleem, Nilesh Samani, Crina Samarghitean, Alba Sanchis-Juan, Ravishankar B Sargur, Robert N Sarkany, Simon Satchell, Sinisa Savic, Genevieve Sayer, John A Sayer, Laura Scelsi, Andrew M Schaefer, Sol Schulman, Richard Scott, Marie Scully, Claire Searle, Werner Seeger, Arjune Sen, W A Carrock Sewell, Denis Sevres, Neil Shah, Olga Shamardina, Susan E Shapiro, Adam C Shaw, Keith Sibson, Lucy Side, Ilenia Simeoni, Michael A Simpson, Matthew C Sims, Suthesh Sivapalaratnam, Damian Smedley, Katherine R Smith, Kenneth G C Smith, Katie Snape, Nicole Soranzo, Florent Soubrier, Laura Southgate, Olivera Spasic-Boskovic, Simon Staines, Emily Staples, Hannah Stark, Jonathan Stephens, Kathleen E Stirrups, Alex Stuckey, Jay Suntharalingam, Emilia M Swietlik, Petros Syrris, R Campbell Tait, Kate Talks, Rhea Y Y Tan, Jenny C Taylor, John M Taylor, James E Thaventhiran, Andreas C Themistocleous, David Thomas, Ellen Thomas, Moira

J Thomas, Patrick Thomas, Kate Thomson, Adrian J Thrasher, Chantal Thys, Tobias Tilly, Marc Tischkowitz, Catherine Titterton, Cheng-Hock Toh, Ian P Tomlinson, Mark Toshner, Matthew Traylor, Carmen Treacy, Paul Treadaway, Richard Trembath, Salih Tuna, Ernest Turro, Philip Twiss, Tom Vale, Chris Van Geet, Natalie van Zuydam, Anthony M Vandersteen, Marta Vazquez-Lopez, Julie von Ziegenweidt, Anton Vonk Noordegraaf, Annette Wagner, Quinten Waisfisz, Neil Walker, Suellen M Walker, James S Ware, Hugh Watkins, Christopher Watt, Andrew R Webster, Lucy Wedderburn, Wei Wei, Steven B Welch, Julie Wessels, Sarah K Westbury, John-Paul Westwood, John Wharton, Deborah Whitehorn, James Whitworth, Andrew O M Wilkie, Martin R Wilkins, Catherine Williamson, Brian T Wilson, Edwin K S Wong, Nicholas Wood, Yvette Wood, Christopher Geoffrey Woods, Emma R Woodward, Stephen J Wort, Austen Worth, Michael Wright, Katherine Yates, Patrick F K Yong, Timothy Young, Ping Yu, Patrick Yu-Wai-Man, Eliska Zlamalova.

PAH Biobank Enrolling Centers' Investigators: Russel Hirsch, MD; R. James White, MD, PhD; Marc Simon, MD; David Badesch, MD; Erika Rosenzweig, MD; Charles Burger, MD; Murali Chakinala, MD; Thenappan Thenappan, MD; Greg Elliott, MD; Robert Simms, MD; Harrison Farber, MD; Robert Frantz, MD; Jean Elwing, MD; Nicholas Hill, MD; Dunbar Ivy, MD; James Klinger, MD; Steven Nathan, MD; Ronald Oudiz, MD; Ivan Robbins, MD; Robert Schilz, DO, PhD; Terry Fortin, MD; Jeffrey Wilt, MD; Delphine Yung, MD; Eric Austin, MD; Ferhaan Ahmad, MD, PhD; Nitin Bhatt, MD; Tim Lahm, MD; Adaani Frost, MD; Zeenat Safdar, MD; Zia Rehman, MD; Robert Walter, MD; Fernando Torres, MD; Sahil Bakshi, DO; Stephen Archer, MD; Rahul Argula, MD; Christopher Barnett MD; Raymond Benza MD; Ankit Desai MD; Veeranna Maddipati MD.

Acknowledgments: We thank NIHR BioResource volunteers for their participation, and gratefully acknowledge NIHR BioResource centers, NHS Trusts and staff for their contribution. We thank the National Institute for Health Research and NHS Blood and Transplant. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health and Social Care. We thank the research nurses and coordinators at the specialist pulmonary hypertension centers involved in this study. We acknowledge the support of the Imperial NIHR Clinical Research Facility, the Netherlands CardioVascular Research Initiative, the Dutch Heart Foundation, Dutch Federation of University Medical Centres, the Netherlands Organization for Health Research and Development and the Royal Netherlands Academy of Sciences. We thank all the patients and their families who contributed to this research and the Pulmonary Hypertension Association (UK) for their support. We also thank Kathryn Auckland for proofreading the manuscript. We thank contributors, including the Pulmonary Hypertension Centers who collected samples used in this study, as well as patients and their families, whose help and participation made this work possible. Exome sequencing and genotyping data were generated by the Regeneron Genetics Center.

Sources of Funding: The UK National Cohort of Idiopathic and Heritable PAH is supported by the National Institute for Health Research (NIHR), the British Heart Foundation (BHF) (SP/12/12/29836 and SP/18/10/33975), the BHF Cambridge Centre of Cardiovascular Research Excellence, and the UK Medical Research Council (MR/K020919/1), the Dinosaur Trust, BHF Programme grants to RCT (RG/08/006/25302) and NWM (RG/13/4/30107), and the UK NIHR National Institute for Health Research Cambridge Biomedical Research Centre. NWM is a BHF Professor and NIHR Senior Investigator. AL is supported by a BHF Senior Basic Science Research Fellowship (FS/13/48/30453). All research at Great Ormond Street Hospital NHS Foundation Trust and UCL Great Ormond Street Institute of Child Health is made possible by the NIHR Great Ormond Street Hospital Biomedical Research Centre. Samples and/or data from the National Biological Sample and Data Repository for PAH, funded by an NIH investigator-initiated resources grant (R24 HL105333 to WCN), were used in this study.

Disclosures: NWM is a Director and Co-founder of Morphogen-IX. JW received personal fees from Actelion Pharmaceuticals. GK reports personal fees and non-financial support from Actelion Pharmaceuticals, Bayer, GSK, MSD, Boehringer Ingelheim, Novartis, Chiesi and Vitalaire outside the submitted work. CJR declares fees from Actelion Pharmaceuticals and United Therapeutics. AL received support and fees from GSK and Actelion Pharmaceuticals.

References:

1. Tuder RM, Groves B, Badesch DB, Voelkel NF. Exuberant endothelial cell growth and elements of inflammation are present in plexiform lesions of pulmonary hypertension. *Am J Pathol.* 1994;144:275–285.

2. Pullamsetti SS, Savai R, Seeger W, Goncharova EA. Translational Advances in the Field of Pulmonary Hypertension. From Cancer Biology to New Pulmonary Arterial Hypertension Therapeutics. Targeting Cell Growth and Proliferation Signaling Hubs. *Am J Respir Crit Care Med.* 2017;195:425–437.

3. International PPH Consortium, Lane KB, Machado RD, Pauciulo MW, Thomson JR, Phillips JA 3rd, Loyd JE, Nichols WC, Trembath RC. Heterozygous germline mutations in BMPR2, encoding a TGF-beta receptor, cause familial primary pulmonary hypertension. *Nat Genet*. 2000;26:81–84.

4. Trembath RC. Mutations in the TGF-beta type 1 receptor, ALK1, in combined primary pulmonary hypertension and hereditary haemorrhagic telangiectasia, implies pathway specificity. *J Heart Lung Transplant*. 2001;20:175.

5. Chaouat A. Endoglin germline mutation in a patient with hereditary haemorrhagic telangiectasia and dexfenfluramine associated pulmonary arterial hypertension. *Thorax*. 2004;59:446–448.

6. Shintani M, Yagi H, Nakayama T, Saji T, Matsuoka R. A new nonsense mutation of SMAD8 associated with pulmonary arterial hypertension. *J Med Genet*. 2009;46:331–337.

7. Austin ED, Ma L, LeDuc C, Berman Rosenzweig E, Borczuk A, Phillips JA 3rd, Palomero T, Sumazin P, Kim HR, Talati MH, et al. Whole exome sequencing to identify a novel gene (caveolin-1) associated with human pulmonary arterial hypertension. *Circ Cardiovasc Genet*. 2012;5:336–343.

8. Ma L, Roman-Campos D, Austin ED, Eyries M, Sampson KS, Soubrier F, Germain M, Trégouët D-A, Borczuk A, Rosenzweig EB, et al. A Novel Channelopathy in Pulmonary Arterial Hypertension. *N Engl J Med*. 2013;369:351–361.

9. Gräf S, Haimel M, Bleda M, Hadinnapola C, Southgate L, Li W, Hodgson J, Liu B, Salmon RM, Southwood M, et al. Identification of rare sequence variation underlying heritable pulmonary arterial hypertension. *Nat Commun.* 2018;9:1416.

10. Turro E, Astle WJ, Megy K, Gräf S, Greene D, Shamardina O, Allen HL, Sanchis-Juan A, Frontini M, Thys C, et al. Whole-genome sequencing of patients with rare diseases in a national health system. *Nature*. 2020;583:96–102.

11. Greene D, NIHR BioResource, Richardson S, Turro E. A Fast Association Test for Identifying Pathogenic Variants Involved in Rare Diseases. *Am J Hum Genet*. 2017;101:104–114.

12. Zhu N, Pauciulo MW, Welch CL, Lutz KA, Coleman AW, Gonzaga-Jauregui C, Wang J, Grimes JM, Martin LJ, He H, et al. Novel risk genes and mechanisms implicated by exome sequencing of 2572 individuals with pulmonary arterial hypertension. *Genome Med.* 2019;11:69.

13. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med.* 2015;17:405–424.

14. Zhu N, Welch CL, Wang J, Allen PM, Gonzaga-Jauregui C, Ma L, King AK, Krishnan U, Rosenzweig EB, Ivy DD, et al. Rare variants in SOX17 are associated with pulmonary arterial hypertension with congenital heart disease. *Genome Med.* 2018;10:56.

15. FitzGerald G, Botstein D, Califf R, Collins R, Peters K, Van Bruggen N, Rader D. The future of humans as model organisms. *Science*. 2018;361:552–553.

16. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, Collins RL, Laricchia KM, Ganna A, Birnbaum DP, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*. 2020;581:434–443.

17. Eyries M, Montani D, Girerd B, Favrolt N, Riou M, Faivre L, Manaud G, Perros F, Gräf S, Morrell NW, et al. Familial pulmonary arterial hypertension by KDR heterozygous loss of function. *Eur Respir J*. 2020;1902165.

18. Terman BI, Carrion ME, Kovacs E, Rasmussen BA, Eddy RL, Shows TB. Identification of a new endothelial cell growth factor receptor tyrosine kinase. *Oncogene*. 1991;6:1677–1683.

19. Ferrara N, Carver-Moore K, Chen H, Dowd M, Lu L, O'Shea KS, Powell-Braxton L, Hillan KJ, Moore MW. Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature*. 1996;380:439–442.

20. Oladipupo SS, Smith C, Santeford A, Park C, Sene A, Wiley LA, Osei-Owusu P, Hsu J, Zapata N, Liu F, et al. Endothelial cell FGF signaling is required for injury response but not for vascular homeostasis. *Proc Natl Acad Sci U S A*. 2014;111:13379–13384.

21. Tuder RM, Flook BE, Voelkel NF. Increased gene expression for VEGF and the VEGF receptors KDR/Flk and Flt in lungs exposed to acute or to chronic hypoxia. Modulation of gene expression by nitric oxide. *J Clin Invest*. 1995;95:1798–1807.

22. Cho YJ, Han JY, Lee SG, Jeon BT, Choi WS, Hwang YS, Roh GS, Lee JD. Temporal changes of angiopoietins and Tie2 expression in rat lungs after monocrotaline-induced pulmonary hypertension. *Comp Med.* 2009;59:350–356.

23. Tuder RM, Chacon M, Alger L, Wang J, Taraseviciene-Stewart L, Kasahara Y, Cool CD, Bishop AE, Geraci M, Semenza GL, et al. Expression of angiogenesis-related molecules in plexiform lesions in severe pulmonary hypertension: evidence for a process of disordered angiogenesis. *J Pathol.* 2001;195:367–374.

24. Taraseviciene-Stewart L, Kasahara Y, Alger L, Hirth P, Mc Mahon G, Waltenberger J, Voelkel NF, Tuder RM. Inhibition of the VEGF receptor 2 combined with chronic hypoxia causes cell death-dependent pulmonary endothelial cell proliferation and severe pulmonary hypertension. *FASEB J*. 2001;15:427–438.

25. Itokawa T, Nokihara H, Nishioka Y, Sone S, Iwamoto Y, Yamada Y, Cherrington J, McMahon G, Shibuya M, Kuwano M, et al. Antiangiogenic effect by SU5416 is partly attributable to inhibition of Flt-1 receptor signaling. *Mol Cancer Ther*. 2002;1:295–302.

26. Fong TA, Shawver LK, Sun L, Tang C, App H, Powell TJ, Kim YH, Schreck R, Wang X, Risau W, et al. SU5416 is a potent and selective inhibitor of the vascular endothelial growth

factor receptor (Flk-1/KDR) that inhibits tyrosine kinase catalysis, tumor vascularization, and growth of multiple tumor types. *Cancer Res.* 1999;59:99–106.

27. Kasahara Y, Tuder RM, Taraseviciene-Stewart L, Le Cras TD, Abman S, Hirth PK, Waltenberger J, Voelkel NF. Inhibition of VEGF receptors causes lung cell apoptosis and emphysema. *J Clin Invest*. 2000;106:1311–1319.

28. Garcia AA, Hirte H, Fleming G, Yang D, Tsao-Wei DD, Roman L, Groshen S, Swenson S, Markland F, Gandara D, et al. Phase II Clinical Trial of Bevacizumab and Low-Dose Metronomic Oral Cyclophosphamide in Recurrent Ovarian Cancer: A Trial of the California, Chicago, and Princess Margaret Hospital Phase II Consortia. *J Clin Oncol.* 2008;26:76–82.

29. Montani D, Bergot E, Günther S, Savale L, Bergeron A, Bourdin A, Bouvaist H, Canuet M, Pison C, Macro M, et al. Pulmonary Arterial Hypertension in Patients Treated by Dasatinib. *Circulation*. 2012;125:2128–2137.

30. El-Dabh A, Acharya D. EXPRESS: Pulmonary hypertension with dasatinib and other tyrosine kinase inhibitors. *Pulm Circ*. 2019;2045894019865704.

31. Bleyl S, Nelson L, Odelberg SJ, Ruttenberg HD, Otterud B, Leppert M, Ward K. A gene for familial total anomalous pulmonary venous return maps to chromosome 4p13-q12. *Am J Hum Genet*. 1995;56:408–415.

32. Reuter MS, Jobling R, Chaturvedi RR, Manshaei R, Costain G, Heung T, Curtis M, Hosseini SM, Liston E, Lowther C, et al. Haploinsufficiency of vascular endothelial growth factor related signaling genes is associated with tetralogy of Fallot. *Genet Med.* 2019;21:1001–1007.

33. Test VJ, Farber HW, McGoon MD, Parsons L, Channick RN. Pulmonary Arterial Hypertension in the Elderly: Baseline Characteristics and Evaluation of Therapeutics. An Examination of the Reveal Registry. *Am J Respir Crit Care Med*. 2020;201:A2649

34. Hadinnapola C, Bleda M, Haimel M, Screaton N, Swift A, Dorfmüller P, Preston SD, Southwood M, Hernandez-Sanchez J, Martin J, et al. Phenotypic Characterization of Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically With Pulmonary Arterial Hypertension. *Circulation*. 2017;136:2022–2033.

35. Trip P, Girerd B, Bogaard H-J, de Man FS, Boonstra A, Garcia G, Humbert M, Montani D, Vonk-Noordegraaf A. Diffusion capacity and BMPR2 mutations in pulmonary arterial hypertension. *Eur Respir J*. 2014;43:1195–1198.

36. Nathan SD, Barbera JA, Gaine SP, Harari S, Martinez FJ, Olschewski H, Olsson KM, Peacock AJ, Pepke-Zaba J, Provencher S, et al. Pulmonary hypertension in chronic lung disease and hypoxia. *Eur Respir J*. 2019;53.

37. Montani D, Dorfmuller P, Maitre S, Jaïs X, Sitbon O, Simonneau G, Humbert M. [Pulmonary veno-occlusive disease and pulmonary capillary hemangiomatosis]. *Presse Med.* 2010;39:134–143.

American Heart Association.

Circulation: Genomic and Precision Medicine

Table 1. Definitions of labels and the number of unrelated cases and controls in the rare variant association analysis with BeviMed. See paragraph on "Number of PAH domain samples in the analysis" in the Supplemental Material for more details.

Tag	Tag description	Cases	Controls	Excluded relatives
РН	Individuals with mPAP > 25 mmHg	1112	9134	2786
РАН	Patients with one of the following diagnoses: IPAH, HPAH, PVOD, PCH, APAH:CHD-PAH, APAH:CTD-PAH, APAH:HIV-PAH, APAH:PH-PAH	1085	9134	2786
І/НРАН	Patients with a clinical diagnosis of IPAH or HPAH	1036	9134	2786
ІРАН	Patients with a clinical diagnosis of IPAH	972	9134	2785
НРАН	Patients with a clinical diagnosis of HPAH	67	9136	2779
PVOD/PCH	Patients with a clinical diagnosis of PVOD/PCH	20	9136	2778
I/HPAH/PVOD/PCH	Patients with one of the following diagnoses: IPAH, HPAH, PVOD, PCH	1056	9134	2786
FPAH	Patients with one of the following diagnoses: IPAH, HPAH, PVOD, PCH and a positive family history	80	9136	2781 American
АРАН	Patients with one of the following diagnoses: APAH:CHD-PAH, APAH:CTD- PAH, APAH:HIV-PAH, APAH:PH-PAH	29	9136	Heart Association 2778
АРАН: СНД-РАН	Patients with PAH associated with congenital heart disease	17	9136	2778
АРАН: СТД-РАН	Patients with PAH associated with connective tissue disease	10	9136	2778
АРАН: РоРН	Patients with PAH associated with portopulmonary hypertension	1	9136	2778
АРАН: НІV-РАН	Patients with PAH associated with HIV	1	9136	2778
PH-LHD	Patients with pulmonary hypertension associated with left heart disease (Group 2)	7	9136	2778
PH-LD	Patients with pulmonary hypertension associated with lung disease (Group 3)	8	9136	2778
СТЕРН	Chronic thromboembolic pulmonary hypertension (Group 4)	6	9136	2778
PH-multifactorial	Multifactorial pulmonary hypertension (Group 5)	6	9136	2778
young age	Lower age tertile (<40.8 years)	378	9136	2785
middle age	Middle age tertile (40.8 - 58.6 years)	376	9134	2779
old age	Higher age tertile (>58.6 years)	355	9136	2778
low KCO	KCO <50% pred.	152	9136	2778
KCO lower tertile	KCO <60% pred.	211	9136	2778
KCO middle tertile	KCO 60-80% pred.	215	9136	2778
KCO higher tertile	KCO >80% pred.	215	9134	2779

Abbreviations: mPAP - mean pulmonary artery pressure, PH - pulmonary hypertension, PAH - pulmonary arterial hypertension, I/HPAH - Idiopathic/Hereditary Pulmonary Arterial Hypertension, PVOD - Pulmonary veno-occlusive disease, PCH - Pulmonary capillary haemangiomatosis, APAH - Associated Pulmonary Arterial Hypertension, CHD - Congenital Heart Disease, CTD - Connective Heart Disease, LHD - Left Heart Disease, LD - Lung Disease, CTEPH - Chronic Thromboembolic Pulmonary Hypertension, KCO - transfer coefficient for carbon monoxide.

Table 2. BeviMed analysis results. Posterior probabilities and Bayes Factors (BF) of gene-tag associations (prior probability $\pi = 0.001$). The BF is the ratio between the probabilities of the data under H1 and under H0. The observed data are BF times more likely under H1 than under H0, and so the larger the BF, the stronger the support in the data for H1 compared with H0. The "High" category comprises only variants of high impact, including loss-of-function variants and large deletions; the "Moderate" category contains variants of moderate impact, including missense variants or variants of consequence type "non_coding_transcript_exon_variant"; the combined category "Moderate and High" includes both respective consequence types.

Gene Transcript		Tag	log(Bayes Factor)	Posterior probability	Consequence type	Mode of inheritance
BMPR2	ENST00000374580	I/HPAH	265.762	1.000	High	dominant
BMPR2	ENST00000374580	РАН	265.639	1.000	High	dominant
BMPR2	ENST00000374580	I/HPAH/PVOD/PCH	263.481	1.000	High	dominant
BMPR2	ENST00000374580	РН	262.625	1.000	High	dominant
BMPR2	ENST00000374580	young age	149.576	1.000	Moderate and high	dominant
BMPR2	ENST00000374580	НРАН	149.091	1.000	Moderate and high	dominant ocid
BMPR2	ENST00000374580	FPAH	147.822	1.000	Moderate and high	dominant
BMPR2	ENST00000374580	IPAH	144.582	1.000	High	dominant
BMPR2	ENST00000374580	KCO higher tertile	99.923	1.000	High	dominant
BMPR2	ENST00000374580	middle age	63.119	1.000	Moderate and high	dominant
BMPR2	ENST00000374580	KCO middle tertile	52.706	1.000	Moderate and high	dominant
EIF2AK4	ENST00000263791	low KCO	29.741	1.000	Moderate and high	recessive
EIF2AK4	ENST00000263791	KCO lower tertile	26.247	1.000	Moderate and high	recessive
TBX4	ENST00000240335	I/HPAH	23.783	1.000	High	dominant
TBX4	ENST00000240335	I/HPAH/PVOD/PCH	23.549	1.000	High	dominant
TBX4	ENST00000240335	РАН	23.141	1.000	High	dominant
TBX4	ENST00000240335	РН	22.877	1.000	High	dominant
EIF2AK4	ENST00000263791	young age	20.547	1.000	Moderate and high	recessive
TBX4	ENST00000240335	IPAH	19.990	1.000	High	dominant
EIF2AK4	ENST00000263791	I/HPAH/PVOD/PCH	15.718	1.000	Moderate and high	recessive
ACVRL1	ENST00000388922	НРАН	15.501	1.000	Moderate and high	dominant
EIF2AK4	ENST00000263791	РАН	15.407	1.000	Moderate and high	recessive
EIF2AK4	ENST0000263791	РН	15.071	1.000	Moderate and high	recessive

	-					
EIF2AK4	ENST00000263791	PVOD/PCH	14.441	0.999	Moderate and high	recessive
AQP1	ENST00000311813	НРАН	12.075	0.994	Moderate	dominant
EIF2AK4	ENST00000263791	FPAH	11.858	0.993	High	recessive
TBX4	ENST00000240335	young age	11.500	0.990	High	dominant
AQP1	ENST00000311813	I/HPAH	11.466	0.990	Moderate and high	dominant
KDR	ENST00000263923	KCO lower tertile	11.362	0.989	High	dominant
AQP1	ENST00000311813	I/HPAH/PVOD/PCH	11.291	0.988	Moderate and high	dominant
AQP1	ENST00000311813	РАН	11.047	0.984	Moderate and high	dominant
AQP1	ENST00000311813	РН	10.791	0.980	Moderate and high	dominant
AQP1	ENST00000311813	FPAH	10.023	0.958	Moderate	dominant
KDR	ENST00000263923	old age	9.249	0.912	High	dominant
GDF2	ENST00000249598	I/HPAH	9.091	0.899	Moderate and high	dominant
BMPR2	ENST00000374580	old age	8.913	0.881	High	dominant
GDF2	ENST00000249598	I/HPAH/PVOD/PCH	8.775	0.866	Moderate and high	dominant
SOX17	ENST00000297316	young age	8.554	0.839	Moderate and high	dominant
GDF2	ENST00000249598	РАН	8.478	0.828	Moderate and high	dominant
ATP13A3	ENST00000439040	KCO higher tertile	8.035	0.755	High	dominant

and Precision Medicine

Table 3. Gene changes for IPAH patients harboring likely loss-of-function variants in the KDR gene. None of the KDR variants have previously been reported in gnomAD, ExAC or internal controls. HGVSc notations are based on transcript sequence ENST00000263923.4. HGVSp notations are based on the amino acid sequence ENSP00000263923.4.

Cohort			UK			US			
ID	W000229	W000229.d	E003448	W000274	E001392	CUMC-JM161	ССНМС-12- 190	ССНМС-19-023	ССНМС-27-015
Exon	3/30	3/30		22/30	3/30	2/30	3/30	5/30	22/30
HGVSc	c.183G>A	c.183G>A	c.490-1G>A	c.3064C>T	c.183del	c.161+1G>T	c.303C>A	c.658+1G>A	c.3064C>T
HGVSp	p.Trp61Ter	p.Trp61Ter	-	p.Arg1022Ter	p.Trp61CysfsTer16		p.Tyr101Ter		p.Arg1022Ter
Consequence type	stop gained	stop gained	splice acceptor variant	stop gained	frameshift variant	splice donor variant	stop gained	stop gained	stop gained
Shared	PAH(1)	PAH(1)	PAH(1)	PAH(1)	PAH(1)	PAH(1)	PAH(1)	PAH(1)	PAH(1)
gnomAD	NA	NA	NA	NA	NA	NA	NA	NA	NA
CADD PHRED v1.3	38	38	26	37	35	26.4	38	24.3	37
SIFT	2			iai	o io		dia	line	-
PolyPhen	d		EU	5	J -	VIE		. I - E	
GerpN	5.93	5.93	5.75	5.95	5.93	5.83	5.48	5.8	5.95
Ancestry	European	European	European	European	European	East-Asian	European	European	European
Sex	male	female	female	male	female	female	male	female	female
Diagnosis	IPAH	IPAH	IPAH	IPAH	IPAH	APAH-CHD secondary to double outlet RV	IPAH	IPAH	IPAH
Age at diagnosis [years]	71	53	62	67	61	4	72	65	42
WHO FC	2	3	3	3	3	2	NA	NA	NA

Status	dead	alive	alive	alive	dead	alive	alive	alive	alive
Family history	Yes, daughter	Yes, father	No	No	No	No	No	No	No
Comorbidities	hyperlipidemia, HTN, DM type 2	DM type 2, OSA, pulmonary fibrosis	HTN, hypothyroidism	DM type 2	CAD, DM type 2	No	HTN, hyperlipidemia,	HTN, hypothyroidism, OA	Obesity, CAD, DM type 2, hypothyroidism
PVR	16.11	12.12	9.17	4.86	6.69	NA	NA	27.9	9.8
CO [L/min]	3.6	3.3	4.58	5.97	5.23	NA	4.33	1.8	4.6
PAWP [mmHg]	4	5	15	12	9	NA	5	16	15
mPAP [mmHg]	62	45	57	41	44	NA	49	66	60
mRAP [mmHg]	5	13	8	8	3	NA	5	29	14
Smoking history	Never	Never	Never	Ex-smoker	Never	Never	Never	Ex-smoker	Heart AssociNever
KCO [% pred.]	44	40	46	46	55.2	NA	NA	35%*	NA
TLC [% pred.]	NA	69	NA	NA	NA	NA	NA	65%	NA
FVC [% pred.]	115	76	94	91	72.8	92%	NA	83%	NA
FEV1 [% pred.]	116	70	90	83	67.3	85%	NA	77%	NA
SpO2 post [%]	86	87	86	NA	91	NA	NA	NA	NA
SpO2 pre [%]	95	96	97	98	97	NA	NA	NA	NA
6MWD [m]	472	202	422	660	180	NA	380	NA	245

Abbreviations: KDR - Kinase insert domain receptor, WHO FC - World Health Organization functional class, 6MWD - 6-minute walk distance, SpO2 - arterial oxygen saturation, mRAP - mean right atrial pressure, mPAP - mean pulmonary artery pressure, mPAWP - mean pulmonary artery wedge pressure, CO - cardiac output, PVR - pulmonary vascular resistance, FEV1 - forced expiratory volume in 1 sec, FVC - forced vital capacity, KCO - transfer factor coefficient for carbon monoxide, HTN - systemic hypertension, CAD - coronary artery disease, OA - osteoarthritis, DM - diabetes mellitus, OSA - obstructive sleep apnea, ASD - atrial septal defect, VSD - ventricular septal defect. * DLCO % predicted.

Table 4. Comparison of high impact likely loss-of-function variants in KDR in the Human largescale sequencing reference populations gnomAD and TOPMed with the NBR non-PAH controls and PAH cases.

Large-scale sequencing population	High impact LoF variants in <i>KDR</i>	Individuals	Alleles	Frequency
gnomAD (v2.1)*	20	141,456	282,912	0.000071
gnomAD (v3)	10	71,702	143,404	0.000070
TOPMed	5	62,784	125,568	0.000040
NBR non-PAH controls	1	11,889	23,778	0.000042
NBR PAH cases	4	1,122	2,244	0.001783

**KDR* constraint metrics: pLI = 1; o/e = 0.15 (0.09 - 0.25); exp(LoF) = 73; obs(LoF) = 11 Abbreviations: gnomAD - The Genome Aggregation Database, TOPMed - The Trans-Omics for Precision Medicine program, KDR - kinase insert domain receptor, NBR - NIHR BioResource - Rare Diseases, LoF - loss of function

Circulation: Genomic and Precision Medicine

Figure Legends:

Figure 1. Design of the genetic association study. A, Overview of the analytical approach. Using deep phenotyping, data tags were assigned to patients who shared phenotypic features. Rare sequence variants, called from whole-genome sequencing data, were filtered and explained cases were labeled. BeviMed was applied to a set of unrelated individuals to estimate the posterior probability of gene-tag associations. B, Consort diagram summarizing the size of the study cohort. C, Schematic representation of the definition of cases, exemplified by the KCO lower tertile tag. Cases were defined as individuals carrying a particular tag, whereas patients with

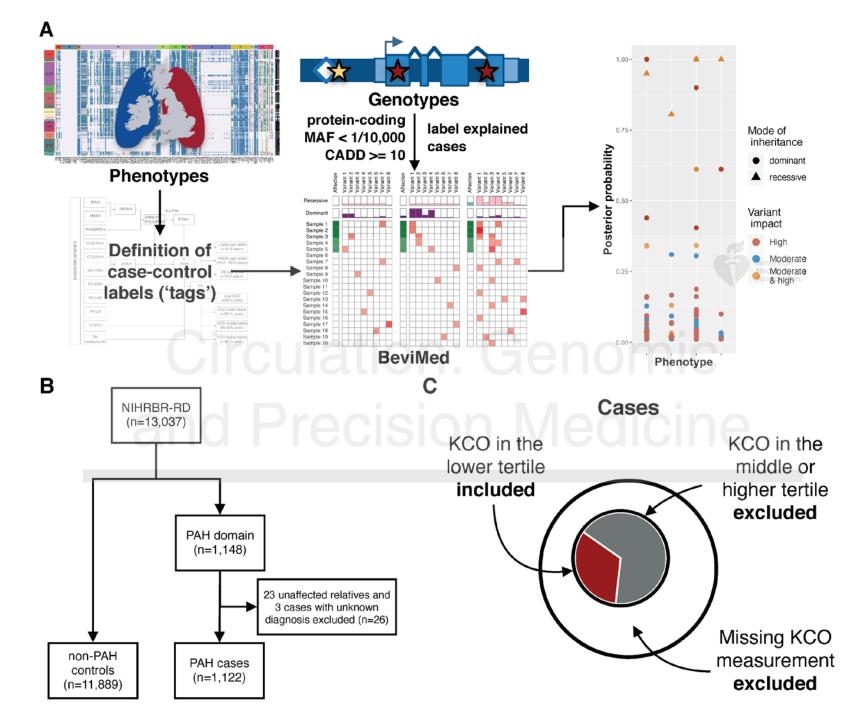
missing information or those without a tag were removed from the gene-tag association testing. Individuals from non-PAH domains served as controls. KCO - transfer coefficient for carbon monoxide, MAF - minor allele frequency.

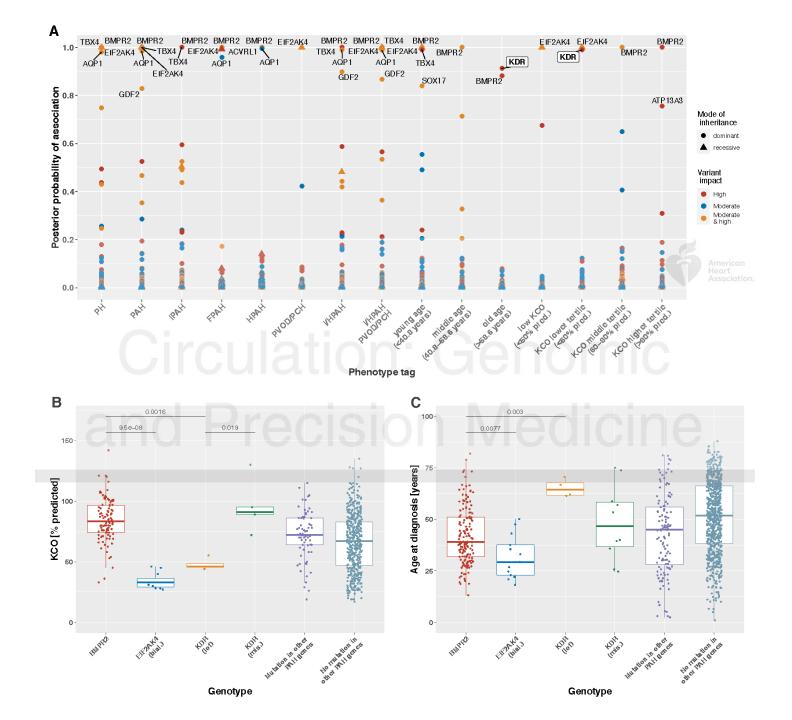
Figure 2. Rare variant association study results revealing established and novel genotypephenotype links. A, Figure showing phenotype tags on the x-axis and corresponding posterior probability of genotype-phenotype association on the y-axis, as calculated by BeviMed. The definitions of the tags are listed in Table 1. Shape and colour of points indicate the mode of inheritance and impact/consequence type of variants driving the association. Box-and-whisker plots showing the distribution of (B) the transfer coefficient for carbon monoxide (KCO) and (C) the age at diagnosis stratified by genotype across the PAH domain. The two-tailed Wilcoxon signed-rank test was used to determine differences in the medians of the distributions, which are indicated by the bars at the top of the figures providing the respective p-values. Abbreviations: bial. - biallelic, lof - loss-of-function, mis. - missense.

Figure 3. Summary of rare single nucleotide variants (SNVs) and small insertions and deletions (indels) identified in the novel PAH candidate gene KDR. A, Only rare predicted deleterious variants in KDR are shown (MAF<1/10,000 and CADD≥10). SNVs and indels are represented by colored lollipops on top of the protein sequence. The domain annotations were retrieved from Uniprot (accession number P35968). Lollipop colors indicate the consequence type and sizes represent the variant frequency within a cohort. Missense variants that are predicted to be deleterious (SIFT) and damaging (PolyPhen-2) are colored in red, otherwise in yellow (i.e. SIFT and PolyPhen-2 disagree). High impact variants are labelled with the respective HGVS notation.

The number of variants by predicted consequence type and cohort is provided in the table. B, Familial segregation of KDR nonsense variant c.183G>A (p.Trp61*) with PAH (i.e. reduced KCO and late onset) from father (W000229) to daughter (W000229.d). Sanger sequencing results are shown in the chromatograms.

Figure 4. Chest computerized tomography (CT) scans of patients carrying high impact KDR mutations. A, Axial image of CT pulmonary angiogram at the level of the right ventricle (RV) moderator band, showing flattening of interventricular septum, leftwards bowing of the interatrial septum and the enlargement of the right atrium (RA) and RV, indicative of RV strain; bilateral pleural effusion, larger on the right side. B, Axial image of a pulmonary CT angiogram demonstrating enlarged pulmonary artery and mild central lung ground-glass opacity (GGO). C, Axial high-resolution CT slice of the chest in the lung window showing a trace of non-specific GGO with a central distribution. D, Coronal image showing the trace of central GGO and enlarged central pulmonary arteries. Axial high-resolution CT slice of the chest in the lung window showing apical subpleural fibrosis (E), and very minor subpleural fibrosis at the lung bases (F). Axial high-resolution CT slice of the chest in the lung window showing subpleural GGO at apical level (G), and mild GGO at mid-thoracic level (H). Patients: E001392 (A, B), E003448 (C, D), W000229 (E, F), W000274 (G,H).





A *KDR* SNVs and indels

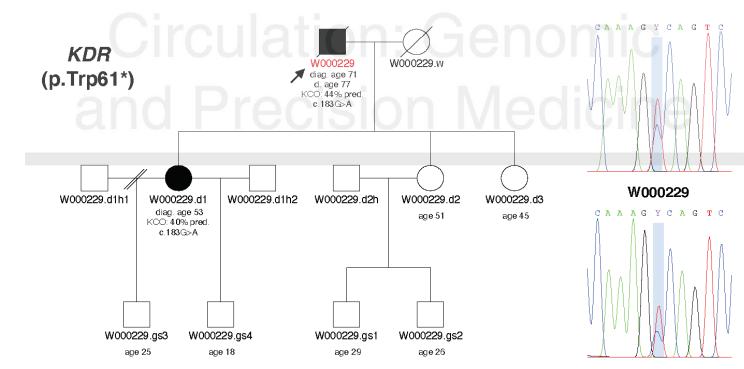
NBR PAH cases p.Trp61Cysfs*16 p.Arg1022* . p.Trp61* •• lg_3 í 532 49 229 295 333 417 677 754 793 834 964 1022 1088 114711791214 1256 1356 c.490-1G>A US Biobank PAH cases p.Arg1022* c.658+1G>A p.Trp101* . Protein tyrosine kinas 101 141 238 299 333 403 438 532 649 749 791 834 961 1022 1059 1147 1216 12701304 1356 c.161+1G>T NBR non-PAH controls p.Glu1206*

664 706 761 813



- Stop gained
- Frameshift
- Splice site
- Missense (SIFT / PolyPhen-2 agree)
- Missense (SIFT / PolyPhen-2 disagree)

KDR variant	N	USBB	
consequence	Cases	Controls	Cases
frameshift	1	0	0
nonsense	2	1	2
splice acceptor	1	Americ HeOt	0
splice donor	0	Associ	ation. 2
missense	13	107	31



906 941

1003

1066 1117 1159 1206 1250 1291

1356

W000229.d1

В

66 112 159

247 321

390 427

497 532

