

Obligatory role for PKC δ in PIP₂-mediated activation of store-operated TRPC1 channels in vascular smooth muscle cells

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Key points

- In vascular smooth muscle cells (VSMCs), activation of Ca²⁺-permeable store-operated channels (SOCs) composed of canonical transient receptor potential channel 1 (TRPC1) subunits mediates Ca²⁺ entry pathways that regulate contraction, proliferation and migration, which are processes associated with vascular disease.
- Activation of TRPC1-based SOCs requires protein kinase C (PKC) activity, which is proposed to phosphorylate TRPC1 proteins to promote channel opening by phosphatidylinositol 4,5-bisphosphate (PIP₂). We investigated the identity of the PKC isoform involved in activating TRPC1-based SOCs in rat mesenteric artery VSMCs.
- TRPC1-based SOCs were reduced by PKC δ inhibitors and knockdown of PKC δ expression. Store depletion induced interactions between TRPC1 and PKC δ and PKC δ -dependent phosphorylation of TRPC1. Furthermore, generation of store-operated interactions between PIP₂ and TRPC1 and activation of TRPC1-based SOCs by PIP₂ required PKC δ .
- These findings reveal that PKC δ activity has an obligatory role in activating TRPC1-based SOCs, through regulating PIP₂-mediated channel opening.

Abstract In vascular smooth muscle cells (VSMCs), stimulation of Ca²⁺-permeable canonical transient receptor potential channel 1 (TRPC1)-based store-operated channels (SOCs) mediates Ca²⁺ entry pathways that regulate cell contraction, proliferation and migration, which are processes associated with vascular disease. It is therefore important to understand how TRPC1-based SOCs are activated. Stimulation of TRPC1-based SOCs requires protein kinase C (PKC) activity, with store-operated PKC-dependent phosphorylation of TRPC1 essential for channel opening by phosphatidylinositol 4,5-bisphosphate (PIP₂). Experimental protocols used to activate TRPC1-based SOCs suggest that the PKC isoform involved requires diacylglycerol (DAG) but is Ca²⁺-insensitive, which are characteristics of the novel group of PKC isoforms (δ , ϵ , η , θ). Hence, the present study examined whether a novel PKC isoform(s) is involved in activating

Miguel Martín-Aragón Baudel obtained his PhD from Edinburgh Napier University under the supervision of Mark Darlison and Amy Poole before moving to St George's, University of London, to work as a postdoctoral research assistant with Anthony Albert. He is currently working at the University of California, Davis, with Manuel Navedo, continuing his research interests in the ion channel-mediated mechanisms driving vascular contraction and with a role in diseases such as hypertension and diabetes.



TRPC1-based SOCs in contractile rat mesenteric artery VSMCs. Store-operated whole-cell cation currents were blocked by Pico145, a highly selective and potent TRPC1/4/5 channel blocker and T1E3, a TRPC1 blocking antibody. PKC δ was expressed in VSMCs, and selective PKC δ inhibitory peptides and knockdown of PKC δ expression with morpholinos oligomers inhibited TRPC1-based SOCs. TRPC1 and PKC δ interactions and phosphorylation of TRPC1 induced by store depletion were both reduced by pharmacological inhibition and PKC δ knockdown. In addition, store-operated PIP₂ and TRPC1 interactions were blocked by PKC δ inhibition, and PKC δ was required for PIP₂-mediated activation of TRPC1 currents. These results identify the involvement of PKC δ in stimulation of TRPC1-based SOCs and highlight that store-operated PKC δ activity is obligatory for channel opening by PIP₂, the probable activating ligand.

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Introduction

Store-operated channels (SOCs) are Ca²⁺-permeable plasmalemmal ion channels activated by depletion of cytosolic endo/sarcoplasmic (ER/SR) Ca²⁺ stores (Baudel *et al.* 2020). In vascular smooth muscle cells (VSMCs), SOCs are stimulated by vasoconstrictors that activate the G α q protein-coupled receptor signalling pathway leading to phospholipase C (PLC) activity, phosphatidylinositol 4,5-bisphosphate (PIP₂) hydrolysis, inositol 1,4,5-trisphosphate (IP₃) generation and IP₃-mediated depletion of SR Ca²⁺ stores. As such SOCs induce Ca²⁺ entry pathways that regulate vasoconstrictor-mediated contraction, proliferation and migration, and are considered drug targets for treatment of vascular diseases such as hypertension and atherosclerosis.

There is substantial evidence that SOCs represent a diverse family of ion channels with differing molecular composition, activation mechanisms and functions (Albert & Large, 2003; Albert *et al.* 2007; 2009; Cheng *et al.* 2013; Prakriya & Lewis, 2015; Ong *et al.* 2015). In native contractile VSMCs, SOCs have relatively low Ca²⁺ permeability, linear current–voltage (*I*–*V*) rectification properties, a single channel conductance of 2–3 pS and are composed of canonical transient receptor potential (TRPC) channels (Trepakova *et al.* 2001; Xu & Beech, 2001; Albert & Large, 2002a; Bergdahl *et al.* 2005; Liu *et al.* 2005a,b; Xu *et al.* 2005a; Albert *et al.* 2006; Saleh *et al.* 2006; 2008; 2009a; Shi *et al.* 2012a, 2014, 2016, 2017a). It is proposed that molecular composition involves a heteromeric TRPC1/C5 template with TRPC1 being the critical subunit for conferring activation by store depletion, hence the term TRPC1-based SOCs (Xu & Beech, 2001; Xu *et al.* 2005a; Saleh *et al.* 2008; Shi *et al.* 2012a). In comparison, synthetic VSMCs that are involved in cell proliferation, migration and growth and are associated with more pathological functions exhibit TRPC1-based SOCs and also SOCs with properties similar to Orai1-based calcium release-activated channels

(CRACs) such as high Ca²⁺ permeability, pronounced inward rectification, single channel conductance in the fS range and composition of Orai1 proteins (Berra-Romani *et al.* 2008; Beech, 2012; Trebak, 2012, Prakriya & Lewis, 2015). It is important to highlight that, in the present study, we examined activation mechanisms of TRPC1-based SOCs in freshly isolated and primary cultured single VSMCs and tissue lysates that have a native contractile phenotype (Shi *et al.* 2016, 2017a) and probably do not involve Orai1 proteins (Shi *et al.* 2017b). These TRPC1-based SOCs are probably important in regulating contractility and switching of VSMCs from contractile to synthetic phenotypes (Berra-Romani *et al.* 2008; Matchkov *et al.* 2012).

How store depletion stimulates TRPC-based SOCs is controversial, especially compared to Orai1-based CRACs for which it is well-established that store depletion induces the ER/SR store Ca²⁺ sensor stromal interaction molecule 1 (STIM1) to oligomerize and translocate to the plasma membrane where it interacts with Orai1 to induce channel assembly and gating (Prakriya & Lewis, 2015). However, there is growing evidence that store depletion also activates TRPC-based SOCs through STIM1-mediated processes, potentially involving direct interactions between TRPC and Orai1 proteins (Liao *et al.* 2014), activation of Orai1-based CRACs as a prerequisite for TRPC1 channel opening (Ambudkar *et al.* 2017), and direct interactions between TRPC and STIM1 proteins (Worley *et al.* 2007; Yuan *et al.* 2009; Lee *et al.* 2014; Asanov *et al.* 2015). In native contractile VSMCs, our recent work indicates that store depletion stimulates TRPC1-based SOCs through a novel STIM1-mediated G α q/PLC β 1 pathway, which probably induces channel opening by regulating interactions between protein kinase C and PIP₂ (Shi *et al.* 2016, 2017a).

There is considerable evidence that protein kinase C (PKC) activity is critical for activation of TRPC1-based SOCs in native contractile VSMCs (Large *et al.* 2009; Albert, 2011). PKC inhibitors prevent activation

of TRPC1-based SOCs and reduce store-operated phosphorylation of TRPC1, and PKC activators stimulate TRPC1-based SOCs (Saleh *et al.* 2008, 2009a; Shi *et al.* 2012a, 2016). Activation of TRPC1-based SOCs and phosphorylation of TRPC1 by physiological vasoconstrictors are also reduced by PKC inhibitors (Albert & Large, 2002b; Saleh *et al.* 2006; 2009b; Shi *et al.* 2012b, 2014, 2016). It is proposed that PKC activity stimulates opening of TRPC1-based SOCs through phosphorylation of TRPC1 proteins to promote PIP₂ binding that acts as the activating ligand (Shi *et al.* 2014, 2016, 2017a). As such, activation of TRPC1-based SOCs by store depletion or PKC activators is inhibited by anti-PIP₂ antibodies and pre-treatment with PI4-kinase inhibitors which deplete PIP₂ levels (Saleh *et al.* 2009a; Shi *et al.* 2012b). In addition, activation of TRPC1-based SOCs by the water soluble PIP₂ analogue diC8-PIP₂ is prevented by PKC inhibitors (Saleh *et al.* 2009a; Shi *et al.* 2012b). These results indicate that interactions between PKC activity and PIP₂ have obligatory roles in activation of TRPC1-based SOCs; PKC cannot activate TRPC1-based SOCs without PIP₂ and vice versa (Baudel *et al.* 2020). The present study further examines the importance of PKC activity in activating TRPC1-based SOCs by investigating the PKC isoform(s) involved.

The PKC family comprises of at least 11 homologous serine/threonine kinases divided into three groups according to their basic structure and activation requirements: conventional PKC isoforms (α , β I, β II and γ) require both Ca²⁺ and diacylglycerol (DAG), novel PKC isoforms (δ , ϵ , η and θ) require DAG but are Ca²⁺-insensitive, and atypical PKC isoforms (ζ , ι and λ) are activated by lipid mediators such as phosphatidylserine and do not require Ca²⁺ or DAG (Salamanca & Khalil, 2005; Ringvold & Khalil, 2017).

Many of these PKC isoforms are expressed in VSMCs and are proposed to regulate several physiological and pathological processes, including those reported to involve a role for TRPC1-based SOCs (Salamanca & Khalil, 2005; Ding *et al.* 2011; Fan *et al.* 2014; Ringvold & Khalil, 2017). In native contractile VSMCs, stimulation of TRPC1-based SOCs and PKC-dependent phosphorylation of TRPC1 by store-depletion requires PLC β 1 activity (Shi *et al.* 2016), DAG activates TRPC1-based SOCs through a PKC-dependent mechanism (Saleh *et al.* 2006; 2008; Large *et al.* 2009; Albert, 2011), and TRPC1-based SOCs are activated by store depleting agents that probably increase (e.g. the SR Ca²⁺-ATPase inhibitor cyclopiazonic acid), decrease (e.g. the cell-impermeable and -permeable high affinity Ca²⁺ chelators BAPTA and BAPTA-AM) or produce little change in [Ca²⁺]_i (e.g. the cell-permeable low affinity Ca²⁺ chelator TPEN). These results suggest that the PKC isoform involved requires DAG, although it is probably Ca²⁺-insensitive; these are characteristics of the novel group of PKC isoforms. These ideas on the identity

of the PKC isoform involved form the basis of the present work, and our findings indicate that the PKC δ novel isoform is the probable candidate involved in activation of TRPC1-based SOCs in native contractile rat mesenteric artery VSMCs.

Methods

Ethical approval

All animal procedures were carried out in accordance with guidelines laid down by St George's, University of London Animal Welfare Committee, and conform with the principles and regulations described by the Service Project Licence: 70/8512, and also to the principles and regulations of *The Journal of Physiology* as described by Grundy (2015). Male Wistar rats (8–12 weeks old) were used for the purpose of the present study. Rats were supplied from Charles River (Margate, UK) and housed and maintained in standard sized plastic cages at the Biological Research Facility at St George's, University of London, under a 12:12 h light/dark photocycle, at 18–20 °C and ~50% relative humidity, with water and laboratory rodent diet (Specialist Dietary Services, UK) available *ad libitum*. Animals were culled by cervical dislocation in accordance with the UK Animals Scientific Procedures Act of 1986 and as revised by European Directive 2010/63/EU.

Cell isolation and tissue lysates

Mesenteric arteries were dissected and cleaned of adherent fat in physiological salt solution containing (mmol L⁻¹): 126 NaCl, 6 KCl, 10 glucose, 11 Hepes, 1.2 MgCl₂, and 1.5 CaCl₂ with pH adjusted to 7.2 with 10 mol L⁻¹ NaOH. Single VSMCs were enzymatically dispersed and tissue lysates prepared as described previously (Shi *et al.* 2016, 2017a).

PKC δ knockdown

Knockdown of PKC δ was performed by transfection of vessel segments with morpholino anti-sense oligonucleotides as described previously (Stott *et al.* 2018). PKC δ morpholino oligomers and a control oligomer containing five mismatched nucleotides (10 μ M; Genetools, Philomath, OR, USA) were mixed with lipofectamine 2000 (Life Technologies, Carlsbad, CA, USA) in Opti-MEM and left at room temperature for 2 h. Mesenteric artery segments were cultured with this mix in Dulbecco's modified Eagle's medium F-12 with 1% penicillin/streptomycin at 37°C for 48 h. Vessel segments were then enzymatically dispersed into single VSMCs or used as tissue lysates as required.

Electrophysiology

Whole-cell and cell-attached patch clamp techniques were used to record TRPC1-based SOCs with an Axopatch 200B amplifier (Axon Instruments, Union City, CA, USA) at room temperature (20–23°C) using bath and patch pipette solutions, data analysis and experimental protocols as described previously (Shi *et al.* 2017a). In experiments investigating the effects of phorbol 12,13-dibutyrate, a phorbol ester and PKC activator, and diC8-PIP₂ on whole-cell cation currents, 750 ms duration voltage ramps from +100 mV to –150 mV were applied every 30 s from a holding potential of 0 mV. The patch pipette solution contained (mM): 126 CsCl, 1.2 MgCl₂, 10 nHepes, 11 glucose, 10 BAPTA, 4.8 CaCl₂, ~100 nM free internal Ca²⁺ concentration as calculated using MaxChelator (<https://somapp.ucdmc.ucdavis.edu/pharmacology/bers/maxchelator>), 1 Na₂ATP, 0.2. NaGTP, pH 7.2 with Tris. The external solution contained (mM): 126 NaCl, 1.5 CaCl₂, 10 Hepes, 11 glucose, 0.1. DIDS, 0.1 niflumic acid and 0.005 niflumic acid, pH 7.2 with NaOH.

Immunoprecipitation and western blotting

Freshly isolated or cultured vessel segments were prepared for immunoprecipitation, one-dimensional protein gel electrophoresis and immunoblotting as described previously (Shi *et al.* 2017a). The primary antibodies used were: mouse anti-PKC ϵ (dilution 1:500; sc-1681; Santa Cruz Biotechnology, Santa Cruz, CA, USA), mouse anti-PKC θ (dilution 1:500; sc-1680; Santa Cruz Biotechnology), mouse anti-PKC η (dilution 1:500; sc-136 036; Santa Cruz Biotechnology) and rabbit anti-PKC δ (dilution 1:1000; ab182126; Abcam, Cambridge, MA, USA). Rabbit anti-TRPC1 antibody (1 μ g mL⁻¹) was generated by GenScript (Piscataway, NJ, USA) using peptide sequences from a previously characterized putative extracellular region (Xu & Beech, 2001). Visualization were performed using anti-rabbit and anti-mouse secondary antibodies conjugated to IRDye 800RD or IRDye 680CW (dilution 1:10 000; Li-Cor Biosciences, Cambridge, UK) as appropriate and the Odyssey Infrared Imaging System (Li-Cor Biosciences). Protein band intensities were measured using Image Studio software (Li-Cor Biosciences) and normalized to smooth muscle actin (dilution 1:2000; #ab5694; Abcam) when quantified.

Immunofluorescence

Single VSMCs were fixed with 4% (w/v) paraformaldehyde for 15 min, cells were treated with 0.1 mol L⁻¹ glycine for 5 min and permeabilized with phosphate-buffered saline (PBS) containing 0.1% (v/v) Triton X-100 (PBS-T) for 15 min at room temperature. Cells were

then incubated with PBS-T containing 1% (w/v) BSA for 1 h at room temperature to block non-specific binding of antibodies. Immunostaining was performed with mouse anti-PKC ϵ (dilution 1:500; sc-1681; Santa Cruz Biotechnology), mouse anti-PKC θ (dilution 1:200; sc-1680; Santa Cruz Biotechnology), rabbit anti-PKC η (dilution 1: 200; ab4134; Abcam), rabbit anti-PKC δ (dilution 1:1000; ab182126; Abcam) overnight at 4°C. Cells were then washed and incubated with a 488 fluorophore-conjugated donkey anti-goat secondary antibody (dilution 1:1000; A-11 055; Alexa Fluor; Thermo Fisher Scientific, Waltham, MA, USA) for 1 h at room temperature. Unbound secondary antibodies were removed by washing with PBS, and nuclei were labelled with 4',6-diamidino-2-phenylindole mounting medium (Sigma, Poole, UK). Control experiments were performed by omitting either primary or secondary antibodies. Cells were imaged using an A1R confocal microscope (Nikon, Tokyo, Japan). PIP₂ immunofluorescence experiments were performed in the presence of saponin with rabbit anti-PIP₂ (sc-53 412, 1:50; Santa Cruz Biotechnology) as described previously (Edimo *et al.* 2016).

Proximity ligation assay

Interactions between different proteins were investigated with a Duolink[®] *in situ* detection kit (Sigma) as described previously (Shi *et al.* 2017a). Single VSMCs were fixed and permeabilized as per immunofluorescence and blocking was performed with Duolink blocking buffer for 1 h at 37°C. Cells were then incubated with appropriate antibodies overnight at 4°C as per immunofluorescence experiments plus mouse anti-P-Ser (dilution 1:100; sc-16B4; Santa Cruz Biotechnology), mouse anti-P-Thr (dilution 1:100; sc-5267; Santa Cruz Biotechnology), mouse anti-TRPC1 (dilution 1:100; sc-133 076; Santa Cruz Biotechnology) and rabbit anti-TRPC5 (T5E3), which was generated by GenScript using peptide sequences from a previously characterized putative extracellular region (1 μ g mL⁻¹) ((Xu & Beech, 2001; Xu *et al.* 2005a,b). The rest of the protocol was conducted in accordance with the manufacturer's instructions. Fluorescent puncta were visualized with an A1R confocal microscope and images were analysed with ImageJ Fiji (<https://fiji.sc>). The mean number of puncta per cell was calculated by counting the number of particles across a z-stack of the cell.

Reagents

Pico145 was a generous gift provided by Robin S Bon and David J Beech (University of Leeds, Leeds, UK). The cell permeable PKC δ inhibitor, δ V1-TAT peptide (RRRQRRKRGY-SFNSYELGSL) was synthesized by Mimotopes (Wirral, UK). PKC δ non-permeable δ PKC₈₋₁₇ peptide inhibitor (SFNSYELGSL) was purchased from

AnaSpec (AnaSpec, EGT Corporate Headquarters, Fremont, CA, USA). PKC ϵ peptide inhibitor was purchased from Santa Cruz Biotechnology. All other drugs were purchased from Sigma or Tocris (Abingdon, UK). Agents were dissolved in distilled H₂O or 0.1% DMSO. DMSO alone had no effect on whole-cell or single channel currents.

Statistical analysis

All statistical analysis was performed using Prism, version 8 (GraphPad Software Inc., San Diego, CA, USA). Data were calculated as the mean \pm SD, with n indicating the number of data points. Data points were generated from at least three different isolated VSMCs or tissue lysate preparations. To compare between two or more I - V relationships, two-way ANOVA with Tukey's multiple comparisons test was used, and differences in means at -80 mV are reported. To compare between two data sets, paired or unpaired t tests were used. $P < 0.05$ was considered statistically significant.

Results

PKC-dependent store-operated currents are composed of TRPC1 subunits in rat mesenteric artery VSMCs

In our first series of experiments, we confirmed that PKC-dependent TRPC1-based SOCs are functionally expressed in native contractile VSMCs from freshly isolated rat mesenteric arteries using a highly selective and potent TRPC1/C4/C5 channel blocker Pico145 (Rubaiy *et al.* 2017), an externally-acting TRPC1 blocking antibody

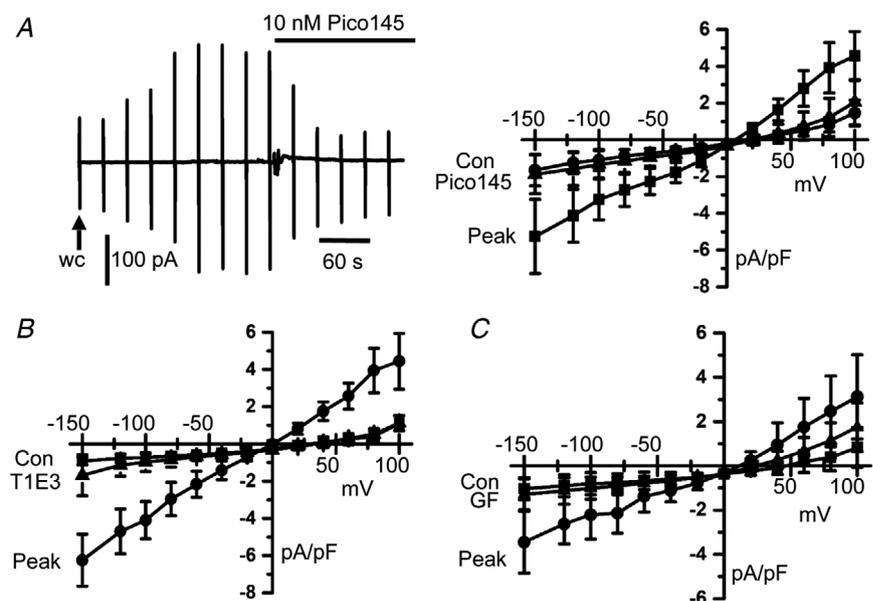
T1E3 (Xu & Beech, 2001, Xu *et al.* 2005b) and the pan-PKC isoform selective inhibitor GF109203X. Figure 1A shows that passive depletion of internal Ca²⁺ stores, following cell dialysis with a patch pipette solution containing high concentrations of BAPTA and no added Ca²⁺, induced whole-cell cation currents that had a relatively linear I - V relationship, an E_{rev} between 0 mV and +20 mV, with a mean control amplitude of -1.15 ± 0.62 pA pF⁻¹ at -80 mV which increased to a mean peak amplitude of -2.74 ± 0.88 pA pF⁻¹ ($n = 6$, $p = 0.0015$) and reduced to -0.89 ± 0.35 pA pF⁻¹ by bath application of Pico145 ($n = 6$, $p = 0.0002$, two-way ANOVA, Tukey's multiple comparisons test). Figure 1B and C show that bath application of T1E3 and GF109203X also inhibited store-operated whole-cell currents, with mean peak amplitudes at -80 mV reduced respectively from -2.96 ± 0.92 pA pF⁻¹ to -0.84 ± 0.43 pA pF⁻¹ ($n = 6$, $p = 0.0001$) and from -2.14 ± 0.92 pA pF⁻¹ to -0.91 ± 0.45 pA pF⁻¹ ($n = 7$, $p = 0.0051$, two-way ANOVA, Tukey's multiple comparisons test).

There is considerable evidence that heteromeric TRPC1/C5 molecular structures compose TRPC1-based SOCs in VSMCs from different rabbit, mouse and human vascular beds (see Introduction). In proximity ligation assay (PLA) studies, Figure 2 shows that fluorescent puncta with a mean number per cell of 51.5 ± 12.6 ($n = 14$) produced using anti-TRPC1 and anti-TRPC5 antibodies were present at (or close to) the plasma membrane in freshly isolated rat mesenteric artery VSMCs.

These findings clearly confirm that PKC-dependent TRPC1-based SOCs are functionally expressed in native contractile rat mesenteric artery VSMCs, similar to previous studies in VSMCs from different rabbit, mouse and human vascular preparations (see Introduction).

Figure 1. TRPC1 compose PKC-dependent SOCs in native contractile VSMCs

A, representative trace and mean I - V relationships showing that the development of store-operated whole-cell currents from control to peak levels in freshly isolated rat mesenteric artery VSMCs following obtaining whole-cell configuration (wc) was inhibited by bath application of Pico145. Vertical deflections represent currents evoked by voltage ramps from +100 mV to -150 mV (750 ms duration) every 30 s from a holding potential of 0 mV. B and C, Mean data showing that bath applications of T1E3 ($1 \mu\text{g mL}^{-1}$) and GF109203X ($3 \mu\text{M}$) inhibited store-operated whole-cell currents.



PKC δ is the dominant novel PKC isoform in VSMCs

The PKC isoform involved in activating TRPC1-based SOCs in VSMCs requires DAG but is Ca²⁺ insensitive and is probably a member of the novel PKC isoform family (see Introduction). We therefore investigated the expression of novel PKC isoforms in tissue lysates from freshly isolated rat mesenteric arteries. We used brain lysates as positive controls where novel PKC isoforms have been previously identified (Fleegal *et al.* 2005; Popp *et al.* 2006; Callender & Newton, 2017; Wang *et al.* 2019; Yang *et al.* 2019).

Figure 3*a* shows PKC δ expression with relatively low levels or little expression of PKC ϵ , PKC η and PKC θ were found in tissue lysates from rat mesenteric arteries. Using the same anti-PKC novel isoform antibodies, expression of PKC δ , PKC ϵ , PKC η and PKC θ was present in brain lysates. In addition, Figure 3*b* shows that immunocytochemical studies revealed PKC δ staining at (or close to) the plasma membrane of VSMCs with little staining recorded for PKC ϵ , PKC η and PKC θ isoforms.

PKC δ activity is essential for activation of TRPC1-based SOCs

In our next series of experiments, we examined whether pharmacological inhibition of PKC δ and knockdown of PKC δ expression using morpholino technology resulted in an anticipated decrease in activation of TRPC1-based SOCs. We focused on the PKC δ isoform based on the expression studies detailed above and previous data on the expression and function of PKC δ in vascular smooth muscle (Salamanca & Khalil, 2005; Ringvold & Khalil, 2017).

Figure 4*A* reveals that store-operated whole-cell TRPC1-based currents were inhibited by bath application of the cell-permeable PKC δ inhibitor δ V1-TAT, with mean peak amplitude reduced from -6.18 ± 2.34 pA pF⁻¹ to -1.26 ± 0.57 pA pF⁻¹ at -80 mV ($n = 6$, $p = 0.0001$, two-way ANOVA, Tukey's multiple comparisons test).

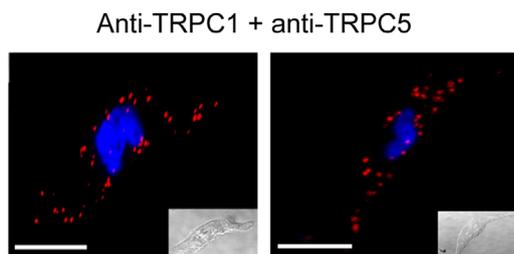


Figure 2. Expression of TRPC1-TRPC5 interactions in native contractile VSMCs

Representative PLA images from two different freshly isolated rat mesenteric artery VSMCs showing the association between TRPC1 and TRPC5. These images, and other PLA images, represent puncta observed across a z-stack of each cell. Scale bar = 10 μ m.

Figure 4*B* also shows that following inclusion of the cell-impermeable PKC δ peptide inhibitor δ PKC δ_{8-17} in the patch pipette solution store depletion induced a small whole-cell TRPC1-based current with mean control amplitude increasing from -0.97 ± 0.27 pA pF⁻¹ to a mean peak amplitude of -2.01 ± 1.1 pA pF⁻¹ at -80 mV ($n = 6$, $p = 0.0902$, two-way ANOVA, Tukey's multiple comparisons test).

Because expression of PKC ϵ was also observed in VSMCs (Fig. 3) and other studies have reported the expression of PKC ϵ in the vasculature (Salamanca & Khalil, 2005; Ringvold & Khalil, 2017), we investigated the potential contribution of this PKC isoform in activating store-operated TRPC1-based SOCs. Figure 4*C* shows that, following inclusion of a PKC ϵ -specific peptide inhibitor in the patch pipette solution, store depletion activated whole-cell TRPC1 currents with a mean control amplitude increasing from -0.84 ± 0.21 pA pF⁻¹ to -3.39 ± 1.42 pA pF⁻¹ at -80 mV ($n = 5$, $p = 0.0006$), which was subsequently inhibited by bath application of δ V1-TAT to -0.99 ± 0.56 pA pF⁻¹ ($n = 5$, $p = 0.0012$, two-way ANOVA, Tukey's multiple comparisons test).

Figure 5*A* shows that incubation of freshly isolated rat mesenteric artery segments with anti-sense morpholino oligomers against PKC δ for 48 h significantly decreased protein expression of PKC δ by $\sim 50\%$ compared to a mismatched oligomer ($n = 3$, $p = 0.0202$, unpaired t test). Figure 5*B* shows that, in VSMCs isolated from vessel segments expressing mismatched (scrambled) oligomers, store depletion induced whole-cell TRPC1 currents with relative linear rectification and a mean control amplitude at -80 mV that increased from -1.07 ± 0.68 pA pF⁻¹ to a mean peak amplitude of -2.57 ± 0.89 pA pF⁻¹ ($n = 9$, $p = 0.0003$, two-way ANOVA, Tukey's multiple comparisons test), which were similar to currents recorded from freshly isolated VSMCs (Figs 1 and 4). By contrast, Figure 5*B* shows that, in VSMCs expressing specific PKC δ oligomers, store depletion activated small whole-cell TRPC1 currents, with the control mean amplitude at -80 mV increasing from -1.05 ± 0.59 pA pF⁻¹ to a mean peak amplitude of -1.36 ± 0.69 pA pF⁻¹ ($n = 10$, $p = 0.8240$, two-way ANOVA, Tukey's comparisons test). As such, mean peak amplitudes of store-operated TRPC1 current at -80 mV were greatly reduced in VSMCs expressing specific PKC δ compared to scrambled oligomers ($p = 0.0051$, two-way ANOVA, Tukey's comparisons test).

Store depletion induces TRPC1 and PKC δ interactions and PKC δ -dependent phosphorylation of TRPC1

The above pharmacological and molecular evidence indicates that PKC δ activity is essential for activation

of TRPC1-based SOCs in native contractile VSMCs. Therefore, we investigated whether store depletion induced PKC δ and TRPC1 interactions and PKC δ -dependent phosphorylation of TRPC1 in freshly isolated rat mesenteric artery tissue lysates and VSMCs.

Immunoprecipitation with anti-TRPC1 antibodies followed by immunoblotting with an anti-PKC δ antibody revealed that interactions between these two

proteins are absent in unstimulated tissue but occur following incubation with BAPTA-AM or TPEN (Fig. 6A). Furthermore, the PLA studies shown in Figure 6B confirmed that interactions between TRPC1 and PKC δ were absent in unstimulated VSMCs, whereas robust puncta at (or close to) the plasma membrane were induced between TRPC1 and PKC δ following pre-treatment with BAPTA-AM and TPEN.

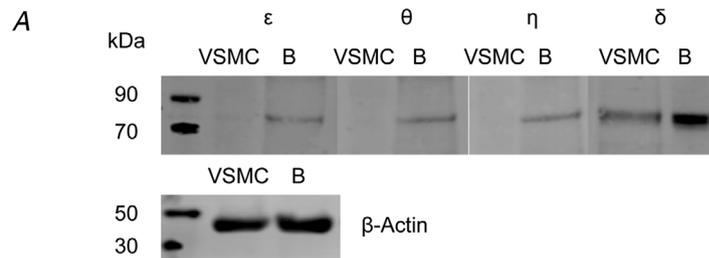


Figure 3. Expression of novel PKC isoforms in native contractile VSMCs
 A, representative western blots showing expression of the novel PKC isoforms PKC ϵ , PKC θ , PKC η and PKC δ in rat mesenteric artery VSMCs and brain (B) protein lysates. B, representative immunocytochemical images of rat mesenteric VSMCs immunolabelled with the same anti-novel PKC isoform antibodies used in (A). Scale bar = 10 μ m.

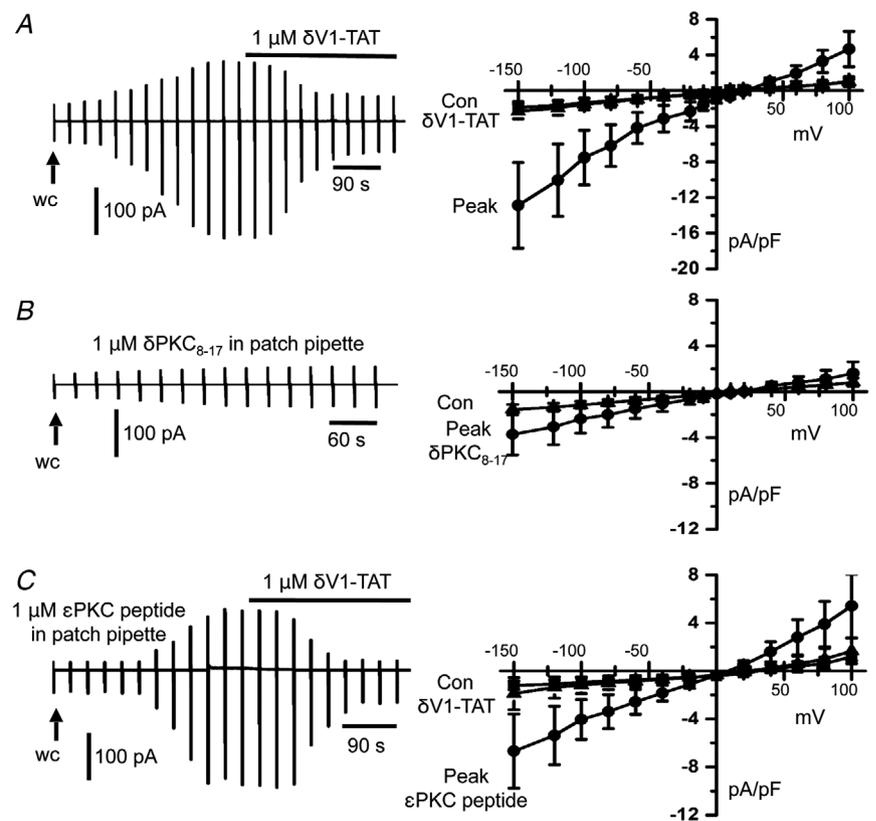
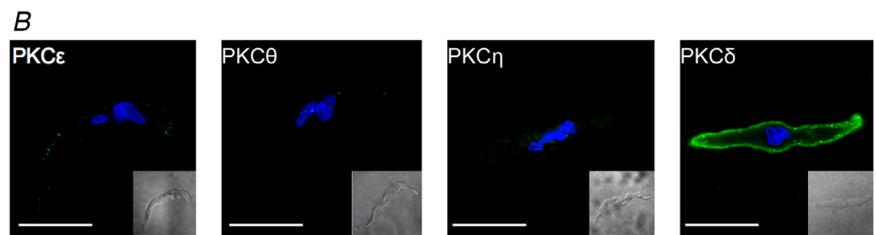


Figure 4. Pharmacological PKC δ inhibitors reduce TRPC1-based SOCs
 A, representative trace and mean I - V relationship showing that the peak amplitude of store-operated whole-cell TRPC1 currents in freshly isolated rat mesenteric artery VSMCs was reduced by bath application of δ V1-TAT. B, a small store-operated whole-cell TRPC1 current was induced following inclusion of δ PKC $_{8-17}$ peptide inhibitor in the patch pipette solution. C, inclusion of ϵ PKC peptide inhibitor in the patch pipette appeared to have little effect on the development of store-operated whole-cell TRPC1 currents, which were subsequently inhibited by bath application of δ V1-TAT.

PLA experiments performed with a mixture of anti-phosphorylated serine/threonine and anti-TRPC1 antibodies revealed a low number of fluorescent puncta in unstimulated freshly isolated VSMCs, which is indicative of a low level of basal TRPC1 phosphorylation. Figure 7A shows that the mean number of puncta per cell was increased following pre-treatment with BAPTA-AM from 6.1 ± 3.9 ($n = 19$) to 20.5 ± 9.9 ($n = 30$, $p = 0.0001$) and inhibited by coapplication of δ V1-TAT to 8.3 ± 4.2 ($n = 25$, $p = 0.0001$, unpaired t test). In addition, Figure 7B shows that transfection of vessel segments with specific PKC δ morpholino oligomers also reduced the BAPTA-AM-induced increase in mean puncta number compared to scrambled oligomers from 22.4 ± 10.4 ($n = 30$) to 7.2 ± 4.9 ($n = 25$, $p = 0.0001$, unpaired t test). Importantly, Figure 7A and B shows a similar mean number of puncta using mismatched oligomers

compared to non-transfected freshly isolated VSMCs under unstimulated (8.7 ± 4.3 , $n = 26$ vs. 6.1 ± 3.9 , $n = 19$, $p = 0.049$) or BAPTA-AM-treated conditions (22.4 ± 10.4 , $n = 30$ vs. 20.5 ± 9.9 , $n = 30$, $p = 0.4653$, unpaired t test), indicating that the transfection process had little effect on this mechanism.

Taken together, these results indicate that store depletion induces interactions between TRPC1 and PKC δ in VSMCs, and that these interactions cause PKC δ -dependent phosphorylation of TRPC1.

PKC δ mediates store-operated interactions between PIP $_2$ and TRPC1, as well as PIP $_2$ -evoked TRPC1 currents

In our previous work, we identified that PIP $_2$ has an obligatory role in the activation of TRPC1-based SOCs

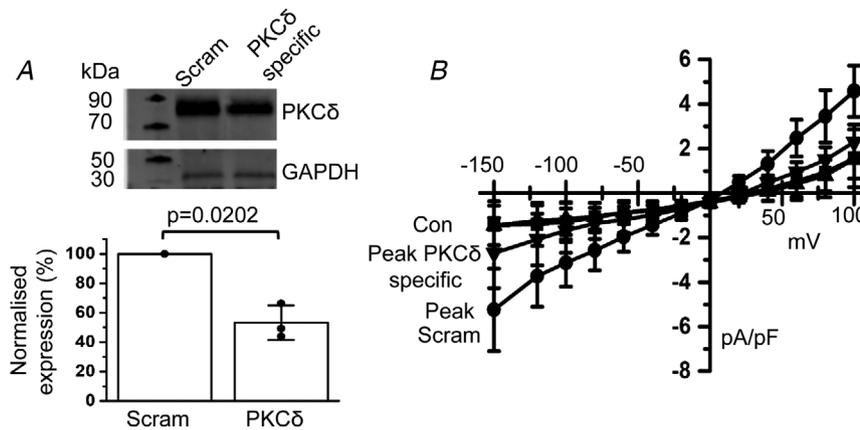


Figure 5. Knockdown of PKC δ expression reduces activation of TRPC1-based SOCs

A, representative western blots and mean data showing that PKC δ -specific morpholino oligomers reduced PKC δ expression in rat mesenteric artery protein lysates compared to scrambled/mismatched oligomers (Scram). Band intensity was normalized to β -GAPDH to calculate percentage expression (vs. scrambled, $n = 3$, $p = 0.0202$, unpaired t test). B, mean I - V relationships showing that the mean peak amplitude of store-operated whole-cell TRPC1 currents was reduced in the presence of PKC δ -specific ($n = 10$) compared to scrambled ($n = 9$) morpholino oligomers.

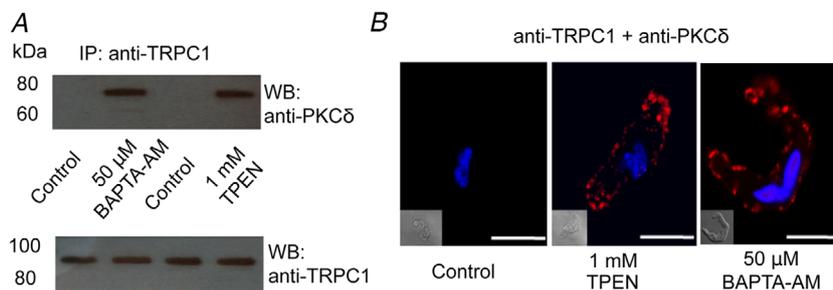


Figure 6. Store depletion induces associations between TRPC1 and PKC δ in native contractile VSMCs

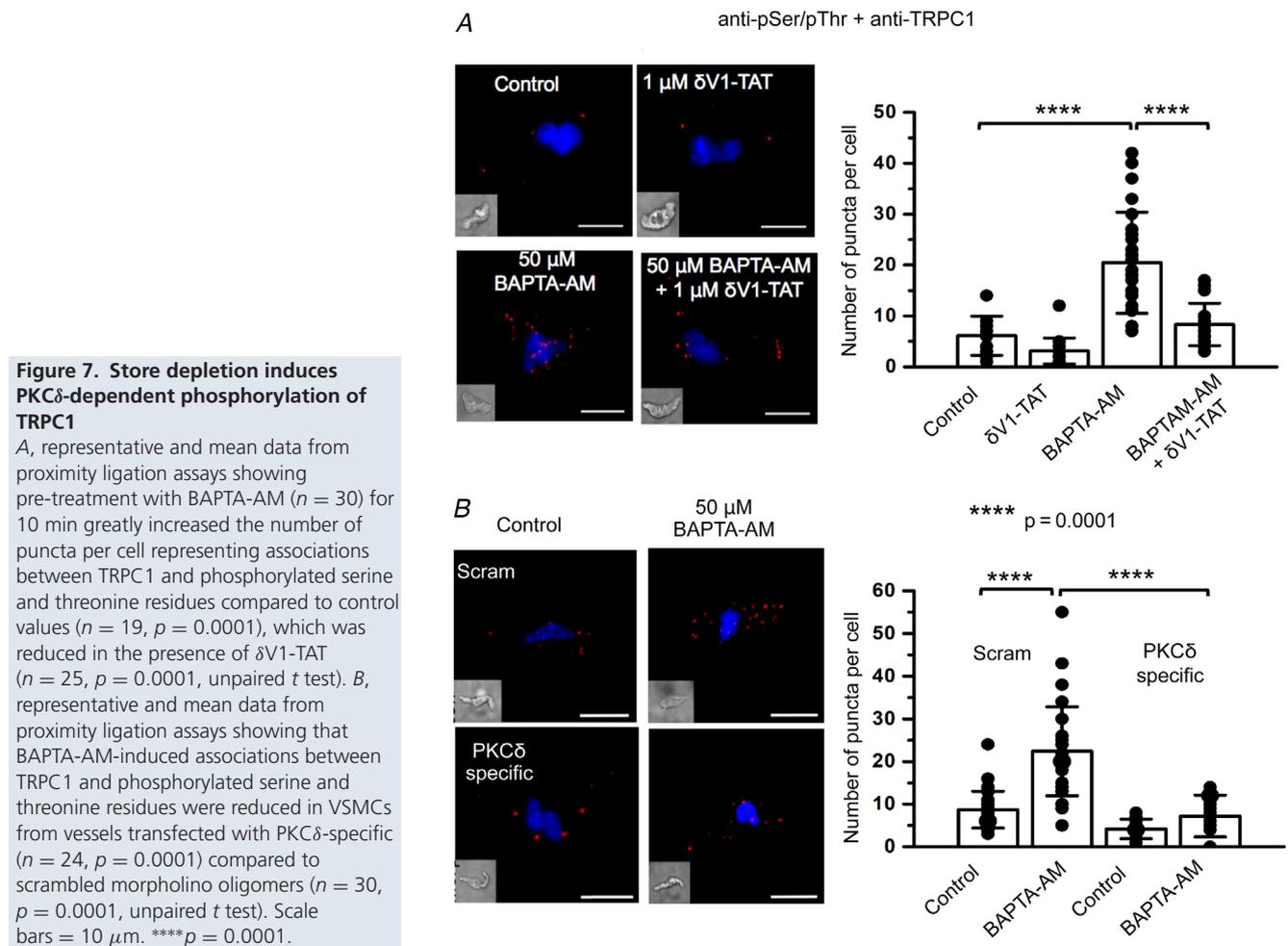
A, representative western blots with anti-TRPC1 and anti-PKC δ antibodies following immunoprecipitation of freshly isolated rat mesenteric artery tissue lysates with an anti-TRPC1 antibody show that pre-treatment with the BAPTA-AM and TPEN induced associations between TRPC1 and PKC δ but did not alter TRPC1 expression. B, representative images of proximity ligation assays from freshly isolated rat mesenteric artery VSMCs showing a low level of association between TRPC1 and PKC δ in unstimulated VSMCs, which was greatly increased following pre-treatment with TPEN and BAPTA-AM for 10 min. Scale bars = 10 μ m.

in VSMCs, potentially as the activating ligand, and that this process requires PKC-dependent phosphorylation of TRPC1 (see Introduction). Therefore, we investigated whether store-operated interactions between PIP₂ and TRPC1 require PKC δ activity using immunocytochemistry and PLA, and also whether PKC δ is essential for activation of store-operated whole-cell TRPC1 currents by PIP₂.

In freshly isolated rat mesenteric artery VSMCs, Figure 8A shows that PIP₂ immunostaining was highly expressed at (or close to) the plasma membrane in unstimulated cells and BAPTA-AM and δ V1-TAT had no obvious effect on this distribution. With respect to the staining produced with anti-PIP₂ antibodies being related to endogenous PIP₂, Figure 8A shows that pre-treatment with high concentrations of wortmannin (20 μ M), which inhibits PI-4/PI-5 kinases to reduce PIP₂ recycling and promote PIP₂ depletion, produced a considerable reduction in the PIP₂ signal (Suh & Hille, 2005; Saleh *et al.* 2009a; Shi *et al.* 2014). In control experiments, TRPC1 immunostaining was unaffected by BAPTA-AM, δ V1-TAT or wortmannin treatment (Fig. 8A). We next adapted this

immunostaining protocol for PLA experiments. Figure 8B reveals that there was a low level of interactions between PIP₂ and TRPC1 in unstimulated VSMCs, which were increased by BAPTA-AM from a mean puncta number per cell of 7.6 ± 5.2 ($n = 23$) to 34.6 ± 14.1 ($n = 24$, $p = 0.0001$) and inhibited in the presence of δ V1-TAT or following pre-treatment of wortmannin, respectively, to 6.5 ± 2.9 ($n = 20$, $p = 0.0001$) and 6.1 ± 3.7 ($n = 19$, $p = 0.0001$, unpaired t test).

In freshly isolated VSMCs, Figure 9A shows that bath application of the phorbol ester 12,13-dibutyrate (PDBu) increased whole-cell non-selective cation currents from a mean control amplitude of -0.71 ± 0.13 pA pF⁻¹ to a mean peak amplitude of -2.12 ± 0.8 pA pF⁻¹ at -80 mV ($n = 8$, $p = 0.0001$, two-way ANOVA, Tukey's comparisons test). By contrast, inclusion of diC8-PIP₂ in the patch pipette solution had no effect on whole-cell cation currents (-80 mV, -0.75 ± 0.18 vs. -1.0 ± 0.08 , $n = 5$, $p = 0.6151$, two-way ANOVA, Tukey's comparisons test). Figure 9B shows that dialysing cells with diC8-PIP₂ in the patch pipette solution increased PDBu-activated whole-cell cation currents, with mean peak amplitude at -80 mV



increased from -2.12 ± 0.8 pA pF⁻¹ ($n = 8$) (Fig. 9A) to -2.88 ± 0.78 pA pF⁻¹ at -80 mV ($n = 7$, $p = 0.0291$, unpaired t test). Figure 9B also shows that PDBu-evoked whole-cell cation currents induced in the presence of diC8-PIP₂ were inhibited by bath applications of T1E3 and δ V1-TAT, respectively, to -1.13 ± 0.24 pA pF⁻¹ ($n = 5$, $p = 0.0001$) and -0.84 ± 0.38 pA pF⁻¹ at -80 mV ($n = 7$, $p = 0.0001$ two-way ANOVA, Tukey's comparisons test) indicating that these currents were mediated by TRPC1 and required PKC δ activity.

Noradrenaline-evoked TRPC1 channels require PKC δ

The present study clearly demonstrates that the PKC δ isoform is important for activation of TRPC1 channels by agents that deplete internal Ca²⁺ stores. Therefore, in the final series of experiments, we investigated the physiological significance of these findings by examining whether PKC δ is involved in activation of TRPC1 channels by the vasoconstrictor and α_1 -adrenoceptor agonist methoxamine (MO). Figure 10 shows that, in unstimulated VSMCs, there was a low level of

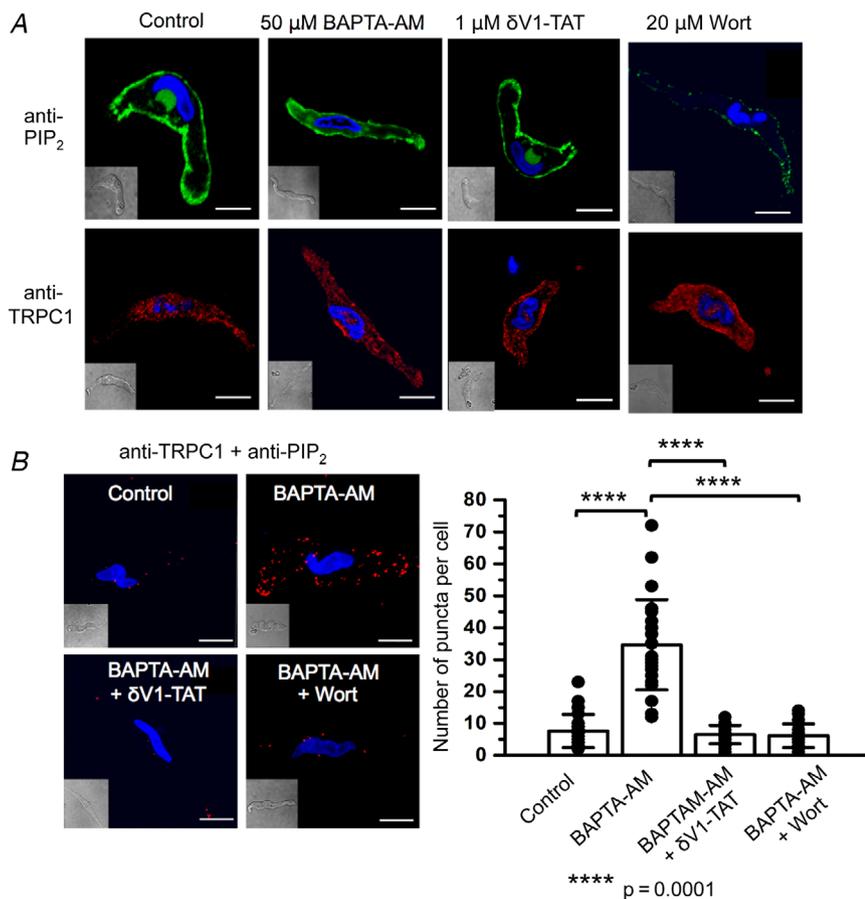


Figure 8. Store-operated interactions between TRPC1 and PIP₂ require PKC δ

A, representative images from freshly isolated rat mesenteric VSMCs showing expression of PIP₂ (green) and TRPC1 (red) following pretreatment with BAPTA-AM for 10 min, wortmannin (Wort) for 20 min and δ V1-TAT for 10 min. **B**, representative images and mean data from PLA experiments showing that control interactions between TRPC1 and PIP₂ ($n = 23$) were greatly increased by pre-treatment with BAPTA-AM for 10 min ($n = 23$, $p = 0.0001$), and this was reduced by coapplication with δ V1-TAT ($n = 20$, $p = 0.0001$) and wortmannin ($n = 19$, $p = 0.0001$, unpaired t test). Scale bars = 10 μ m.

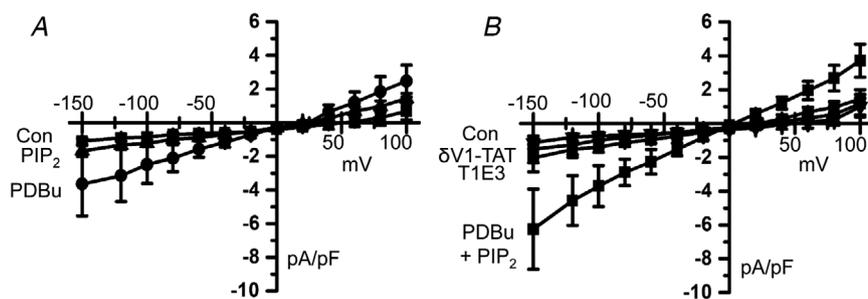


Figure 9. PIP₂ increases PDBu-induced whole-cell TRPC1 currents

A, mean I - V relationships from freshly isolated rat mesenteric artery VSMCs showing that bath application of PDBu but not inclusion of diC8-PIP₂ in the patch pipette solution induced a whole-cell current. **B**, mean I - V relationships showing that, following cell dialysis with diC8-PIP₂, PDBu-evoked whole-cell currents had a greatly increased peak amplitude compared to currents induced by PDBu alone (**A**) and were inhibited by T1E3 and δ V1-TAT.

spontaneous 2 pS TRPC1-based channel activity in cell-attached patches held at -80 mV, and that mean peak open probability (NP_o) was increased following bath application of MO from 0.14 ± 0.07 to 0.51 ± 0.2 ($n = 6$, $p = 0.0035$) and was significantly reduced by subsequent coapplication of $\delta V1$ -TAT to 0.11 ± 0.08 ($n = 6$, $p = 0.0029$, paired t test).

Discussion

The present study reveals that activity of PKC δ , a member of the novel PKC isoform subgroup, is essential for activation of TRPC1-based channels evoked by store depletion and α_1 -adrenoceptor stimulation in native contractile rat mesenteric artery VSMCs. It is proposed that PKC δ -dependent phosphorylation of TRPC1 is required for TRPC1 channel activation by PIP $_2$, the probable activating ligand. These results provide further evidence that PKC activity and PIP $_2$ are obligatory for activation of TRPC1-based SOCs in VSMCs, ion channels implicated in regulating contraction, proliferation and migration that are processes linked to hypertension and atherosclerosis. As such, regulating PKC δ activity and PIP $_2$ levels may comprise therapeutic targets for these and other vascular diseases.

TRPC1-based SOCs require PKC activity

Our results show that the well-established protocol for inducing store depletion, the inclusion of a high concentration of BAPTA in the patch pipette solution, evoked a whole-cell non-selective cation current with a relatively linear I - V relationship and an E_{rev} of between 0 mV and $+20$ mV in freshly isolated rat mesenteric artery VSMCs, which is similar to currents previously recorded in rabbit and mouse VSMCs (Shi *et al.* 2012a, 2014, 2016, 2017a,b). Considerable evidence obtained using pharmacological agents, antibodies as blocking agents and TRPC1 knockout mice indicates that these store-operated currents are mediated by a heteromeric TRPC1/TRPC5 molecular template (Xu & Beech, 2001; Xu *et al.* 2005a; Saleh *et al.* 2008; Shi *et al.* 2012a). In further support of this proposal, the present study shows

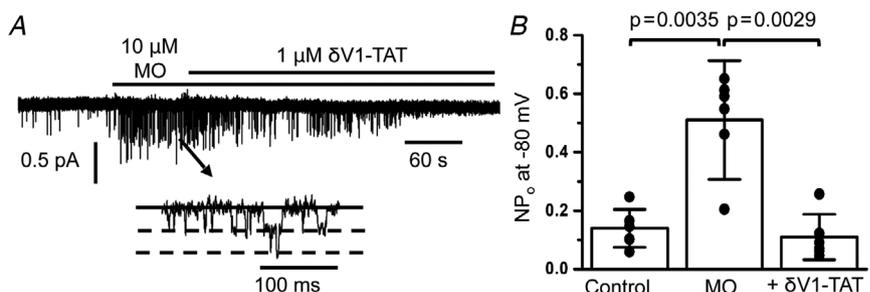
that store-operated whole-cell currents were inhibited by Pico145, a highly selective and potent TRPC1/C4/C5 channel blocker (Rubaiy *et al.* 2017) and T1E3, a TRPC1 antibody raised against an extracellular pore region and known to act as a blocking agent (Xu & Beech, 2001; Xu *et al.* 2005b). In addition, using PLA, the present study reveals that TRPC1 and TRPC5 co-localize within <40 nm of each other at (or close to) the plasma membrane of VSMCs. It should be noted that the electrophysiological and pharmacological properties of these store-operated currents in freshly isolated VSMCs exhibiting a contractile phenotype are considerably different from those of Orai1-based CRACs (Prakriya & Lewis, 2015), indicating that Orai1 proteins are probably not involved, as reported previously (Shi *et al.* 2017b).

Previous studies show that PKC activity and PKC-mediated phosphorylation of TRPC1 are essential for activation of TRPC1-based SOCs in native contractile VSMCs (Albert & Large, 2002b; Saleh *et al.* 2006, 2008; Large *et al.* 2009; Albert, 2011; Shi *et al.* 2012a, 2014, 2016). The present study provides further support for an excitatory role for PKC, with the pan-PKC isoform selective inhibitor GF109203X inhibiting store-operated whole-cell TRPC1 currents in freshly isolated rat mesenteric artery VSMCs. Therefore, identifying the specific PKC isoform responsible for activating TRPC1-based SOCs was the main aim of the present study.

PKC δ is the PKC isoform responsible for activation of TRPC1-based SOCs and store-operated TRPC1 phosphorylation

The PKC family is subdivided into conventional PKC (α , β and γ), novel PKC (δ , ϵ , η and θ) and atypical PKC (ζ and ι/κ) isoforms (Salamanca & Khalil, 2005; Ringvold & Khalil, 2017). Given that DAG analogues activate TRPC1 channels through a PKC-dependent mechanism in VSMCs (Saleh *et al.* 2006; 2008; Large *et al.* 2009; Albert, 2011), and that the Ca $^{2+}$ store depletion protocols used to activate TRPC1-based SOCs probably increase, decrease or do not alter [Ca $^{2+}$] $_i$, the PKC isoform(s) responsible is probably belongs to the novel subfamily (see Introduction).

Figure 10. Methoxamine-evoked TRPC1 channel activity is mediated by PKC δ
A and B, representative trace and mean open probability (NP_o) data showing that bath application of methoxamine evoked TRPC1 channel activity in cell-attached patches held at -80 mV ($n = 6$, $p = 0.0035$), which was reduced by coapplication of $\delta V1$ -TAT ($n = 6$, $p = 0.0029$, paired t test).



Western blotting and immunocytochemical analysis demonstrated that PKC δ is expressed in native contractile rat mesenteric artery VSMCs and was distributed at (or close to) the plasma membrane. There was greater expression of PKC δ compared to other novel PKC isoforms in VSMCs and tissue lysates; however, caution is warranted when suggesting that PKC δ is the dominant novel PKC isoform in VSMCs because expression variations may be linked to potential differences between the affinity and specificity of the anti-novel PKC isoform antibodies used.

Inhibition of PKC δ activity via well-characterized selective PKC δ peptide inhibitors and knockdown of PKC δ expression using morpholino oligomers resulted in a significant reduction in the development and peak amplitude of store-operated whole-cell TRPC1 currents. Moreover, immunoprecipitation and PLA studies revealed that store depletion induced interactions between TRPC1 and PKC δ . Importantly, PLA experiments showed that store depletion induced puncta formation between anti-TRPC1 and anti-phosphorylated serine and threonine antibodies that was greatly reduced by PKC δ peptide inhibitors and PKC δ selective morpholino oligomers, which indicates that store depletion induces PKC δ -dependent phosphorylation of TRPC1. Taken together, these findings provide strong evidence for the PKC δ isoform being essential in the activation of TRPC1-based SOCs in native contractile VSMCs.

PKC δ has also been reported to participate in store-operated Ca²⁺ entry in airway smooth muscle (Gao *et al.* 2012), proliferation and migration of VSMCs (Ding *et al.* 2011; Fan *et al.* 2014), development of the myogenic response (Kashihara *et al.* 2008), and membrane insertion of TRPM4 channels and associated vasoconstriction (Crnich *et al.* 2010; Garcia *et al.* 2011). Interestingly, TRPC1 has also been associated with many of these vascular functions (Earley & Brayden, 2015) and, accordingly, in the future, it may be useful to investigate the functional association between TRPC1 and PKC δ activities in the vasculature. By contrast to the results outlined above, PKC activity has an inhibitory action on Orai1-based CRACs with PKC inhibitors including GF109203X and knockdown of PKC β isoform increasing Orai1 activity (Kawasaki *et al.* 2010), which further indicates that Orai1 proteins or Orai1-based CRACs are not involved in the activation of TRPC1-based SOCs in native contractile VSMCs.

PKC δ activity is required for PIP₂-mediated activation of TRPC1 channels

We have previously described the obligatory requirement of PIP₂ in the activation of TRPC1-based SOCs and proposed that PKC-dependent phosphorylation of TRPC1 is required for this phospholipid to act as the activating ligand (Large *et al.* 2009; Saleh *et al.* 2009a,b; Albert,

2011; Shi *et al.* 2012a,b, 2014). In the present study, using PLA, store depletion induces interactions between PIP₂ and TRPC1 at (or close to) the plasma membrane of VSMCs that are reduced by PKC δ peptide inhibitor. To provide evidence that endogenous PIP₂ is involved in these results, we showed that a high concentration of wortmannin (20 μ M) that depletes PIP₂ levels by inhibiting PI-4/PI-5 kinase-mediated PIP₂ synthesis (Suh & Hille, 2005) reduced PIP₂ immunostaining and store-operated associations between TRPC1 and PIP₂. It should be noted that a high concentration of wortmannin also inhibits myosin light chain kinase (MLCK) and PI(3) kinase, although previous findings have shown that lower concentrations (<1 μ M) inhibiting MLCK and PI-3 kinase but not depleting PIP₂ levels had no effect on activation of TRPC1 channel activity in VSMCs (Saleh *et al.* 2009a; Shi *et al.* 2014). The PKC activator PDBu activated whole-cell TRPC1 currents that were increased in amplitude when the water soluble PIP₂ analogue diC8-PIP₂ was included in the patch pipette solution and were inhibited by a PKC δ peptide inhibitor. Cell dialysis with diC8-PIP₂ failed to activate any whole-cell currents on its own, as reported previously (Albert *et al.* 2008), which indicates that the effects of PDBu and PIP₂ are not merely additive but probably represent synergism between the two molecules. Taken together, these results re-enforce the idea that PIP₂ is the activating ligand of TRPC1-based SOCs, and that this requires PKC δ -dependent phosphorylation of TRPC1.

The proposed excitatory roles of PKC and PIP₂ on TRPC1-based SOCs are opposite to the action of these molecules in the activation of non-TRPC1-containing channels (e.g. TRPC3/C6/C7 channels) in native contractile VSMCs (Venkatachalam *et al.* 2003; Albert & Large, 2004; Large *et al.* 2009; Shi *et al.* 2010; Albert, 2011). This subgroup of TRPC channels, known as receptor-operated channels (ROCs), is activated by receptor-mediated generation of DAG, which leads to channel opening via PKC independent mechanisms, with subsequent PKC activity induced by DAG producing channel inhibition. Interestingly, stimulation of TRPC1-based SOCs is proposed to inhibit TRPC6-based SOCs through inducing Ca²⁺ influx and PKC activity, which suggests the involvement of a conventional PKC isoform (Shi *et al.* 2010). A distinct role of PIP₂ on TRPC3/C6/C7-based ROCs is less clear, with both inhibitory and excitatory actions on channel activity being proposed (Albert *et al.* 2008; Imai *et al.* 2012). These findings further indicate that TRPC1-based SOCs and TRPC3/C6/C7-based ROCs form distinct channel structures with differing activation mechanisms involving differing PKC isoforms, and probably distinct functions in VSMCs.

Computation predication of potential PKC δ -dependent phosphorylation sites within the TRPC1 sequence using GPS 3.0 (Xue *et al.* 2008) reveals five intracellular serine residues, with Ser619 and Ser752 at the C-terminal domain

being of significance in that both of these sites are close to a known PIP₂-binding domain (Kwon *et al.* 2007). Hence, it is possible that PKC δ -dependent phosphorylation of one or both sites increases PIP₂ binding to TRPC1, leading to channel opening. Other studies have demonstrated that protein kinase A, protein kinase G and calmodulin kinase II have inhibitory actions on TRPC1-based SOCs in VSMCs (Liu *et al.* 2005a; Albert *et al.* 2006; Chen *et al.* 2011), and therefore TRPC1 phosphorylation by these kinases might inhibit TRPC1-based SOCs by reducing TRPC1 and PIP₂ interactions. Similar roles for kinase activities with respect to modulating lipid–protein interactions are well-established in the regulation of K⁺ channel subtypes (Logothetis *et al.* 2015).

One important aspect is how PKC δ activity is stimulated by store depletion. Our proposed hypothesis based on previous work indicates that store depletion induces STIM1–TRPC1 interactions, which stimulate a Gq-PLC β 1 pathway that generates DAG and PKC activity, leading to PKC-dependent phosphorylation of TRPC1 (Baudel, 2020). PKC-dependent phosphorylation of TRPC1 leads to a dissociation of interactions between TRPC1 and the PIP₂ binding protein myristoylated alanine-rich C-kinase substrate (MARCKS), with the latter molecule then releasing PIP₂ into the local environment, allowing it to act as the channel activating ligand. The present study indicates that the PKC δ isoform is probably an essential PKC isoform in this process and this will be important to investigate in detail in future studies.

Physiological relevance of PKC δ -mediated TRPC1 channels

TRPC1 channel activity induced by the α_1 -adrenoceptor agonist and vasoconstrictor methoxamine was reduced by a PKC δ peptide inhibitor, which suggests that a similar store-operated activation process involving PKC δ may be used by physiological stimulants of TRPC1 channels. This is further supported by findings showing that PLC β 1 and STIM1 knockdown both significantly inhibit noradrenaline-evoked TRPC1 channel activity (Shi *et al.* 2016, 2017b) and noradrenaline induces an increase in PKC-dependent phosphorylation of TRPC1 and stimulates MARCKS to release PIP₂, which is then available to interact with TRPC1 (Shi *et al.* 2014). It was previously proposed that vasoconstrictor agents may also activate TRPC1-based SOCs independently of store depletion (Albert & Large, 2002b; Saleh *et al.* 2006; Large *et al.* 2009; Saleh *et al.* 2009b; Shi *et al.* 2010, 2012b; Albert, 2011), which suggests that TRPC1 channels may also act as ROCs. The current and previous findings certainly suggest that a significant contribution to physiological stimulation of TRPC1 channels probably occurs via a store-operated pathway.

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Additional information

Data availability statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Competing interests

The authors declare that they have no competing interests.

Author contributions

MMAB, JS, WL and AA all contributed to the conception or design of the work, as well as the analysis of data or the interpretation of data for the work, and were involved in drafting the work or revising it critically for important intellectual content. MMAB and JS were involved in the acquisition of data. All authors approved final version of the manuscript submitted for publication and agree to be accountable for all aspects of the work. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

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Keywords

PKC, PIP, store-operated channels, TRPC1, vascular smooth muscle

Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Statistical Summary Document