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- 3 Title: Arylacetamide deacetylase (AADAC) gene polymorphism and HIV infection
- 4 affect the exposure of Rifapentine: a population pharmacokinetics analysis.

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35 ABSTRACT

36 Rifapentine is a rifamycin used to treat tuberculosis. As for rifampicin, plasma exposures of 37 rifapentine are associated with treatment response. While concomitant food intake and HIV infection explain part of the pharmacokinetic variability associated with rifapentine, few 38 studies have evaluated the contribution of genetic polymorphisms. We evaluated the effects 39 40 of functionally significant polymorphisms of the genes encoding OATP1B1, PXR, CAR, and AADAC on rifapentine exposure. Two studies evaluating novel regimens amongst Southern 41 African patients with drug-susceptible pulmonary tuberculosis were included in this analysis. 42 In RIFAQUIN, rifapentine was administered in the continuation phase of antituberculosis 43 treatment in 1200mg once-weekly or 900mg twice-weekly doses. In Daily-RPE 450 or 44 45 600mg were given daily during the intensive-phase of treatment. Nonlinear mixed-effects modelling was used to describe the pharmacokinetics of rifapentine and to identify significant 46 covariates. A total of 1144 drug-concentration measurements, from 326 patients, were 47 included in the analysis. Pharmacogenetic information was available for 162 patients. A one-48

compartment model with first-order elimination and transit compartment absorption 49 described the data well. In a typical patient (body weight of 56kg, fat-free-mass of 45kg), the 50 values of clearance and volume of distribution were 1.33L/h and 25L, respectively. Patients 51 carrying the AA variant (65.4%) of AADAC rs1803155 were found to have 10.4% lower 52 clearance. HIV+ infected patients had 21.9% lower bioavailability. Once weekly doses of 53 1200 mg were associated reduced clearance (-13.2%), compared to more frequently 54 administered doses. Bioavailability was 23.3% lower amongst patients participating in the 55 Daily-RPE study compared to RIFAQUIN. This is the first study to report the effect of 56 AADAC rs1803155AA on rifapentine clearance. The observed increase in exposure is modest 57 and unlikely to be of clinical relevance. The difference in bioavailability between the two 58 59 studies is probably related to the different food concomitant to the dose. HIV coinfected patients had lower rifapentine exposures. 60

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62 INTRODUCTION

Rifamycins play a key role in the multidrug treatment of tuberculosis. Their sterilizing 63 activity is exposure-dependent (1-3). Rifapentine, was approved by the Food and Drug 64 Administration (FDA) in 1998 for the treatment of pulmonary tuberculosis (3,4). Rifapentine 65 pharmacokinetics are influenced by age, weight, dosing pattern, human immunodeficiency 66 virus (HIV) infection, and sex (5,6). Rifapentine is less rapidly absorbed than rifampicin, 67 68 with peak plasma concentrations reached within 5 hours. Concomitant food markedly increases its absorption; the extent of rifapentine absorption increased by 33-86% when given 69 with meals (7). Rifapentine has a half-life of approximately 12 hours in humans (8,9). With 70 71 its long half-life and excellent sterilizing activity, rifapentine is an attractive alternative to 72 rifampicin and is increasingly used to treat active tuberculosis and latent infection. However, there is marked interpatient variability in rifamycin pharmacokinetics (10). The primary metabolic pathways for rifapentine involve deacetylation to the primary enzymatic metabolite 25-desacetyl rifapentine, which is mediated by human arylacetamide deacetylase (*AADAC*) and non-enzymatic hydrolysis resulting in formation of the secondary metabolites 3-formyl rifapentine and 3-formyldesacetyl rifapentine (11). Protein binding of rifapentine is estimated to be about 98% (3,12). Like other rifamycin's, rifapentine induces its own metabolism (9).

Previously published data indicate that single nucleotide polymorphisms (SNPs) in the solute 79 carrier organic anion transporter 1B1 (SLCO1B1) gene encoding the OATP1B1 80 transmembrane receptor affect rifampicin concentrations (13,14). SLCO1B1 rs4149032 C>T 81 polymorphism, found in 70% of South Africans with tuberculosis living in Cape Town, was 82 associated with 20% and 28% reductions in rifampicin bioavailability in heterozygotes and 83 homozygotes, respectively (14). Rifamycins are also substrates of the drug efflux pump P-84 glycoprotein coded for by the polymorphic ABCB1 gene (15) and are metabolized mainly by 85 polymorphic human arylacetamide deacetylase (AADAC) (16). Human rifamycin exposures 86 are also modulated by the pregnane X receptor (PXR) and constitutive and rostane (CAR)87 nuclear receptors (17). Since the development of resistance to rifamycins and their 88 bactericidal effects are related to rifamycin concentrations, SNPs substantially influencing 89 90 rifamycin concentrations may be of therapeutic importance. Little is known about the pharmacogenetic correlates of rifapentine pharmacokinetics, which may potentially help in 91 92 finding the optimal dose of rifapentine. Therefore, the aim of this study was to determine the effect of polymorphisms of SLCO1B1, PXR, CAR, and AADAC on rifapentine 93 pharmacokinetics. 94

95 **RESULTS**

A total of 326 patients were included in the study and contributed a total of 1151 96 concentrations-time points. Only 7 concentrations were below the LLOQ and were omitted 97 from the analysis. The median body weight and age of the study participants were 56 kg and 98 32 years respectively. All demographic characteristics are summarized in Table 1. 99

The population pharmacokinetics of rifapentine was well described by a one-compartment 100 model with first-order elimination and transit compartment absorption. Fat-free-mass(FFM) 101 was found to be the best size descriptor for clearance (ΔOFV 93 points, p<0.001 when 102 103 including FFM for allometric scaling on clearance and 23 points better than using body weight) and total body weight for volume of distribution (ΔOFV 20, p<0.001). The 104 105 absorption of rifapentine was described using a series of transit compartments, which 106 significantly improved the model with respect to simple first-order absorption (ΔOFV 421, p < 0.001). In a typical patient (46 kg FFM and 56 kg weight), the values of clearance and 107 volume of distribution were 1.33 L/h and 25 litres. Final parameter estimates (shown in Table 108 109 3) were in agreement with the previously published results (6,18) and a VPC of the final model is shown in Figure 1. 110

111 Of 326 patients, pharmacogenetic data was available for 162 (49.7%) all of whom were enrolled from South African sites. The distribution of genotype and allele frequencies are 112 presented in Table 2. SLCO1B1 rs2306283 and AADAC rs1803155 variant alleles were 113 114 found in 82% of patients whereas the NR112 rs2472677 and NR112 rs1523130 variant alleles existed at a low overall frequency of 33.5% and 16.4% respectively. In keeping with our 115 previous findings among South Africans in Cape Town (14), the SLCO1B1 rs4149032 variant 116 117 allele frequency was found to be 0.75 (Table 2).

118 After screening and inclusion of genetic information (and imputation of missing genotype 119 with a mixture model), patients homozygous for AADAC rs1803155 AA polymorphism were Downloaded from http://aac.asm.org/ on January 25, 2019 by guest

found to have 10.4% lower clearance of rifapentine compared to subjects that were 120 rs1803155 GG or GA (ΔOFV 6.2, p=0.013). Initially the three categories of rs1803155 (AA, 121 GA, GG) were analysed as separate groups to estimate the respective effects of GA and GG. 122 However, the estimated effects were similar for GG & GA, and when combined the model 123 goodness of fit was not affected. Using the principle of parsimony, we decided to use the 124 125 simpler model, as the effects of GG and GA were not statistically significant. The other pharmacogenetic variants did not affect the pharmacokinetic parameters. 126

127 Patients infected with HIV infection were found to have 21.9% lower bioavailability (ΔOFV 42, p < 0.001). The patients who were treated with high 1200 mg doses of rifapentine tended 128 to have clearance reduced by 13.2% compared to other dose groups (ΔOFV 17, p<0.001). 129 130 The pharmacokinetic differences between the two studies were explored and it was found that the bioavailability of rifapentine in the Daily RPE study was 23.3% lower than in the 131 RIFAQUIN study (Δ OFV 59, p<0.001). The pharmacogenetic covariates other than AADAC 132 rs1803155 polymorphism didn't have significant effects on the pharmacokinetic parameters. 133

DISCUSSION 134

The present study is the first to investigate the influence of various plausible physiologically-135 136 relevant candidate gene polymorphisms on rifapentine pharmacokinetics. We developed a population pharmacokinetic model of rifapentine, which was consistent with previous reports, 137 and tested the effect of genotype information on the pharmacokinetic parameters. We showed 138 that the AADAC rs1803155 polymorphism is associated with rifapentine clearance. Subjects 139 carrying the AA genotype had 10.4% lower clearance than those carrying AG or GG, thus 140 leading to increased rifapentine exposure. The low clearance due to this polymorphism is 141 142 consistent with previous studies reporting decreased activity of AADAC activity due to the presence of the variant allele (19). The majority of patients in our study had the AADAC 143

rs1803155 AA variant allele which occurred at a frequency of 0.82, and 65% were 144 homozygous for the single nucleotide polymorphism, which could, in part, account for the 145 relatively high rifapentine exposures described. The polymorphism occurs at lower 146 frequencies of 0.50 to 0.64 in European American, African American, Korean, and Japanese 147 populations (19). Another study identified lower rifapentine concentrations in black Africans 148 149 but the influence of pharmacogenetic factors, which might account for the difference in the genotype frequencies between the populations, was not explored (20), whereas Sloan et al., 150 who explored the influence of AADAC gene polymorphisms on rifampicin pharmacokinetics 151 152 in Malawian patients, did not identify a significant relationship (21). The prevalence of variant genotypes is different between African ethnic groups and may be the reason for this 153 154 contrasting effect. As only 3 of 162 patients had rs1803155 GG, no meaningful separate 155 estimate of clearance for this genotype could be obtained. In further attempts to explain variability in rifapentine pharmacokinetics, we explored the effects of several polymorphisms 156 of drug transporters and transcriptional regulators. The choice of polymorphisms was based 157 158 on those previously described to affect drug disposition, and also by previous pharmacogenetic studies conducted on rifampicin. Interestingly, we could not detect the 159 effect SLCO1B1 rs4149032 polymorphism on pharmacokinetics of rifapentine, even with a 160 carrier, no carrier approach. The frequency of SLCO1B1 in our cohort was 0.75, which is in 161 agreement with previous finding in South African patients from the Cape Town region. 162 Similarly, we did not find a statistically significant effect associated with SLCO1B1 163 rs2306283, which existed in our study population at a frequency of 0.82. SLCO1B1 164 polymorphisms have been reported to be associated with low rifampicin levels (13,14) and 165 the lack of effect on rifapentine may suggest differences in the ADME of the two drugs. It 166 167 may be that this transporter does not play a major role in the pharmacokinetics of rifapentine, or that the variant allele is associated with greater induction by rifampicin. We did not 168

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observe an effect due to polymorphisms of the transcriptional regulators. This could be due to
activation of PXR or CAR by rifapentine, which may have overridden any constitutive
effects.

Additionally, we found that HIV infected patients have lower bioavailability of rifapentine. While the association of HIV infection with antituberculosis drug exposures is inconsistent, our findings for rifapentine are consistent with recent studies (22-24). The data available was not sufficient to identify potential drug-drug interactions with the various antiretroviral drugs prescribed concomitantly.

Patients in the higher dose group (1200 mg given once weekly) had increased exposure in the current study contrary to the findings by Savic *et al.*, which describes a decrease in the bioavailability of rifapentine with increased dose (6). The reduced dosing frequency in this group, may have led to reduced auto-induction and thus increased exposure.

181 Previous reports demonstrate that exposure to rifamycins is reduced in males due to a higher 182 FFM: body weight ratio (25). The study by Langdon et al. described a 35% reduction in the 183 clearance of 25-desacetyl rifapentine amongst females (5). In the present analysis, as 184 allometric scaling with FFM accounted for the variability associated with sex, we did not 185 observe any outstanding effects of sex. There was a difference in bioavailability between the two studies included in this analysis. This may be due to differences in food intake with the 186 dose. Rifapentine absorption is strongly enhanced when it is administered with food (7). The 187 finding that the Daily RPE study had a lower bioavailability may arise from the fact that 188 189 meals with the dose were not standardized, in contrast to the RIFAQUIN study where a standard meal was provided throughout the study. 190

191 To conclude, our study is the first to show that the *AADAC* rs1803155 (AA) genotype is 192 associated with lower rifapentine clearance, leading to increased rifapentine exposure. This

effect should be confirmed in a larger independent analysis. The pharmacogenetic association 193 was modest compared to the study effect, which is likely linked to differences in the pattern 194 of food use across the studies and highlights the importance of food recommendations both 195 when the drug is used in a programmatic setting and when its pharmacokinetics is 196 investigated. Additionally, we found that rifapentine exposure was lower in HIV infected 197 198 patients, a finding consistent with previous studies and warranting further investigation to assess whether dose adjustment strategies should be considered. Lastly, patients dosed with 199 200 1200 mg once weekly doses had lower clearance, possibly as a result of less pronounced 201 autoinduction.

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MATERIALS AND METHODS 203

Study population: This analysis was performed on patients diagnosed with pulmonary TB 204 205 from two clinical studies: The Phase III RIFAQUIN study (ISRCTN44153044) (26) and twostage activity-safety study of daily rifapentine (27), hereinafter "Daily RPE" 206 (NCT00814671). A subset of participants from these studies provided their consent to assess 207 208 the effect of genetic polymorphisms of nuclear receptors, drug metabolizing enzymes, and 209 drug transporters on the pharmacokinetics of rifapentine.

210 The RIFAQUIN study included two experimental arms in which patients were dosed with 211 daily moxifloxacin, rifampicin, pyrazinamide, and ethambutol for 2 months followed by a continuation phase with either 4 months of once weekly 1200 mg rifapentine together with 212 213 400 mg moxifloxacin, or 2 months of 400 mg moxifloxacin twice weekly with 900 mg of 214 rifapentine. The RIFAQUIN study was conducted at sites in the Western Cape and Gauteng regions of South Africa and in Harare, Zimbabwe. The doses of rifapentine and moxifloxacin 215 216 were taken with 240 mL of water 15 minutes after a light meal of 2 hard-boiled eggs with

bread. During the 4th month of treatment, blood samples were drawn for determination of 217 218 plasma rifapentine concentrations. The pharmacokinetic assessment involved rich (with a 219 pre-dose and samples at 1, 2, 3, 5, 7, 10, 12, 26, and 50 h after dosing) or sparse sampling (samples drawn around 2, 5, and 24 or 48 h after dosing). 220

221 The Daily RPE study was open-label and had two experimental arms. Patients with pulmonary tuberculosis were randomized to 450 or 600 mg rifapentine daily, which replaced 222 223 600 mg rifampicin during the intensive phase of standard therapy. The study participants 224 were recruited in the Western Cape, South Africa. The patients were advised to take the required Rifapentine dose with food, but no standardised meal was provided during the study 225 226 and no accurate details about food intake with the dose were recorded. Pharmacokinetic 227 sampling was performed at approximately one month after starting therapy and samples were obtained either with intensive (with samples pre-dose and at 0.75, 1.5, 3.5, 5, 12, and 24 h 228 after dose), or sparse sampling (0.5-2 h and 5-8 h after dose). Separate written informed 229 230 consent for the pharmacogenetic study was obtained from participants retrospectively. The pharmacogenetic study was reviewed and approved the Research Ethics Committee of the 231 University of Cape Town and the University of the Witwatersrand. 232

233 Drug determination: Plasma rifapentine concentrations were determined with a validated 234 liquid chromatography-tandem mass spectrometry assay developed in the Division of Clinical 235 Pharmacology, University of Cape Town. Samples were processed with a protein precipitation extraction method using rifaximin as internal standard, followed by high 236 performance liquid chromatography with MS/MS detection using an AB SCIEX API 3200 237 238 instrument. The analyte and internal standard were monitored at mass transitions of the protonated precursor ions m/z 877.3 and m/z 786.3 to the product ions m/z 845.4 and m/z 239 754.1 for rifapentine and rifaximin, respectively. The calibration curves fit quadratic 240 241 (weighted by 1/concentration) regressions over the ranges 0.156 - 40.0 mg/L for rifapentine.

The accuracies for the rifapentine assay were 103.9%, 102.8%, and 97.5% at the low, medium, and high QC levels, respectively, during inter-batch validation. The lower limit of quantification (LLOQ) was 0.156 mg/L.

245 **SNP genotyping:** Genomic DNA was extracted from 200 µL whole blood using QIAamp DNA Mini Kit (Qiagen, Inc., Valencia, California) in accordance with the manufacturer's 246 protocol. DNA was quantified spectrophotometrically using NanoDrop (Thermo Fisher 247 Scientific Inc., Wilmington, Delaware) before storage at -20°C. Genotyping was performed 248 by real-time polymerase chain reaction (PCR) on a DNA Engine Chromo4 system (Bio-Rad 249 Laboratories, Inc., Hercules, California). The PCR protocol involved an initial denaturation 250 step at 95°C for 15 min, followed by 50 cycles of amplification at 95°C for 15 s and final 251 annealing at 60°C for 1 min. TaqMan Genotyping Master Mix and assays for SLCO1B1 252 rs2306283 (SNP ID: C 1901697 20), SLC01B1 rs4149032 (C 1901709 10), NR112 253 rs2472677 (C_26079845_10), NR112 rs1523130 (C_9152783_20), and AADAC rs1803155 254 (C 8911003 1) were obtained from Thermo Fisher Scientific (Waltham, Massachusetts). 255 Allelic discrimination plots and genotype assignments were performed using Opticon 256 Monitor, version 3.1 from Bio-Rad Laboratories. 257

Pharmacokinetic analysis: Rifapentine plasma concentration-time data was analysed using a 258 259 nonlinear mixed-effects model implemented in NONMEM 7.4.2 (28). The execution of runs 260 was through Perl-speaks-NONMEM, Pirana and graphical diagnostics were created using Xpose 4.6.0 and R (29,30). Estimation of typical population pharmacokinetic parameters, 261 along with their random inter-individual (IIV) and inter-occasion (IOV) variability was 262 263 performed using first-order conditional estimation method with ε - η interaction (FOCE INTER). A lognormal distribution was assumed for IIV and IOV and a combined additive 264 and proportional model for the residual unexplained variability (RUV) was evaluated. 265 266 Various structural models were tested including one or two-compartment distribution with

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first-order elimination and first-order absorption with or without lag time or transit 267 compartment absorption (31). The influence of genetic polymorphisms on rifapentine 268 269 pharmacokinetics for patients with unknown genotype was identified using mixture modelling (32). The effect of the genotype was first tested using method EXTRA which 270 271 estimates the association only for the patients with available genetic information but also 272 estimating an additional covariate effect for the unknown genotype. Subsequently, the MIX 273 method to impute values using mixture modelling was applied to include the patient with 274 unknown genotype to strengthen the robustness of the findings (32). Model selection was based on changes in the NONMEM objective function value (ΔOFV), and visual inspection 275 276 of conditional weighted residuals (CWRES) versus time, visual predictive checks (33), and basic goodness of fit plots (GOF). During model development, physiological plausibility and 277 278 the precision of the parameter estimates were also considered. The model parameters of the 279 final model were evaluated for their precision using sampling importance resampling method (SIR) (34). 280

Allometric scaling was applied on clearance (CL), and volume of distribution (V) to adjust 281 for the effect of body size, according to Anderson and Holford (35). Fat-free mass (FFM), 282 and fat mass (FAT) were tested as alternative size predictors through allometric scaling 283 284 instead of total body weight (35,36). After the inclusion of allometric scaling, potential 285 demographic, study site specific and pharmacogenetic covariates were screened inspecting 286 parameter versus covariate plots and then tested in the model using drops in objective 287 function value (assumed to be χ -square distributed and thus using 3.84 points drop as 288 significant at p < 0.05 for the inclusion of a single parameter), while scrutinising the 289 physiological plausibility of the effect (37).

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432 **Table 1:** <u>Demographic and clinical characteristics of patients</u>

Demographic & Clinical	Daily RPE 450 mg	Daily RPE 600 mg	RIFAQUIN 900 mg	RIFAQUIN 1200 mg	Overall (N=326)
Characteristics	(N=44)	(N=41)	(N=116)	(N=125)	
No. of PK samples	166	130	416	432	1144
Sex (male/female)	(33/11)	(32/9)	(72/44)	(81/44)	(218/108)
Number of HIV+ patients	6 (13.6%)	7 (17.1%)	30 (25.9%)	16(12.8%)	59(18.1%)
Median age, range (yrs)	29 (19-61)	29 (18-63)	31 (19-64)	34 (19- 80)	32 (18-80)
Median weight in kg (range)	55 (45-79)	55 (45-94)	55 (38-77)	57 (38- 78)	56 (38-94)
Median FFM in kg (range)	47 (32-58)	47 (32-56)	45 (27-62)	45 (27-60)	45 (27-62)

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Table 2: Observed genotype and allele frequency of Single nucleotide polymorphisms in the study (N=162)

Genotype	Genotype frequency (%)			Allele frequencies	
SLCO1B1 A>G	AA	AG	GG	A	G
rs2306283	8 (4.94)	43 (26.5)	111 (68.5)	0.18	0.82
SLCO1B1 C>T	CC	СТ	TT	С	Т
rs4149032	15 (9.26)	52 (32.1)	95 (58.6)	0.25	0.75
*NR1I2 C>T	CC	СТ	TT	С	Т
rs2472677	71 (44.1)	72(44.7)	18 (11.2)	0.67	0.34
NR1I2 T>C	TT	СТ	CC	Т	С
rs1523130	116 (71.6)	39 (24.1)	7 (4.3)	0.84	0.16
AADAC G>A	GG	GA	AA	G	А
rs1803155	3 (1.85)	53 (32.7)	106 (65.4)	0.18	0.82

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Parameter	Estimate	95% CI	IIV ⁺ or IOV ⁺⁺	95% CI
CL ^a (L/hr)	1.33	1.14;1.54	23.0%+	17.7;28.6
$V^{a}\left(L ight)$	25	21.9;28.4	12.8%+	8.8;17.4
ka (hr ⁻¹)	0.814	0.568;1.26	48.9%++	36.4;59.8
MTT (hr)	1.47	1.20;1.78	37.4%++	28.3;48.6
NN	10.2	6.70;14.0	-	-
F	1 FIXED		20.3%**	14.9;26.4
Proportional residual error (%)	9.56	7.09;13.2	-	-
Additive residual error (mg/L)	0.247	0.143-0.401	-	-
HIV+ effect on F (%)	-21.9	-33.2; -6.64	-	-
Group on 1200 mg dose in RIFAQUIN study on CL (%)	-13.2	-22.8; -4.36	-	-
Daily RPE study on F (%)	-23.3	-35.6: -9.25	-	-
AADAC rs1803155 (AA) effect on CL (%)	-10.4	-17.3; -3.53	-	-

462 CL-oral clearance; V-apparent volume of distribution in the central compartment; k_a -first-

463 order absorption rate constant; MTT- absorption mean transit time; NN- number of
464 hypothetical transit compartments; F- oral bioavailability; HIV+ - Human immunodeficiency

- a The typical values of clearance and volume of distribution reported for a patient with body 467 weight 56 kg and FFM of 46 kg. 468 469 95% CI of parameter estimates obtained with Sampling importance resampling (SIR) n=1000 470 of the final model.
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- Figure 1: Visual predictive check (VPC) for the final rifapentine population pharmacokinetic 473

virus positive; AADAC- arylacetamide deacetylase. IIV- inter-individual variability, and

IOV- inter-occasional variability are expressed as percent coefficient of variation (% CV).



474 model in log scale, stratified according to different dose groups in the analysis.



476 observed plasma concentration. The shaded areas are the 95% confidence intervals for the

477 same percentiles, obtained from re-simulations of the same trial.