Phenotypic Characterization of *EIF2AK4*Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically With Pulmonary Arterial Hypertension

Editorial, see p 2034

BACKGROUND: Pulmonary arterial hypertension (PAH) is a rare disease with an emerging genetic basis. Heterozygous mutations in the gene encoding the bone morphogenetic protein receptor type 2 (*BMPR2*) are the commonest genetic cause of PAH, whereas biallelic mutations in the eukaryotic translation initiation factor 2 alpha kinase 4 gene (*EIF2AK4*) are described in pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis. Here, we determine the frequency of these mutations and define the genotype-phenotype characteristics in a large cohort of patients diagnosed clinically with PAH.

METHODS: Whole-genome sequencing was performed on DNA from patients with idiopathic and heritable PAH and with pulmonary veno-occlusive disease/ pulmonary capillary hemangiomatosis recruited to the National Institute of Health Research BioResource–Rare Diseases study. Heterozygous variants in *BMPR2* and biallelic *EIF2AK4* variants with a minor allele frequency of <1:10 000 in control data sets and predicted to be deleterious (by combined annotation-dependent depletion, PolyPhen-2, and *sorting intolerant from tolerant* predictions) were identified as potentially causal. Phenotype data from the time of diagnosis were also captured.

RESULTS: Eight hundred sixty-four patients with idiopathic or heritable PAH and 16 with pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis were recruited. Mutations in *BMPR2* were identified in 130 patients (14.8%). Biallelic mutations in *EIF2AK4* were identified in 5 patients with a clinical diagnosis of pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis. Furthermore, 9 patients with a clinical diagnosis of PAH carried biallelic *EIF2AK4* mutations. These patients had a reduced transfer coefficient for carbon monoxide (Kco; 33% [interquartile range, 30%–35%] predicted) and younger age at diagnosis (29 years; interquartile range, 23–38 years) and more interlobular septal thickening and mediastinal lymphadenopathy on computed tomography of the chest compared with patients with PAH without *EIF2AK4* mutations. However, radiological assessment alone could not accurately identify biallelic *EIF2AK4* mutation carriers. Patients with PAH with biallelic *EIF2AK4* mutations had a shorter survival.

CONCLUSIONS: Biallelic *EIF2AK4* mutations are found in patients classified clinically as having idiopathic and heritable PAH. These patients cannot be identified reliably by computed tomography, but a low Kco and a young age at diagnosis suggests the underlying molecular diagnosis. Genetic testing can identify these misclassified patients, allowing appropriate management and early referral for lung transplantation.

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Clinical Perspective

What Is New?

- One percent of patients with a clinical diagnosis of pulmonary arterial hypertension (PAH) carry biallelic EIF2AK4 mutations.
- Patients diagnosed clinically with PAH who had a transfer coefficient for carbon monoxide (Kco) <50% predicted and age of diagnosis <50 years were more likely to carry biallelic *EIF2AK4* mutations. The diagnostic yield for genetic testing in this group was 53%.
- Radiological assessment was unable to distinguish reliably between these patients and patients with idiopathic PAH.
- Histology from these patients may show predominately pulmonary arteriopathy, with subtle involvement of the pulmonary veins and capillaries.
- Patients with PAH with biallelic EIF2AK4 mutations had a worse prognosis compared with other patients with PAH.

What Are the Clinical Implications?

- Younger patients diagnosed with idiopathic PAH but with a low Kco have a high frequency of biallelic EIF2AK4 mutations.
- Such patients should be reclassified as having pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis.
- Similar to patients with pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis, these patients have a poor prognosis compared with other patients with PAH.
- The spectrum of radiological and histological changes associated with biallelic EIF2AK4 mutations is wider than previously assumed. The presence of only subtle or infrequent features associated with pulmonary veno-occlusive disease may lead to misclassification of these patients as having PAH.
- Genetic testing allows early identification of these patients, facilitating appropriate management.

ulmonary arterial hypertension (PAH) is a heterogeneous and rare disorder that can be classified into idiopathic and heritable forms, associated with an underlying condition such as connective tissue disease or congenital heart disease or related to specific drugs and toxins. ^{1,2} In addition, pulmonary veno-occlusive disease (PVOD) and pulmonary capillary hemangiomatosis (PCH) are even rarer forms of pulmonary hypertension that are grouped together with PAH under the current classification system.²

Clinical features described in patients with PVOD/PCH include a low transfer coefficient for carbon monoxide (Kco) and oxygen desaturation on exertion, as well as the presence of centrilobular ground glass opacification, interlobular septal thickening, and mediastinal lymph-

adenopathy on high-resolution computed tomography (CT) of the lung parenchyma.^{3,4} However, these clinical and radiological features have also been reported in idiopathic PAH.^{5–7} Consequently, the clinical distinction between PVOD/PCH and idiopathic PAH can be challenging. It has been estimated that 10% of patients with PVOD/PCH are misdiagnosed as having idiopathic PAH.^{8,9} The diagnosis of PVOD/PCH is often confirmed only postmortem or from explanted lungs by histology.

The histological features of PVOD/PCH typically include pulmonary venous obstructions and pulmonary capillary proliferation, although the distribution of these changes within the lung can be heterogeneous. 10,11 Pulmonary artery smooth muscle hypertrophy and intimal hyperplasia, similar to the changes observed in other forms of PAH, may also be present. Furthermore, pulmonary venous changes have been reported in cases of idiopathic PAH, patients with scleroderma-associated PAH, and those with *BMPR2* mutations to varying extents. 12,13

A major advance in the molecular diagnosis of PVOD/PCH was the finding of biallelic mutations in the gene encoding the eukaryotic translation initiation factor 2 alpha kinase 4 (*EIF2AK4*) in both familial (100%) and sporadic (20% to 25%) cases of PVOD/PCH.14,15 El-F2AK4 is an activator of the integrated stress response pathway and responds to environmental stresses, including amino acid deprivation, by phosphorylating the α subunit of eukaryotic translation initiation factor 2.11,16,17 These discoveries suggest that EIF2AK4 mutations are specific to PVOD/PCH and that finding biallelic EIF2AK4 mutations in a patient with pulmonary hypertension would be diagnostic of PVOD/PCH. Patients with PVOD/PCH have a poor prognosis and risk fatal pulmonary edema with the use of pulmonary artery vasodilator therapies. 4,18-20 Consequently, early and accurate diagnosis is vital to guide clinical management.

Heterozygous mutations in the gene encoding the bone morphogenetic protein type 2 receptor (*BMPR2*) are the most common genetic cause of PAH. They are found in ≈17% of individuals with idiopathic PAH and 82% with a family history of the disease.²¹ However, mutations in *BMPR2* have also been reported in patients with histologically proven PVOD.^{4,22–24} Thus, considerable uncertainty remains as to what extent the finding of *EIF2AK4* or *BMPR2* mutations reliably predicts the clinical phenotype and response to therapy in a population of patients with PAH.

Here, we report the genetic and phenotypic characteristics of patients assessed for *BMPR2* and *EIF2AK4* mutations through whole-genome sequencing within a large cohort (n=880) of patients with PAH recruited to the National Institute of Health Research (NIHR) BRIDGE study (BioResource–Rare Diseases) (Table I in the online-only Data Supplement). The frequency of mutations in other previously reported genes associated with PAH will be reported in a future publication. In this study, we identified and characterized patients with a clinical

and radiological diagnosis of idiopathic PAH who were found to possess biallelic EIF2AK4 mutations. These patients had a low Kco and were diagnosed at a younger age compared with patients with idiopathic PAH without mutations in these genes. We show that, in common with patients diagnosed clinically with PVOD/PCH, patients with PAH with biallelic EIF2AK4 mutations have a shorter survival. We conclude that clinical assessment alone is inadequate for the accurate diagnosis of PVOD/PCH. Clinical genetic testing in younger patients presenting clinically with PAH but with a low Kco will allow appropriate classification, leading to better risk stratification and management of these patients.

METHODS

Ethics Approval and Consent

UK patients (621 [70.6%]) were recruited prospectively to the BRIDGE study and provided written informed consent for genetic analysis and the capture of clinical data (BRIDGE study 13/EE/0325). In addition, the study included patients recruited retrospectively from non-UK centers (191 [21.7%]) and deceased UK patients (68 [7.7%]) if they had signed local tissue bank consent forms allowing genetic sequencing.

Explanted lung tissue from an individual undergoing lung transplantation for end-stage PAH was collected under Papworth Hospital Research Tissue Bank ethics (08/H0304/56).

Recruitment and Patients

The BRIDGE study is a prospective study recruiting both prevalent and incident patients with selected rare diseases. Recruitment to the BRIDGE PAH study started in January 2013, and the last patient included in this analysis was recruited on June 15, 2016. Patients with idiopathic PAH, heritable PAH, PVOD, and PCH, diagnosed according to international guidelines at specialist pulmonary hypertension centers in the United Kingdom, the Netherlands, and France, were recruited (Figure 1 and Table II in the online-only Data Supplement).² This included 14 patients with confirmed mutations in BMPR2.

Throughout this article, we classify patients recruited to the study as having idiopathic PAH or familial PAH on the

basis of the absence or presence of a family history of the disease. The term heritable PAH does not distinguish between patients with sporadic PAH with a mutation and patients with a mutation who have a family history. Therefore, the term heritable PAH is used only when referring to previous publications and guidelines.

Patients with other rare diseases and their unaffected relatives recruited to the BRIDGE study (Table III in the online-only Data Supplement) acted as control subjects without PAH for the genetic analysis.

Whole-Genome Sequencing and Variant Calling

Next-generation sequencing with 100– to 150–base pair (bp) paired-end sequencing was performed on DNA libraries created from genomic DNA with Illumina HiSeq 2500 and HiSeq X (Illumina Inc, San Diego, CA).

Reads were aligned against the Genome Reference Consortium human genome (build 37), and variants were called with the Isaac Aligner and Variant Caller (version 2, Illumina Inc). Variants in BMPR2 and EIF2AK4 were extracted and annotated with the Ensembl Variant Effect Predictor version 84.25 Deletions (resulting in the loss of >50 bp) were identified by applying Isaac Copy Number Variant Caller (Canvas, Illumina) and Isaac Structural Variant Caller (Manta, Illumina). Further information is provided in the online-only Data Supplement.

Likely causal variants were identified on the basis of minor allele frequency and predicted deleteriousness. Variants were considered further if they had a minor allele frequency of <1 in 10000 in unrelated BRIDGE control subjects without PAH and the ExAC database.²⁶ The rare variants that passed the minor allele frequency filtering were then assessed for deleteriousness. Variants were considered pathogenic on the basis of a combined annotation-dependent depletion score of ≥15 and PolyPhen-2 or sorting intolerant from tolerant predictions not classified as benign or tolerated, respectively.^{27–29}

Overrepresentation Analyses

For comparison of variant frequencies between disease and control groups, only variants from unrelated individuals were used. The PRIMUS software package was used to identify

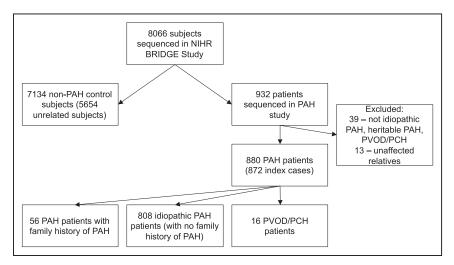


Figure 1. Subjects recruited to the National Institute of Health Research (NIHR) BRIDGE study (BioResource-Rare Diseases) and the clinical diagnostic categories of patients with pulmonary arterial hypertension (PAH) included in this study.

PVOD/PCH indicates pulmonary venoocclusive disease/pulmonary capillary hemangiomatosis.

nonrelated individuals among both BRIDGE control subjects without PAH and patients with PAH.³⁰ The number of unrelated control subjects was maximized by including either patients with other rare diseases or their unaffected relatives. The frequency of rare and predicted deleterious heterozygous *EIF2AK4* variants in PAH index cases was also compared with publically available information in the ExAC database (http://exac.broadinstitute.org).²⁶ This analysis provides the maximum estimate of the frequency of heterozygous *EIF2AK4* variants in the ExAC database because variants in ExAC were assumed not to be in a compound heterozygous state.

Phenotypic Data Capture and CT Assessment

Paper and electronic patient records of patients with PAH were reviewed to capture demographic and phenotypic variables from the time of diagnosis and follow-up. Survival data for UK patients were obtained from recruiting centers through the NHS National Spine and local databases. Anonymized information was captured securely online with the free OpenClinica software, adapted for data capture specific to PAH.

CT images of the chest, when available, were reviewed independently by 2 cardiothoracic radiologists (A.S. and N.S.) with specialist imaging experience in pulmonary hypertension who were blinded to the underlying diagnoses with a customized proforma. Further information is provided in the supplemental materials and Tables IV and V in the online-only Data Supplement.

Statistical Analysis

Statistical analysis was performed in R (www.r-project.org). Further information is provided in the online-only Data Supplement.

Semiparametric Cox proportional hazard models were used to assess survival between groups with the survival package in R. Time from diagnosis to both death and death or transplantation was assessed. Age at diagnosis and sex were used as covariates in the models. To avoid immortal time bias arising from the inclusion of retrospectively recruited patients and prevalent patients, a sensitivity analysis was conducted. In this analysis, only prospectively recruited patients from the UK were included, and patients entered the risk set only from the time they consented to the study. Further information is provided in the online-only Data Supplement.

RESULTS Study Patients

Whole-genome sequencing was performed on 932 patients recruited to the NIHR BRIDGE PAH study and 7134 control subjects without PAH recruited to other NIHR BRIDGE study cohorts. Fifty-two patients were excluded from further analysis because they did not have a clinical diagnosis of idiopathic PAH, heritable PAH, PVOD, or PCH (Figure 1). The remaining 880 patients (of whom 872 were defined as unrelated index cases)

consisted of 16 patients (1.8%) with a clinical diagnosis of PVOD/PCH, 56 (6.4%) with PAH and a family history of the disease (referred to as familial PAH), and 808 (91.8%) with idiopathic PAH and no known family history. One of the 16 patients with a clinical diagnosis of PVOD/PCH had an affected sister, whereas the remainder had the sporadic form of the disease.

BMPR2 Mutations in the PAH Cohort

Rare and predicted deleterious *BMPR2* mutations (single-nucleotide variants, indels, and larger deletions) were found in 41 patients (73.2%) with familial PAH and 89 patients (11.0%) with idiopathic PAH. No *BMPR2* mutations were found in patients with a clinical diagnosis of PVOD/PCH.

Rare and Predicted Deleterious *EIF2AK4*Variants in the PAH Cohort

Sixty-nine rare and predicted deleterious *EIF2AK4* single-nucleotide variants and indels were present in the NIHR BRIDGE study. No large deletions were found that affected the *EIF2AK4* gene locus. The variants are summarized in Table VI in the online-only Data Supplement. Five of the 16 patients (31.3%) with clinically diagnosed PVOD/PCH carried biallelic *EIF2AK4* mutations (2 homozygotes and 3 compound heterozygotes).

Twenty-five *EIF2AK4* variants were also found in 19 patients (2.2%) diagnosed clinically with PAH, in whom there was no clinical suspicion of PVOD/PCH (5 homozygotes, 4 compound heterozygotes, and 10 heterozygotes; Table VII in the online-only Data Supplement). One of these patients with a homozygous *EIF2AK4* mutation (c.3097C>T creating a premature stop codon) had a sister who had died of PAH. There was no reported family history of PVOD/PCH.

The remaining rare *EIF2AK4* variants were found in a heterozygous state in 36 control subjects (0.5%). Four of these variants appeared in >1 control subject without PAH, and none were shared with patients with PAH.

Overrepresentation of Rare Heterozygous *EIF2AK4* Variants in Patients With Idiopathic PAH Compared With Control Subjects

The proportion of patients with a clinical diagnosis of idiopathic PAH carrying heterozygous rare *EIF2AK4* variants (1.2%) was significantly greater than the percentage of control subjects without PAH (0.5%; P=0.030). A similar overrepresentation in patients with idiopathic PAH was observed compared with allele frequencies in the ExAC database (0.6%; P=0.042). Two patients with idiopathic PAH with heterozygous rare

EIF2AK4 variants also carried a rare and predicted deleterious BMPR2 mutation.

Phenotype of Patients With a Clinical Diagnosis of PAH and Biallelic *EIF2AK4* Mutations

Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations presented at a younger age (median, 29 years; interquartile range, 23–38 years) compared with patients without mutations in the PAH associated genes (51 years; IQR, 37–65 years; *P*=0.024; Table 1). Mean pulmonary artery pressure, cardiac output, and pulmonary vascular resistance were not significantly different between patients with PAH with biallelic *EIF2AK4* mutations and the other groups. As previously reported, hemodynamic variables were significantly worse in patients with *BMPR2* mutations compared with those without any mutations in these genes.

The patients with PAH with biallelic *EIF2AK4* mutations exhibited a reduced Kco (33% [IQR, 30%–35%] predicted) compared with *BMPR2* mutation carriers (81% [IQR, 73%–92%] predicted; *P*<0.001) and patients with PAH with no identified mutation (71% [IQR, 51%–85%] predicted; *P*=0.001). Patients with PAH with biallelic *EIF2AK4* mutations had no obstructive or restrictive deficit on spirometry. These differences remained after the exclusion of patients with abnormal

spirometry in the other groups (forced expiratory volume in 1 second of expiration $[FEV_1]$ <80% or forced vital capacity [FVC] <80%; Table VIII in the online-only Data Supplement).

Digital clubbing was overrepresented among patients with biallelic *EIF2AK4* mutations diagnosed clinically with PAH (42%; *P*=0.002). Eleven percent of patients with a clinical diagnosis of PVOD were clubbed.

Only 1 patient with a heterozygous rare and predicted deleterious *EIF2AK4* variant (c.2516T>C) had a reduced Kco (54% predicted) with normal spirometry (FEV₁, 102% predicted; FVC, 98% predicted; and total lung capacity, 100% predicted). There was mild paraseptal emphysema on thoracic CT (<5% of the lung parenchyma affected). This patient, a 44-year-old white man diagnosed with idiopathic PAH, also carried a rare and deleterious *BMPR2* splice acceptor mutation (c.853-2A>G).

We questioned whether Kco was a predictor of biallelic *EIF2AK4* mutations in the wider cohort. However, among patients with PAH with no mutations and normal spirometry (n=255), a reduced Kco (<50% predicted) was present in 65 patients (25.5%). In these patients with a reduced Kco and preserved spirometry, 90.8% were >50 years old at diagnosis, and 69.2% had a history of coronary artery disease, left ventricular dysfunction, or cardiovascular risk factors (diabetes mellitus, systemic hypertension, or hyperlipidemia).

Table 1. Phenotypic Summary of *EIF2AK4* Variant Carriers: Patients With a Clinical Diagnosis of PAH and Biallelic *EIF2AK4* Mutations Were Younger at Diagnosis and Had a Significantly Reduced Kco Compared With Other Groups

	PAH Patients With BMPR2 Mutations*	PAH Patients With No Mutations in PAH- Associated Genes	PAH Patients With EIF2AK4 Heterozygous Variants	PAH Patients With Biallelic <i>EIF2AK4</i> Mutations	Patients With PVOD/PCH	P Value
n	130	704	8	9	16	
Age, y	39 (31–52)	51 (37–65)	49 (36–67)	29 (23–38)	57 (41–69)	<0.001
Female, n (%)	85 (65.4)	494 (70.2)	7 (87.5)	4 (44.4)	9 (56.2)	0.180
White, n (%)	108 (83.1)	551 (78.5)	5 (62.5)	2 (22.2)	13 (81.2)	0.002
Digital clubbing, n (%)	6 (9.7)	10 (3.4)	0 (0)	3 (42.9)	1 (11.1)	0.002
BMI, kg/m²	28 (24–33)	28 (24–33)	26 (23–28)	24 (20–27)	27 (24–31)	0.216
mPAP, mmHg	57 (51–69)	52 (44–61)	44 (42–52)	52 (46–65)	48 (40–58)	<0.001
CO, L/min	3 (3–4)	4 (3–5)	3 (3–5)	5 (3–6)	4 (3–4)	<0.001
PVR, WU	15 (11–20)	10 (7–14)	9 (6–10)	9 (8–13)	10 (9–12)	<0.001
Vasoresponders, n (%)	0 (0)	28 (17.5)	0 (0)	0 (0)	0 (0)	0.011
FEV ₁ , % predicted	90 (78–99)	84 (72–95)	83 (71–94)	94 (85–100)	85 (70–95)	0.031
FVC, % predicted	97 (86–109)	95 (82–106)	96 (75–98)	100 (86–119)	97 (81–103)	0.310
KCO, % predicted	81 (73–92)	71 (51–85)	81 (72–95)	33 (30–35)	37 (32–47)	<0.001
Resting S _A O ₂ , %	96 (94–97)	96 (93–97)	98 (98–98)	91 (90–94)	94 (91–95)	0.010
S _A O ₂ after walk test, %	94 (90–97)	92 (85–96)	94 (84–96)	78 (75–82)	88 (85–89)	<0.001

BMI indicates body mass index; CO, cardiac output; FEV₁, forced expiratory volume in 1 second; FVC, forced vital capacity; Kco, transfer coefficient for carbon monoxide; mPAP, mean pulmonary artery pressure; PAH, pulmonary arterial hypertension; PCH, pulmonary capillary hemangiomatosis; PVOD, pulmonary veno-occlusive disease; PVR, pulmonary vascular resistance; and S_aO₃, arterial oxygen saturation.

^{*}Also includes the 2 patients with a heterozygous EIF2AK4 variant and a BMPR2 variant. Data presented as median (interquartile range) unless indicated. Percentages were calculated from the number of patients for whom data were available as the denominator.

Given the high prevalence of a low Kco with preserved spirometry in the wider cohort, we restricted an analysis to patients <50 years of age who at the time of diagnosis had normal spirometry (n=164). Even in this group, a significant proportion (n=15, 9.1%) had a Kco <50% predicted (Figure 2). Eight of these 15 patients carried biallelic *EIF2AK4* mutations. One patient with biallelic *EIF2AK4* mutations was 70 years of age at diagnosis and subsequently did not meet this cutoff.

Among patients with normal spirometry, the presence of a Kco <50% predicted and age at diagnosis <50 years had a high sensitivity (0.889) and specificity (0.977) for identifying patients who carry biallelic *El-F2AK4* mutations; the positive predictive value was low (0.533). Nevertheless, in terms of the diagnostic yield, although genetic testing for biallelic *ElF2AK4* mutations in the entire cohort of patients diagnosed clinically with PAH yielded a 1% detection rate, the presence of biallelic *ElF2AK4* mutations in patients with PAH with a Kco <50% predicted with normal spirometry and <50 years of age at diagnosis was 53%.

CT Features of EIF2AK4 Mutation Carriers

Centrilobular ground glass opacification extent, mediastinal lymphadenopathy, and interlobular septal thickening are considered suggestive of PVOD/PCH. However, we found subtle or gross centrilobular ground glass opacification in 38% of patients diagnosed clinically with PAH and carrying no mutations (n=21) and 67% of patients with PAH with *BMPR2* mutations (n=21). This was not significantly different compared with patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations (86%, n=7) and patients with a clinical diagnosis of PAH and biallelic *EIF2AK4*

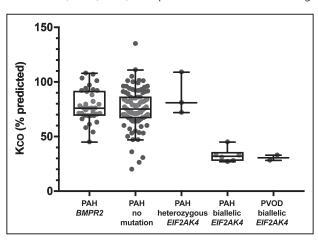


Figure 2. The transfer coefficient for carbon monoxide (Kco) is influenced by genotype in pulmonary arterial hypertension (PAH).

Patients with forced expiratory volume in 1 second of expiration <80% predicted and forced vital capacity <80% predicted and diagnosed with PAH or pulmonary veno-occlusive disease (PVOD)/pulmonary capillary hemangiomatosis after 50 years of age were excluded from the plot.

nosis of PVOD (50%, n=14). Gross interlobular septal thickening and mediastinal lymphadenopathy were significantly more frequent among patients with PAH and biallelic *EIF2AK4* mutations (29% and 57%, respectively) and those with PVOD (64% and 79%) compared with patients with PAH and no mutation (5% and 0%) or *BMPR2* mutations (5% and 10%). A radiological suspicion of PVOD/PCH was raised in 71% of those with PVOD, 57% of patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations, 14% of patients with PAH with no mutation, and 5% of those with *BMPR2* mutations (Table 2).

A further CT analysis comparing patients with biallelic EIF2AK4 mutations (with a clinical diagnosis of PVOD/PCH or PAH; n=11) and those with a clinical diagnosis of PVOD but not carrying biallelic EIF2AK4 mutations (n=10) was made (Table IX in the online-only Data Supplement). Patients with biallelic *EIF2AK4* mutations were younger at diagnosis (27 years; IQR, 23–34 years) compared with those with PVOD and no EIF2AK4 mutations (68 years; IQR, 64-72 years; P=0.001). The patients with biallelic EIF2AK4 mutations also had a lower Kco (32% [IQR, 29%–33%] predicted) compared with patients with PVOD and no EIF2AK4 mutations (41.4% [IQR, 37%–54%] predicted; *P*=0.013). Centrilobular ground glass opacification appeared more extensive in those with biallelic EIF2AK4 mutations (82%) compared with those without a mutation (10%; P=0.012). However, pleural effusions were more common among those without a mutation (40%) compared with patients with biallelic EIF2AK4 mutations (0%; P=0.035). This may suggest that patients with biallelic EIF2AK4 mutations have a distinct radiological phenotype compared with patients with PVOD and no biallelic EIF2AK4 mutations.

Response to Pulmonary Artery Vasodilator Therapies

The response to pulmonary artery vasodilator therapies at 1 and 3 years was assessed for patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations and the other patients with PAH included in the CT analysis. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations did not improve their functional class at either 1 or 3 years after diagnosis, unlike the other PAH groups (Table X in the online-only Data Supplement).

Histological Features of a Biallelic EIF2AK4 Mutation Carrier

The explanted lungs of 1 patient diagnosed with idiopathic PAH but found to have a homozygous *EIF2AK4* missense mutation (c.1795G>C, p.G599R) were as-

Table 2. Radiological Features and Consensus Radiological Diagnosis of Patients With PAH in the CT Substudy

	Group	Patients With PAH With <i>BMPR2</i> Mutations (n=21), n (%)	Patients With PAH With No Mutations in the Previously Reported PAH Genes (n=21), n (%)	Patients With PAH With Heterozygous EIF2AK4 Variants (n=4), n (%)	Patients With PAH With Biallelic EIF2AK4 Mutations (n=7), n (%)	Patients With PVOD (n=14), n (%)	P Value
Centrilobular	None	7 (33.3)	13 (61.9)	2 (50.0)	1 (14.3)	7 (50.0)	
ground glass opacification	Subtle	12 (57.1)	5 (23.8)	0 (0.0)	2 (28.6)	3 (21.4)	0.122
density	Present	2 (9.5)	3 (14.3)	2 (50.0)	4 (57.1)	4 (28.6)	
Centrilobular	None	8 (38.1)	13 (61.9)	2 (50.0)	1 (14.3)	8 (57.1)	
ground glass opacification	<5%	0 (0.0)	3 (14.3)	0 (0.0)	1 (14.3)	1 (7.1)	
extent	5%-25%	2 (9.5)	0 (0.0)	1 (25.0)	2 (28.6)	1 (7.1)	0.077
	25%-50%	2 (9.5)	4 (19.0)	0 (0.0)	0 (0.0)	2 (14.3)	0.077
	50%-75%	5 (23.8)	1 (4.8)	0 (0.0)	2 (28.6)	0 (0.0)	
	75%-100%	4 (19.0)	0 (0.0)	1 (25.0)	1 (14.3)	2 (14.3)	
Interlobular septal thickening	None	17 (81.0)	18 (85.7)	4 (100.0)	5 (71.4)	4 (28.6)	
	Subtle	3 (14.3)	2 (9.5)	0 (0.0)	0 (0.0)	1 (7.1)	0.001
	Present	1 (4.8)	1 (4.8)	0 (0.0)	2 (28.6)	9 (64.3)	
Mediastinal	None	19 (90.5)	21 (100.0)	4 (100.0)	3 (42.9)	3 (21.4)	<0.001
lymphadenopathy	Present	2 (9.5)	0 (0.0)	0 (0.0)	4 (57.1)	11 (78.6)	<0.001
Pleural effusion	None	17 (81.0)	21 (100.0)	3 (75.0)	7 (100.0)	10 (71.4)	0.048
	Small	4 (19.0)	0 (0.0)	1 (25.0)	0 (0.0)	4 (28.6)	0.048
Neovascularity	None	12 (57.1)	18 (85.7)	4 (100.0)	6 (85.7)	13 (92.9)	0.077
	Present	9 (42.9)	3 (14.3)	0 (0.0)	1 (14.3)	1 (7.1)	0.077
CT diagnosis	PAH	20 (95.2)	18 (85.7)	3 (75.0)	3 (42.9)	4 (28.6)	
	Possible PVOD/PCH	1 (4.8)	3 (14.3)	1 (25.0)	4 (57.1)	10 (71.4)	

CT indicates computed tomography; PAH, pulmonary arterial hypertension; and PVOD, pulmonary veno-occlusive disease.

sessed. The predominant histological feature was pulmonary arterial vasculopathy. The pulmonary arteries predominantly showed concentric and eccentric intimal fibrosis. No plexiform lesions were observed. Although infrequent, there was some fibrosis of the septal veins and venules, some of which were nearly completely occluded. Although there was evidence of capillary congestion, no capillary hemangiomatosis was observed (Figure 3). The missense variant carried by this patient was not reported in the ExAC database, occurs in a conserved area of the genome (Genomic Evolutionary Rate Profiling score, 5.5), and was predicted to be deleterious (combined annotation-dependent depletion score, 32; PolyPhen-2 prediction of "probably damaging [1]," sorting intolerant from tolerant prediction of "deleterious [0]"). The same homozygous mutation was also found in a second unrelated patient with a clinical diagnosis of idiopathic PAH.

Impact of Genotype on Survival

Eight hundred fifty-eight patients were included in the Cox proportional hazards model (Table XI and Figure I in the online-only Data Supplement). Patients diagnosed clinically as having PAH with biallelic EIF2AK4 mutations had a shorter survival time from diagnosis compared with the BMPR2 mutation carriers (P<0.001) and those without any variants in PAH-associated genes (P<0.001). Age (P<0.001) and sex (P=0.001) also had a significant effect on survival, with male sex and older age at diagnosis associated with shorter survival in the model. Similar results were obtained in the assessment of time to death or transplantation (Table XII in the online-only Data Supplement). In the sensitivity analysis, including only prospectively recruited UK patients, only 2 events occurred in the biallelic EIF2AK4 group. Thus, no significant difference was observed in mortality between patients diagnosed clinically as having PAH with biallelic EIF2AK4 mutations and patients with BMPR2 mutations (P=0.215) or patients without any variants in PAH-associated genes (P=0.282; Table XIII in the onlineonly Data Supplement).

DISCUSSION

This is the first study to analyze the frequency of El-F2AK4 rare variation in a large cohort of patients with PAH and to make detailed phenotypic and radiological assessments. Previously, the presence of biallelic El-F2AK4 mutations was reported in patients with a clear

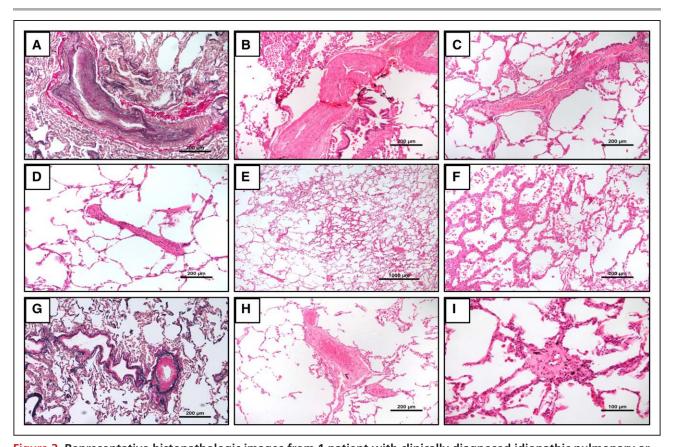


Figure 3. Representative histopathologic images from 1 patient with clinically diagnosed idiopathic pulmonary arterial hypertension (PAH) but found to have a rare (not reported in the ExAC database) and predicted deleterious (combined annotation-dependent depletion score, 32) homozygous *EIF2AK4* missense variant (c.1795G>C). The patient was of Pakistani origin and did not have a family history of PAH or pulmonary veno-occlusive disease (PVOD). At presentation, he was 22 years old and had a reduced transfer coefficient for carbon monoxide (Kco; 31% predicted) despite preserved spirometry. High-resolution computed tomography of his chest showed subtle but extensive (50%–75% involvement) ground glass opacification. No interlobular septal thickening or mediastinal lymphadenopathy was observed. No suspicion of PVOD/pulmonary capillary hemangiomatosis (PCH) was raised from the radiological appearances. Histopathology was reviewed by 2 independent pathologists, each confirming the predominant histological pattern to be one of pulmonary arterial vasculopathy. The pulmonary arteries showed eccentric and concentric intimal fibrosis and medial hypertrophy (A and B), as well as some lesions with features of recanalized thrombus (C). Several concentrically muscularized arterioles were also observed (D). No complex plexiform lesions were present. There was patchy thickening of the alveolar septa with capillary

congestion and pigmented intra-alveolar macrophages similar to PCH (E and F). Venous remodeling was difficult to trace and

infrequent but present. Fibrous thickening of the intima in septal veins (G and I) and a microvessel (H).

clinical diagnosis of PVOD/PCH and a large kindred and a single family with a possible diagnosis of PAH.^{20,31,32} As expected, we identified a high frequency of biallelic *EIF2AK4* mutations in patients with a clear clinical presentation of PVOD/PCH. However, we also found biallelic *EIF2AK4* mutations in patients with a clinical diagnosis of PAH.

The discovery of biallelic *EIF2AK4* mutations in PVOD/PCH raised the possibility of rapid molecular diagnosis in the majority of patients with familial and up to 25% of patients with sporadic PVOD/PCH.^{14,15} In the present study, the presence of biallelic *EIF2AK4* mutations was associated with a poor prognosis, even in patients who have a clinical diagnosis of PAH and who did not develop pulmonary edema in response to pulmonary

artery vasodilator therapies. Therefore, early identification of these patients through genetic testing may prompt early referral for lung transplantation similar to patients with clinically diagnosed PVOD/PCH.¹⁸

The presence of biallelic *EIF2AK4* mutations in patients with a clinical diagnosis of PAH raises the question whether *EIF2AK4* mutations can cause classic idiopathic PAH or whether there are cases of PVOD/PCH caused by *EIF2AK4* mutations that are wrongly classified even by expert centers. We further show that phenotypic, radiological, and histological assessments can be difficult to interpret. The presence of subtle or infrequent features may lead to an incorrect diagnosis of PAH in patients with biallelic *EIF2AK4* mutations. This study suggests that patients with pathogenic biallelic

EIF2AK4 mutations may present with a spectrum of phenotypic, radiological, and histological features that can overlap with PAH.

Patients with PAH with biallelic EIF2AK4 mutations demonstrated a reduced Kco despite normal spirometry, which is characteristic of patients with PVOD/PCH. The reduced Kco likely reflects widespread reduction in alveolar gas exchange caused by endothelial proliferation and patchy thickening of the blood-gas barrier by the process of capillary hemangiomatosis. Ultrastructural thickening of the capillary basement membrane may also play a role.33 In keeping with previous reports in PVOD/PCH, we also show that patients with PAH with biallelic mutations in EIF2AK4 are younger at diagnosis than patients with either BMPR2 mutations or no known mutation. 14,20 However, the presence of these characteristic features has a low positive predictive value for the identification of patients with biallelic EIF2AK4 mutations.

In contrast to previous descriptions of patients with PVOD, none of the patients with clinically diagnosed PAH and biallelic *EIF2AK4* mutations developed pulmonary edema in response to pulmonary artery vasodilator therapies. For example, intravenous prostanoids were used in 50% of these patients. In patients with classic PVOD, pulmonary edema with intravenous prostanoids has been reported in up to 44% of patients after a median treatment duration of just 9 days.⁴ Presumably, the extent and severity of the pulmonary venous involvement in these patients might underlie the differing responses to prostanoids.

It is generally considered that high-resolution CT imaging is a useful noninvasive test to assist in the diagnosis of suspected PVOD/PCH.¹¹ Although there was an increased prevalence of mediastinal lymphadenopathy and interlobular septal thickening in patients with PAH with biallelic EIF2AK4 mutations, we found that radiological features at the time of diagnosis could not accurately determine the underlying genotype.⁶ The differing radiological features of all patients with biallelic EIF2AK4 mutations compared with patients with PVOD without mutations is of interest. This may reflect differences between the younger-onset genetic cases of PVOD compared with the predominantly older group of patients without EIF2AK4 mutations in whom other nongenetic factors such as exposure to inorganic solvents may play an important role.34

Histological examination (usually postmortem or from explanted lungs) is often considered essential for diagnostic confirmation of PVOD/PCH but may be confounded by the heterogeneous nature of vascular pathology.³⁵ Surgical biopsy of the lung in patients with severe PAH is contraindicated, and a limitation of this study is that lung tissue from only 1 patient with biallelic *EIF2AK4* mutations was available for analysis. This patient had a rare and predicted deleterious homozy-

gous missense mutation in *EIF2AK4*. The predominant feature on assessment of the explanted lung tissue was pulmonary arteriopathy, as usually seen in PAH. Although only infrequent, fibrosis of the septal venules and the possible presence of siderophages in the alveolar space were observed. These features are found in patients with PVOD/PCH. This case supports the hypothesis that patients with biallelic *EIF2AK4* mutations may present with a spectrum of venous and arterial involvement.

There are increasing reports of phenotypic, radiological, and histological similarities between PAH and PVOD/PCH.^{6,12,13} Tenorio et al³¹ reported a homozygous missense mutation in EIF2AK4 in a large kindred of Iberian Romani with apparent heritable PAH. This kindred is likely to have PVOD/PCH because these diagnoses were not confirmed histologically and PVOD was suspected in half the patients. More recently, Best et al³² also report 2 sisters with apparent heritable PAH-carrying biallelic *EIF2AK4* mutations. These patients also had a reduced Kco but had not had high-resolution CT assessment of their lung parenchyma, which may have altered their clinical diagnosis. Taken together, these previous reports are compatible with the findings in this larger cohort that patients with a clinical presentation of idiopathic or heritable PAH may in fact have underlying PVOD/PCH as determined by genetic analysis.

A strength of this study is the centralized reporting of radiographic features. However, the data collection was retrospective and incomplete in some cases. Assessing rare diseases such as PAH and PVOD/ PCH with a prospective study recruiting incident cases would take a prohibitively long time. This is especially true for the assessment of survival and response to therapy. In this study including prevalent and retrospectively recruited patients, we demonstrated a worse prognosis in patients with a clinical diagnosis of PAH and biallelic *EIF2AK4*. However, the inclusion of prevalent and retrospectively recruited patients can introduce bias such as immortal time bias, when there are long periods between diagnosis and enrollment in the study. The effect of immortal time bias and other confounders such as the inclusion of prevalent and incident cases can be difficult to predict. All groups are likely to include patients who died before study enrollment and thus would not feature in any analysis. When we attempted to eliminate these sources of bias in a sensitivity analysis restricted to prospectively recruited patients from the United Kingdom, the study did not have sufficient power to show a difference in survival between different genotypes. Further studies of survival and response to therapy are needed to definitively show whether misclassified patients with PAH with biallelic *EIF2AK4* mutations and patients with classic PVOD with these mutations have a similarly poor prognosis.

The genetic architecture of idiopathic and heritable PAH remains to be fully elucidated. Ongoing analysis of whole-genome sequence data in our cohort is likely to reveal novel rare variation underlying this condition. Mutations in BMPR2 account for ≈17% of cases of idiopathic PAH, and other known PAH genes account for ≈1% to 2% of all cases. 21,36 In the present study, BMPR2 mutations were found in 11% of patients without a family history of PAH. It is worth noting that patients with the sporadic form of the disease with no reported family history represent a higher burden of BMPR2 mutations (n=89) compared with those with a family history (n=49). This has important implications for clinical genetic testing in patients with sporadic and familial disease.

In previous studies, mutations in both *EIF2AK4* alleles are required to cause PVOD and PCH.14,15 In autosomal recessive disorders, it is unusual for the heterozygous state to manifest the disease phenotype, and heterozygous EIF2AK4 variants thus would not be expected to be pathogenic. In this study, we found a significant overrepresentation of heterozygous rare and predicted deleterious EIF2AK4 variants in patients with PAH compared with control subjects and report 2 patients with rare variants in both BMPR2 and EIF2AK4. Recently, the possibility that heterozygous *EIF2AK4* variants influence the penetrance of BMPR2 mutations has been raised in a single family with PAH.³⁷ Further studies are required to determine whether heterozygous *EIF2AK4* variants contribute to pathogenesis in PAH.

CONCLUSIONS

We demonstrate that biallelic EIF2AK4 mutations are found in patients diagnosed clinically with idiopathic and familial PAH. These patients may have subtle features suggestive of PVOD/PCH on close inspection and are likely to have underlying PVOD/PCH. The spectrum of phenotypic, radiological, and histological features found in patients with biallelic EIF2AK4 mutations made by current clinical assessments is wider and less clear-cut than previously recognized. This may lead to misclassification of patients as having PAH rather than PVOD and hinders accurate risk stratification. Ascertaining the EIF2AK4 mutation status of patients through clinical genetic testing provides additional information to aid risk stratification and to guide management. In a young patient presenting with apparent PAH, the presence of a low Kco with normal spirometry strongly suggests the presence of underlying biallelic EIF2AK4 mutations. Patients with an apparent clinical diagnosis of PAH and biallelic EIF2AK4 mutations have a worse prognosis compared with patients with BMPR2 mutations and those without these mutations. Clinical genetic testing should aid identification of this high-risk

group and facilitate early referral for lung transplantation and appropriate management.

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FOOTNOTES

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Phenotypic Characterization of *EIF2AK4* Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically With Pulmonary Arterial Hypertension

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- m	University of Cambridge/Cambridge University	
Geoff Woods	Hospitals NHS Foundation Trust	UK
NAC CONTRACTOR OF	University of Cambridge/Cambridge University	1117
Marc Tischkowitz	Hospitals NHS Foundation Trust	UK
Richard Sandford	University of Cambridge/Cambridge University	UK
	Hospitals NHS Foundation Trust	UK
РАН	University of Cambridge/Cambridge University	
Nicholas Morrell	University of Cambridge/Cambridge University Hospitals	UK
	•	
Stefan Gräf	University of Cambridge	UK

	Department of Medicine, University of	
Marta Bleda	Cambridge	UK
	Department of Medicine, University of	
Charaka Hadinnapola	Cambridge	UK
	Department of Medicine, University of	
Matthias Haimel	Cambridge	UK
	Cambridge University Hospitals NHS	
Simon Holden	Foundation Trust	UK
	Department of Medicine, University of	
Jennifer Martin	Cambridge	UK
Sonia Ali	Imperial and Hammersmith	UK
Harm Boggard	VU University Medical Center, Amsterdam	Netherlands
Colin Church	Golden Jubilee National Hospital	UK
Paul Corris	Newcastle Freeman	UK
Gerry Coghlan	Royal Free	UK
Amanda Creaser-Myers	Sheffield CRF, Royal Hallamshire	UK
Victoria Cookson	GOSH	UK
Rosa DaCosta	Royal Brompton	UK
Natalie Dormand	Royal Brompton	UK
Pavandeep K Ghataorhe	Imperial and Hammersmith	UK
Simon Gibbs	Imperial and Hammersmith	UK
Alan Greenhalgh	Newcastle Freeman	UK
Marc Humbert	University of South Paris	France
Anna Huis in't Veld	VU University Medical Center, Amsterdam	Netherlands
Fiona Kennedy	Golden Jubilee National Hospital	UK
David Kiely	Sheffield CRF, Royal Hallamshire	UK
Allan Lawrie	Sheffield CRF, Royal Hallamshire	UK
Rob Mackenzie Ross	Bath	UK
Rajiv Machado	University of Lincoln	UK
Larahmie Masati	Imperial and Hammersmith	UK
Sharon Meehan	Imperial and Hammersmith	UK

Shahin Moledina	GOSH	UK
Shokri Othman	Imperial and Hammersmith UK	
Andrew Peacock	Golden Jubilee National Hospital UK	
Joanna Pepke-Zaba	Papworth Hospital UK	
Val Pollock	Golden Jubilee National Hospital	UK
Gary Polwarth	Papworth Hospital	UK
Christopher J Rhodes	Imperial and Hammersmith	UK
Kevin Rue-Albrecht	Imperial and Hammersmith	UK
Gwen Schotte	VU University Medical Center, Amsterdam	Netherlands
Debbie Shipley	Newcastle Freeman	UK
Laura Southgate	Kings College, London	UK
Respiratory Nurse Specialists	Bath	UK
Jay Suntharalingam	Bath	UK
Yvonne Tan	Royal Free UK	
Mark Toshner	Papworth Hospital	UK
	Department of Medicine, University of	
Carmen Treacy	Cambridge	UK
Richard Trembath	Kings College, London	UK
Anton Vonk Noordegraaf	VU University Medical Center, Amsterdam Nether	
Ivy Wanjiku	Imperial and Hammersmith	UK
John Wharton	Imperial and Hammersmith	UK
Martin Wilkins	Imperial and Hammersmith	UK
John Wort	Royal Brompton	UK
John Wharton	Imperial and Hammersmith	UK
PID		
Kenneth Smith	University of Cambridge	UK
Taco Kuijpers	Emma Children's Hospital, Amsterdam UCL Great Ormond Street Institute of Child	Netherlands
Adrian Thrasher	Health	UK
James Thaventhiran	University of Cambridge	UK

Matthew Brown	University of Cambridge	UK
Hana Lango Allen	University of Cambridge	UK
Ilenia Simeoni	University of Cambridge	UK
	University of Cambridge/Cambridge University	
Emily Staples	Hospitals NHS Foundation Trust	UK
Crina Samarghitean	University of Cambridge	UK
Hana Alachkar	Salford Royal NHS Foundation	UK
Richard Antrobus	University Hospitals Birmingham	UK
Gururaj Arumugakani	Leeds Teaching Hopsital	UK
	UCL Great Ormond Street Institute of Child	
Chiara Bacchelli	Health	UK
Helen Baxendale	Papworth Hospital	UK
Claire Bethune	Plymouth Hopsital	UK
	UCL Great Ormond Street Institute of Child	
Shahnaz Bibi	Health	UK
	UCL Great Ormond Street Institute of Child	
Claire Booth	Health	UK
Michael Browning	Leicester Royal Infirmary	UK
Siobhan Burns	Royal Free Hospital	UK
	Cambridge University Hospitals NHS	
Anita Chandra	Foundation Trust	UK
Nichola Cooper	Imperial College Healthcare NHS Trust	UK
Coulty Do to	Cambridge University Hospitals NHS	1.117
Sophie Davies	Foundation Trust	UK
Lisa Devlin	Royal Hospitals Belfast	UK
Rainer Doffinger	University of Cambridge	UK
Elizabeth Drewe	Nottingham University Hospitals NHS Trust	UK
David Edgar	Royal Hospitals Belfast	UK
William Egner	Sheffield Teaching Hospitals	UK
Rohit Ghurye	Barts Health NHS Trust	UK

	UCL Great Ormond Street Institute of Child	
Kimberley Gilmour	Health	UK
Sarah Goddard	University Hospitals of North Midlands	UK
Pavel Gordins	Hull & East Yorkshire Hospitals NHS Trust	UK
Sofia Grigoriadou	Barts Health NHS Trust	UK
Scott Hackett	Birmingham Heartlands	UK
	Royal Hospital for Children, NHS Greater	
Rosie Hague	Glasgow and Clyde	UK
	Epsom & St Helier University Hospitals NHS	
Grant Hayman	Trust	UK
Archana Herwadkar	Salford Royal NHS Foundation	UK
Aarnoud Huissoon	Birmingham Heartlands	UK
Stephen Jolles	University Hospital Wales	UK
Peter Kelleher	Imperial College Healthcare NHS Trust	UK
	Cambridge University Hospitals NHS	
Dinakantha Kumararatne	Foundation Trust	UK
Sara Lear	Norforlk & Norwich University Hospital	UK
Hilary Longhurst	Barts Health NHS Trust	UK
Lorena Lorenzo	Barts Health NHS Trust	UK
	UCL Great Ormond Street Institute of Child	
Jesmeen Maimaris	Health	UK
	Cambridge University Hospitals NHS	
Ania Manson	Foundation Trust	UK
Elizabeth McDermott	Nottingham University Hospitals NHS Trust	UK
	Gartnavel General Hospital, NHS Greater	
Sai Murng	Glasgow and Clyde	UK
Sergey Nejentsev	University of Cambridge	UK
Sadia Noorani	Sandwell and West Birmingham Hospitals	UK
Eric Oksenhendler	Hopital St Louis, Paris	France
Mark Ponsford	University Hospital Wales	UK

	UCL Great Ormond Street Institute of Child	
Waseem Qasim	Health	UK
Isabella Quinti	Sapienza Universita di Roma	Italy
Alex Richter	University Hospitals Birmingham	UK
Ravishankar Sargur	Sheffield Teaching Hospitals	
Sinisa Savic	Leeds Teaching Hopsital	UK
Suranjith Seneviratne	Royal Free Hospital	
Carrock Sewell	Scunthorpe General Hospital	UK
Hans Stauss	Royal Free Hospital	UK
	Gartnavel General Hospital, NHS Greater	
Moira Thomas	Glasgow and Clyde	UK
Steve Welch	Birmingham Heartlands	UK
	Cambridge University Hospitals NHS	
Lisa Willcocks	Foundation Trust	UK
Nigel Yeatman	Barts Health NHS Trust	UK
Patrick Yong	Frimley Park Hospital	UK

SUPPLEMENTAL MATERIAL:

Phenotypic characterisation of *EIF2AK4* mutation carriers in a large cohort of patients diagnosed clinically with pulmonary arterial hypertension

Hadinnapola et al.

Supplemental Methods:

Whole genome sequencing

Genomic DNA was extracted from whole blood samples prior to assessment of concentration by Qubit, and quality by gel electrophoresis. After fragmentation of DNA into 200bp fragments (Covaris E220, Covaris Inc, Woburn, USA) DNA libraries were created using Tru SeqDNA LT Prep kit (Illumina Inc, San Diego, USA). The libraries underwent next generation sequencing using 100-150 base pair paired-end sequencing using Illumina HiSeq 2500 and HiSeq X (Illumina Inc, San Diego, USA).

Variant calling

Reads were aligned against the Genome Reference Consortium human genome (build 37) (GRCh37) and variants were called using the Issac Aligner and Variant Caller respectively (version 2, Illumina Inc.). Genebuilds for *BMPR2* and *EIF2AK4* genes were based on Ensembl v75. Variants from these genes were extracted and annotated using Ensembl's Variant Effect Predictor (VEP) v84 ¹. VEP was also used to annotate data from the Exome Aggregation Consortium's (ExAC) database ².

Deletions (resulting in the loss of more than 50bp) were identified by applying Isaac copy number variant caller (Canvas, Illumina) and Isaac Structural Variant Caller (Manta, Illumina).

To be called by both Canvas and Manta deletions required a reciprocal overlap of \geq 20%. Overlapping deletions represented in the Zarrei dataset with a reciprocal overlap of \geq 50% and deletions with a non-PAH BRIDGE control frequency of more than 1 in 1,000 were excluded 3 .

Analysis of computed tomographic images of the chest

CT images of the chest, where available, were reviewed independently by 2 cardiothoracic radiologists (AS and NS), with specialist imaging experience in pulmonary hypertension, blinded to the underlying diagnoses using a customised proforma (Supplemental Table 4). In addition to CT scans of patients with EIF2AK4 mutations or with a clinical diagnosis of PVOD in the cohort, CT scans of patients from Papworth Hospital and the Royal Hallamshire Hospital with normal spirometry (FEV₁ > 80% predicted and FVC > 80% predicted) and either BMPR2 mutations (n=21) or no variants in the known PAH genes (n=21) were analysed (Supplemental Table 5). A consensus read was undertaken for individual CT features and a mutually agreed overall radiological diagnosis was recorded.

Histology

The explanted lung tissue of one patient with a clinical diagnosis of idiopathic PAH and biallelic *EIF2AK4* mutations was available for further analysis. Four micrometre (µm) tissue sections were cut from formalin-fixed paraffin wax embedded blocks from the explanted lung tissue. Representative sections from each lobe of both lungs were stained with Elastic-Van Gieson and Haemotoxylin and Eosin stains. Two expert histopathologists examined the sections independently by light microscopy.

Statistical analysis

Statistical analysis was performed in R (www.r-project.org).

Differences between groups of categorical variables were assessed using the Fisher Exact test. Where one of the variables was an ordinal the Cochran-Armitage test was applied using the chisq_test function from the "coin" package ⁴. Differences in continuous variables were assessed using the Mann–Whitney U test (2 comparator groups) and the Kruskal-Wallis test (3 or more comparator groups). Post-hoc pairwise comparisons were performed using Dunn's Test for multiple testing.

Semi-parametric Cox-proportional hazards models were used to assess survival between groups using the "survival" package in R ⁵. Survival time from diagnosis to death and diagnosis to death or transplantation was assessed. Patients were censored at the date of transplantation for the primary survival analysis. Age at diagnosis and gender were used as covariates in the models.

The proportional hazards assumptions were tested by assessing Schoenfeld residuals over log time 6 . The goodness of fit of the model was assessed by plotting the log of cumulative hazard of Cox-Snell residuals against the log of time and confirming the simple regression has 0 intercept and slope of 1 7 .

The inclusion of retrospectively recruited and prevalent patients in a survival analysis assessing time from diagnosis to death/transplantation can cause immortal time bias. The immortal time is the period between diagnosis and enrolment in the study and so patients

had to have survived till this point. Patients with worse prognosis diagnosed at a similar time may not have survived long enough to enrol in the study. To further explore this potential bias, a sensitivity analysis was performed including only on UK patients recruited prospectively to the study. In this multivariate Cox-proportional hazards model, the survival period was defined as the time period from date of diagnosis to date of death and patients only entered the risk set after enrolment into the study (consent date).

Supplemental Tables

Supplemental Table 1. NIHR BioResource – Rare Diseases Collaboration. See spreadsheet.

Centre	Principle	Clinicians and research staff
	Investigator	
Freeman Hospital, Newcastle,	Paul A Corris	Alan Greenhalgh, Debbie Shipley,
UK		Margaret Day
Golden Jubilee National	Andrew	Colin Church, Val Irvine, Fiona Kennedy
Hospital, Glasgow, UK	Peacock	
Great Ormond Street	Shahin	Victoria Cookson
Hospital, London, UK	Moledina	
Hammersmith Hospital and	Martin R	Simon Gibbs, John Wharton, Sonia Ali,
Imperial College, London, UK	Wilkins	Larahmie Masati, Sharon Meehan, Ivy
		Wanjiku, Shokri Othman
Papworth Hospital,	Joanna Pepke-	Mark Toshner, Gary Polwarth
Cambridge, UK	Zaba	
Royal Brompton Hospital,	Stephen J Wort	Rosa DaCosta, Natalie Dormand, Alice
London, UK		Parker
Royal Free Hospital, London,	Gerry Coghlan	Yvonne Tan, Dipa Ghedia
UK		
Royal Hallamshire Hospital,	David G Kiely	Robin Condliffe, Amanda Creaser-Myers,
Sheffield, UK		Stephen Roney, Sara Walker
Royal United Hospitals Bath	Jay	Robert MacKenzie Ross, Mark Grover, Ali
NHS Foundation Trust, Bath,	Suntharalingam	Grove, Jill Peel, Ann Coy
UK		
University of South Paris	Marc Humbert	David Montani, Florent Soubrier, Barbara
		Girerd, Mélanie Eyries
VU University Medical Center,	Anton Vonk	Harm Bogaard, Anna Huis in't Veld, Gwen
Amsterdam, Netherlands	Noordegraaf	Schotte, Ale Struiksma
Supplemental Table 2. Specialist pulmonary hypertension centres participating in the study		

Recruiting cohorts	n	
Genomics England	1965	
Specialist Pathology: Evaluating Exomes in	1356	
Diagnostics		
Primary Immune Disorders	1299	
Bleeding and Platelet Disorders	1004	
Pulmonary Arterial Hypertension	932	
Multiple Primary Malignant Tumours	376	
Hypertrophic Cardiomyopathy	187	
Cerebral Small Vessel Diseases	183	
Steroid Resistant Nephrotic Syndrome	161	
Intrahepatic Cholestasis of Pregnancy	140	
Stem Cell & Myeloid Disorders	132	
Primary Membranoproliferative Glomerulonephritis	128	
Neuropathic Pain Disorder	114	
Leber Hereditary Optic Neuropathy	59	
Control	15	
Ehlers-Danlos Syndromes 15		
Supplemental Table 3. NIHR BioResource - Rare Diseases Study		
recruiting cohorts and GEL		

Parameter	Response
ID	
Date of birth	
Unenhanced CT	(Y/N)
СТРА	(Y/N)
HRCT	(Y/N)
Expiratory CT	(Y/N)
Pulmonary artery diameter (cm)	
Aorta diameter (cm)	
Ground glass opacification centrilobular pattern DENSITY	(None / Subtle / Present)
Ground glass centrilobular pattern EXTENT	(0, <5%, 5-25, 25-50, >50)
Ground glass DISTRIBUTION	(central (C)/peripheral (P)/zonal (Z) or diffuse (D))
Non-specific mosaic pattern / GGO	
Neovascularity vessels	(Y/N)
Arterio-venous malformations	(Y/N)
Bronchial arteries	(Y/N)
Largest bronchial artery size	
Interlobular septal thickening	(None, Subtle, Present)
Mediastinal lymphadenopathy	(Y/N)
Emphysema	(Y/N) and % of parenchyma involved
Fibrosis	(Y/N) and % of parenchyma involved
Pleural effusion	(Y/N)
Air trapping	(Y/N)
Comments	
Likely diagnosis	Any suspicion of PVOD or PCH / PAH
Supplemental Table 4. Proforma used in analysis of CT scans	

Group	n
PAH patients with BMPR2 variants	21
PAH patients with biallelic <i>EIF2AK4</i>	7
variants	,
PVOD patients	14
PAH patients with heterozygous EIF2AK4	4
variants	4
PAH patients with no variants in the	21
previously reported PAH genes	21

Supplemental Table 5. CT scans of patients with PVOD and patients with PAH carrying biallelic *EIF2AK4* mutations were reassessed by radiologists blinded to the diagnosis. For comparison CT scans of PAH patients with normal spirometry (FEV $_1$ > 80 % predicted and FVC > 80 % predicted) who either had no mutations in the previously reported PAH genes or carried *BMPR2* mutations were assessed.

			Su	pplemental	Table 6. Pa	age 1/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.292C>G	missense variant	p.L98V	0	1	0.00001656	probably damaging (0.999)	deleterious (0)	25.7	Heterozygous variant
BRIDGE control	c.354_355delTG	frameshift variant	p.C118Wfs*7	0	2	Not found in ExAC			35	Heterozygous variant
BRIDGE control	c.745C>T	stop gained & splice region variant	p.R249*	0	1	0.00007451			39	Heterozygous variant
BRIDGE control	c.746G>A	missense variant & splice region variant	p.R249Q	0	1	2.48E-05	probably damaging (0.999)	deleterious (0.02)	34	Heterozygous variant
BRIDGE control	c.767G>T	missense variant	p.C256F	0	1	1.66E-05	possibly damaging (0.904)	deleterious (0.02)	28.4	Heterozygous variant
BRIDGE control	c.985G>A missense p.E329K variant		p.E329K	0	1	Not found in ExAC	probably damaging (0.981)	deleterious (0.01)	34	Heterozygous variant
BRIDGE control	c.1153dupG	frameshift variant	p.V385Gfs*30	0	1	0.00003308			32	Heterozygous variant
BRIDGE control	c.1190T>A	missense variant	p.l397N	0	1	Not found in ExAC	possibly damaging (0.67)	deleterious (0)	32	Heterozygous variant

			S	upplement	al Table 6.	Page 2/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.1215C>G	stop gained	p.Y405*	0	2	Not found in ExAC			29.4	Heterozygou variant
BRIDGE control	c.1331A>G	missense variant	p.Y444C	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	28.7	Heterozygou variant
BRIDGE control	c.1345C>T	missense variant	p.R449C	0	1	0.00001654	probably damaging (1)	deleterious (0)	35	Heterozygou variant
BRIDGE control	c.2249T>A	missense variant & splice region variant	p.L750Q	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	28	Heterozygou variant
BRIDGE control	c.2298delG	frameshift variant	p.N767Tfs*24	0	1	Not found in ExAC			28.3	Heterozygou variant
BRIDGE control	c.2720A>T	missense variant	p.Y907F	0	4	1.66E-05	probably damaging (1)	deleterious (0)	31	Heterozygou variant
BRIDGE control	c.2828C>T	missense variant	p.T943M	0	1	0.00003311	probably damaging (1)	deleterious (0)	34	Heterozygou variant
BRIDGE control	c.3104_3106delT CT	inframe deletion	p.F1035del	0	1	Not found in ExAC			22	Heterozygou variant

			Sı	upplementa	Table 6. Pa	age 3/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.3217C>T	missense variant	p.R1073C	0	1	0.0000166	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.3223T>G	missense variant	p.F1075V	0	1	0.0000083	probably damaging (0.997)	deleterious (0)	32	Heterozygous variant
BRIDGE control	c.3344C>T	missense variant	p.P1115L	0	1	8.26E-06	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.3358-3C>T	splice region variant & intron variant	p.NA	0	1	Not found in ExAC			17.15	Heterozygous variant
BRIDGE control	c.3406C>T	stop gained & splice region variant	p.R1136*	0	1	Not found in ExAC			40	Heterozygous variant
BRIDGE control	c.3430A>T	missense variant	p.R1144W	0	1	0.0000248	probably damaging (1)	deleterious (0)	33	Heterozygous variant
BRIDGE control	c.3986T>C	missense variant	p.F1329S	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	33	Heterozygous variant

			Su	pplemental	Table 6. Pa	age 4/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.3992T>C	missense variant	p.F1331S	0	1	8.28E-06	possibly damaging (0.872)	deleterious (0.01)	28.4	Heterozygous variant
BRIDGE control	c.4039G>A	missense variant	p.A1347T	0	1	8.28E-05	probably damaging (1)	deleterious (0)	34	Heterozygous variant
BRIDGE control	c.4388_4389+12 delAGGTAAAGAC GTCA	splice donor variant & coding sequence variant & intron variant	p.NA	0	1	Not found in ExAC			36	Heterozygous variant
BRIDGE control	c.4397C>A	missense variant	p.S1466Y	0	2	Not found in ExAC	probably damaging (0.988)	deleterious (0)	33	Heterozygous variant
BRIDGE control	c.4729G>A	missense variant & splice region variant	p.V1577M	0	1	Not found in ExAC	probably damaging (0.999)	deleterious (0)	29.6	Heterozygous variant
BRIDGE control	c.4751dupT	frameshift variant	p.L1585lfs*11	0	1	Not found in ExAC			34	Heterozygous variant
BRIDGE control	c.4920_4931delT AGAGATGACTA	inframe deletion	p.R1641_Y1644 del	0	1	Not found in ExAC			23	Heterozygous variant

			Sı	upplemental	Table 6. Pa	ige 5/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
РАН	c.44C>T	missense variant	p.P15L	1	0	8.32E-06	unknown (0)	deleterious low confidence (0.03)	23.5	Heterozygous variant
PAH	c.220G>A	missense variant	p.D74N	1	0	1.66E-05	possibly damaging (0.954)	deleterious (0)	32	Heterozygous variant
PAH	c.1072_1073dup GT	frameshift variant	p.V359*	1	0	Not found in ExAC			32	Heterozygous variant
PAH	c.1660G>T	missense variant & splice region variant	p.D554Y	1	0	Not found in ExAC	probably damaging (0.966)	deleterious (0)	28	Heterozygous variant
PAH	c.2446C>T	stop gained	p.Q816*	1	0	Not found in ExAC	,		41	Heterozygous variant
PAH	c.2516T>C	missense variant	p.I839T	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	28.9	Heterozygous variant
PAH	c.3218G>T	missense variant	p.R1073L	1	0	Not found in ExAC	probably damaging (0.995)	deleterious (0.01)	35	Heterozygous variant
PAH	c.3604C>T	missense variant	p.H1202Y	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	29.7	Heterozygous variant

			Su	pplemental	Table 6. Pa	age 6/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.3711_3713del GAG	inframe deletion	p.R1238del	1	0	0.0000083			21.6	Heterozygous variant
PAH	c.3722A>G	missense variant	p.E1241G	1	0	Not found in ExAC	probably damaging (0.971)	deleterious (0)	27.2	Heterozygous variant
PAH	c.4646G>A	missense variant	p.R1549H	1	0	0.0000910	probably damaging (0.998)	deleterious (0.01)	35	Heterozygous variant
РАН	c.145-2A>G	splice acceptor variant	p.NA	1	0	Not found in ExAC			23.9	Additional second (likely trans) variant identified
PAH	c.257+4A>C	splice region variant & intron variant	p.NA	1	0	8.28E-06			15.5	Additional second (likely trans) variant identified
PAH	c.1392delT	frameshift variant	p.R465Vfs*38	1	0	2.48E-05			35	Additional second (likely trans) variant identified
PAH	c.1739dupA	frameshift variant	p.R581Efs*9	1	0	Not found in ExAC			35	Additional second (likely trans) variant identified

			Su	pplemental	Table 6. Pa	ige 7/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.1820T>G	missense variant & splice region variant	p.V607G	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	27.3	Additional second (likely trans) variant identified
PAH	c.2727C>G	missense variant	p.S909R	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	33	Additional second (likely trans) variant identified
PAH	c.2827A>G	missense variant	p.T943A	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	26.4	Additional second (likely trans) variant identified
PAH	c.2841delG	frameshift variant	p.1948Sfs*35	1	0	Not found in ExAC			35	Additional second (likely trans) variant identified
PAH	c.3055_3064delC TGACCAACG	frameshift variant	p.L1019Wfs*9	1	0	Not found in ExAC			36	Additional second (likely trans) variant identified
PAH	c.3097C>T	stop gained	p.Q1033*	3	0	8.24E-06			45	Additional second (likely trans) variant identified

			Su	pplemental	Table 6. Pa	ige 8/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.3325G>A	missense variant	p.G1109R	1	0	0.0000082	probably damaging (1)	deleterious (0.02)	35	Additional second (likely trans) variant identified
PAH	c.3884T>G	missense variant	p.L1295R	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	32	Additional second (likely trans) variant identified
PAH	c.4400dupT	frameshift variant	p.E1468Rfs*14	1	0	Not found in ExAC			36	Additional second (likely trans) variant identified
PAH	c.4418_4421delC AGA	frameshift variant	p.T1473Rfs*17	1	0	0.0000083			36	Additional second (likely trans) variant identified
PAH	c.4769delT	frameshift variant	p.L1590*	1	0	0.0000083			33	Additional second (likely trans) variant identified
PAH	c.281dupA	frameshift variant	p.N94Lfs*8	2	0	Not found in ExAC			35	Homozygous variant
PAH	c.1159_1160delC T	frameshift variant	p.L387Cfs*27	2	0	Not found in ExAC			29.6	Homozygous variant

			Sı	upplemental	Table 6. Pa	ige 9/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.1795G>C	missense variant	p.G599R	4	0	Not found in ExAC	probably damaging (1)	deleterious (0)	32	Homozygous variant
PAH	c.3097C>T	stop gained	p.Q1033*	3	0	8.24E-06			45	Homozygous variant
PAH	c.3605A>T	missense variant	p.H1202L	2	0	Not found in ExAC	probably damaging (1)	deleterious (0)	31	Homozygous variant
PAH	c.4392dupT	frameshift variant & splice region variant	p.K1465*	2	0	Not found in ExAC			35	Homozygous variant

	Supplemental Table 7. Page 1/4																		
Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVSc	Consequence type	EIF2AK4 genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
22		Duitiah	c.3884T>G	missense variant	Clist			F2	2.2	2	07	110	22	V	Possible		PDE5i +	Na	
23	М	British	c.3055_30 64delCTGA CCAACG	frameshift variant	C Het			52	3.3	3	97	119	33	Yes	PVOD / PCH		ERA + IV Prostanoid	No	
48	М	Other	c.4400dup T	frameshift variant	C Het			46	6.4	3	116	120	45	No	CT not available		ERA + PDE5i +	No	
40	IVI	Other	c.1739dup A	frameshift variant	Criet			40	0.4	3	110	120	43	NO	for analysis		inhaled Prostanoid	NO	
			c.2827A>G	missense variant											CT not				
38	38 I F I	Other Asian	c.4418_44 21delCAGA	frameshift variant	C Het			40	4.5	2				No	available for analysis		ERA + PDE5i	No	
			c.145- 2A>G	splice acceptor variant											anaiysis				

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

	Supplemental Table 7. Page 2/4																		
Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVSc	Consequence type	EIF2AK4 genotype	BMPR2 mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ¹ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
			c.1392del T	frameshift variant											Possible		PDE5i +		
70	F	British	c.257+4A >C	splice region variant & intron variant	C Het			76	6.6	З	101	127	33	Unk	PVOD / PCH		ERA + inhaled Prostanoid	No	
36	F	Indian	c.3605A> T	missense variant	Hom			44	2.7	3	73	83	40	Yes	Possible PVOD / PCH		ERA + PDE5i + inhaled Prostanoid	No	
22	М	Pakistani	c.1795G> C	missense variant	Hom			65	3.0	3	92	93	31	Yes	РАН		ERA + PDE5i + IV Prostanoid	No	Yes
29	Δ	Pakistani	c.3097C> T	stop gained	Hom			50	4.9	3	99	107	27	Unk	РАН	Sister died from PAH	PDE5i	No	
18	М	Not stated	c.1159_1 160delCT	frameshift variant	Hom			92		3	86	82	28	No	Possible PVOD / PCH		ERA + IV Prostanoid	No	
25	F	Pakistani	c.1795G> C	missense variant	Hom			57	5.6	3	82	87	33	No	РАН		PDE5i + ERA	No	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with EIF2AK4 variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

	Supplemental Table 7. Page 3/4																			
Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVSc	Consequence type	EIF2AK4 genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed	
24	F	Not	c.2446C> T	stop gained	Het (both on	ed (both		60	60	5.2	3	96	97	81	Unk	CT not available	Father and sister	Unk	Unk	
24	1	stated	c.3218G> T	missense variant	same allele) *						3	30	3			for analysis	died of PAH	J	5	
39	F	British	c.1072_1 073dupG T	frameshift variant	Het			54	3.0	2	87	98	72	No	CT not available for analysis		ERA	No		
40	F	British	c.44C>T	missense variant	Het		c.4303- 50delT	43	5.6	2	99	96	109	Unk	Possible PVOD / PCH		ERA	No		
44	М	British	c.2516T> C	missense variant	Het	c.853- 2A>G (splice acceptor variant)	c.361- 180A>G	53	3.8	3	102	98	54	Unk	РАН		PDE5i + ERA	No		
25	F	British	c.3722A> G	missense variant	Het					3	53	49	41	No	CT not available for analysis		PDE5i + ERA + IV Prostanoid	No		

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown, *maternally inherited

	Supplemental Table 7. Page 4/4																		
Age (years)	Gender	Ethnicity	EIF2AK4 variant HGVSc	Consequence type	EIF2AK4 genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
66	F	Not stated	c.4646G> A	missense variant	Het			44	2.1	3	79	100		Unk	РАН		PDE5i + ERA	No	
72	М	British	c.1660G> T	missense variant & splice region variant	Het			30	2.8	3				No	РАН		IV Prostanoid	No	
59	F	Other	c.3711_3 713delGA G	inframe deletion	Het			41	3.4	3	68	68	95	Unk	РАН		ERA + PDE5i	No	
48	F	British	c.3604C>	missense variant	Het	c.2695C>T (stop gained)		57	4.4	4	90	100	61	Unk	РАН		PDE5i + ERA	No	
70	F	Other White	c.220G>A	missense variant	Het			42	5.4	2				Unk	CT not available for analysis		ERA	Unk	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with EIF2AK4 variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

	Supplemental Table 8. Page 1/2									
	PAH patients with BMPR2 mutations *	PAH patients with no mutations in PAH associated genes	PAH patients with EIF2AK4 heterozygous variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	р				
n	64	255	3	7	5					
Age (years)	42 [31 - 52]	53 [39 - 67]	39 [32 - 40]	25 [23 - 38]	63 [27 - 76]	<0.001				
Gender (n female [%])	45 [70.3%]	179 [70.2%]	3 [100%]	2 [28.6%]	4 [80%]	0.161				
Ethnicity (n white Caucasian [%])	50 [78.1%]	226 [88.6%]	2 [66.7%]	2 [28.6%]	4 [80%]	<0.001				
Digital clubbing (n [%])	5 [13.2%]	3 [2.2%]	0 [0%]	2 [40%]	0 [0%]	0.004				
ВМІ	28 [25 - 33]	27 [24 - 31]	24 [24 - 25]	24 [21 - 27]	27 [24 - 32]	0.202				

Supplemental Table 8. Phenotype summary of patients with preserved spirometry (FEV₁ > 80 % predicted and FVC > 80 % predicted). PAH patients with biallelic *EIF2AK4* mutations are still younger at diagnosis and have a significantly reduced KCO compared to other groups.

mPAP – mean pulmonary artery pressure, CO – cardiac output, PVR – pulmonary vascular resistance, FEV₁ – forced expiratory volume in 1 second, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide, BMI – body mass index. * Also includes the 2 patients with heterozygous *EIF2AK4* variants and a *BMPR2* mutation. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.

		Supplemental	Table 8. Page 2/2			
	PAH patients with <i>BMPR2</i> mutations *	PAH patients with no mutations in PAH associated genes	PAH patients with EIF2AK4 heterozygous variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	р
mPAP (mmHg)	56 (15)	51 (18)	54 (8)	57 (20)	57 (7)	0.00
CO (L/min)	3 [3 - 4]	4 [3 - 5]	5 [4 - 5]	5 [4 - 6]	3 [3 - 3]	<0.00
PVR (WU)	14 [10 - 18]	10 [7 - 14]	8 [7 - 9]	9 [8 - 15]	14 [11 - 19]	<0.0
Vasoresponders (n [%])	0 [0%]	18 [21.7%]	0 [0%]	0 [0%]		0.01
FEV ₁ (%pred)	97 [88 - 102]	93 [87 - 101]	96 [92 - 97]	97 [89 - 100]	98 [94 - 106]	0.52
FVC (%pred)	102 [96 - 113]	103 [96 - 112]	97 [96 - 98]	107 [90 - 120]	109 [101 - 113]	0.70
KCO (%pred)	80 [71 - 93]	68 [46 - 84]	81 [76 - 95]	33 [30 - 33]	33 [28 - 37]	<0.0
Resting S _A O ₂ (%)	96 [94 - 98]	96 [93 - 98]	98 [98 - 99]	91 [90 - 92]	95 [91 - 95]	0.02
S _A O ₂ post walk test (%)	95 [90 - 98]	91 [85 - 96]	94 [87 - 96]	80 [75 - 84]	85 [85 - 88]	<0.0

Supplemental Table 8. Phenotype summary of patients with preserved spirometry (FEV₁ > 80 % predicted and FVC > 80 % predicted). PAH patients with biallelic *EIF2AK4* mutations are still younger at diagnosis and have a significantly reduced KCO compared to other groups.

mPAP – mean pulmonary artery pressure, CO – cardiac output, PVR – pulmonary vascular resistance, FEV₁ – forced expiratory volume in 1 second, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide, BMI – body mass index. * Also includes the 2 patients with heterozygous *EIF2AK4* variants and a *BMPR2* mutation. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.

Supplemental Table 9. Page 1/2									
Group		All biallelic <i>EIF2AK4</i> mutation carriers	PVOD with no EIF2AK4 mutation	р					
n		11	10						
Age (years)		26.8 [22.5 - 34.3]	68.3 [63.9 - 72.1]	0.001					
Gender (n female [%])		6 [54.5%]	5 [50.0%]	1.000					
Ethnicity (n white Caucasian [%])		5 [45.5%]	9 [90.0%]	0.063					
mPAP (mmHg)		52 [47 - 63]	48 [42 - 57]	0.342					
PCWP (mmHg)		11 [7.5 - 12]	11.5 [9.0 – 12.2]	0.560					
FEV ₁ (% pred)		93.1 [82.8 - 98.5]	79.0 [72.3 – 91.0]	0.236					
FVC (% pred)		95.5 [84.6 - 108.5]	96.0 [73.0 – 101.0]	0.720					
KCO (% pred)		32.0 [28.7 – 33.0]	41.4 [36.8 – 54.0]	0.013					
Contribution and the second states	None	2 [18.2%]	6 [60.0%]						
Centrilobular ground glass opacification density	Subtle	2 [18.2%]	3 [30.0%]	0.012					
opacification density	Present	7 [63.6%]	1 [10.0%]						

Supplemental Table 9. Phenotypic and radiological characteristics of biallelic *EIF2AK4* mutation carriers compared to patients with a clinical diagnosis of PVOD and no *EIF2AK4* mutation.

mPAP – mean pulmonary artery pressure, PCWP – pulmonary capillary wedge pressure, FEV_1 – forced expiratory volume 1 s, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide. Data presented as median [IQR] unless stated.

Supplemental Table 9. Page 2/2									
Group		All biallelic <i>EIF2AK4</i> mutation carriers	PVOD with no EIF2AK4 mutation	р					
	None	2 [18.2%]	7 [70.0%]						
	<5%	1 [9.1%]	1 [10.0%]						
Centrilobular ground glass	5-25%	2 [18.2%]	1 [10.0%]	0.007					
opacification extent	25-50%	1 [9.1%]	1 [10.0%]	0.007					
	50-75%	2 [18.2%]	0 [0.0%]						
	75-100%	3 [27.3%]	0 [0.0%]						
	None	7 [63.6%]	2 [20.0%]						
Interlobular septal thickening	Subtle	0 [0.0%]	1 [10.0%]	0.068					
	Present	4 [36.4%]	7 [70.0%]						
Mediastinal	None	4 [36.4%]	2 [20.0%]						
lymphadenopathy	Present	7 [63.6%]	8 [80.0%]	0.635					
Diamed offusion	None	11 [100.0%]	6 [60.0%]	0.025					
Pleural effusion	Small	0 [0.0%]	4 [40.0%]	0.035					
Nie europeulo witeu	None	10 [90.9%]	9 [90.0%]	1 000					
Neovascularity	Present	1 [9.1%]	1 [10.0%]	1.000					
CT diagnosis	PAH	4 [36.4%]	3 [30.0%]						
CT diagnosis	Possible PVOD/PCH	7 [63.6%]	7 [70.0%]						

Supplemental Table 9. Phenotypic and radiological characteristics of biallelic *EIF2AK4* mutation carriers compared to patients with a clinical diagnosis of PVOD and no *EIF2AK4* mutation.

mPAP - mean pulmonary artery pressure, PCWP - pulmonary capillary wedge pressure, FEV_1 - forced expiratory volume 1 s, FVC - forced vital capacity, KCO - transfer coefficient for carbon monoxide. Data presented as median [IQR] unless stated.

Group	Time to assessment 1 (days)	n	Change in 6mwd (m)	Change in FC	Time to assessment 2 (days)	n	Change in 6mwd (m)	Change in FC	Number on prostanoid therapy before the 2 nd assessment [%]
PAH <i>BMPR2</i>	357 [314 - 386]	21	+69 [20 - 100]	-1 [-11]	1120 [1055 - 1174]	18	+45 [31 - 115]	-1 [-10.5]	5 [23%]
PAH biallelic EIF2AK4	358 [335 -388]	9	+28 [-13 - 77]	0 [-1 - 0]	1102 [1090 – 1112]	5	+62 [-8 - 132]	0 [0 - 0]	1 [10%]
PAH no mutation	387 [340 - 414]	16	+81 [61 - 151]	-1 [-1 - 0]	1118 [1105 - 1159]	9	+104 [20 - 144]	-1 [-1 - 0]	4 [17%]
р	0.295		0.343	0.039	0.730		0.748	0.044	0.816

Supplemental Table 10. Response to pulmonary artery vasodilator therapies at 1 and 3 years after diagnosis compared to baseline. 6mwd - six-minute walk test distance, FC - functional class. Drop in number of patients between assessment 1 and 2 due to death, transplantation or lack of sufficient follow up time. Data presented as median [IQR] unless stated.

Variable	Hazard Ratio [95% confidence interval]	р
PAH BMPR2 mutation*	0.148 [0.055 - 0.396]	<0.001
PAH no mutation*	0.179 [0.073 - 0.440]	<0.001
PVOD*	0.393 [0.075 - 2.065]	0.27
Age at diagnosis	1.043 [1.033 - 1.053]	<0.001
Male gender	1.631 [1.222 - 2.179]	<0.001

Supplemental Table 11. Cox proportional hazards model assessing time to death. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations had an increased risk of death compared to other PAH patients. Number of patients = 858. Events = 194.

* compared to the PAH biallelic *EIF2AK4* mutation carriers

Variable	Hazard Ratio [95% confidence interval]	р
PAH <i>BMPR2</i> mutation*	0.175 [0.066 - 0.462]	<0.001
PAH no mutation*	0.203 [0.083 - 0.501]	<0.001
PVOD*	0.840 [0.222 - 3.193]	0.798
Age at diagnosis	1.036 [1.027 - 1.046]	<0.001
Male gender	1.542 [1.165 - 2.042]	0.002

Supplemental Table 12. Cox proportional hazards model assessing time to death or transplantation. Number of patients = 858. Events = 208.

Variable	Hazard Ratio [95% confidence interval]	р
PAH BMPR2 mutation*	0.376 [0.080 - 1.763]	0.215
PAH no mutation*	0.456 [0.109 - 1.905]	0.282
PVOD*	1.029 [0.133 - 7.953]	0.978
Age at diagnosis	1.034 [1.020 - 1.046]	<0.001
Male gender	1.515 [1.000 - 2.296]	0.051

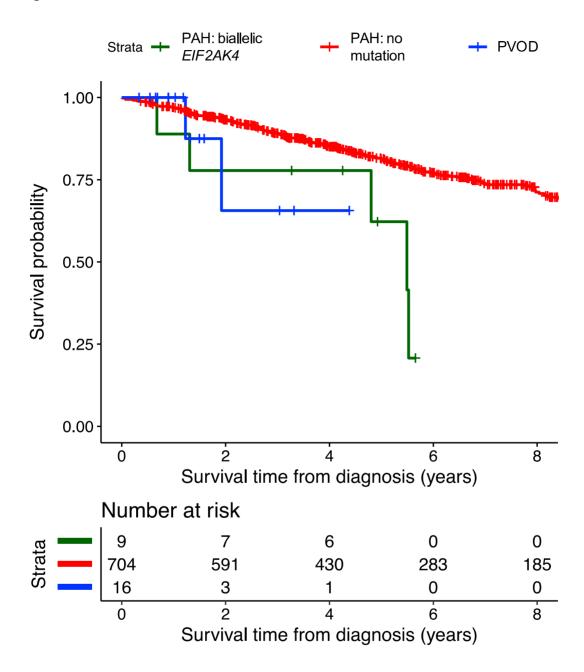
Supplemental Table 13. Sensitivity analysis including only prospectively recruited UK patients. Cox proportional hazards model assessing time to death. Number of patients = 608. Events = 95.

^{*} compared to the PAH biallelic EIF2AK4 mutation carriers

^{*} compared to the PAH biallelic *EIF2AK4* mutation carriers

Supplemental Figures

Figure S1



Supplemental Figure Legends:

Figure S1: Kaplan – Meier survival curves showing survival time (time to death) for patients with a clinical diagnosis of PAH or PVOD.

Supplemental References

- 1. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P and Cunningham F. The Ensembl Variant Effect Predictor. *Genome Biol.* 2016;17:122.
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