Genetic studies of birth weight give biological insights into links with adult disease

Momoko Horikoshi^{1,2,*}, Robin N Beaumont^{3,*}, Felix R Day^{4,*}, Nicole M Warrington^{5,6,*}, Marjolein N Kooijman^{7,8,9,*}, Juan Fernandez-Tajes^{1,*}, Bjarke Feenstra¹⁰, Natalie R van Zuydam^{1,2}, Kyle J Gaulton^{1,11}, Niels Grarup¹², Jonathan P Bradfield¹³, David P Strachan¹⁴, Ruifang Li-Gao¹⁵, Tarunveer S Ahluwalia^{12,16,17}, Eskil Kreiner-Møller¹⁶, Rico Rueedi^{18,19}, Leo-Pekka Lyytikäinen^{20,21}, Diana L Cousminer^{22,23,24}, Ying Wu²⁵, Elisabeth Thiering^{26,27}, Carol A Wang⁶, Christian T Have¹², Jouke-Jan Hottenga²⁸, Natalia Vilor-Tejedor^{29,30,31}, Peter K Joshi³², Eileen Tai Hui Boh³³, Ioanna Ntalla^{34,35}, Niina Pitkänen³⁶, Anubha Mahajan¹, Elisabeth M van Leeuwen⁸, Raimo Joro³⁷, Vasiliki Lagou^{1,38,39}, Michael Nodzenski⁴⁰, Louise A Diver⁴¹, Krina T Zondervan^{1,42}, Mariona Bustamante^{29,30,31,43}, Pedro Marques-Vidal⁴⁴, Josep M Mercader⁴⁵, Amanda J Bennett², Nilufer Rahmioglu¹, Dale R Nyholt⁴⁶, Ronald Ching Wan Ma^{47,48,49}, Claudia Ha Ting Tam⁴⁷, Wing Hung Tam⁵⁰, CHARGE Consortium Hematology Working Group, Santhi K Ganesh⁵¹, Frank JA van Rooij⁸, Samuel E Jones³, Po-Ru Loh^{52,53}, Katherine S Ruth³, Marcus A Tuke³, Jessica Tyrrell^{3,54}, Andrew R Wood³, Hanieh Yaghootkar³, Denise M Scholtens⁴⁰, Lavinia Paternoster^{55,56}, Inga Prokopenko^{1,57}, Peter Kovacs⁵⁸, Mustafa Atalay³⁷, Sara M Willems⁸, Kalliope Panoutsopoulou⁵⁹, Xu Wang³³, Lisbeth Carstensen¹⁰, Frank Geller¹⁰, Katharina E Schraut³², Mario Murcia^{31,60}, Catharina EM van Beijsterveldt²⁸, Gonneke Willemsen²⁸, Emil V R Appel¹², Cilius E Fonvig^{12,61}, Caecilie Trier^{12,61}, Carla MT Tiesler^{26,27}, Marie Standl²⁶, Zoltán Kutalik^{19,62}, Sílvia Bonas-Guarch⁴⁵, David M Hougaard^{63,64}, Friman Sánchez^{45,65}, David Torrents^{45,66}, Johannes Waage¹⁶, Mads V Hollegaard^{63,64,‡}, Hugoline G de Haan¹⁵, Frits R Rosendaal¹⁵, Carolina Medina-Gomez^{7,8,67}, Susan M Ring^{55,56}, Gibran Hemani^{55,56}, George McMahon⁵⁶, Neil R Robertson^{1,2}, Christopher J Groves², Claudia Langenberg⁴, Jian'an Luan⁴, Robert A Scott⁴, Jing Hua Zhao⁴, Frank D Mentch¹³, Scott M MacKenzie⁴¹, Rebecca M Reynolds⁶⁸, William L Lowe Jr⁶⁹, Anke Tönjes⁷⁰, Michael Stumvoll^{58,70}, Virpi Lindi³⁷, Timo A Lakka^{37,71,72}, Cornelia M van Duijn⁸, Wieland Kiess⁷³, Antje Körner^{58,73}, Thorkild IA Sørensen^{55,56,74,75}, Harri Niinikoski^{76,77}, Katja Pahkala^{36,78}, Olli T Raitakari^{36,79}, Eleftheria Zeggini⁵⁹, George V Dedoussis³⁵, Yik-Ying Teo^{33,80,81}, Seang-Mei Saw^{33,82}, Mads Melbye^{10,83,84}, Harry Campbell³², James F Wilson^{32,85}, Martine Vrijheid^{29,30,31}, Eco JCN de Geus^{28,86}, Dorret I Boomsma²⁸, Haja N Kadarmideen⁸⁷, Jens-Christian Holm^{12,61}, Torben Hansen¹², Sylvain Sebert^{88,89}, Andrew T Hattersley³, Lawrence J Beilin⁹⁰, John P Newnham⁶, Craig E Pennell⁶, Joachim Heinrich^{26,91}, Linda S Adair⁹², Judith B Borja^{93,94}, Karen L Mohlke²⁵, Johan G Eriksson^{95,96,97}, Elisabeth E Widén²², Mika Kähönen^{98,99}, Jorma S Viikari^{100,101}, Terho Lehtimäki^{20,21}, Peter Vollenweider⁴⁴, Klaus Bønnelykke¹⁶, Hans Bisgaard¹⁶, Dennis O Mook-Kanamori^{15,102,103}, Albert Hofman^{7,8}, Fernando Rivadeneira^{7,8,67}, André G Uitterlinden^{7,8,67}, Charlotta Pisinger¹⁰⁴, Oluf Pedersen¹², Christine Power¹⁰⁵, Elina Hyppönen^{105,106,107}, Nicholas J Wareham⁴, Hakon Hakonarson^{13,23,108}, Eleanor Davies⁴¹, Brian R Walker⁶⁸, Vincent WV Jaddoe^{7,8,9}, Marjo-Riitta Jarvelin^{88,89,109,110}, Struan FA Grant^{13,23,108,111}, Allan A Vaag^{83,112}, Debbie A Lawlor^{55,56}, Timothy M Frayling³, George Davey Smith^{55,56}, Andrew P Morris^{1,113,114,§}, Ken K Ong^{4,115,§}, Janine F Felix^{7,8,9,§}, Nicholas J Timpson^{55,56,§}, John RB Perry^{4,§}, David M Evans^{5,55,56,§}, Mark I McCarthy^{1,2,116,§}, Rachel M Freathy^{3,55,§}, on behalf of the Early Growth Genetics (EGG) Consortium

- 1. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK.
- 2. Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Oxford, UK.
- 3. Institute of Biomedical and Clinical Science, University of Exeter Medical School, Royal Devon and Exeter Hospital, Exeter, UK.
- 4. MRC Epidemiology Unit, University of Cambridge School of Clinical Medicine, Cambridge, UK.
- 5. The University of Queensland Diamantina Institute, Translational Research Institute, Brisbane, Australia.
- 6. School of Women's and Infants' Health, The University of Western Australia, Perth, Australia.

- 7. The Generation R Study Group, Erasmus MC, University Medical Center Rotterdam, the Netherlands.
- 8. Department of Epidemiology, Erasmus MC, University Medical Center Rotterdam, the Netherlands.
- 9. Department of Pediatrics, Erasmus MC, University Medical Center Rotterdam, the Netherlands.
- 10. Department of Epidemiology Research, Statens Serum Institute, Copenhagen, Denmark.
- 11. Department of Pediatrics, University of California San Diego, La Jolla, California, USA.
- 12. The Novo Nordisk Foundation Center for Basic Metabolic Research, Section of Metabolic Genetics, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark.
- 13. Center for Applied Genomics, The Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA.
- 14. Population Health Research Institute, St George's University of London, London, Cranmer Terrace, UK.
- 15. Department of Clinical Epidemiology, Leiden University Medical Center, Leiden, the Netherlands.
- 16. COPSAC, Copenhagen Prospective Studies on Asthma in Childhood, Herlev and Gentofte Hospital, University of Copenhagen, Copenhagen, Denmark.
- 17. Steno Diabetes Center, Gentofte, Denmark.
- 18. Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland.
- 19. Swiss Institute of Bioinformatics, Lausanne, Switzerland.
- 20. Department of Clinical Chemistry, Fimlab Laboratories, Tampere, Finland.
- 21. Department of Clinical Chemistry, University of Tampere School of Medicine, Tampere, Finland.
- 22. Institute for Molecular Medicine, Finland (FIMM), University of Helsinki, Helsinki, Finland.
- 23. Division of Human Genetics, The Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA.
- 24. Department of Genetics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA.
- 25. Department of Genetics, University of North Carolina, Chapel Hill, NC, USA.
- 26. Institute of Epidemiology I, Helmholtz Zentrum München- German Research Center for Environmental Health, Neuherberg, Germany.
- 27. Division of Metabolic and Nutritional Medicine, Dr. von Hauner Children's Hospital, University of Munich Medical Center, Munich, Germany.
- 28. Netherlands Twin Register, Department of Biological Psychology, VU University, Amsterdam, the Netherlands.
- 29. ISGlobal, Centre for Research in Environmental Epidemiology (CREAL), Barcelona, Spain.
- 30. Universitat Pompeu Fabra (UPF), Barcelona, Spain.
- 31. CIBER de Epidemiología y Salud Pública (CIBERESP), Spain.
- 32. Usher Institute for Population Health Sciences and Informatics, University of Edinburgh, Edinburgh, Scotland, UK.
- 33. Saw Swee Hock School of Public Health, National University of Singapore, National University Health System, Singapore, Singapore.
- 34. William Harvey Research Institute, Barts and the London School of Medicine and Dentistry, Queen Mary University of London, London, UK.
- 35. Department of Nutrition and Dietetics, School of Health Science and Education, Harokopio University, Athens, Greece.
- 36. Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, Finland.
- 37. Institute of Biomedicine, Physiology, University of Eastern Finland, Kuopio, Finland.
- 38. KUL University of Leuven, Department of Neurosciences, Leuven, Belgium.
- 39. Translational Immunology Laboratory, VIB, Leuven, Belgium.

- 40. Department of Preventive Medicine, Division of Biostatistics, Feinberg School of Medicine, Northwestern University, Chicago, USA.
- 41. Institute of Cardiovascular & Medical Sciences, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, UK.
- 42. Endometriosis CaRe Centre, Nuffield Department of Obstetrics & Gynaecology, University of Oxford, Oxford, UK.
- 43. Center for Genomic Regulation (CRG), Barcelona, Spain.
- 44. Department of Internal Medicine, Internal Medicine, Lausanne University Hospital (CHUV), Lausanne, Switzerland.
- 45. Joint BSC-CRG-IRB Research Program in Computational Biology, Barcelona Supercomputing Center, Barcelona, Spain.
- 46. Institute of Health and Biomedical Innovation, Queensland University of Technology, Queensland, Australia.
- 47. Department of Medicine and Therapeutics, The Chinese University of Hong Kong, Hong Kong, Hong Kong, China.
- 48. Li Ka Shing Institute of Health Sciences, The Chinese University of Hong Kong, Hong Kong, Hong Kong, China.
- 49. Hong Kong Institute of Diabetes and Obesity, The Chinese University of Hong Kong, Hong Kong, China.
- 50. Department of Obstetrics and Gynaecology, The Chinese University of Hong Kong, Hong Kong, Hong Kong, China.
- 51. Cardiovascular Medicine, Department of Internal Medicine, University of Michigan, Ann Arbor, Michigan, USA.
- 52. Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, Massachusetts, USA.
- 53. Program in Medical and Population Genetics, Broad Institute of Harvard and MIT, Cambridge, Massachusetts, USA.
- 54. European Centre for Environment and Human Health, University of Exeter, Truro, UK.
- 55. Medical Research Council Integrative Epidemiology Unit at the University of Bristol, Bristol, UK.
- 56. School of Social and Community Medicine, University of Bristol, Bristol, UK.
- 57. Department of Genomics of Common Disease, School of Public Health, Imperial College London, London, UK.
- 58. IFB Adiposity Diseases, University of Leipzig, Leipzig, Germany.
- 59. Wellcome Trust Sanger Institute, Hinxton, Cambridgeshire, UK.
- 60. FISABIO–Universitat Jaume I–Universitat de València, Joint Research Unit of Epidemiology and Environmental Health, Valencia, Spain.
- 61. The Children's Obesity Clinic, Department of Pediatrics, Copenhagen University Hospital Holbæk, Holbæk, Denmark.
- 62. Institute of Social and Preventive Medicine, Lausanne University Hospital (CHUV), Lausanne, Switzerland.
- 63. Danish Center for Neonatal Screening, Statens Serum Institute, Copenhagen, Denmark.
- 64. Department for Congenital Disorders, Statens Serum Institute, Copenhagen, Denmark.
- 65. Computer Sciences Department, Barcelona Supercomputing Center, Barcelona, Spain.
- 66. Institució Catalana de Recerca i Estudis Avançats (ICREA), Barcelona, Spain.
- 67. Department of Internal Medicine, Erasmus MC, University Medical Center Rotterdam, the Netherlands.
- 68. BHF Centre for Cardiovascular Science, University of Edinburgh, Queen's Medical Research Institute, Edinburgh, Scotland, UK.
- 69. Department of Medicine, Division of Endocrinology, Metabolism, and Molecular Medicine, Feinberg School of Medicine, Northwestern University, Chicago, USA.
- 70. Medical Department, University of Leipzig, Leipzig, Germany.

- 71. Department of Clinical Physiology and Nuclear Medicine, Kuopio University Hospital, Kuopio, Finland.
- 72. Kuopio Research Institute of Exercise Medicine, Kuopio, Finland.
- 73. Pediatric Research Center, Department of Women's & Child Health, University of Leipzig, Leipzig, Germany.
- 74. Novo Nordisk Foundation Center for Basic Metabolic Research and Department of Public Health, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark.
- 75. Institute of Preventive Medicine, Bispebjerg and Frederiksberg Hospital, The Capital Region, Copenhagen, Denmark.
- 76. Department of Pediatrics, Turku University Hospital, Turku, Finland.
- 77. Department of Physiology, University of Turku, Turku, Finland.
- 78. Paavo Nurmi Centre, Sports and Exercise Medicine Unit, Department of Physical Activity and Health, Turku, Finland.
- 79. Department of Clinical Physiology and Nuclear Medicine, Turku University Hospital, Turku, Finland.
- 80. Department of Statistics and Applied Probability, National University of Singapore, Singapore, Singapore.
- 81. Life Sciences Institute, National University of Singapore, Singapore, Singapore.
- 82. Singapore Eye Research Institute, Singapore, Singapore.
- 83. Department of Clinical Medicine, Copenhagen University, Copenhagen, Denmark.
- 84. Department of Medicine, Stanford School of Medicine, Stanford, California, USA.
- 85. MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, Scotland, UK.
- 86. EMGO Institute for Health and Care Research, VU University and VU University Medical Center, Amsterdam, the Netherlands.
- 87. Department of Large Animal Sciences, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark.
- 88. Center for Life Course Health Research, Faculty of Medicine, University of Oulu, Oulu, Finland.
- 89. Biocenter Oulu, University of Oulu, Finland.
- 90. School of Medicine and Pharmacology, Royal Perth Hospital Unit, The University of Western Australia, Perth, Australia.
- 91. Institute and Outpatient Clinic for Occupational, Social and Environmental Medicine, Inner City Clinic, University Hospital Munich, Ludwig Maximilian University of Munich, Munich, Germany.
- 92. Department of Nutrition, University of North Carolina, Chapel Hill, NC, USA.
- 93. USC-Office of Population Studies Foundation, Inc., University of San Carlos, Cebu City, Philippines.
- 94. Department of Nutrition and Dietetics, University of San Carlos, Cebu City, Philippines.
- 95. National Institute for Health and Welfare, Helsinki, Finland.
- 96. Department of General Practice and Primary Health Care, University of Helsinki and Helsinki University Hospital, Helsinki, Finland.
- 97. Folkhälsan Research Center, Helsinki, Finland.
- 98. Department of Clinical Physiology, Tampere University Hopital, Tampere, Finland.
- 99. Department of Clinical Physiology, University of Tampere School of Medicine, Tampere, Finland.
- 100. Division of Medicine, Turku University Hospital, Turku, Finland.
- 101. Department of Medicine, University of Turku, Turku, Finland.
- 102. Department of Public Health and Primary Care, Leiden University Medical Center, Leiden, the Netherlands.
- 103. Epidemiology Section, BESC Department, King Faisal Specialist Hospital and Research Centre, Riyadh, Saudi Arabia.

- 104. Research Center for Prevention and Health Capital Region, Center for Sundhed, Rigshospitalet Glostrup, Copenhagen University, Glostrup, Denmark.
- 105. Population, Policy and Practice, UCL Institute of Child Health, University College London, London, UK.
- 106. Centre for Population Health Research, School of Health Sciences, and Sansom Institute, University of South Australia, Adelaide, Australia.
- 107. South Australian Health and Medical Research Institute, Adelaide, Australia.
- 108. Department of Pediatrics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA.
- 109. Department of Epidemiology and Biostatistics, MRC–PHE Centre for Environment & Health, School of Public Health, Imperial College London, London, UK.
- 110. Unit of Primary Care, Oulu University Hospital, Oulu, Finland.
- 111. Division of Endocrinology, The Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA.
- 112. Department of Endocrinology, Rigshospitalet, Copenhagen, Denmark.
- 113. Department of Biostatistics, University of Liverpool, Liverpool, UK.
- 114. Estonian Genome Center, University of Tartu, Tartu, Estonia.
- 115. Department of Paediatrics, University of Cambridge, Cambridge, UK.
- 116. Oxford National Institute for Health Research (NIHR) Biomedical Research Centre, Churchill Hospital, Oxford, UK.

^{*}These authors contributed equally to this work.

[§]These authors jointly directed this work.

[†]Deceased.

2 reproducibly associated with future risk of adult metabolic diseases including type 2 diabetes (T2D) and cardiovascular disease¹. These lifecourse associations have often been attributed to the 3 4 impact of an adverse early life environment. We performed a multi-ancestry genome-wide 5 association study (GWAS) meta-analysis of BW in 153,781 individuals, identifying 60 loci where 6 foetal genotype was associated with BW ($P < 5 \times 10^{-8}$). Overall, ~15% of variance in BW could be 7 captured by assays of foetal genetic variation. Using genetic association alone, we found a strong inverse genetic correlation between BW and systolic blood pressure (r_e =-0.22, P=5.5x10⁻¹³), T2D 8 9 (r_g=-0.27, P=1.1x10⁻⁶) and coronary artery disease (r_g=-0.30, P=6.5x10⁻⁹) and, in large cohort data sets, that genetic factors were the major contributor to the negative covariance between BW and 10 11 future cardiometabolic risk. Pathway analyses indicated that the protein products of genes within 12 BW-associated regions were enriched for diverse processes including insulin signalling, glucose 13 homeostasis, glycogen biosynthesis and chromatin remodelling. There was also enrichment of 14 associations with BW in known imprinted regions (P=1.9x10⁻⁴). We have demonstrated that 15 lifecourse associations between early growth phenotypes and adult cardiometabolic disease are in 16 part the result of shared genetic effects and have highlighted some of the pathways through which

Birth weight (BW) is influenced by both foetal and maternal factors and in observational studies is

- 17 these causal genetic effects are mediated.
- 18

1

We combined GWAS data for BW in 153,781 individuals representing multiple ancestries from 37
 studies across three components (Extended Data 1, Supplementary Table 1): (i) 75,891 individuals

of European ancestry from 30 studies; (ii) 67,786 individuals of European ancestry from the UK

22 Biobank; and (iii) 10,104 individuals of diverse ancestries (African American, Chinese, Filipino,

23 Surinamese, Turkish and Moroccan) from six studies. Within each study, BW was z-score

24 transformed separately in males and females after excluding non-singletons and premature births

and adjusting for gestational age where available. Genotypes were imputed using reference panels

from the 1000 Genomes (1000G)² or combined 1000G and UK10K Project³ (Supplementary Table 2).
 We performed quality control assessments to confirm that the distribution of BW was consistent

We performed quality control assessments to confirm that the distribution of BW was consistent
 across studies, irrespective of the data collection protocol, and confirmed that self-reported BW in

29 UK Biobank showed genetic and phenotypic associations consistent with those seen for measured
 30 BW in other studies⁴ (Methods).

31

We identified 60 loci (59 autosomal) associated with BW at genome-wide significance ($P < 5 \times 10^{-8}$) in either the European ancestry or trans-ancestry meta-analyses (**Figure 1a, Extended Data 2**,

34 **Supplementary Data; Methods**). At lead SNPs, we observed no heterogeneity in allelic effects

35 between the three study components (Cochran's *Q* statistic *P*>0.00083) (**Supplementary Table 3**).

36 Fifty-three of these loci were novel in that the lead SNP mapped >2Mb away from, and was

statistically independent (EUR r^2 <0.05) of, the seven previously-reported BW signals⁵, all of which

38 were confirmed in this larger analysis (Supplementary Table 4). Approximate conditional analysis in

39 the European ancestry data indicated that three of these novel loci (near *ZBTB7B, HMGA1* and

40 *PTCH1*) harboured multiple distinct association signals attaining genome-wide significance

- 41 (Methods; Supplementary Table 5, Extended Data 3).
- 42

The lead variants for most signals mapped to non-coding sequence, and at only two loci, *ADRB1* (rs7076938; r^2 =0.99 with *ADRB1* G389R) and *NRIP1* (rs2229742, R448G) did the association data

45 point to likely causal non-synonymous coding variants (**Supplementary Table 6; Methods**). Lead

46 SNPs for all but two loci (those mapping near *YKT6-GCK* and *SUZ12P1-CRLF3*) were common (minor

allele frequency (MAF) \geq 5%) with individually modest effects on BW (β =0.020-0.053 per allele,

48 equivalent to 10 to 26g). This was despite much improved coverage of low-frequency variants in this

49 study (compared to previous HapMap 2 imputed meta-analyses⁵) reflecting imputation from larger,

50 and more complete, reference panels (Extended Data 4). Indeed, all but five of the common variant

association signals were tagged by variants (EUR r^2 >0.6) in the HapMap 2 reference panel

52 (Supplementary Tables 4 and 5), indicating that most of the novel discovery in the present study 53 was driven by increased sample size⁵. Fine-mapping analysis yielded 14 regions within which fewer 54 than ten variants contributed to the locus-specific "credible set" accounting for >99% of the 55 posterior probability of association (Methods; Supplementary Table 7). Collectively, these credible 56 set variants showed enrichment for overlap with DNasel hypersensitivity sites, particularly those 57 generated by ENCODE, from foetal (4.2-fold, 95% CI [1.8-10.7]) and neonatal tissues (4.9 [1.8-11.0]) 58 (Supplementary Table 8; Methods). The greatest refinement was at YKT6-GCK, where the credible 59 set included only the low frequency variant rs138715366, which maps intronic to YKT6. 60 In combination, the 62 distinct genome-wide significant signals at the 59 autosomal loci, explained 61 62 2.0% (standard error (SE) 1.1%) of variance in BW (Supplementary Table 9; Methods), similar in magnitude to that attributable to sex or maternal body mass index (BMI)⁵. However, the variance in 63 64 BW captured collectively by all autosomal genotyped variants on the array was considerably larger, 65 estimated at 15.1% (SE=0.9) in UK Biobank (Methods). These figures are consistent with a long tail of 66 genetic variants of smaller effects contributing to variation in BW. 67

68 Associations between foetal genotype and BW could result from indirect effects of the maternal 69 genotype (r with foetal genotype ≈ 0.5) influencing BW via the intrauterine environment. However, 70 two lines of evidence indicated that variation in the foetal genome was the predominant driver of 71 the BW associations. First, an analysis of the global contribution of maternal vs. foetal genetic 72 variation, using a maternal-GCTA model^b in 4,382 mother-child pairs, estimated that the child's genotype (σ_c^2 =0.24, SE=0.11) makes a larger contribution to BW variance than either the mother's 73 74 genotype (σ_M^2 =0.04, SE=0.10), or the covariance between the two (σ_{CM} =0.04, SE=0.08). Second, 75 when we compared, at each of the 60 loci, the point estimates of the BW effect size dependent on 76 maternal genotype (as measured in up to 68,254 women') with those dependent on foetal genotype 77 (using European ancestry data from 143,677 individuals in the present study), foetal variation had greater impact than maternal at 93% of loci (55/60; binomial $P=1\times10^{-11}$) (Supplementary Table 10, 78 79 Extended Data 5 and 6; Methods). Notably, the association at MTNR1B, one of the five loci at which the maternal effect size exceeded the foetal (maternal: β =0.048, *P*=5.1x10⁻¹⁵; foetal: β =0.023, 80 P=2.9x10⁻⁸), is consistent with prior evidence pointing towards a maternal effect: MTNR1B variation 81 is associated with hyperglycaemia and impaired glucose tolerance^{8,9} and these in turn have been 82 shown to be causally related to offspring BW^{10,11}. Power to disentangle maternal and foetal 83 84 contributions using analyses of foetal genotype conditional on maternal genotype was constrained 85 by the limited sample size available (n=12,909 mother-child pairs) (Supplementary Table 11). 86

These analyses collectively provide compelling evidence that foetal genotype has a substantial
impact on early growth, as measured by BW. We sought to use these genetic associations to
understand the causal relationships between the observed BW-disease associations and to
characterise the underlying processes responsible.

91

92 To quantify the shared genetic contribution to BW and other health-related traits, we estimated 93 their genetic correlations using LD Score regression¹² (**Methods**). BW (in European ancestry samples) showed strong positive genetic correlations with anthropometric and obesity-related traits including 94 birth length ($r_g=0.81$, $P=2.0x10^{-44}$), and in adults, height ($r_g=0.41$, $P=4.8x10^{-52}$), waist circumference 95 $(r_g=0.18, P=3.9 \times 10^{-10})$ and BMI $(r_g=0.11, P=7.3 \times 10^{-6})$. In contrast, BW showed inverse genetic 96 97 correlations with indicators of adverse metabolic and cardiovascular health including coronary artery disease (CAD r_g =-0.30, P=6.5x10⁻⁹), systolic blood pressure (SBP r_g =-0.22, P=5.5x10⁻¹³) and T2D (r_g =-98 99 0.27, $P=1.1 \times 10^{-6}$) (Figure 2, Supplementary Table 12). These correlations between BW and adult 100 cardiometabolic phenotypes are of similar magnitude, although directionally-opposite, to the reported genetic correlations between adult BMI and those same cardiometabolic outcomes¹². 101 102 These findings support observational associations between paternal T2D and lower BW⁴, and

establish more generally that the observed lifecourse associations between early growth and adult disease, at least in part, reflect the impact of shared genetic variants that influence both directly. In

an effort to estimate the extent of genetic contribution to these lifecourse associations, we first

106 focused on data from UK Biobank (n=57,715). For many of the traits for which data were available,

- 107 genetic variation significantly contributed to the lifecourse relationship between BW and adult
- 108 phenotypes, and in some cases appeared to be the major source of covariance between the traits.
- For example, we estimated that 85% (95% CI=70%-99%) of the negative covariance between BW and
 SBP was explained by shared genetic associations captured by directly genotyped SNPs
- 111 (Supplementary Table 13). For continuous cardiometabolic measures, including lipids and fasting
- 112 glycaemia, for which measures are not currently available in UK Biobank, we turned to the Northern
- 113 Finland Birth Cohort (n=5,009), and obtained similar results (**Supplementary Table 13**). However,
- these estimates are limited, not only by wide confidence intervals, but also by the assumption of a
- linear relationship between BW and each of the phenotypes and inability to explicitly model
- 116 maternal genotypic effects. In other words, the negative genetic correlations between BW and
- 117 cardiometabolic traits may not exclusively reflect genetic effects mediated directly through the
- offspring, but also effects mediated by maternal genotype acting indirectly via perturbation of the *in*
- 119 utero environment. Nevertheless, these estimates indicate that a substantial proportion of the 120 variance in cardiometabolic risk that covaries with BW can be attributed to the effects of common
- variance in cardiometabolic risk that covaries with BW can be attributed to the effects of commongenetic variation.
- 122
- 123 To elucidate the biological pathways and processes underlying regulation of foetal growth, we first
- 124 performed gene set enrichment analysis of our BW GWAS analysis using MAGENTA¹³ (**Methods**).
- 125 Twelve pathways reached study-wide significance (FDR<0.05), including pathways involved in
- 126 metabolism (insulin signalling, glycogen biosynthesis, cholesterol biosynthesis), growth (IGF-
- signalling, growth hormone pathway) and development (chromatin remodelling) (**Extended Data**
- **7a**). Similar pathways were detected in a complementary analysis where we interrogated empirical
- protein-protein interaction (PPI) data (collated within Inweb¹⁴) identifying 13 PPI network modules
- 130 with marked (z-score>5) enrichment for BW-association scores (Extended Data 7b and 8a,b;
- **Methods**). The proteins within these modules were themselves enriched for diverse processes
- 132 related to metabolism, growth and development (**Extended Data 8a,b**).
- 133
- We also observed enrichment of BW association signals across the set of 77 imprinted genes defined 134 by the Genotype-Tissue Expression (GTEx) project¹⁵ (*P*=1.9 x 10⁻⁴; Extended Data 7a, Supplementary 135 136 Table 14). Such enrichment is consistent with the "parental conflict" hypothesis regarding the allocation of maternal resources to the foetus¹⁶. Although the role of imprinted genes in foetal 137 growth is described in animal models and rare human disorders¹⁷, our result is the first large-scale, 138 139 systematic demonstration of their contribution to normal variation in BW. Of the 60 genome-wide 140 significant loci, two (INS-IGF2, RB1) fall within (or near) imprinted regions (Figure 1b), with a 141 noteworthy third signal at *DLK1* (previously foetal antigen-1; *P*=5.6x10⁻⁸). Parent-of-origin specific 142 analyses to further investigate these individual loci (comparing heterozygote vs. homozygote BW 143 variance in 57,715 unrelated individuals and testing BW associations with paternal vs. maternal 144 alleles in 4,908 mother-child pairs; see Methods) proved, despite these sample sizes, to be
- 145 underpowered (Extended Data 9, Supplementary Tables 15 and 16).
- 146
- 147 Many of the genome-wide signals for BW detected here are also established genome-wide
- association signals for a wide variety of cardiometabolic traits (Figure 3). These include the BW
- signals near CDKAL1, ADCY5, HHEX/IDE and ANK1 (also genome-wide significant for T2D), NT5C2 and
- 150 *ADRB1* (BP/CAD/BMI). We used two approaches to understand whether this pattern of adult trait
- association represented a generic property of BW-associated loci, or reflected heterogeneous
- 152 mechanisms linking BW to adult disease.
- 153

- 154 First, we applied unsupervised hierarchical clustering (**Methods**) to the non-BW trait association
- statistics for the 60 significant BW loci. The resultant heatmap indicates the heterogeneity of
- between-locus effect-size across the range of adult traits (Figure 3, Supplementary Table 17). For
- example, it shows that the associations between BW-raising alleles and increased adult height
- 158 concentrated amongst a subset of loci including *HHIP* and *GNA12*, and highlights particularly strong
- associations with lipid traits for variants at the *TRIB1* and *MAFB* loci.
- 160
- 161 Second, we constructed trait-specific "point-of-contact" (PoC) PPI networks from proteins
- represented in both the global BW PPI network and equivalent PPI networks generated for each of the adult traits (**Methods; Extended Data 8c-e**). We reasoned that these PoC PPI networks would be
- 164 enriched for the specific proteins mediating the observed links between BW and adult traits. To
- 165 highlight processes implicated in specific BW-trait associations, we overlaid these PoC PPI with the
- 166 50 pathways over-represented in the global BW PPI network. These analyses revealed, for example,
- 167 that proteins in the Wnt canonical signalling pathway were only detected in the PoC PPI network for
- 168 BP traits. We can use these PPI overlaps to highlight the specific transcripts within BW GWAS loci
- that are likely to mediate the mechanistic links. For example, the overlap between the Wnt signalling
- pathway and the PoC PPI network for the intersection of BW and BP-related traits implicates *FZD9* as
 the likely effector gene at the *MLXIPL* BW locus (Extended Data 8d, Supplementary Table 6).
- 172
- We focused our more detailed investigation of the mechanistic links between early growth and adulttraits on two phenotypic areas: arterial BP and T2D/glycaemia.
- 175

176 Across both the overall GWAS and specifically among the 60 significant BW loci, most BW-raising 177 alleles were associated with reduced BP (Figures 2, 3): the strongest inverse associations were seen for the loci near NT5C2, FES, NRIP1, EBF1 and PTH1R. However, we also observed locus-specific 178 heterogeneity in the genetic relationships between BP and BW: the SBP-raising allele at ADRB1¹⁸ is 179 associated with higher, rather than lower, BW (Extended Data 10a). When we considered the 180 reciprocal relationship, i.e. the effects on BW of BP-raising alleles at 30 reported loci for SBP^{18,19}, 181 182 there was an excess of associations (5/30 with lower BW at P<0.05; P=0.0026; Extended Data 10a). 183 To dissect maternal and foetal genotype effects at these loci, we tested the impact on BW of a risk score generated from 30 SBP SNPs, restricted to the untransmitted maternal haplotype score²⁰ in a 184 185 set of 5,201 mother-child pairs. Analysis of these loci indicated that maternal genotype effects on 186 intrauterine environment are likely to contribute to the inverse genetic correlation between SBP and 187 BW (Methods; Supplementary Table 18), and was consistent with the results of a wider study of 188 >30,000 women which demonstrated associations between a maternal genetic score for SBP (conditional on foetal genotype) and lower offspring BW¹⁰. 189

190

191 The BP-raising allele with the largest BW-lowering effect maps to the NT5C2 locus (index variant for 192 BW, rs74233809, r^2 =0.98 with index variant for BP, rs11191548¹⁹) and is also associated with lower adult BMI (r^2 =0.99 with rs11191560²¹). The BW-lowering allele at rs74233809 is a proxy for a 193 194 recently-described functional variant in the nearby CYP17A1 gene $(r^2=0.92 \text{ with rs}138009835)^{22}$. The 195 CYP17A1 gene encodes the cytochrome P450c17 α enzyme, CYP17²³, which catalyses key steps in 196 steroidogenesis that determine the balance between mineralocorticoid, glucocorticoid and 197 androgen synthesis. This variant is known to alter transcriptional efficiency in vitro and is associated with higher urinary tetrahydroaldosterone excretion²². CYP17A1 is expressed in foetal adrenal glands 198 and testes from early gestation²⁴ as well as in the placenta²⁵. These data implicate variation in 199 CYP17A1 expression as a contributor to the observational association between low BW and adult 200 hypertension²⁶. 201

202

When we examined 45 loci associated with CAD²⁷, the inverse genetic correlation between CAD and BW was concentrated amongst the five CAD loci with primary BP associations. This suggests that genetic determinants of BP play a leading role in mediating the lifecourse associations between BW
 and CAD (Extended Data 10b,e).

207

208 The LD score regression analyses demonstrated overall inverse genetic correlation between lower 209 BW and elevated risk of T2D (Figure 2). However, the locus specific heatmap indicates a 210 heterogeneous pattern across individual loci (Figure 3). To explore this further, we tested the 84 reported T2D loci²⁸ for association with BW. Some T2D risk alleles (such as at ADCY5, CDKAL1 and 211 HHEX-IDE) were strongly associated with lower BW, whilst others (e.g. ANK1 and MTNR1B) were 212 associated with higher (Extended Data 10c). This was in contrast with the BW effects of 422 known 213 214 height loci²⁹ (Extended Data 10d), which showed a strong positive correlation consistent with the 215 overall genetic correlation between height and BW, indicating that the growth effects of many height loci start prenatally and persist into adulthood. 216 217 218 The contrasting associations of T2D risk-alleles with both higher and lower BW are likely to reflect 219 the differential impacts across loci of variation in the maternal and foetal genomes. Observational data link paternal diabetes with lower offspring BW⁴ indicating that the inheritance of T2D risk 220 alleles by the foetus tends, in line with the LD score regression analysis, to reduce growth. These 221 relationships are consistent with the precepts of the "foetal insulin hypothesis"³⁰ and reflect the 222 223 potential for reduced insulin secretion and/or signalling to lead to both reduced foetal growth and, 224 many decades later, enhanced predisposition to T2D. In line with this, the paternal transmitted 225 haplotype score generated from the 84 T2D risk variants was associated with lower BW (P=0.045) in 226 5,201 mother-child pairs (Methods; Supplementary Table 18). In contrast, maternal diabetes is observationally associated with higher offspring BW⁴, reflecting the impact of maternal 227 228 hyperglycaemia to stimulate foetal insulin secretion. The contribution of genotype-dependent 229 maternal hyperglycaemia to BW is in line with the observation that some T2D risk alleles (most notably at MTNR1B) are associated with higher BW⁷, and with evidence, from a recent study, that a 230 maternal genetic score for T2D (conditional on foetal genotype) was associated with higher offspring 231 232 BW¹⁰. Thus, both maternal and foetal genetic effects connect BW to later T2D risk, albeit acting in 233 opposing directions. When we categorised T2D loci using a classification of physiological function 234 derived from their effects on related glycaemic and anthropometric traits³¹, we found that T2D-risk 235 alleles associated with lower BW were those typically characterised by reduced insulin processing 236 and secretion without detectable changes in fasting glucose (the "Beta Cell" cluster in Extended 237 Data 10f).

238

239 The YTK6 signal at rs138715366 is notable, not only because the genetic data indicates that a single 240 low-frequency non-coding variant is driving the association signal (see above) but because of the 241 proximity of this signal to GCK. Rare coding variants in glucokinase are causal for a form of 242 monogenic hyperglycaemia and lead to large reductions in BW when parental alleles are passed to 243 their offspring³². In addition, common non-coding variants nearby are implicated in T2D-risk and fasting hyperglycaemia^{33,34}. However, the latter variants are conditionally independent of 244 rs138715366 (Supplementary Table 19) and show no comparable association with lower BW. Either 245 246 rs138715366 acts through effector transcripts other than GCK, or the impact of the low-frequency 247 SNP near YKT6 on GCK expression involves tissue- and/or temporal-specific variation in regulatory 248 impact.

- In conclusion, we have identified 60 genetic loci associated with BW and used these to gain insights
 into the aetiology of foetal growth and into well-established, but until now poorly understood,
 lifecourse disease associations. The evidence that the relationship between early growth and later
- 253 metabolic disease has an appreciable genetic component contrasts with, but is not necessarily
- incompatible with, the emphasis on adverse early environmental events highlighted by the Foetal
- 255 Origins Hypothesis¹. As we have shown, these genetic effects reflect variation in both the foetal and

- the maternal genome: the impact of the latter on the offspring's predisposition to adult disease
- 257 could be mediated, at least in part, through perturbation of the antenatal and early life
- 258 environment. Future mechanistic and genetic studies should support reconciliation between these
- alternative, but complementary, explanations for the powerful lifecourse associations that exist
- 260 between events in early life and predisposition to cardiometabolic disease several decades later.
- 261
- 262
- 263

264	REF	ERENCES
265		
266 267	1.	(2004).
268	2.	The 1000 Genomes Project Consortium, Abecasis GR, Auton A, Brooks LD, DePristo MA, et al. An
269	2	integrated map of genetic variation from 1,092 human genomes. <i>Nature</i> 491 , 56-65 (2012).
270 271	3.	identifies rare variants in health and disease. <i>Nature</i> 526 , 82-90 (2015).
272	4.	Tyrrell JS, Yaghootkar H, Freathy RM, Hattersley AT, Frayling TM. Parental diabetes and
273		birthweight in 236 030 individuals in the UK biobank study. <i>Int J Epidemiol</i> 42 , 1714-1723
274	-	(2013). Harikashi M. Yashastkar II. Maak Karamari DO, Savia II. Taal IID, et s/ New lasi sessisted
275	5.	HORIKOSNI IVI, Yagnootkar H, Mook-Kanamori DU, Sovio U, Taai HR, et di. New loci associated
276		with birth weight identify genetic links between intrauterine growth and adult height and
277	c	metabolism. Nat Genet 45 , 76-82 (2013).
278	6.	Eaves LJ, Pourcain BS, Smith GD, York TP, Evans Divi. Resolving the effects of maternal and
279 280		Behav Genet 44 , 445-455 (2014).
281	7.	Feenstra B, Beaumont RN, Cavadino A, Tyrrell J, McMahon G, et al. Maternal genome-wide
282		association study identifies a fasting glucose variant associated with offspring birth weight. (in
283		preparation) doi: http://dx.doi.org/10.1101/034207
284	8.	Prokopenko I, Langenberg C, Florez JC, Saxena R, Soranzo N, et al. Variants in MTNR1B influence
285		fasting glucose levels. Nat Genet 41, 77-81 (2009).
286	9.	Kwak SH, Kim SH, Cho YM, Go MJ, Cho YS, et al. A genome-wide association study of gestational
287		diabetes mellitus in Korean women. <i>Diabetes</i> 61 , 531-541 (2012).
288	10.	Tyrrell J, Richmond RC, Palmer TM, Feenstra B, Rangarajan J, et al. Genetic evidence for causal
289		relationships between maternal obesity-related traits and birth weight. JAMA 315, 1129-1140
290		(2016).
291	11.	Landon MB, Spong CY, Thom E, Carpenter MW, Ramin SM, et al. A multicenter, randomized trial
292		of treatment for mild gestational diabetes. <i>N Engl J Med</i> 361 , 1339-1348 (2009).
293	12.	Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J, et al. LD Score regression distinguishes
294		confounding from polygenicity in genome-wide association studies. <i>Nat Genet</i> 47 , 291-295
295		(2015).
296	13.	Segrè AV, DIAGRAM Consortium, MAGIC investigators, Groop L, Mootha VK, et al. Common
297		inherited variation in mitochondrial genes is not enriched for associations with type 2 diabetes
298		or related glycemic traits. <i>PLoS Genet</i> 6 , e1001058 (2010).
299	14.	Lage K, Karlberg EO, Størling ZM, Olason PI, Pedersen AG, <i>et al</i> . A numan phenome-interactome
300		(2007)
301	4 5	(2007).
302	15.	imprinting agrees diverse adult human tissues. Canoma Res 25 , 027,026 (2015)
303	10	Imprinting across diverse adult numan tissues. Genome Res 25 , 927-936 (2015).
304 205	10.	Amorican Naturalist 124 , 147, 155 (1080)
202	17	American Naturalist 134 , 147-155 (1969).
207	17.	Genet 15, 517, 520 (2014)
202	10	Johnson T. Gount TP. Newhouse SI. Padmanabhan S. Tomaszowski M. <i>et al.</i> Blood pressure loci
309	10.	identified with a gene-centric array Am I Hum Genet 89 688-700 (2011)
310	1٩	International Consortium for Blood Pressure Genome-Wide Association Studies Ebret GR
311	т <i>э</i> .	Munroe PB Rice KM Bochud M Johnson AD et al Genetic variants in novel nathways influence
317		blood pressure and cardiovascular disease risk Nature 478 103-100 (2011)
312		sion pressure and cardiovascular discuse risk. Mature 476 , 105 105 (2011).

- 20. Zhang G, Bacelis J, Lengyel C, Teramo K, Hallman M, *et al.* Assessing the Causal Relationship of
 Maternal Height on Birth Size and Gestational Age at Birth: A Mendelian Randomization
 Analysis. *PLoS Med* 12, e1001865 (2015).
- 21. Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, *et al.* Genetic studies of body mass index yield
 new insights for obesity biology. *Nature* 518, 197-206 (2015).
- Diver LA, MacKenzie SM, Fraser R, McManus F, Freel EM, *et al.* Common Polymorphisms at the
 CYP17A1 locus associate with steroid phenotype: support for blood pressure GWAS signals at
 this locus. *Hypertension* 67, 724-732 (2016).
- Picado-Leonard J, Miller WL. Cloning and sequence of the human gene for P450c17 (steroid 17
 alpha-hydroxylase/17,20 lyase): similarity with the gene for P450c21. DNA 6, 439-448 (1987).
- Pezzi V, Mathis JM, Rainey WE, Carr BR. Profiling transcript levels for steroidogenic enzymes in
 fetal tissues. *J Steroid Biochem Mol Biol* 87, 181-189 (2003).
- 25. Escobar JC, Patel SS, Beshay VE, Suzuki T, Carr BR. The human placenta expresses CYP17 and
 generates androgens *de novo. J Clin Endocrinol Metab* **96**, 1385-1392 (2011).
- Reynolds RM, Walker BR, Phillips DI, Dennison EM, Fraser R, *et al.* Programming of
 hypertension: associations of plasma aldosterone in adult men and women with birth weight,
 cortisol, and blood pressure. *Hypertension* 53, 932-936 (2009).
- 27. CARDIoGRAMplusC4D Consortium, Deloukas P, Kanoni S, Willenborg C, Farrall M, *et al.* Large scale association analysis identifies new risk loci for coronary artery disease. *Nat Genet* 45, 25 33 (2013).
- 28. DIAbetes Genetics Replication And Meta-analysis (DIAGRAM) Consortium; Asian Genetic
 Epidemiology Network Type 2 Diabetes (AGEN-T2D) Consortium; South Asian Type 2 Diabetes
 (SAT2D) Consortium; Mexican American Type 2 Diabetes (MAT2D) Consortium; Type 2 Diabetes
 Genetic Exploration by Nex-generation sequencing in muylti-Ethnic Samples (T2D-GENES)
 Consortium, Mahajan A, *et al.* Genome-wide trans-ancestry meta-analysis provides insight into
 the genetic architecture of type 2 diabetes susceptibility. *Nat Genet* 46, 234-244 (2014).
- Wood AR, Esko T, Yang J, Vedantam S, Pers TH, *et al*. Defining the role of common variation in
 the genomic and biological architecture of adult human height. *Nat Genet* 46, 1173-1186
 (2014).
- 30. Hattersley AT, Tooke JE. The fetal insulin hypothesis: an alternative explanation of the
 association of low birthweight with diabetes and vascular disease. *Lancet* 353, 1789-1792
 (1999).
- 345 31. Dimas AS, Lagou V, Barker A, Knowles JW, Mägi R, *et al.* Impact of type 2 diabetes susceptibility
 variants on quantitative glycemic traits reveals mechanistic heterogeneity. *Diabetes* 63, 21582171 (2014).
- 348 32. Hattersley AT, Beards F, Ballantyne E, Appleton M, Harvey R, *et al.* Mutations in the glucokinase
 349 gene of the fetus result in reduced birth weight. *Nat Genet* **19**, 268-270 (1998).
- 33. Morris AP, Voight BF, Teslovich TM, Ferreira T, Segrè AV, *et al.* Large-scale association analysis
 provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet* 44, 981-990 (2012).
- 34. Dupuis J, Langenberg C, Prokopenko I, Saxena R, Soranzo N, *et al*. New genetic loci implicated in
 fasting glucose homeostasis and their impact on type 2 diabetes risk. *Nat Genet* 42, 105-116
 (2010).
- 356
- 357

- 358 METHODS
- 359

Ethics statement. All human research was approved by the relevant institutional review boards and
 conducted according to the Declaration of Helsinki. All participants provided written informed
 consent. Ethical approval for the study was obtained from the ALSPAC Ethics and Law Committee
 and the Local Research Ethics Committees.

364

Study-level analyses. Within each study, BW was collected from a variety of sources, including measurements at birth by medical practitioners, obstetric records, medical registers, interviews with the mother and self-report as adults (Supplementary Table 1). BW was z-score transformed, separately in males and females. Individuals with extreme BW (>5 standard deviations (SD) from the sex-specific study mean), monozygotic or polyzygotic siblings, or preterm births (gestational age <37 weeks, where this information was available) were excluded from downstream association analyses (Supplementary Table 1).

372 Within each study, stringent quality control of the GWAS genotype scaffold was undertaken, prior to imputation (Supplementary Table 2). Each scaffold was then pre-phased and imputed^{35,36} 373 up to reference panels from the 1000 Genomes Project Consortium² or 1000G and UK10K Project 374 Consortium³ (Supplementary Table 2). Association of BW with each variant passing established 375 376 GWAS quality control filters³⁷ was tested in a linear regression framework, under an additive model 377 for the allelic effect, after adjustment for study-specific covariates, including gestational age, where 378 available (Supplementary Table 2). Where necessary, population structure was accounted for by adjustment for axes of genetic variation from principal components analysis³⁸ and subsequent 379 genomic control correction³⁹, or inclusion of a genetic relationship matrix in a mixed model⁴⁰ 380 381 (Supplementary Table 2). We calculated the genomic control inflation factor (λ) in each study to 382 confirm that study-level population structure was accounted for prior to meta-analysis. 383

384 Preparation, quality control and genetic analysis in UK Biobank samples. UK Biobank phenotype data were available for 502,655 participants⁴¹. All participants in the UK Biobank were asked to recall 385 386 their BW, of which 279,971 did so at either the baseline or follow-up assessment visit. Of these, 387 7,686 participants reported being part of multiple births and were excluded from downstream 388 analyses. Ancestry checks, based on self-reported ancestry, resulted in the exclusion of 8,998 389 additional participants reported not to be white European. Of those individuals reporting BW at 390 baseline and follow-up assessments, 393 were excluded because the two reported values differed by 391 more than 0.5 kg. For those reporting different values (≤0.5 kg) between baseline and follow-up, we 392 took the baseline measure forward for downstream analyses. We then excluded 36,716 individuals 393 reporting values <2.5 kg or >4.5 kg as implausible for live term births before 1970. In total 226,178 394 participants had data relating to BW that matched these inclusion criteria.

395 Genotype data from the May 2015 release were available for a subset of 152,249 396 participants from UK Biobank. In addition to the quality control metrics performed centrally by UK 397 Biobank, we defined a subset of "white European" ancestry samples using a K-means (K=4) 398 clustering approach based on the first four genetically determined principal components. A 399 maximum of 67,786 individuals (40,425 females and 27,361 males) with genotype and valid BW 400 measures were available for downstream analyses. We tested for association with BW, assuming an 401 additive allelic effect, in a linear mixed model implemented in BOLT-LMM⁴² to account for cryptic 402 population structure and relatedness. Genotyping array was included as a binary covariate in all 403 models. Total chip heritability (i.e. the variance explained by all autosomal polymorphic genotyped SNPs passing quality control) was calculated using Restricted Maximum Likelihood (REML) 404 implemented in BOLT-LMM⁴². We additionally analysed the association between BW and directly 405 406 genotyped SNPs on the X chromosome: for this analysis, we used 57,715 unrelated individuals with 407 BW available and identified by UK Biobank as white British. We excluded SNPs with evidence of 408 deviation from Hardy-Weinberg Equilibrium ($P < 1 \times 10^{-6}$), MAF<0.01 or overall missing rate >0.015,

resulting in 19,423 SNPs for analysis in Plink v1.07 (http://pngu.mgh.harvard.edu/purcell/plink/)⁴³,
 with the first five ancestry principal components as covariates.

In both the full UK Biobank sample and our refined sample, we observed that BW was
 associated with sex, year of birth and maternal smoking (*P*<0.0015, all in the expected directions),
 confirming more comprehensive previous validation of self-reported BW⁴. We additionally verified
 that BW associations with lead SNPs at seven established loci⁵ based on self-report in UK Biobank
 were consistent with those previously published.

416

417 European ancestry meta-analysis. The European ancestry meta-analysis consisted of two

418 components: (i) 75,891 individuals from 30 GWAS from Europe, USA and Australia; and (ii) 67,786

419 individuals of white European origin from UK Biobank. In the first component, we combined sex-

420 specific BW association summary statistics across studies in a fixed-effects meta-analysis,

421 implemented in GWAMA⁴⁴ and applied a second round of genomic control³⁹ (λ_{GC} = 1.001).

422 Subsequently, we combined association summary statistics from this component with UK Biobank in 423 a European ancestry fixed-effects meta-analysis, implemented in GWAMA⁴⁴. Variants failing GWAS

424 quality control filters in UK Biobank, reported in less than 50% of the total sample size in the first

425 component, or with MAF<0.1%, were excluded from the European ancestry meta-analysis. We meta-

426 analysed the X-Chromosome UK Biobank results (19,423 SNPs) with corresponding results from the

- European ancestry meta-analysis using fixed effects *P*-value based meta-analysis in METAL⁴⁵ (max
 N=99,152).
- 429 We were concerned that self-reported BW as adults in UK Biobank would not be comparable 430 with that obtained from more stringent collection methods used in other European ancestry GWAS. 431 In addition, UK BioBank lacked information on gestational age for adjustment, which could have an 432 impact on difference in strength of association compared to the results obtained from other 433 European ancestry GWAS. However, we observed no evidence of heterogeneity in BW allelic effects 434 at lead SNPs between the two components of European ancestry meta-analysis, using Cochran's Q statistic⁴⁶, implemented in GWAMA⁴⁴, after Bonferroni correction (*P*>0.00083) (**Supplementary** 435 Table 3). We tested for heterogeneity in allelic effects between studies within the European 436 437 component using Cochran's Q. At loci demonstrating evidence of heterogeneity, we confirmed that 438 association signals were not being driven by outlying studies by visual inspection of forest plots. We 439 performed sensitivity analyses to assess the impact of covariate adjustment (gestational age and
- 440 population structure) on heterogeneity.
- 441 We were also concerned that overlap of individuals (duplicated or related) between the two 442 components of the European ancestry meta-analysis might lead to false positive association signals. We performed bivariate LD Score regression¹² using the two components of the European ancestry 443 444 meta-analysis and observed a genetic covariance intercept of 0.0156 (SE 0.0058), indicating a 445 maximum of 1,119 duplicate individuals. Univariate LD Score regression¹² of the European ancestry 446 meta-analysis estimated the intercept as 1.0426, which may indicate population structure or 447 relatedness that is not adequately accounted for in the analysis. To assess the impact of this inflation 448 on the European ancestry meta-analysis, we expanded the standard errors of BW allelic effect size 449 estimates and re-calculated association P-values. On the basis of this adjusted analysis, the lead SNP 450 only at MTNR1B dropped below genome-wide significance (rs10830963, P=5.5x10⁻⁸).
- 451

452 Trans-ancestry meta-analysis. The trans-ancestry meta-analysis combined the two European 453 ancestry components with an additional 10,104 individuals from six GWAS from diverse ancestry 454 groups: African American, Chinese, Filipino, Surinamese, Turkish and Moroccan. Within each GWAS, we first combined sex-specific BW association summary statistics in a fixed-effects meta-analysis, 455 implemented in GWAMA⁴⁴ and applied a second round of genomic control³⁹. Subsequently, we 456 combined association summary statistics from the six non-European GWAS and the two European 457 458 ancestry components in a trans-ancestry fixed-effects meta-analysis, implemented in GWAMA⁴⁴. 459 Variants failing GWAS quality control filters in UK Biobank, reported in less than 50% of the total

sample size in the first component, or with MAF <0.1%, were excluded from the trans-ancestry meta-analysis. We tested for heterogeneity in allelic effects between ancestries using Cochran's Q^{46} .

462

463 **Prioritising candidate genes in each BW locus.** We combined a number of approaches to prioritise 464 the most likely candidate gene(s) in each BW locus. Expression quantitative trait loci (eQTLs) were obtained from the Genotype Tissue Expression (GTEx) Project⁴⁷, the GEUVADIS Project⁴⁸ and eleven 465 other studies⁴⁹⁻⁵⁹ using HaploReg v4⁶⁰. We interrogated coding variants for each BW lead SNP and its 466 proxies (EUR r^2 >0.8) using Ensembl⁶¹ and HaploReg. Their likely functional consequences were 467 predicted by SIFT⁶² and PolyPhen2⁶³. Biological candidacy was assessed by presence in significantly 468 enriched gene set pathways from MAGENTA analyses (see below for details). We extracted all genes 469 470 within 300 kb of all lead BW SNPs and searched for connectivity between any genes using STRING⁶⁴. 471 If two or more genes between two separate BW loci were connected, they were given an increased 472 prior for both being plausible candidates. We also applied protein-protein interaction (PPI) analysis 473 (see below for details) to all genes within 300 kb of each lead BW SNPs and ranked the genes based 474 on the score for connectivity with the surrounding genes.

475

476 Approximate conditional analysis. We searched for multiple distinct BW association signals in each 477 of the established and novel loci, defined as 1Mb up- and down-stream of the lead SNP from the 478 trans-ancestry meta-analysis, through approximate conditional analysis. We applied GCTA⁶⁵ to 479 identify "index SNPs" for distinct association signals attaining genome-wide significance (P<5x10⁻⁸) in 480 the European ancestry meta-analysis using a reference sample of 5,000 individuals of white British 481 origin, randomly selected from UK Biobank, to approximate patterns of linkage disequilibrium (LD) 482 between variants in these regions. Note that we performed approximate conditioning on the basis of 483 only the European ancestry meta-analysis because GCTA cannot accommodate LD variation between 484 diverse populations.

485

486 **Fine-mapping analyses.** We sought to leverage LD differences between populations contributing to 487 the trans-ancestry meta-analysis and to take advantage of the improved coverage of common and 488 low-frequency variation offered by 1000G or 1000G and UK10K combined imputation to localise 489 variants driving each distinct association signal achieving locus-wide significance. For each distinct signal, we used MANTRA⁶⁶ to construct 99% credible sets of variants⁶⁷ that together account for 99% 490 491 of the posterior probability of driving the association. MANTRA incorporates a prior model of 492 relatedness between studies, based on mean pair-wise allele frequency differences across loci, to 493 account for heterogeneity in allelic effects (Supplementary Table 3). MANTRA has been 494 demonstrated, by simulation, to improve localisation of causal variants compared with either a fixed- or random-effects trans-ancestry meta-analysis^{66,68}. 495

For loci with only one signal of association, we used MANTRA to combine summary statistics from the six non-European GWAS and the two European ancestry components. However, for loci with multiple distinct association signals, we used MANTRA to combine summary statistics from approximate conditioning for the two European components, separately for each signal.

500 For each distinct signal, we calculated the posterior probability that the *j*th variant, π_{Cj} , is 501 driving the association, given by 502

503

 $\pi_{\mathsf{C}j} = \frac{\Lambda_j}{\sum_k \Lambda_k},$

504 505 where the summation is over all variants mapping within the (conditional) meta-analysis across the 506 locus. In this expression, Λ_j is the Bayes' factor (BF) in favour of association from the MANTRA 507 analysis. A 99% credible set⁶⁷ was then constructed by: (i) ranking all variants according to their BF, 508 Λ_j ; and (ii) including ranked variants until their cumulative posterior probability exceeds 0.99. 509

510 Genomic annotation. We used genomic annotations of DNasel hypersensitive sites (DHS) from the ENCODE⁶⁹ project and protein coding genes from GENCODE⁷⁰. We filtered cell types that are cancer 511 cell lines (karyotype 'cancer' from https://genome.ucsc.edu/ENCODE/cellTypes.html), and merged 512 513 data from multiple samples from the same cell type. This resulted in 128 DHS cell-type annotations, 514 as well as 4 gene-based annotations (coding exon, 5UTR, 3UTR and 1kb upstream of TSS). First, we 515 tested for the effect of each cell type DHS and gene annotation individually using the Bayes Factors for all variants in the 62 credible sets using fgwas⁷¹. Second, we categorised the annotations into 516 'genic', 'foetal DHS', 'embryonic DHS', 'stem cell DHS', 'neonatal DHS' and 'adult DHS' based on the 517 518 description fields from ENCODE, and tested for the effect of each category individually as described 519 above using fgwas. Third, we then tested the effect of each category by including all categories in a 520 joint model using fgwas. For each of the three analyses, we obtained the estimated effects and 95% 521 confidence intervals (CI) for each annotation, and considered an annotation enriched if the 95% CI 522 did not overlap zero.

523

Evaluation of imputation quality of low-frequency variant at the YKT6-GCK locus. At the YKT6-GCK
 locus, the lead SNP (rs138715366) is of low-frequency in European ancestry populations
 (MAF=0.92%) and even rarer in other ancestry groups (MAF=0.23% in African Americans, otherwise
 monomorphic) and is not present in the HapMap reference panel⁷². To assess the accuracy of
 imputation for this low-frequent variant, we genotyped rs138715366 in the Northern Finland Birth
 Cohort (NFBC) 1966 (Supplementary Table 1). Of the 5,009 samples in the study, 4,704 were
 successfully imputed and genotyped (or sequenced) for rs138715366. The overall concordance rate

between imputed and directly assayed genotypes was 99.8% and for directly assayed heterozygotecalls was 75.0%.

533

546

Estimation of genetic variance explained. Variance explained was calculated using the REML
 method implemented in GCTA⁷³. We considered the variance explained by two sets of SNPs: (i) lead

536 SNPs of all 62 distinct association signals at the 59 established and novel autosomal BW loci

537 identified in the European-specific or trans-ancestry meta-analyses; (ii) lead SNPs of 55 distinct

association signals at the 52 novel autosomal BW loci (Extended Data 2 and Supplementary Table

539 7). Variance explained was calculated in samples of European ancestry in the Hyperglycemia and

Adverse Pregnancy Outcome (HAPO) study⁷⁴ (independent of the meta-analysis) and two studies that were part of the European ancestry meta-analysis: NFBC1966 and Generation R

542 (**Supplementary Table 1**). In each study, the genetic relationship matrix was estimated for each set 543 of SNPs and was tested individually against BW (males and females combined) with study specific 544 covariates. These analyses provided an estimate and standard error for the variance explained by 545 each of the given sets of SNPs.

547 Examining the relative effects on BW of maternal and foetal genotype at the 60 identified loci. We 548 performed four sets of analyses:

- (i) We used GWAS data from 4,382 mother-child pairs in the Avon Longitudinal Study of
 Parents and Children (ALSPAC) study to fit a "maternal-GCTA model"⁶ to estimate the
 extent to which the maternal genome might influence offspring BW independent of the
 foetal genome. The m-GCTA model uses genome-wide genetic similarity between
 mothers and offspring to partition the phenotypic variance in BW into components due
 to the maternal genotype, the child's genotype, the covariance between the two and
 environmental sources of variation;
- We compared associations with BW of the foetal versus maternal genotype at each of
 the 60 BW loci. The maternal allelic effect on offspring BW was obtained from a
 maternal GWAS meta-analysis of 68,254 European mothers from the EGG Consortium
 (n=19,626)⁷ and the UK Biobank (n=48,628). In the UK Biobank, mothers were asked to
 report the BW of their first child. Women of European ancestry with genotype data

- 568 (iii) We additionally conducted analyses in 12,909 mother-child pairs from nine contributing
 569 studies: at each of the 60 loci, we compared the effect of the foetal genotype on BW
 570 adjusted for sex and gestational age, with and without adjustment for maternal
 571 genotype. We reciprocally compared the association between the maternal genotype
 572 and BW with and without adjustment for foetal genotype;
- (iv) We used the method of Zhang et al²⁰ to test associations between BW and the maternal untransmitted, maternal transmitted and paternal transmitted haplotype score of 422 height SNPs²⁹, 30 SBP SNPs^{18,19} and 84 T2D SNPs²⁸ in 5,201 mother-child pairs from the ALSPAC study.

578 LD Score Regression. The use of LD Score regression to estimate the genetic correlation between two traits/diseases has been described in detail elsewhere⁷⁵. Briefly, "LD Score" is a measure of how 579 much genetic variation each variant tags; if a variant has a high LD Score then it is in high LD with 580 581 many nearby polymorphisms. Variants with high LD Scores are more likely to contain more true 582 signals and hence provide more chance of overlap with genuine signals between GWAS. The LD 583 score regression method uses summary statistics from the GWAS meta-analysis of BW and the other traits of interest, calculates the cross-product of test statistics at each SNP, and then regresses the 584 cross-product on the LD Score. Bulik-Sullivan et al⁷⁵ show that the slope of the regression is a 585 586 function of the genetic covariance between traits:

587

577

$$E(z_{1j}z_{2j}) = \frac{\sqrt{N_1N_2}\rho_g}{M}l_j + \frac{\rho N_s}{\sqrt{N_1N_2}}$$

588 where N_i is the sample size for study *i*, ρ_a is the genetic covariance, *M* is the number of SNPs in the 589 reference panel with MAF between 5% and 50%, I_i is the LD score for SNP *j*, N_s quantifies the number of individuals that overlap both studies, and ρ is the phenotypic correlation amongst the N_s 590 overlapping samples. Thus, if there is sample overlap (or cryptic relatedness between samples), it will only affect the intercept from the regression (i.e. the term $\frac{\rho N_s}{\sqrt{N_1 N_2}}$) and not the slope, and hence 591 592 593 estimates of the genetic covariance will not be biased by sample overlap. Likewise, population 594 stratification will affect the intercept but will have minimal impact on the slope (i.e. intuitively since 595 population stratification does not correlate with linkage disequilibrium between nearby markers). 596 Summary statistics from the GWAS meta-analysis for traits and diseases of interest were 597 downloaded from the relevant consortium website. The summary statistics files were reformatted for LD Score regression analysis using the munge sumstats.py python script provided on the 598 599 developer's website (https://github.com/bulik/ldsc). For each trait, we filtered the summary statistics to the subset of HapMap 3 SNPs⁷⁶, as advised by the developers, to ensure that no bias was 600 601 introduced due to poor imputation quality. Summary statistics from the European-specific BW meta-602 analysis were used because of the variable LD structure between ancestry groups. Where the sample 603 size for each SNP was included in the results file this was flagged using --N-col; if no sample size was 604 available then the maximum sample size reported in the reference for the GWAS meta-analysis was 605 used. SNPs were excluded for the following reasons: MAF<0.01; ambiguous strand; duplicate rsID; 606 non-autosomal SNPs; reported sample size less than 60% of the total available. Once all files were

reformatted, we used the ldsc.py python script, also on the developers' website, to calculate the
 genetic correlation between BW and each of the traits and diseases. The European LD Score files

that were calculated from the 1000G reference panel and provided by the developers were used for

- 610 the analysis. Where multiple GWAS meta-analyses had been conducted on the same phenotype (i.e.
- over a period of years), the genetic correlation with BW was estimated using each set of summary
 statistics and presented in **Supplementary Table 12**. The phenotypes with multiple GWAS included
- 613 height, BMI, waist-hip ratio (adjusted for BMI), total cholesterol, triglycerides, HDL and LDL. The
- 614 estimate of the genetic correlation between the multiple GWAS meta-analyses on the same
- 615 phenotype were comparable and the later GWAS had a smaller standard error due to the increased
- sample size, so only the genetic correlation between BW and the most recent meta-analyses were
- 617 presented in **Figure 2**.

In the published GWAS for BP¹⁹ the phenotype was adjusted for BMI. Caution is needed 618 when interpreting the genetic correlation between BW and BMI-adjusted SBP due to the potential 619 620 for collider bias⁷⁷. Since BMI is associated with both BP and BW, it is possible that the use of a BP 621 genetic score adjusted for BMI might bias the genetic correlation estimate towards a more negative 622 value. To verify that the inverse genetic correlation with BW (r_e =-0.26, SE=0.05, P=6.5x10⁻⁹) was not 623 due to collider bias caused by the BMI adjustment of the phenotype, we obtained an alternative 624 estimate using UK Biobank GWAS data for SBP that was unadjusted for BMI and obtained a similar result (r_{e} =-0.22, SE=0.03, P=5.5x10⁻¹³). The SBP phenotype in UK Biobank was prepared as follows. 625 Two BP readings were taken at assessment, approximately 5 minutes apart. We included all 626 627 individuals with an automated BP reading (taken using an automated Omron BP monitor). Two valid 628 measurements were available for most participants (averaged to create a BP variable, or 629 alternatively a single reading was used if only one was available). Individuals were excluded if the 630 two readings differed by more than 4.56 SD. BP measurements more than 4.56 SD away from the mean were excluded. We accounted for BP medication use by adding 15 mmHg to the SBP measure. 631 632 BP was adjusted for age, sex and centre location and then inverse rank normalised. We performed the GWAS on 127,698 individuals of British descent using BOLT-LMM⁴², with genotyping array as 633 634 covariate.

635

636 Estimating the proportion of the BW-adult traits covariance attributable to genotyped SNPs. We 637 estimated the phenotypic, genetic and residual correlations as well as the genetic and residual covariance between BW and several quantitative traits/disease outcomes in UK Biobank using 638 directly genotyped SNPs and the REML method implemented in BOLT-LMM⁴². The traits examined 639 640 included T2D, SBP, DBP, CAD, height, BMI, weight, waist-hip ratio, hip circumference, waist 641 circumference, obesity, overweight, age at menarche, asthma, and smoking. Where phenotypes 642 were not available (e.g. serum blood measures are not currently available in UK Biobank), we 643 obtained estimates using the NFBC1966 study (for correlations/covariance between BW and 644 triglycerides, Total-, HDL- and LDL-cholesterol, fasting glucose and fasting insulin). In the UK Biobank 645 analysis, we used 57,715 unrelated individuals with BW available and identified by UK Biobank as 646 white British. SNPs with evidence of deviation from Hardy-Weinberg Equilibrium ($P < 1 \times 10^{-6}$), 647 MAF<0.05 or overall missing rate >0.015 were excluded, resulting in 328,928 SNPs for analysis. We 648 included the first five ancestry principal components as covariates. In the NFBC1966 analysis, 5,009 649 individuals with BW were enrolled. Genotyped SNPs that passed quality control (Supplementary 650 Table 2) were included, resulting in 324,895 SNPs for analysis. The first three ancestry principal 651 components and sex were included as covariates.

652

Gene set enrichment analysis. Meta-Analysis Gene-set Enrichment of variaNT Associations
 (MAGENTA) was used to explore pathway-based associations using summary statistics from the
 trans-ancestry meta-analysis. MAGENTA implements a gene set enrichment analysis (GSEA) based
 approach, as previously described¹³. Briefly, each gene in the genome is mapped to a single index

657 SNP with the lowest *P*-value within a 110 kb upstream and 40 kb downstream window. This *P*-value,

658 representing a gene score, is then corrected for confounding factors such as gene size, SNP density 659 and LD-related properties in a regression model. Genes within the HLA-region were excluded from analysis due to difficulties in accounting for gene density and LD patterns. Each mapped gene in the 660 661 genome is then ranked by its adjusted gene score. At a given significance threshold (95th and 75th percentiles of all gene scores), the observed number of gene scores in a given pathway, with a 662 663 ranked score above the specified threshold percentile, is calculated. This observed statistic is then 664 compared to 1,000,000 randomly permuted pathways of identical size. This generates an empirical GSEA P-value for each pathway. Significance was attained when an individual pathway reached a 665 666 false discovery rate (FDR) < 0.05 in either analysis. In total, 3,216 pre-defined biological pathways 667 from Gene Ontology, PANTHER, KEGG and Ingenuity were tested for enrichment of multiple modest 668 associations with BW. The MAGENTA software was also used for enrichment testing of custom gene 669 sets.

670

671 **Protein-Protein interaction network analyses.** We used the integrative Protein-Interaction-

Network-Based Pathway Analysis (iPINBPA) method⁷⁸. Briefly, we generated gene-wise *P*-values
 from the trans-ancestry meta-analysis using VEGAS2⁷⁹, which map the SNPs to genes and account for

from the trans-ancestry meta-analysis using VEGAS2⁷⁹, which map the SNPs to genes and account for
 possible cofounders, such as LD between markers. The empirical gene-wise *P*-values are calculated

675 using simulations from the multivariate normal distribution. Those that were nominally significant

 $(P \le 0.01)$ were selected as "seed genes", and were collated within high confidence version of

677 inweb3¹⁴, to weight the nodes in the network following a guilt-by-association approach. In a second step, a network score was defined by the combination of the z-scores derived from the gene-wise P-678 values with node weights using the Liptak-Stouffer method⁸⁰. A heuristic algorithm was then applied 679 680 to extensively search for modules enriched in genes with low P-values. The modules were further 681 normalised using a null distribution of 10,000 random networks. Only those modules with z-score >5 682 were selected. Finally, the union of all modules constructed a BW-overall PPI network. Both the 683 proteins on the individual modules and on the overall BW-PPI were interrogated for enrichment in 684 Gene Ontology Terms (Biological Processes) using a Hypergeometric test. Terms were considered as 685 significant when adjusted P-value, following Benjamini-Hochberg procedure, was below 0.05.

- 686
 687 Point of contact analyses (PoC). The same methodology described above was applied to 16 different
 688 adult traits resulting in a number of enriched modules per trait. Different modules for each trait
 689 were combined in a single component and the intersection between these trait-specific components
 690 and the BW component was calculated. This intersection is defined as the PoC network. We used the
 691 resulting PoC networks in downstream analyses to interrogate which set of proteins connects BW
 692 variation and adult trait variation via pathways enriched in the overall BW analysis.
- 693

Parent-of-origin specific associations. We first searched for evidence of parent of origin effects in
 the UK Biobank samples by comparing variance between heterozygotes and homozygotes using
 Quicktest⁸¹. In this analysis, we used only unrelated individuals identified genetically as of white
 British origin (n=57,715). Principal components were generated using these individuals and the first
 five were used to adjust for population structure as covariates in the analysis, in addition to a binary
 indicator for genotyping array.

700 We also examined 4,908 mother-child pairs in ALSPAC and determined the parental origin of the alleles where possible⁸². Briefly, the method uses mother-child pairs to determine the parent of 701 702 origin of each allele. For example, if the mother/child genotypes are AA/Aa, the child's 703 maternal/paternal allele combination is A/a. For the situation where both mother and child are 704 heterozygous, the child's maternal/paternal alleles cannot be directly specified. However, the 705 parental origin of the alleles can be determined by phasing the genotype data and comparing 706 maternal and child haplotypes. We then tested these alleles for association with BW adjusting for 707 sex and gestational age.

Statistical power in these currently available sample sizes is insufficient to rule out
widespread parent-of-origin effects across the regions tested. Using the mean beta (0.034SD) and
MAF (0.28) of the identified loci, we estimate that we would need at least 200,000 unrelated
individuals or 70,000 mother-child pairs for 80% power to detect parent-of-origin effects at
P<0.00085.

713

714 Hierarchical clustering of BW loci. To explore the different patterns of association between BW and 715 other anthropometric/metabolic/endocrine traits and diseases, we performed hierarchical clustering analysis. The lead SNP (or proxy, EUR r^2 >0.6) at the 60 BW loci was queried in publicly available 716 GWAS meta-analysis datasets or in GWAS result obtained through collaboration⁸³. Results were 717 718 available for 53 of those loci and the extracted z-score (allelic effect/SE, Supplementary Table 17) 719 was aligned to the BW-raising allele. We performed 2D clustering by trait and by locus. We 720 computed the Euclidean distance amongst z-scores of the extracted traits/loci and performed 721 complete hierarchical clustering implemented in the pvclust package (http://www.sigmath.es.osaka-722 u.ac.jp/shimo-lab/prog/pvclust/) in R v3.2.0 (http://www.R-project.org/). Clustering uncertainty was 723 measured by multiscale bootstrap resampling estimated from 1,000 replicates. We used α =0.05 to 724 define distinct clusters and, based on the bootstrap analysis, calculated the Calinski index to identify 725 the number of well-supported clusters (cascadeKM function, Vegan package, http://CRAN.R-726 project.org/package=vegan). Clustering was visualised by constructing dendrograms and a heatmap. 727 Separately from the hierarchical clustering analysis, we queried the lead SNP at EPAS1 in a 728 GWAS of haematological traits⁸⁴ because variation at that locus has previously been implicated in BW and adaptation to hypoxia at high altitudes in Tibetans^{85,86} (**Supplementary Table 17**). 729 730 731 732 **ADDITIONAL REFERENCES FOR METHODS** 733 734 35. Marchini J, Howie B. Genotype imputation for genome-wide association studies. Nat Rev Genet 735 **11**, 499-511 (2010). 736 36. Howie B, Fuchsberger C, Stephens M, Marchini J, Abecasis GR. Fast and accurate genotype 737 imputation in genome-wide association studies through pre-phasing. Nat Genet 44, 955-959 738 (2012). 739 37. Winkler TW, Day FR, Croteau-Chonka DC, Wood AR, Locke AE, et al. Quality control and conduct 740 of genome-wide association meta-analyses. Nat Protoc 9, 1192-1212 (2014). 741 38. Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, et al. Principal components 742 analysis corrects for stratification in genome-wide association studies. Nat Genet 38, 904-909 743 (2006).744 39. Devlin B, Roeder K. Genomic control for association studies. *Biometrics* 55, 997-1004 (1999). 745 40. Kang HM, Sul JH, Service SK, Zaitlen NA, Kong SY, et al. Variance component model to account 746 for sample structure in genome-wide association studies. Nat Genet 42, 348-354 (2010). 747 41. Allen NE, Sudlow C, Peakman T, Collins R. UK Biobank. UK biobank data: come and get it. Sci 748 Transl Med 6, 224ed4 (2014). 749 42. Loh PR, Tucker G, Bulik-Sullivan BK, Vilhjálmsson BJ, Finucane HK, et al. Efficient Bayesian 750 mixed-model analysis increases association power in large cohorts. Nat Genet 47, 284-290 751 (2015). 752 43. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR et al. PLINK: a toolset for whole-753 genome association and population-based linkage analysis. Am J Hum Genet 81, 559-575 754 (2007).755 44. Mägi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. BMC 756 *Bioinformatics* **11**, 288 (2010).

45. Willer CJ, Li Y, and Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* **26**, 2190–2191 (2010). 759 46. Ioannidis J, Patsopoulos NA, Evangelou E. Heterogeneity in meta-analyses of genome-wide 760 association investigations. PLoS One 2, e0000841 (2007). 761 47. GTEx Consortium. Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis: 762 multitissue gene. Science 348, 648-660 (2015). 763 48. Lappalainen T, Sammeth M, Friedländer MR, 't Hoen PA, Monlong J, et al. Transcriptome and 764 genome sequencing uncovers functional variation in humans. Nature 501, 506-511 (2013). 765 49. Montgomery SB, Sammeth M, Gutierrez-Arcelus M, Lach RP, Ingle C, et al. Transcriptome 766 genetics using second generation sequencing in a Caucasian population. Nature 464, 773-777 767 (2010). 768 50. Schadt EE, Molony C, Chudin E, Hao K, Yang X, et al. Mapping the genetic architecture of gene 769 expression in human liver. PLoS Biol 6, e107 (2008). 770 51. Gibbs JR, van der Brug MP, Hernandez DG, Traynor BJ, Nalls MA, et al. Abundant quantitative 771 trait loci exist for DNA methylation and gene expression in human brain. PLoS Genet 6, 772 e1000952 (2010). 773 52. Stranger BE, Nica AC, Forrest MS, Dimas A, Bird CP, et al. Population genomics of human gene 774 expression. Nat Genet 9, 1217-1224 (2007). 775 53. Li Q, Stram A, Chen C, Kar S, Gayther S, et al. Expression QTL-based analyses reveal candidate 776 causal genes and loci across five tumor types. Hum Mol Genet 23, 5294-5302 (2014). 777 54. Westra HJ, Peters MJ, Esko T, Yaghootkar H, Schurmann C, et al. Systematic identification of 778 trans eQTLs as putative drivers of known disease associations. Nat Genet 45, 1238-1243 (2013). 779 55. Zou F, Chai HS, Younkin CS, Allen M, Crook J, et al. Brain expression genome-wide association 780 study (eGWAS) identifies human disease-associated variants. PLoS Genet 8, e1002707 (2012). 781 56. Hao K, Bossé Y, Nickle DC, Paré PD, Postma DS, et al. Lung eQTLs to help reveal the molecular 782 underpinnings of asthma. PLoS Genet 8, e1003029 (2012). 783 57. Koopmann TT, Adriaens ME, Moerland PD, Marsman RF, Westerveld ML, et al. Genome-wide 784 identification of expression quantitative trait loci (eQTLs) in human heart. PLoS One 9, e97380 785 (2014). 786 58. Fairfax BP, Humburg P, Makino S, Naranbhai V, Wong D, et al. Innate immune activity 787 conditions the effect of regulatory variants upon monocyte gene expression. Science 343, 788 1246949 (2014). 789 59. Grundberg E, Adoue V, Kwan T, Ge B, Duan QL, et al. Global analysis of the impact of 790 environmental perturbation on cis-regulation of gene expression. PLoS Genet 7, e1001279 791 (2011). 792 60. Ward LD and Kellis M. HaploReg: a resource for exploring chromatin states, conservation, and 793 regulatory motif alterations within sets of genetically linked variants. Nucleic Acids Res 40, 794 D930-934 (2012). 795 61. Flicek P, Amode MR, Barrell D, Beal K, Billis K, et al. Ensembl 2014. Nucleic Acids Res 42, D749-796 755 (2014). 797 62. Kumar P, Henikoff S, Ng P. Predicting the effects of coding non-synonymous variants on protein 798 function using the SIFT algorithm. Nat Protoc 4, 1073-1081 (2009). 799 63. Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, et al. A method and server for 800 predicting damaging missense mutations. Nat Methods 7, 248-249 (2010). 801 64. Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, et al. STRING v10: protein-protein 802 interaction networks, integrated over the tree of life. Nucleic Acids Res 43, D447-452 (2015). 803 65. Yang J, Ferreira T, Morris AP, Medland SE, Genetic Investigation of ANthropometric Traits 804 (GIANT) Consortium, et al. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. Nat Genet 44, 369-375 (2012). 805 806 66. Morris AP. Transethnic meta-analysis of genome-wide association studies. Genet Epidemiol 35, 807 809-822 (2011).

- 808 67. Wellcome Trust Case Control Consortium, Maller JB, McVean G, Byrnes J, Vukcevic D, *et al.*809 Bayesian refinement of association signals for 14 loci in 3 common diseases. *Nat Genet* 44,
 810 1294-1301 (2012).
- 68. Wang X, Chua HX, Chen P, Ong RT, Sim X, *et al.* Comparing methods for performing trans-ethnic
 meta-analysis of genome-wide association studies. *Hum Mol Genet* 22, 2303-2311 (2013).
- 813 69. ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human
 814 genome. *Nature* 489, 57-74 (2012).
- 815 70. Harrow J, Frankish A, Gonzalez JM, Tapanari E, Diekhans M, *et al.* GENCODE: the reference
 816 human genome annotation for The ENCODE Project. *Genome Res* 22, 1760-1774 (2012).
- 817 71. Pickrell JK. Joint analysis of functional genomic data and genome-wide association studies of 18
 818 human traits. *Am J Hum Genet* 94, 559–573 (2014).
- The International HapMap 3 Consortium. Integrating common and rare genetic variation in
 diverse human populations. *Nature* 467, 52-58 (2010).
- 73. Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, *et al.* Common SNPs explain a large
 proportion of the heritability for human height. *Nat Genet* 42, 565-569 (2010).
- Vrbanek M, Hayes MG, Armstrong LL, Morrison J, Lowe LP *et al*. The chromosome 3q25
 genomic region is associated with measures of adiposity in newborns in a multi-ethnic genomewide association study. *Hum Mol Genet* 22, 3583-3596 (2013).
- 826 75. Bulik-Sullivan B, Finucane HK, Anttila V, Gusev A, Day FR, *et al*. An atlas of genetic correlations
 827 across human diseases and traits. *Nat Genet* 47, 1236-1241 (2015).
- 76. The International HapMap Consortium. A second generation haplotype map of over 3.1 million
 SNPs. *Nature* 449, 851-861 (2007).
- Aschard H, Vilhjálmsson BJ, Joshi AD, Price AL, Kraft P. Adjusting for heritable covariates can
 bias effect estimates in genome-wide association studies. *Am J Hum Genet* **96**, 329-339 (2015).
- 832 78. Wang L, Mousavi P, Baranzini SE. iPINBPA: an integrative network-based functional module
 833 discovery tool for genome-wide association studies. *Pac Symp Biocomput* 255-266 (2015).
- Mishra A and Macgregor S. VEGAS2: Software for More Flexible Gene-Based Testing. *Twin Res Hum Genet* 18, 86-91 (2015).
- 836 80. Whitlock MC. Combining probability from independent tests: the weighted Z-method is superior
 837 to Fisher's approach. *J Evol Biol* 18, 1368-1373 (2005).
- 81. Hoggart, CJ, Venturini G, Mangino M, Gomez F, Ascari G, *et al.* Novel Approach Identifies SNPs
 in SLC2A10 and KCNK9 with Evidence for Parent-of-Origin Effect on Body Mass Index. *PLoS Genet* 10, 1–12 (2014).
- 82. Wang S, Yu Z, Miller RL, Tang D & Perera FP. Methods for detecting interactions between
 imprinted genes and environmental exposures using birth cohort designs with mother-offspring
 pairs. Hum Hered **71**, 196–208 (2011).
- 844 83. Painter JN, Anderson CA, Nyholt DR, Macgregor S, Lin J, *et al*. Genome-wide association study 845 identifies a locus at 7p15.2 associated with endometriosis. *Nat Genet* **43**, 51-54 (2011).
- 84. Ganesh SK, Zakai NA, van Rooij FJ, Soranzo N, Smith AV, Nalls MA, *et al.* Multiple loci influence
 847 erythrocyte phenotypes in the CHARGE Consortium. *Nat Genet* 41, 1191-1198 (2009).
- 848 85. Xu XH, Huang XW, Qun L, Li YN, Wang Y, *et al*. Two functional loci in the promoter of EPAS1
 849 gene involved in high-altitude adaptation of Tibetans. *Sci Rep* 4, 7465 (2014).
- 86. Huerta-Sánchez E, Jin X, Asan, Bianba Z, Peter BM, *et al.* Altitude adaptation in Tibetans caused
 by introgression of Denisovan-like DNA. *Nature* 512, 194-197 (2014).
- 87. Kato N, Loh M, Takeuchi F, Verweij N, Wang X, *et al*. Trans-ancestry genome-wide association
 study identifies 12 genetic loci influencing blood pressure and implicates a role for DNA
 methylation. *Nat Genet* 47, 1282-1293 (2015).
- 855
- 856
- 857

ENDNOTES

Supplementary Information is available in the online version of the paper.

Acknowledgements Full grant supports and acknowledgements can be found in the Supplementary Information.

Author Contributions

Core analyses and writing: M.H., R.N.B., F.R.D., N.M.W., M.N.K., J.F-T., N.R.v.Z., K.J.G., A.P.M., K.K.O., J.F.F., N.J.T., J.R.P., D.M.E., M.I.M., R.M.F. Statistical analysis in individual studies (lead analysts in italics): M.H., R.N.B., F.R.D., N.M.W., M.N.K., B.F., N.G., J.P.B., D.P.S., R.L-G., T.S.A., E.K-M., R.R., L-P.L., D.L.C., Y.W., E.T., C.A.W., C.T.H., J-J.H., N.V-T., P.K.J., E.T.H.B., I.N., N.P., A.M., E.M.v.L., R.J., V. Lagou, M.N., J.M.M., S.E.J., P-R.L., K.S.R., M.A.T., J.T., A.R.W., H.Y., D.M.S., I.P., K. Panoutsopoulou, X.W., L.C., F.G., K.E.S., M. Murcia, E.V.R.A., Z.K., S.B.-G., F.S., D.T., J.W., C.M-G., N.R.R., E.Z., G.V.D., Y-Y.T., H.N.K., A.P.M., J.F.F., N.J.T., J.R.P., D.M.E., R.M.F. GWAS look-up in unpublished datasets: K.T.Z., N.R., D.R.N., R.C.W.M., C.H.T.T., W.H.T., S.K.G., F.J.v.R. Sample collection and data generation in individual studies: F.R.D., M.N.K., B.F., N.G., J.P.B., D.P.S., R.L-G., R.R., L-P.L., J-J.H., I.N., E.M.v.L., M.B., P.M-V., A.J.B., L.P., P.K., M.A., S.M.W., F.G., C.E.v.B., G.W., E.V.R.A., C.E.F., C.T., C.M.T., M. Standl, Z.K., M.V.H., H.G.d.H., F.R.R., C.M-G., S.M.R., G.H., G.M., N.R.R., C.J.G., C.L., J.L., R.A.S., J.H.Z., F.D.M., W.L.L.Jr, A.T., M. Stumvoll, V. Lindi, T.A.L., C.M.v.D., A.K., T.I.S., H.N., K. Pahkala, O.T.R., E.Z., G.V.D., S-M.S., M. Melbye, H.C., J.F.W., M.V., J-C.H., T.H., L.J.B., J.P.N., C.E.P., L.S.A., J.B.B., K.L.M., J.G.E., E.E.W., M.K., J.S.V., T.L., P.V., K.B., H.B., D.O.M-K., F.R., A.G.U., C. Pisinger, O.P., N.J.W., H.H., V.W.J., S.F.G., A.A.V., D.A.L., G.D.S., K.K.O., J.F.F., N.J.T., J.R.P., M.I.M. Functional follow-up experiment: L.A.D., S.M.M., R.M.R., E.D., B.R.W. Individual study design and principal investigators: J.P.B., I.N., M.A., F.D.M., W.L.L.Jr, A.T., M. Stumvoll, V. Lindi, T.A.L., C.M.v.D., W.K., A.K., T.I.S., H.N., K. Pahkala, O.T.R., G.V.D., Y-Y.T., S-M.S., M. Melbye, H.C., J.F.W., M.V., E.J.d.G., D.I.B., H.N.K., J-C.H., T.H., A.T.H., L.J.B., J.P.N., C.E.P., J.H., L.S.A., J.B.B., K.L.M., J.G.E., E.E.W., M.K., J.S.V., T.L., P.V., K.B., H.B., D.O.M-K., A.H., F.R., A.G.U., C. Pisinger, O.P., C. Power, E.H., N.J.W., H.H., V.W.J., M-R.J., S.F.G., A.A.V., T.M.F., A.P.M., K.K.O., N.J.T., J.R.P., M.I.M., R.M.F.

Author Information

Correspondence and requests for materials should be addressed to mark.mccarthy@drl.ox.ac.uk and r.freathy@ex.ac.uk. Reprints and permissions information is available at www.nature.com/reprints.

Disclosures

Krina Zondervan has a scientific collaboration with Bayer HealthCare Ltd. and Population Diagnostics Inc.

Grants and funding supports for studies

The Academy of Finland [41071, 1114194, 117787, 120315, 121584, 124282, 126925, 129287, 129378, 134309, 206374, 218029, 24300796, 251360, 267561, 276861, 286284 and EGEA-project]; Althingi (the Icelandic Parliament); the American Diabetes Association; the Arthritis Research UK; the Augustinus Foundation; Baylor Medical College [N01- HC-55016]; the Becket Foundation; Biobanking and Biomolecular Resources Research Infrastructure (BBMRI-NL); Biocenter Oulu, University of Oulu, Finland; Biomedical Research Council, Singapore (BMRC 06/1/21/19/466); the British Heart Foundation [SP/13/2/30111]; the C.G. Sundell Foundation; Cambridge Institute for Medical Research (CIMR); the Canadian Institutes of Health Research [MOP-82893]; the Cancer Research UK [SP2024-0201 and SP2024-0204]; the Capital Region Research Foundation; Center for Medical Systems

Biology (CMSB); the Chief Scientist Office of the Scottish Government [CZB/4/276 and CZB/4/710]; the Children's Hospital of Philadelphia [Institute Development Award]; Chinese University of Hong Kong (CUHK) [Faculty of Medicine Outstanding Fellowship]; the city of Kuopio; Conselleria de Sanitat Generalitat Valenciana; the Cotswold Foundation [Research Development Award]; Daniel B. Burke Endowed Chair for Diabetes Research; Danish Centre for Health Technology Assessment; Danish Council for Independent Research; Danish Innovation Foundation [0603-00484B and 0603-00457B]; Danish Diabetes Association; Danish Heart Foundation; Danish Innovation Foundation; the Danish National Research Foundation; Danish Pharmaceutical Association; Danish Pharmacists' Fund; Danish Research Council; DHFD (Diabetes Hilfs- und Forschungsfonds Deutschland); Diabetes and Inflammation Laboratory; Diabetes Research Foundation of Finland; EFSD/Lilly research fellowship; the Egmont Foundation; Emil Aaltonen Foundation; Erasmus Medical Center, Rotterdam, the Erasmus University Rotterdam; European Commission [ENGAGE (HEALTH-F4-2007-201413), Framework Programme 5 (QLG2-CT-2002-01254), Framework Programme 6 (018996, 018947 (LSHG-CT-2006-01947) and LSHG-CT-2006-018947), Framework Programme 7 (FP7/2007-2013), H2020-633595 DynaHEALTH action, Beta-JUDO n° 279153 and DG XII]; European Research Council (ERC Advanced, 230374); European Science Foundation (ESF, EU/QLRT-2001-01254); European Union (European Social Fund - ESF); Faculty of Biology and Medicine of Lausanne; Finnish Cardiac Research Foundation; Finnish Cultural Foundation; Finnish Foundation of Cardiovascular Research, Finnish Innovation Fund Sitra; Finnish Ministry of Education and Culture; FIS-FEDER [03/1615, 04/1112, 04/1509, 04/1931, 05/1052, 05/1079, 06/1213, 07/0314, 09/02647, 11/00178, 11/01007, 11/02591, 13/02032, 13/1944, 14/00891, 14/01687, 97/0588, 00/0021-2, PI041436, PI061756, PI081151 and PS0901958], Foundation for Paediatric Research of Finland; French Ministry of Research; Fundació La Marató de TV3; Gene-diet Interactions in Obesity project (GENDINOB); Generalitat de Catalunya [CIRIT 1999SGR 00241], Genetic Association Information Network (GAIN); the Genetic Laboratory of the Department of Internal Medicine, Erasmus MC; German Diabetes Association; German Research Council [DFG - SFB 1052 "Obesity mechanisms"; A01, B01, B03, C01 and SPP 1629 TO 718/2-1]; German Research Foundation Collaborative Research Center [CRC1085]; GlaxoSmithKline; Greek national funds: Heracleitus II; Health Fund of the Danish Health Insurance Societies; Hjartavernd (the Icelandic Heart Association); the Ib Henriksen Foundation; the Impact of our Genomes on Individual Treatment Response in Obese Children (TARGET); the Indo-Danish bilateral project, Genetics and Systems Biology of Childhood Obesity in India and Denmark (BioChild); Instituto de Salud Carlos III [Red INMA G03/176 and CB06/02/0041]; Integrated Research and Treatment Center (IFB) Adiposity Diseases [01EO1001]; the Italian Ministry of Health [ICS 110.1RS97.71]; Johns Hopkins University [N01-HC-55020]; the Juho Vainio Foundation; the Juvenile Diabetes Research Foundation International (JDRF); Kuopio University Hospital [5031343]; Kuopio, Tampere and Turku University Hospital Medical Funds [X51001]; Leiden University Medical Center; Leiden University, Research Profile Area Vascular and Regenerative Medicine; the Lundbeck Foundation; the March of Dimes Birth Defects Foundation; Medical Research Council [G0000934, G0500539, G0600705, G0601261, G9502233, MC U106179471, PrevMetSyn and MRC Doctoral Training Grant Scholarship]; Ministry of Education and Culture of Finland; Ministry of Education, Culture and Science of the Netherlands; Ministry of Health of Denmark; Ministry of Health, Welfare and Sport of the Netherlands; Ministry of Internal Affairs and Health of Denmark; Ministry of Social Affairs and Health of Finland; Ministry of Youth and Families of the Netherlands; MRC Integrative Epidemiology Unit at the University of Bristol (MC_UU_12013/1-9); MRC Human Genetics Unit; Municipality of Rotterdam; National Center for Advancing Translational Sciences [CTSI grant UL1TR000124]; National Heart, Lung and Blood Institute (NHLBI) [5R01HL087679, STAMPEED program (1RL1MH083268-01), HHSN268201100005C, HHSN268201100006C, HHSN2682 01100007C, HHSN268201100008C, HHSN268201100009C, HHSN268201100010C, HHSN268201100011C and HHSN268 201100012C, HHSN268201200036C, HHSN268200800007C, N01HC55222, N01HC85079, N01HC85080, N01HC85081, N01HC85082, N01HC85083, N01HC8508, U01HL080295, R01HL087652, R01HL105756, R01HL103612, R01HL120393, N01-HC-25195 and N02-HL-6-4278]; National Health

and Medical Research Council of Australia [003209, 403981 and 572613]; National Human Genome Research Institute (NHGRI) [U01HG 004402]; National Institute of Aging (NIA) [Intramural Research Program, R01AG023629, N01-AG-916413, N01-AG-821336, 263 MD 9164 13 and 263 MD 821336]; NIA/NIH [AG000932-2]; National Institute of Allergy and Infectious Diseases (NIAID); National Institute of Child Health and Human Development (NICHD); National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) [DK063491]; National Institutes of Health (NIH) [1RC2MH089995-01; DK056350, DK078150, DK099820, ES10126, HD34242, HD34243, HG004415, HL085144, RR20649, R01D0042157-01A, R01DK092127-01; R01HD056465, TW05596, U01DK062418, U01HG004423, U01HG004438, U01HG004446, R01HL087641, R01HL59367, R01HL086694, UL1RR025005 and N01-AG-12100]; National Institute for Health Research Cambridge Biomedical Research Centre; National Institute of Mental Health (NIMH) [MH081802, U24 MH068457-06]; National Institute of Neurological Disorders and Stroke (NINDS); National Medical Research Foundation, Singapore (NMRC/0975/2005); NBIC/BioAssist/RK(2008.024); the Netherlands Consortium for Healthy Aging (NCHA); the Netherlands Genomics Initiative (NGI); the Netherlands Organisation for Scientific Research and the Russian Foundation for Basic Research [NWO-RFBR 047.017.043]; the Netherlands Organisation for Scientific Research (NWO) [NWO/ZonMw; NWO Genomics; NWO: MagW/ZonMW grants 400-05-717, 480-04-004, 481-08-011, 451- 04-034, 463-06-001, 904-61-090, 904-61-193, 912-10-020, 916-76-125, 985-10-002, Addiction-31160008 Middelgroot-911-09-032 and Spinozapremie 56-464-14192, 175.010.2005.011, 911-03-012]; Neuroscience Campus Amsterdam (NCA); Novo Nordisk Foundation; Novo Nordisk Inc.; the Paavo Nurmi Foundation; Paulo Foundation; the Region Zealand Health and Medical Research Foundation; Research Committee of the Kuopio University Hospital Catchment Area; Research Foundation of Copenhagen County; Research Grant Council General Research Fund [CU473408, CU471713]; Research Institute for Diseases in the Elderly (RIDE) [014-93-015]; Robert Dawson Evans Endowment; the Royal Society; Rutgers University Cell and DNA Repository; the Sigrid Juselius Foundation; Social Insurance Institution of Finland; Spanish Government [SEV-2011-00067]; Spanish Ministry of Science and Innovation [SAF2008-00357]; Spanish National Genotyping Centre (CEGEN-Barcelona); Special Governmental Grants for Health Sciences Research, Turku University Hospital; Swiss National Science Foundation [33CSCO-122661, 33CS30-139468 and 33CS30-148401]; Tampere Tuberculosis Foundation; Turku University Foundation; University of Bristol; University of Cambridge; University of Minnesota [N01-HC-55019]; University of Mississippi Medical Center [N01-HC- 55021]; University of North Carolina [N01-HC-55018]; University of North Carolina at Chapel Hill [N01-HC-55015]; University of Texas Houston [N01-HC-55017]; the VU University's Institute for Health and Care Research (EMGO+); Wellcome Trust [068545/Z/02, 076113/B/04/Z, 079895, 090532, 098017, 098051, 098381, 102215/2/13/2, GR069224]; Yrjö Jahnsson Foundation.

FIGURE LEGENDS

Figure 1 | Manhattan and quantile-quantile (QQ) plots of the trans-ancestry meta-analysis for birth weight. a, Manhattan (main panel) and QQ (top right) plots of genome-wide association results for birth weight from trans-ancestry meta-analysis of up to 153,781 individuals. The association Pvalue (on -log₁₀ scale) for each of up to 22,185,636 SNPs (y axis) is plotted against the genomic position (NCBI Build 37; x axis). Association signals that reached genome-wide significance ($P < 5 \times 10^{-8}$) are shown in green if novel and pink if previously reported. In the QQ plot, the black dots represent observed P-values and the grey line represents expected P-values under the null distribution. The red dots represent observed P-values after excluding the previously identified signals described in Extended Data 2. b, Manhattan (main panel) and QQ (top right) plots of trans-ethnic GWAS metaanalysis for BW highlighting the reported imprinted regions described in Supplementary Table 14. Novel association signals that reached genome-wide significance ($P < 5 \times 10^{-8}$) and mapped to imprinted regions are shown in green. Genomic regions outside imprinted regions are shaded in grey. SNPs in the imprinted regions are shown in light blue or dark blue, if they are on chromosome with odd number or even number, respectively. In the QQ plot, the black dots represent observed P values and the grey lines represent expected P-values and their 95% confidence intervals under the null distribution for the SNPs within the imprinted regions.

Figure 2 | Genome-wide genetic correlation between birth weight and a range of traits and diseases in later life. Genetic correlation (r_g) and corresponding standard error between birth weight and the traits displayed on the *x* axis are estimated using LD Score regression¹². The genetic correlation estimates (r_g) are colour coded according to their intensity and direction (red for positive and blue for negative correlation). HC=head circumference, WHR=waist-hip ratio, WHR (adj BMI)=waist-hip ratio adjusted for body mass index, BMI=body mass index, Pubertal growth=standardized difference in height between age 8 and adult height, Total growth=standardized difference in height at age 14 and adult height, CAD=coronary artery disease, DBP=diastolic blood pressure, SBP=systolic blood pressure*, Chol=total cholesterol, TG=triglycerides, LDL=low-density lipoprotein, HDL=high-density lipoprotein, T2D=type 2 diabetes, HOMA-B=homeostatic model assessment of beta-cell function, HOMA-IR=homeostatic model assessment of insulin resistance, HbA1C=Hemoglobin A1c, LSBMD=lumbar spine bone mineral density, FNBMD=femoral neck bone mineral density, Edu Att=educational attainment, ADHD=attention deficit hyperactivity disorder, Cigarettes/Day=number of cigarettes per day. See Supplementary Table 12 for references for each of the traits and diseases displayed here.

Figure 3 | Hierarchical clustering of birth weight loci based on similarity of overlap with adult diseases, metabolic and anthropometric traits. For the lead SNP at each of the birth weight loci (listed on y axis), the z-score aligned to the birth weight-raising allele is obtained from publicly available GWAS results for various diseases and traits (listed on x axis) and are displayed in the heatmap. Of the 60 birth weight loci, z-score for lead SNP (or proxy) at 7 loci were not available (see Supplementary Table 17). Positive z-scores in red indicate the BW-raising allele increases the value of the trait concerned, and negative z-scores in blue show that it is associated with decrease of the trait value of interest. Birth weight loci and traits are clustered according to the Euclidean distance amongst z-score of the loci/traits (see Methods). White cross with a black circle in the middle signifies that BW locus is significantly ($P < 5 \times 10^{-8}$) associated with the trait of interest in the publicly available GWAS look-up (Supplementary Table 17). If the BW locus does not attain $P < 5 \times 10^{-8}$ in the GWAS look-up, but is reported significant elsewhere, it is marked with plain white cross. For example, ADRB1 (lead BW variant rs7076938) was not significantly associated with various measurements of blood pressure in Ehret *et al.*¹⁹ from which the z-scores were extracted, but was reported as blood pressure locus by Johnson et al.¹⁸ and Kato et al.⁸⁷. BMI=body mass index, CAD=coronary artery disease, Chol=total cholesterol, DBP=diastolic blood pressure, HC=head

circumference, HDL=high-density lipoprotein, LDL=low-density lipoprotein, SBP=systolic blood pressure, TG=triglycerides, T2D=type 2 diabetes, WHRadjBMI=waist-hip ratio adjusted for BMI.

EXTENDED DATA LEGENDS

Extended Data 1 | Flow chart of the study design.

Extended Data 2 | Sixty loci associated with birth weight (*P*<5x10⁻⁸) in European ancestry metaanalysis of up to 143,677 individuals and/or trans-ancestry meta-analysis of up to 153,781 individuals.

Extended Data 3 | Regional plots for multiple distinct signals at three birth weight loci, *ZBTB7B* (a), *HMGA1* (b) and *PTCH1* (c). Regional plots for each locus are displayed from: the unconditional European-specific meta-analysis of up to 143,677 individuals (left); the approximate conditional meta-analysis for the primary signal after adjustment for the index variant for the secondary signal (middle); and the approximate conditional meta-analysis for the secondary signal after adjustment for the index variant for the primary signal (right). Directly genotyped or imputed SNPs are plotted with their association *P*-values (on a -log₁₀ scale) as a function of genomic position (NCBI Build 37). Estimated recombination rates (blue lines) are plotted to reflect the local LD structure around the index SNPs and their correlated proxies. SNPs are coloured in reference to LD with the particular index SNP according to a blue to red scale from $r^2 = 0$ to 1, based on pairwise r^2 values estimated from a reference of 5,000 individuals of white British origin, randomly selected from the UK Biobank.

Extended Data 4 | Absolute effect size and minor allele frequency of lead SNPs at 60 known and novel birth weight loci from the trans-ancestry meta-analysis of up to 153,781 individuals. The effect of the lead SNP (absolute value of beta, *y* axis) is given as a function of minor allele frequency (*x* axis) for known birth weight loci in pink and novel loci in green. Error bars are proportional to the standard error of the effect size. The dashed line indicates 80% power to detect association at genome-wide significance level for the sample size in trans-ancestry meta-analysis. All birth weight loci were common except for two: at the *YKT6-GCK* locus, the index SNP (rs138715366) was of low-frequency in European ancestry populations (MAF=0.92%), and even rarer in other ancestry groups (MAF=0.23% in African Americans, otherwise monomorphic). Similarly, at the *SUZ12P1-CRLF3* locus, the index SNP (rs144843919) was of low-frequency in European and African American ancestry studies (MAF of 3.5% and 4.0%, respectively), and absent from other ancestry groups. None of the variants attaining genome-wide significance in European ancestry or trans-ancestry meta-analyses at these two loci was present in HapMap, suggesting that these two association signals would not have been identified without imputation up to the denser reference panels from 1000G/UK10K, irrespective of the available sample size.

Extended Data 5 | Comparison of foetal effect sizes and maternal effect sizes at 60 known and novel birth weight loci (continues to Extended Data 6). These plots illustrate that in large GWAS of BW, foetal effect size estimates are larger than maternal effect size estimates at 55/60 identified loci (binomial *P*=1x10⁻¹¹), suggesting that most of the associations are likely to be driven by the foetal genotype. In conditional analyses that modelled the effects of both maternal and foetal genotypes (n=12,909 mother-child pairs), confidence intervals around the estimates were wide, precluding inference about the likely contribution of maternal vs. foetal genotype at individual loci. For each birth weight locus, the following six effect sizes (with 95% CI) are shown, all aligned to the birth weight-raising allele in the overall trans-ancestry meta-analysis: foetal_GWAS = foetal allelic effect on birth weight (from European ancestry meta-analysis of up to n=143,677 individuals); foetal_unadjusted = foetal allelic effect on birth weight (unconditioned in n=12,909 mother-child pairs); foetal_adjusted = foetal effect conditioned on maternal genotype in n=12,909 mother-child pairs); maternal_GWAS = maternal allelic effect on offspring birth weight (from meta-analysis of up to n=68,254 European mothers)⁷; maternal_unadjusted = maternal allelic effect on offspring birth weight (unconditioned in n=12,909 mother-child pairs); maternal_gives = maternal allelic effect on offspring birth weight (unconditioned in n=12,909 mother-child pairs); maternal_gives = maternal allelic effect on offspring birth weight (from meta-analysis of up to n=68,254 European mothers)⁷; maternal_unadjusted = maternal allelic effect on offspring birth weight (unconditioned in n=12,909 mother-child pairs); maternal_adjusted = maternal effect

conditioned on foetal genotype (in n=12,909 mother-child pairs). The 60 birth weight loci are ordered by chromosome and position (Supplementary Tables 10 and 11).

Extended Data 6 | Comparison of foetal effect sizes and maternal effect sizes at 60 known and novel birth weight loci (continued from Extended Data 5). The scatterplot illustrates the difference between the foetal (*x* axis) and maternal (*y* axis) effect sizes in the overall maternal vs. foetal GWAS results.

Extended Data 7 | Gene set enrichment analysis and protein-protein interaction (PPI) analysis. Two complementary analyses of the overall GWAS summary data identified enrichment of BW associations in biological pathways related to metabolism, growth and development: Table (a) shows the top results (FDR<0.05 at the 95th percentile enrichment threshold) from a total of 3216 biological pathways tested for enrichment of multiple modest associations with BW. Additionally, results are presented for custom sets of imprinted genes. In Table (b) are the results of a complementary analysis of empirical PPI data, displaying the top 10 most significant pathways enriched for BW-association scores.

Extended Data 8 | Protein-Protein Interaction (PPI) Network analysis. a, illustrates the global largest component birth weight (BW) PPI network containing 13 modules. **b,** the histogram shows the null distribution of z-scores of BW PPI networks based on 10,000 random networks and where the z-scores for the 13 birth weight modules (M1-13) lie. For each module, the two most significant GO terms are depicted. **c,** heatmap showing the enrichment for the top 50 biological processes overrepresented in the global BW PPI network of the trait-specific "point of contact" (PoC) PPI networks. **d-e,** trait-specific PoC PPI networks composed of proteins that are shared in both the global birth weight PPI networks generated using the same pipeline for each of the adult traits: **d,** canonical Wnt signalling pathway enriched for PoC PPI between BW and blood pressure (BP)-related phenotypes; and **e,** regulation of insulin secretion pathway enriched for PoC between BW and type 2 diabetes (T2D)/fasting glucose (FG). Red nodes are those that are present in PoC for BW and traits of interest; blue nodes correspond to the pathway nodes; purple nodes are those present in both the pathway and PoC. Large nodes correspond to genes in BW loci (within 300kb from the lead SNP), and are orange when they overlap the enriched pathway and the PoC, and have black border if they, amongst all BW loci, have a stronger (top 5) association with at least one of the pairing adult traits.

Extended Data 9 | Quantile-Quantile (QQ) plots of (a) variance comparison between heterozygotes and homozygotes analysis in 57,715 UK Biobank samples and (b) parent-of-origin specific analysis in 4,908 ALSPAC mother-child pairs at 59 autosomal birth weight loci plus *DLK1*. Despite the large sample size, especially of unrelated individuals, these analyses were underpowered to detect parent-of-origin-specific effects at the identified loci. **a**, QQ plot from the Quicktest⁸¹ analysis comparing the BW variance of heterozygotes with homozygotes in 57,715 UK Biobank samples. **b**, QQ plot from the parent-of-origin specific analysis testing the association between birth weight and maternally transmitted vs. paternally transmitted alleles in 4,908 mother-child pairs from the ALSPAC study (Methods, Supplementary Tables 15 and 16). In both panels, the black dots represent lead SNPs at 59 identified autosomal birth weight loci and a further sub-genome-wide significant signal for birth weight near *DLK1* (rs6575803; *P*=5.6x10⁻⁸). The grey lines represent expected *P* values and their 95% confidence intervals under the null distribution for the 60 SNPs.

Extended Data 10 | Summary of previously reported loci for systolic blood pressure (SBP, a), coronary artery disease (CAD, b, e), type 2 diabetes (T2D, c, f) and adult height (d) and their effect on birth weight. a-d, Effect sizes (left *y* axis) of previously reported 30 SBP loci^{18,19}, 45 CAD loci²⁷, 84 T2D loci²⁸ and 422 adult height loci²⁹ are plotted against changes in birth weight z-score (*x* axis). Effect sizes are aligned to the adult trait-raising allele. The colour of each dot indicates birth weight association *P* value: red, $P < 5 \times 10^{-8}$; orange, $5 \times 10^{-8} \le P < 0.001$; yellow, $0.001 \le P < 0.01$; white, $P \ge 0.01$. The superimposed grey frequency histogram shows the number of SNPs (right *y* axis) in each category of birth weight effect size. **e**, Effect sizes (with 95% CI) on BW of 45 known CAD loci are plotted arranged in the order of CAD effect size from highest to lowest, separating out the known SBP loci and **f**, those of 32 known T2D loci are plotted, subdivided by previously reported categories derived from detailed adult physiological data³¹. Heterogeneity in BW effect sizes between five T2D loci groups with different mechanistic categories was substantial ($P_{het}=1.2\times 10^{-9}$). In pairwise comparisons, the "beta cell" group of variants differed from the other four groups: fasting hyperglycaemia ($P_{het} = 3\times 10^{-11}$), insulin resistance ($P_{het} = 0.002$), proinsulin ($P_{het} = 0.78$) and unclassified ($P_{het} = 0.02$) groups. All of the birth weight effect sizes plotted in the forest plots are aligned to the trait (or risk)-raising allele.



7 8 9 Chromosome



Figure 1



Figure 2



Figure 3