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# **Supplemental Information**

# Recombinant neutralizing secretory IgA

# antibodies for preventing mucosal acquisition

# and transmission of SARS-CoV-2

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## **Supplemental Information**

### **Supplemental Figures**

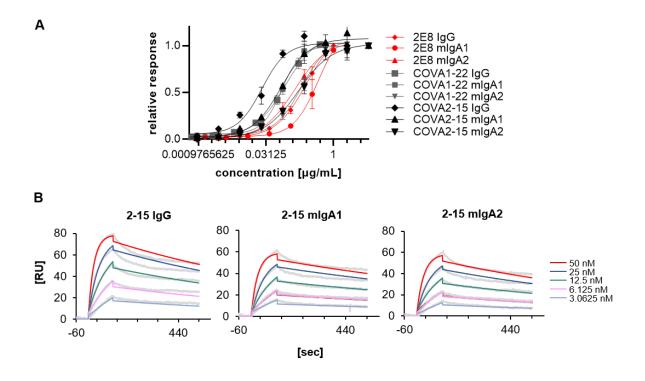


Figure S1: Binding of affinity purified COVA2-15, COVA1-22, 2-15 and 2E8 IgG, monomeric IgA1 and IgA2 to SARS-CoV-2 spike protein. (A) Binding of COVA1-22, 2E8 and COVA2-15 variants to ELISA plates coated with 100 ng/well full-length trimeric spike protein (Wuhan, kind gift from Dr. Svend Kjaer, the Crick Institute). Each displayed value is the mean  $\pm$  SD from three independent measurements. (B) Binding kinetics of 2-15 variants to RBD were obtained by SPR spectroscopy in multi-cycle kinetic experiments. An anti-His antibody was immobilized on an CM5 chip, RBD-His was captured (20 RU), and 5 different concentrations of the respective mAb were injected. The obtained curves were fitted with a 1:1 binding model. Data shown are from one experiment representative of at least two technical repeats. The calculated  $K_D$  values are 3.75 nM for 2-15 mIgA1, 2.88 nM for 2-15 mIgA2 and 1.16 nM for 2-15 IgG.

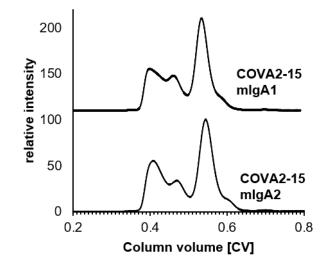


Figure S2: Normalized size-exclusion chromatograms of affinity-purified monomeric IgA1 and IgA2 variants of COVA2-15 from infiltrated *N. benthamiana*  $\Delta$ XT/FT leaves. Values were normalized based on the highest signal of each chromatogram.

2

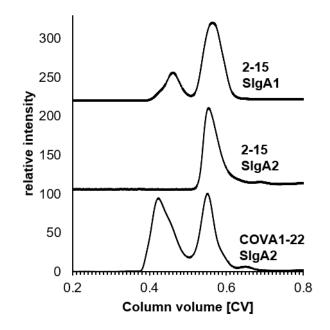


Figure S3: Normalized size-exclusion chromatograms of affinity-purified 2-15 and COVA1-22 IgA1 and IgA2 which were co-expressed with the joining chain and secretory component in *N. benthamiana* ΔXT/FT leaves to obtain multimeric secretory IgA. Values were normalized based on the highest signal of each chromatogram

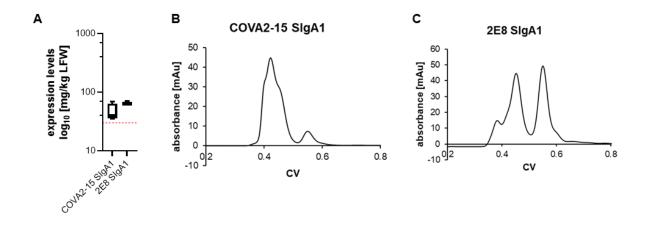
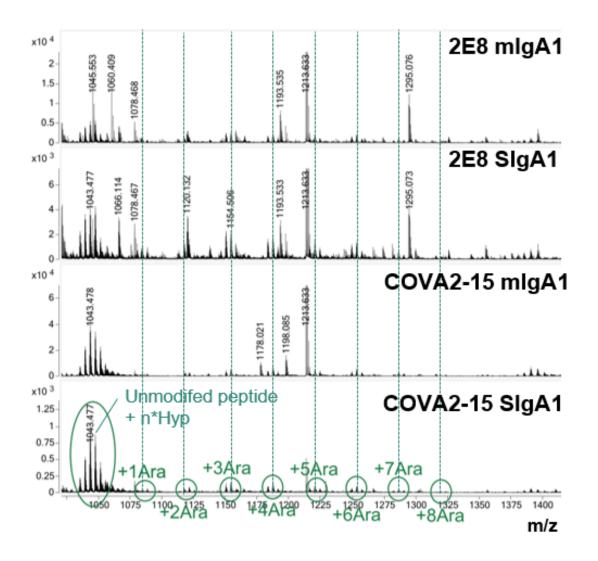
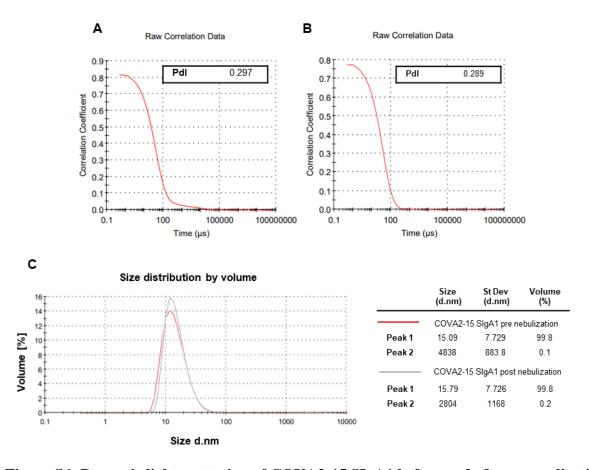


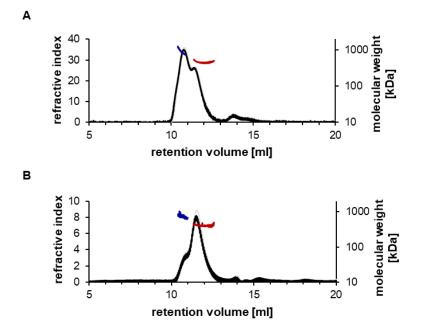
Figure S4: COVA2-15 and 2E8 secretory IgA1 were transiently expressed in *N.benthamiana* plants in pilot-scale 500 g LFW batches. (A) Expression levels were quantified by sandwich ELISA in crude leaf extracts. SIgA antibodies were detected using antisecretory component antibodies for all SIgA variants. Quantification data represent the mean of two technical repeats of three independent biological repeats  $\pm$  SD. Normalized size-exclusion chromatograms of affinity-purified COVA2-15 SIgA1 (B) and 2E8 SIgA1 (C) from infiltrated *N. benthamiana*  $\Delta$ XT/FT leaves.



**Figure S5**: **O-glycosylation profiles of** *N. benthamiana* Δ**XT/FT derived recombinant COVA2-15 and 2E8 monomeric and secretory IgA.** Mass spectra of the hinge region peptide are shown and glycosylated peaks are indicated as arabinoses (Ara) and hydroxyprolines (Hyp).



**Figure S6: Dynamic light scattering of COVA2-15 SIgA1 before and after aerosolization using the Omron MicroAir U22 portable mesh nebulizer**. The mean diameter and the homogeneity were measured using a Malvern Zetasizer nano-ZS (Malvern Instruments Ltd., Worcestershire, UK) at 25°C. Each sample was measured in triplicates. The homogeneity of the antibody solution was determined by the polydispersity index (PdI) from the raw correlation before (A) and after (B) nebulisation. Size distribution was calculated by the volume report (C).



**Figure S7: Size-exclusion chromatography light-scattering of COVA2-15 SIgA1 (A) and 2E8 SIgA1 (B) before and after nebulization**. Overlays of the elution profiles of COVA2-15 SIgA1 (nebulization concentration: 0.5 mg/mL) and 2E8 SIgA1 (nebulization concentration: 0.1 mg/mL) before (grey) and after (black) aerosolization are shown. The depicted molar masses of the tetrameric (blue) and dimeric (red) SIgA species were derived from lightscattering and are as follows in the formulation/the condensate: COVA2-15 SIgA1: 958/974 kDa, 468/470 kDa; 2E8 SIgA1:728/786 kDa, 409/410 kDa.

### **Supplemental Tables**

Table S1: Transient expression levels of monoclonal IgG and IgA antibodies in *N*. *benthamiana*  $\Delta$ XT/FT. IgG, monomeric and secretory IgA1 and IgA2 versions of 4 different mAbs recognizing the SARS-CoV-2 spike proteins were transiently expressed in plants. Expression levels were quantified by sandwich ELISA in crude leaf extracts. Detection of monomeric IgA and IgG variants was with either HRP-labeled anti-kappa (COVA2-15) or anti-lambda light chain (2E8, COVA1-22, 2-15) antibodies. SIgA antibodies were detected using anti-secretory component antibodies for all SIgA variants. Quantification data represent the mean of two technical repeats of three independent biological repeats  $\pm$  SD.

mAb	expression leve	els [	mg/kg LFW]
COVA2-15 IgG	134.90	±	30.78
COVA2-15 mlgA1	112.50	±	10.12
COVA2-15 mlgA2	103.30	±	12.69
COVA2-15 SIgA1	33.65	±	5.99
COVA2-15 SIgA2	45.49	±	15.90
2E8 lgG	100.10	±	17.79
2E8 mlgA1	74.57	±	10.91
2E8 mlgA2	45.40	±	12.65
2E8 SIgA1	35.71	±	8.41
2E8 SIgA2	23.07	±	7.48
2-15 lgG	52.48	±	8.89
2-15 mlgA1	74.57	±	10.91
2-15 mlgA2	45.40	±	12.65
2-15 SIgA1	10.03	±	2.95
2-15 SIgA2	1.77	±	0.95
COVA1-22 IgG	74.84	±	20.00
COVA1-22 mlgA1	79.47	±	11.11
COVA1-22 mlgA2	10.89	±	2.76
COVA2-15 SIgA2	6.36	±	0.41
COVA1-22 SIgA1	0.29	±	0.34

Table S2: Purification yields of monoclonal IgG and SIgA antibodies produced in N.benthamiana  $\Delta XT/FT$  after affinity chromatography and size-exclusion chromatography.Concentration in the pooled and concentrated SEC-fractions were quantified via A280 nm

absorbance. Data represent the mean of three obtained from three independent purifications from 100 g leaf fresh weight  $\pm$  SD.

mAb	purification yield [mg/kg LFW]						
COVA2-15 lgG	36.20	±	6.20				
COVA2-15 SIgA1	25.90	±	0.90				
COVA2-15 SIgA2	32.00	±	4.00				
2E8 lgG	38.10	±	8.10				
2E8 SIgA1	19.75	±	0.75				
2E8 SIgA2	10.20	±	0.60				

Table S3: Quantification of the relative abundance (%) of *N*-glycans detected on the heavy chain of IgG and IgA isotypes produced in *N. benthamiana*  $\Delta$ XT/FT. *N*-glycans are abbreviated according to the ProGlycAn system (www.proglycan.com). The symbols for the monosaccharides are drawn according to the nomenclature from the Consortium for Functional Glycomics.

2E8 mAb		mlg	gA1	Slg	gA1		mlg	gA2			Slg	JA2		lgG
		NLT	NVS	NLT	NVS	NVT	NLT	NIT	NVS	NVT	NLT	NIT	NVS	NST
non glyc.		0.2	38.3	0.6	36.1	0.0	0.5	0.0	19.9	0.0	0.8	0.0	31.6	11.3
GnGn		12.8	3.2	7.1	3.5	14.1	6.1	7.6	4.5	18.4	4.6	11.6	5.2	60.7
MGn	-2	23.1	3.4	15.5	4.2	18.4	15.1	21.0	3.4	16.2	10.8	21.5	3.8	22.5
MM	2	11.1	3.8	9.3	4.9	28.7	7.2	26.7	4.8	21.4	5.1	20.0	4.2	1.0
GnGnF	20- <b>1</b>	0.0	4.1	0.0	1.0	2.6	0.0	2.5	3.7	5.9	0.0	3.0	1.1	0.7
MGnF	• <b>;•</b> ••	0.0	1.7	0.0	0.5	4.8	0.0	6.6	1.3	3.5	0.0	5.8	0.5	1.4
MMF	<b>3-4</b>	0.0	0.0	0.0	0.0	5.5	0.0	8.8	0.0	3.5	0.0	5.4	0.0	0.0
Man4	фот	2.7	2.4	3.1	2.1	2.0	2.7	3.8	2.3	2.7	2.7	3.6	1.0	0.0
Man5	277	4.1	4.3	3.4	4.2	2.3	4.5	4.1	1.6	2.5	4.1	4.2	4.1	0.9
Man6	1200	11.0	5.0	12.8	5.3	4.1	10.4	1.9	6.6	4.1	9.1	2.4	5.8	0.0
Man7	- See	6.5	12.6	8.0	10.9	6.6	10.0	3.7	23.5	8.2	8.6	4.5	14.4	0.0
Man8	3	16.5	12.0	18.1	15.2	7.6	20.6	7.8	18.8	9.5	21.8	10.6	16.6	0.3
Man9	- 19-1	12.0	6.5	22.0	9.0	3.2	23.0	2.6	7.6	4.1	32.3	3.8	9.2	0.2
Man4Gn	÷	0.0	1.5	0.0	1.5	0.0	0.0	2.1	1.0	0.0	0.0	2.4	1.2	1.2
Man5Gn	Ş	0.0	1.1	0.0	1.6	0.0	0.0	0.8	1.1	0.0	0.0	1.1	1.2	0.0
CaVA2 45	~ ^ h	ml	~ ^ 4	614		I	m			1	614			1
CoVA2-15	mAb	<u>mlg</u>						3A2	NIVO			jA2	NIVO	IgG
	mAb	NLT	NVS	NLT	NVS	NVT	NLT	NIT	NVS	NVT	NLT	NIT	NVS	NST
non glyc.		NLT 0.5	NVS 38.5	NLT 0.9	NVS 37.8	0.0	NLT 0.3	NIT 0.0	67.6	0.0	NLT 0.7	NIT 0.0	48.6	NST 20.5
non glyc. GnGn		NLT 0.5 17.1	NVS 38.5 5.6	NLT 0.9 7.8	NVS 37.8 3.8	0.0 16.6	NLT 0.3 10.2	NIT 0.0 13.3	67.6 5.1	0.0 17.3	NLT 0.7 2.6	NIT 0.0 8.4	48.6 1.7	NST 20.5 43.5
<u>non glyc.</u> GnGn MGn	0 	NLT 0.5 17.1 19.3	NVS 38.5 5.6 13.9	NLT 0.9 7.8 10.6	NVS 37.8 3.8 6.3	0.0 16.6 14.7	NLT 0.3 10.2 17.1	NIT 0.0 13.3 21.2	67.6 5.1 9.9	0.0 17.3 14.3	NLT 0.7 2.6 3.6	NIT 0.0 8.4 11.8	48.6 1.7 2.3	NST 20.5 43.5 21.3
<b>non glyc.</b> GnGn MGn MM	0 	NLT 0.5 17.1 19.3 10.2	NVS 38.5 5.6 13.9 12.1	NLT 0.9 7.8 10.6 4.5	NVS 37.8 3.8 6.3 1.9	0.0 16.6 14.7 11.2	NLT 0.3 10.2 17.1 8.0	NIT 0.0 13.3 21.2 23.4	67.6 5.1 9.9 1.7	0.0 17.3 14.3 11.0	NLT 0.7 2.6 3.6 1.7	NIT 0.0 8.4 11.8 4.0	48.6 1.7 2.3 0.5	NST 20.5 43.5 21.3 3.1
non glyc. GnGn MGn MM GnGnF	0 -0 5 0-4	NLT 0.5 17.1 19.3 10.2 0.0	NVS 38.5 5.6 13.9 12.1 4.3	NLT 0.9 7.8 10.6 4.5 0.0	NVS 37.8 3.8 6.3 1.9 1.0	0.0 16.6 14.7 11.2 3.6	NLT 0.3 10.2 17.1 8.0 0.0	NIT 0.0 13.3 21.2 23.4 2.7	67.6 5.1 9.9 1.7 3.8	0.0 17.3 14.3 11.0 3.6	NLT 0.7 2.6 3.6 1.7 0.0	NIT 0.0 8.4 11.8 4.0 0.9	48.6 1.7 2.3 0.5 0.2	NST 20.5 43.5 21.3 3.1 0.8
non glyc. GnGn MGn MM GnGnF MGnF	00 00 00-4 00-4	NLT 0.5 17.1 19.3 10.2 0.0 0.0	NVS 38.5 5.6 13.9 12.1 4.3 1.4	NLT 0.9 7.8 10.6 4.5 0.0 0.0	NVS 37.8 3.8 6.3 1.9 1.0 0.5	0.0 16.6 14.7 11.2 3.6 2.5	NLT 0.3 10.2 17.1 8.0 0.0 0.0	NIT 0.0 13.3 21.2 23.4 2.7 4.7	67.6 5.1 9.9 1.7 3.8 1.2	0.0 17.3 14.3 11.0 3.6 1.9	NLT 0.7 2.6 3.6 1.7 0.0 0.0	NIT 0.0 8.4 11.8 4.0 0.9 1.3	48.6 1.7 2.3 0.5 0.2 0.3	NST 20.5 43.5 21.3 3.1 0.8 1.3
non glyc. GnGn MGn MM GnGnF MGnF MMF	00 -0 00 00-4 -00-4 00-4 00-4	NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0	0.0 16.6 14.7 11.2 3.6 2.5 1.5	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 0.0	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2	67.6 5.1 9.9 1.7 3.8 1.2 0.0	0.0 17.3 14.3 11.0 3.6 1.9 1.0	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 0.0	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4	48.6 1.7 2.3 0.5 0.2 0.3 0.0	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0
non glyc. GnGn MGn MM GnGnF MGnF MMF Man4	10 -0 10-4 -0-4 -0-4 -0-4 -0-4	NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0 1.6	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 0.0 2.4	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 1.4	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0
non glyc. GnGn MGn MM GnGnF MGnF MMF Man4 Man5	00 -0 00 00-4 -0-4 -0 00 00 00 00	NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7 1.8	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2 2.0	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1 2.8	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0 1.6 3.8	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4 3.2	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 0.0 2.4 3.5	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7 3.0	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5 1.2	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0 5.2	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 0.0 1.4 2.6	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3 6.6	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1 2.6	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0 3.2
non glyc. GnGn MGn GnGnF MGnF MMF Man4 Man5 Man6	00 -0 00 00-4 -0-4 -0 00 00 00 00	NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7 1.8 5.4	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2 2.0 2.0	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1 2.8 8.7	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0 1.6 3.8 5.1	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4 3.2 4.8	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 0.0 2.4 3.5 5.3	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7 3.0 2.0	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5 1.2 0.9	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0 5.2 6.1	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 1.4 2.6 4.9	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3 6.6 6.2	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1 2.6 3.8	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0 3.2 0.0
non glyc. GnGn MGn MM GnGnF MGnF MMF Man4 Man5 Man6 Man7		NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7 1.8 5.4 4.8	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2 2.0 2.0 9.3	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1 2.8 8.7 6.3	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0 1.6 3.8 5.1 12.8	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4 3.2 4.8 11.3	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 0.0 2.4 3.5 5.3 7.0	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7 3.0 2.0 2.7	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5 1.2 0.9 2.3	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0 5.2 6.1 14.3	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 1.4 2.6 4.9 4.8	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3 6.6 6.2 6.2 6.2	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1 2.6 3.8 10.0	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0 3.2 0.0 0.7
non glyc. GnGn MGn MM GnGnF MGnF MMF Man4 Man5 Man6 Man7 Man8	00 -0 00 00-4 -0-4 -0 00 00 00 00	NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7 1.8 5.4 4.8 23.4	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2 2.0 2.0 9.3 6.0	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1 2.8 8.7 6.3 23.6	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0 1.6 3.8 5.1 12.8 15.3	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4 3.2 4.8 11.3 21.0	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 2.4 3.5 5.3 7.0 21.2	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7 3.0 2.0 2.7 11.7	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5 1.2 0.9 2.3 2.0	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0 5.2 6.1 14.3 16.2	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 1.4 2.6 4.9 4.8 19.1	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3 6.6 6.2 6.2 6.2 25.4	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1 2.6 3.8 10.0 15.5	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0 3.2 0.0 0.7 1.7
non glyc. GnGn MGn MM GnGnF MGnF MMF Man4 Man5 Man6 Man7 Man8 Man9		NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7 1.8 5.4 4.8 23.4 15.8	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2 2.0 2.0 9.3 6.0 2.4	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1 2.8 8.7 6.3 23.6 32.7	NVS 37.8 6.3 1.9 1.0 0.5 0.0 1.6 3.8 5.1 12.8 15.3 8.5	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4 3.2 4.8 11.3 21.0 7.4	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 2.4 3.5 5.3 7.0 21.2 25.0	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7 3.0 2.0 2.7 11.7 5.5	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5 1.2 0.9 2.3 2.0 1.9	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0 5.2 6.1 14.3 16.2 6.1	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 1.4 2.6 4.9 4.8 19.1 58.5	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3 6.6 6.2 6.2 6.2 25.4 23.1	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1 2.6 3.8 10.0 15.5 12.5	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0 3.2 0.0 0.7 1.7 2.1
non glyc. GnGn MGn MM GnGnF MGnF MMF Man4 Man5 Man6 Man7 Man8		NLT 0.5 17.1 19.3 10.2 0.0 0.0 0.0 0.0 1.7 1.8 5.4 4.8 23.4	NVS 38.5 5.6 13.9 12.1 4.3 1.4 0.0 1.2 2.0 2.0 9.3 6.0	NLT 0.9 7.8 10.6 4.5 0.0 0.0 0.0 2.1 2.8 8.7 6.3 23.6	NVS 37.8 3.8 6.3 1.9 1.0 0.5 0.0 1.6 3.8 5.1 12.8 15.3	0.0 16.6 14.7 11.2 3.6 2.5 1.5 2.4 3.2 4.8 11.3 21.0	NLT 0.3 10.2 17.1 8.0 0.0 0.0 0.0 2.4 3.5 5.3 7.0 21.2	NIT 0.0 13.3 21.2 23.4 2.7 4.7 5.2 2.7 3.0 2.0 2.7 11.7	67.6 5.1 9.9 1.7 3.8 1.2 0.0 0.5 1.2 0.9 2.3 2.0	0.0 17.3 14.3 11.0 3.6 1.9 1.0 3.0 5.2 6.1 14.3 16.2	NLT 0.7 2.6 3.6 1.7 0.0 0.0 0.0 1.4 2.6 4.9 4.8 19.1	NIT 0.0 8.4 11.8 4.0 0.9 1.3 0.4 4.3 6.6 6.2 6.2 6.2 25.4	48.6 1.7 2.3 0.5 0.2 0.3 0.0 1.1 2.6 3.8 10.0 15.5	NST 20.5 43.5 21.3 3.1 0.8 1.3 0.0 0.0 3.2 0.0 0.7 1.7

Table S4: Quantification of the relative abundance [%] of *N*-glycans detected on the secretory component and joining chain of IgA isotypes produced in *N. benthamiana*  $\Delta XT/FT$ . *N*-glycans are abbreviated according to the ProGlycAn system (www.proglycan.com). The symbols for the monosaccharides are drawn according to the nomenclature from the Consortium for Functional Glycomics.

2E8 mAb	SIgA1					SIgA2					
			S	С		JC		S	С		JC
		NDT	NYT	NGT	NVT	NIS	NDT	NYT	NGT	NVT	NIS
non glycos	ylated	0.0	6.3	0.0	0.0	0.0	0.0	6.2	0.0	0.0	0.0
GnGn	2	9.1	20.5	0.0	14.2	1.7	10.1	18.6	0.0	11.2	2.1
GnM	•;•••	23.7	27.4	0.0	36.3	3.1	22.8	21.4	0.0	35.5	4.0
MM		18.9	45.8	0.0	45.9	5.4	24.1	53.9	0.0	49.8	5.7
GnGnF	20- <b>1</b>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
MGnF	• <b>:</b> ‡	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
MMF	5- <b>1</b>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Man4		0.0	0.0	0.0	0.0	9.4	0.0	0.0	0.0	0.0	8.9
Man5	200	0.0	0.0	3.8	0.0	11.6	0.0	0.0	4.3	0.0	16.8
Man6		0.0	0.0	8.5	0.0	14.9	0.0	0.0	10.2	0.0	14.5
Man7	- 19-m -	12.4	0.0	26.1	0.0	30.4	9.0	0.0	26.4	0.0	27.6
Man8	3	26.6	0.0	41.2	0.0	15.7	21.9	0.0	38.0	9.1	13.4
Man9	<u>}</u>	9.3	0.0	19.9	0.0	7.7	12.0	0.0	19.1	23.7	7.0
Man9+Hex		0.0	0.0	0.5	0.0	0.0	0.0	0.0	2.0	0.0	0.0
MU		0.0	0.0	0.0	3.5	0.0	0.0	0.0	0.0	3.4	0.0

CoVA2-15 n			SIgA1					SIgA2			
			S	C		JC		S	С		JC
		NDT	NYT	NGT	NVT	NIS	NDT	NYT	NGT	NVT	NIS
non glycosy	/lated	0.0	11.1	0.0	0.0	0.0	0.0	19.4	0.0	0.0	0.0
GnGn		12.9	17.4	0.0	12.7	0.9	4.9	13.4	0.0	12.1	1.1
GnM	•;•••	22.3	20.2	0.0	30.9	1.6	12.8	20.6	0.0	32.2	2.0
MM	200	27.0	51.3	0.0	53.6	4.6	19.8	46.6	0.0	53.3	4.2
GnGnF	2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
MGnF	• <b>**</b> •	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
MMF	2- <b>-</b>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Man4		0.0	0.0	0.0	0.0	7.4	0.0	0.0	0.0	0.0	5.8
Man5	200	0.0	0.0	3.3	0.0	13.9	0.0	0.0	3.2	0.0	10.0
Man6		0.0	0.0	7.6	0.0	11.9	0.0	0.0	7.5	0.0	10.2
Man7	- 19	8.7	0.0	24.4	0.0	22.1	11.9	0.0	21.7	0.0	20.2
Man8	3	22.4	0.0	37.8	0.0	17.2	36.4	0.0	33.2	12.9	20.8
Man9	9	6.7	0.0	26.7	0.0	20.4	14.2	0.0	32.8	22.3	25.9
Man9+Hex	32	0.0	0.0	0.3	0.0	0.0	0.0	0.0	1.5	0.0	0.0
MU		0.0	0.0	0.0	2.8	0.0	0.0	0.0	0.0	2.5	0.0

# Table S5: Modifications of the hinge-region of plant-produced COVA2-15 and 2E8 monomeric and secretory IgA1. Relative quantification (%) of hydroxyprolines and arabinose chains in the hinge-region tryptic peptide.

	2E8 mlgA1	2E8 SIgA1	COVA2-15 mlgA1	COVA2-15 SIgA1
0 hyP + no Ara	7.3	8.2	2.4	3.5
1 hyP + no Ara	9.5	11.6	7.1	10.4
2 hyP + no Ara	12.6	14.5	15.2	21.4
3 hyP + no Ara	11.7	11.6	16.6	21.4
4 hyP + no Ara	5.3	4.8	11.3	13.6
5 hyP + no Ara	1.5	1.4	5.8	6.0
4 hyP + 1 Ara	2.0	1.6	1.5	1.1
5 hyP + 1 Ara	2.3	2.5	2.1	1.4
6 hyP + 1 Ara	0.8	0.7	0.3	0.3
4 hyP + 2 Ara	2.3	2.2	1.8	1.2
5 hyP + 2 Ara	1.5	2.6	2.7	1.8
6 hyP + 2 Ara	0.4	0.9	1.0	0.2
4 hyP + 3 Ara	3.1	3.8	1.8	1.1
5 hyP + 3 Ara	3.0	3.6	3.7	2.0
6 hyP + 3 Ara	2.7	1.8	1.6	1.1
4 hyP + 4 Ara	1.6	2.3	1.8	1.1
5 hyP + 4 Ara	3.0	3.0	3.7	1.6
6 hyP + 4 Ara	2.1	1.8	1.0	0.8
4 hyP + 5 Ara	2.1	1.9	1.0	0.8
5 hyP + 5 Ara	3.8	2.9	2.5	1.4
6 hyP + 5 Ara	3.1	2.0	2.2	0.9
4 hyP + 6 Ara	3.6	2.9	2.0	1.0
5 hyP + 6 Ara	3.2	2.6	2.7	1.7
6 hyP + 6 Ara	1.9	2.0	1.7	0.8
4 hyP + 7 Ara	1.8	1.5	1.0	0.5
5 hyP + 7 Ara	2.2	1.2	1.6	0.9
6 hyP + 7 Ara	1.9	1.7	1.7	0.9
4 hyP + 8 Ara	1.3	0.6	0.3	0.2
5 hyP + 8 Ara	1.4	1.0	1.0	0.4
6 hyP + 8 Ara	1.0	0.8	0.9	0.5

			EC₅₀[nM]					
	lgG	mlgA1	mlgA2	SlgA1	SIgA2			
			COVA2-15					
Wuhan	0.032 ± 0.011	0.054 ± 0.030	0.149 ± 0.050	0.010 ± 0.018	0.017 ± 0.009			
Delta	9.028 ± 0.019	1.951 ± 0.014	2.063 ± 0.068	0.086 ± 0.026	0.542 ± 0.036			
	2E8							
Wuhan	0.271 ± 0.143	0.216 ± 0.030	0.836 ± 0.023	0.002 ± 0.123	0.005 ± 0.068			
Delta	1.071 ± 0.133	0.439 ± 0.029	2.031 ± 0.029	0.023 ± 0.030	0.029 ± 0.028			

**Table S7:** Kinetic parameters of COVA2-15 and 2E8 monomeric IgA mAbs to RBD. Rate

 constants were determined at 5 different concentrations using a 1:1 binding model.

	ka (1/Ms)	kd (1/s)	<i>KD</i> (nM)
COVA2-15 mlgA1	230920.6	0.00027	1.43
COVA2-15 mlgA2	207058.8	0.00035	1.67
2E8 mlgA1	79590.0	0.00100	125.42
2E8 mlgA2	97574.8	0.00085	89.59

**Table S8: ID<sub>50</sub> values of COVA2-15 and 2E8 antibody variants for neutralization of SARS-CoV-2 (England 02/2020).** Neutralisation capacity was measured using a PRNT assay on Vero E6 cells. mAbs were added in serial 1:10 dilutions starting with 10 μg/mL. A positive control (Pos; WHO International Standard of anti-SARS-CoV-2 immunoglobulin, 20/136, NIBSC, UK) was included. IC<sub>50</sub> vales have been determined from duplicates of one representative out of two experiments with similar results using non-linear regression in GraphPad Prism.

	IC50 [ng/mL]					
mAb	value	+error	-error			
Positive control	76.1	24,0	18.2			
CoVA2-15 IgG	53.4	6.9	6.1			
CoVA2-15 mlgA1	29.7	6.9	5.6			
CoVA2-15 SIgA1	13.5	3.6	2.8			
CoVA2-15 mlgA2m(1)	81.2	8.2	7.5			
CoVA2-15 SIgA2m(1)	22.8	5.5	4.4			
2E8 lgG		n.d				
2E8 mlgA1	1707.0	902.8	590.6			
2E8 SIgA1	36.6	37.4	18.5			
2E8 mlgA2m(1)	3210.1	4482.5	1870.5			
2E8 SIgA2m(1)	176.1	149.4	80.8			