iScience

Review

The effect of the delta SARS-CoV-2 variant on maternal infection and pregnancy

Athina Samara,^{1,2,*} Asma Khalil,^{3,4,5} Patrick O'Brien,^{6,7} and Eric Herlenius^{1,2}

SUMMARY

A greater proportion of pregnant women with COVID-19 have mild disease compared with their non-pregnant counterparts. Paradoxically, however, they are at higher risk of developing severe disease, requiring respiratory support and admission to intensive care. The delta SARS-Cov-2 variant is associated with increased risk of hospitalization and morbidity in unvaccinated pregnant populations. However, it is not known whether the worse pregnancy outcomes associated with the delta variant are due to a direct effect of the virus on the pregnancy, or whether this effect is mediated through more severe maternal infection. Here, we synthesize studies of COVID-19 pregnancies, focusing on the different routes of SARS-CoV-2 infection of lung and placenta, and the mechanisms of syncytial formation for each SARS-CoV-2 variant. To delineate COVID-19 complications in pregnant women, future studies should explore whether the delta variant causes greater placental infection compared to other variants and contributes to increased syncytial formation.

INTRODUCTION

Although pregnant women with COVID-19 usually display mild to moderate symptoms when compared to non-pregnant women with the same risk profile, they are at higher risk of developing severe disease, requiring respiratory support and admission to intensive care (ICU) (Knight et al., 2021; Villar et al., 2021; Allotey et al., 2020; Engjom et al., 2022; Donati et al., 2022; Overtoom et al., 2021). The USA Centers for Disease Control and Prevention (CDC) reported an apparent increase in the ratio of COVID-19-associated deaths per 1,000 cases among pregnant women as the delta variant became predominant (Kasehagen et al., 2021). That amounted to 5 vs 25 deaths per 1,000 SARS-CoV-2 infections during pregnancy, in the pre-delta (Mar 2020–Jun 2021) compared to the delta-predominant period (Jul–Oct 2021). Several studies reported that, compared with the other widespread variants, the delta variant is associated with increased risk of hospitalization and morbidity in unvaccinated pregnant populations (Vousden et al., 2021). The CDC reported (DeSisto et al., 2021) that the risk of stillbirth was 2-fold higher in pregnant women with COVID-19 compared with those without the infection.

What is not known, however, is whether the worse pregnancy outcomes associated with the delta variant are due to a direct effect of the virus on the pregnancy, or whether this effect is mediated through more severe maternal infection. The delta variant has been associated with greater viral loads than the alpha variant (Luo et al., 2021), but the pathophysiological mechanisms affecting pregnancy outcomes remain unclear.

EPIDEMIOLOGICAL STUDIES REPORTING AN ASSOCIATION BETWEEN THE DELTA VARIANT AND WORSE MATERNAL INFECTION AND PREGNANCY OUTCOMES

A national UK prospective cohort study showed that, after adjusting for pre-existing medical conditions and sociodemographic variables, the proportion of symptomatic pregnant women hospitalized with moderate to severe COVID-19 increased significantly from the wild type (24%) to alpha (36%) to the delta period (45%) (Vousden et al., 2021). Furthermore, pregnant women admitted during the delta wave had increased risk (compared to those hospitalized during the alpha wave) of having pneumonia and had non-significant increases in the need for respiratory support and ICU admission (Vousden et al., 2021) and death (UKOSS MBRRACE Infographic v13, 2021). A US study also found that the proportion of severe to critical disease resulting in ICU admission was greater in the delta cohort compared to the pre-delta cohort (Seasely



²Astrid Lindgren Children's Hospital, Karolinska University Hospital, Stockholm, Sweden

³Fetal Medicine Unit, St George's Hospital, St George's University of London, London, UK

⁴Vascular Biology Research Centre, Molecular and Clinical Sciences Research Institute, St George's University of London, London, UK

⁵Fetal Medicine Unit, Liverpool Women's Hospital, University of Liverpool, Liverpool, UK

⁶The Royal College of Obstetricians and Gynaecologists, London, UK

⁷University College London Hospitals NHS Foundation Trust, London, UK

*Correspondence: athina.samara@ki.se

https://doi.org/10.1016/j.isci. 2022.104295







et al., 2021). Moreover, in this study, the rates of adverse pregnancy outcomes including cesarean delivery, preterm birth, and neonatal ICU admission were also higher in the delta cohort. Increased morbidity was also observed in pregnancy with COVID-19 during the Delta surge (Adhikari et al., 2022a).

A meta-analysis evaluated the severity of disease caused by the SARS-COV-2 variants of concern (VOCs) in the general population from June 1, 2020 to October 15, 2021. This showed that the beta and delta variants pose a greater risk of hospitalization, ICU admission, and mortality compared to the alpha, gamma, and wild-type variants (Lin et al., 2021). Findings from the UK report suggested that during the periods of alpha and delta variant dominance, COVID-19 was associated with more severe maternal infection and worse pregnancy outcomes compared to the period of wild-type dominance (Vousden et al., 2022). Moreover, recent US reports showed that the delta and omicron variants were associated with increased SARS-CoV-2 infections in pregnancy (Adhikari et al., 2022a; 2022b). Among those, the majority occurred in unvaccinated pregnant women and, after adjusting for prior vaccination, the predominance of the delta variant was associated with increased, and omicron with decreased, severity of illness (Adhikari et al., 2022a; 2022b). The finding that most women admitted with SARS-CoV-2-related symptoms were unvaccinated was also reported (Vousden et al., 2022; Birol Ilter et al., 2022).

TRANSPLACENTAL TRANSFER OF NEUTRALIZING ANTIBODIES AGAINST BOTH WILD TYPE AND DELTA VARIANTS IN VACCINATED WOMEN

Full vaccination during pregnancy is essential but data are limited regarding transplacental antibody transfer after vaccination with the currently available vaccines. A small study (n = 29) evaluating maternal and umbilical cord blood on the day of birth assessed neutralizing antibody levels for both the wild type and delta variant and showed a pronounced reduction for the delta variant (Shen et al., 2022).

SEVERITY OF MATERNAL COVID-19 DOES NOT CONSISTENTLY CORRELATE WITH PLACENTAL SARS-COV-2 VIRAL LOAD

Although vertical transmission of SARS-CoV-2 is rare, has been observed (Zaigham and Andersson, 2020; Kasehagen et al., 2021) and placental pathology might be present even in late term asymptomatic cases (Jaiswal et al., 2021). Several reports of severe placental SARS-CoV-2 infection describe malperfusion and diffuse inflammatory histological changes, including massive perivillous fibrin depositions, necrosis of syncytiotrophoblast, and diffuse chronic intervillositis (Mao et al., 2022; Schwartz et al., 2021; Schwartz and Morotti, 2020). While these severe placental histopathological findings are not unique to maternal COVID-19 (Husen et al., 2021), the severity of maternal COVID-19 has been correlated with placental SARS-CoV-2 viral load (Rangchaikul and Venketaraman, 2021). A histological analysis comparing placentas of 28 uninfected and 85 women with symptomatic SARS-CoV-2 infection in pregnancy showed severe vascular remodeling of the placental arteries, thickened placental vascular walls, and narrowed lumen (Gychka et al., 2021). The vascular remodeling was associated with increased smooth muscle cell proliferation and fibrosis.

Pregnant women with gestational diabetes mellitus are nine times as likely as those without diabetes to be infected with the delta variant and are three times more susceptible than those with cardiovascular disease or hypertension (Mamun and Khan, 2021). Women with gestational diabetes are also more vulnerable to infection with the delta variant than with wild type or alpha variants (Mamun and Khan, 2021). SARS-CoV-2 was more commonly found in placentas of COVID-19-positive mothers with hypertensive disorders of pregnancy (HDP) compared to those without HDP (Fabre et al., 2021). The authors of this study also suggest the possibility that SARS-CoV-2 infection during pregnancy could trigger HDP through persistent placental infection causing placental damage (Fabre et al., 2021), but did not offer a mechanistic explanation.

In a review, Rangchaikul and Venketaraman suggested some parameters that may predispose the pregnant patient to greater susceptibility to and severity of SARS-CoV-2. These include, among others, hampering of cell-mediated immune clearance of SARS-CoV-2, altered immunomodulation by progesterone, coagulation, and complement-associated hyperinflammation (Rangchaikul and Venketaraman, 2021).

A recent report described two cases of intra-uterine fetal demise and a case of severe fetal distress after infection of unvaccinated mothers with the delta variant and mild COVID-19 disease severity (Shook





et al., 2022). All mothers had viremia and high nasopharyngeal viral load, but also evidence of placental infection with the delta variant and features of SARS-CoV-2-induced placentitis. Similarly, another case report of third trimester fetal demise following infection with the delta variant, described an unvaccinated mother with mild COVID-19 symptoms, but the placental analysis showed intervillous inflammation, degeneration, loss of syncytiotrophoblastic layers, and syncytial knots (Guan et al., 2022).

FORMATION OF SYNCYTIA AS A PATHOLOGICAL FEATURE OF COVID-19

Cell-to-cell fusion that leads to the formation of a syncytium is a physiological mechanism that occurs in various cell types, such as myocytes. The same mechanism allows viruses to infect neighboring cells without the exocytosis of free virus, and virus-infected syncytia increase tissue damage. Infectious syncytia are formed by the attachment of virions to cells or cell-to-cell fusion and are typical of coronaviruses (Li et al., 2003; Buchrieser et al., 2020). Formation of syncytia is a central pathological feature of COVID-19. COVID-19 severity also correlates closely with lung damage, and syncytia are often observed in the lungs of patients who have developed fatal pneumonia (Sanders et al., 2021; Braga et al., 2021; Bussani et al., 2020). It has also been demonstrated that SARS-CoV-2-infected syncytia may affect cardiomyocytes (Bailey et al., 2021).

The rate of formation of syncytia by SARS-CoV-2 has been shown to be much faster than by SARS-CoV-1 (Mehta et al., 2020). Of note, this syncytium formation determines both the degree of virulence and induction of the cytokine storm by SARS-CoV-2 (Matsuyama et al., 2020; Xia et al., 2020), as seen with other viruses.

Regarding the size of the syncytia, it has been reported that the alpha and beta variants produced larger syncytia than both the original Wuhan strain and the D614G variant (one of the earliest variants associated with an increased rate of transmission) (Rajah et al., 2021). Another study compared the replication rate and cell-to-cell transmission infection pattern of SARS-CoV-2 variants *in vitro*. Of the cells positive for any variant, at least 43% were distributed as clusters of 2 or more cells, and the delta variant proved to have the highest percentage (>78%) of infected cells organized in clusters, relative to alpha (ca. 59%) or beta (ca. 69%) variants (Al-Beltagi et al., 2021).

There is evidence, therefore, that the delta variant spike protein may increase cell-to-cell fusion (Arora et al., 2021) when compared to the wild type, the alpha, and the omicron variant (Suzuki et al., 2022). Regarding omicron, preliminary animal model studies show that it causes less severe disease accompanied by lower viral load in both the lower and upper respiratory tract (Bentley et al., 2021). Preliminary studies on this variant show reduced ability to induce syncytia in tissue culture, and that it may instead use (Meng et al., 2022) endosomal fusion through cathepsins (Peacock et al., 2022; Willett et al., 2022) (Figure 1).

SARS-COV-2 EFFECT ON PLACENTATION, SYNCYTIALIZATION, AND GAS EXCHANGE

Placentation is a complex multi-step process that enables placental perfusion, during which trophoblast cells adhere, invade, and remodel spiral arteries. Using a process called syncytialization, the cytotrophoblasts fuse with the overlying giant multinucleated syncytiotrophoblasts and form the outer layer of the placental microvilli (Burton and Fowden, 2015; Robbins and Bakardjiev, 2012) rendering the tissue impermeable and enabling mother–child immune tolerance (Alasadi et al., 2019). The cytotrophoblasts may regenerate the syncytiotrophoblast if damaged, but if the process is significantly compromised, pathological conditions such as preeclampsia and intra-uterine fetal growth restriction (FGR) might ensue (Huppertz and Kingdom, 2004; Pötgens et al., 2002; Sankar et al., 2012). Increased numbers of syncytial knots, as seen in the placenta under conditions of hypoxia, hyperoxia, or in the presence of reactive oxygen species (ROS) (Heazell et al., 2007), and in preeclampsia (Redline and Patterson, 1995), were also observed with COVID-19 (Singh et al., 2021; Gao et al., 2021). This was possibly due to maternal vascular malperfusion; the increase in numbers of syncytial knots was positively correlated with disease severity, as is the case with vascular endothelial growth factor (VEGF) expression (Shchegolev et al., 2021).

There may be small amounts of fibrin deposition in normal placentas, which is increased in HDP (Fox, 1967). Massive intervillous fibrin deposition has also been reported as a hallmark of COVID-19 pregnancies. As systemic SARS-CoV-2 infection leads to hypoxia and reduced utero-placental perfusion, it induces focal necrosis and extensive fibrin deposition. This increased amount of intravillous and perivillous fibrin might result from impaired fibrinolytic capacity of the compromised maternal endothelium or from immune





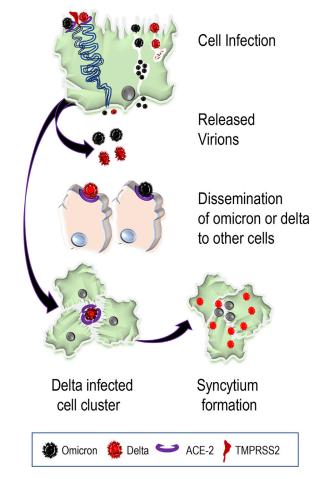


Figure 1. Delta SARS-CoV-2 variant efficiently enters cells via binding to the ACE2 receptor and activating cell membrane fusion using the host cell-surface protease TMPRSS2

This specific mechanism of infection of cells bearing both the ACE2 and TMPRSS2 (double ACE2+ TMPRSS2+ cell infection) allows the delta virions to bypass the endosome and not be destroyed by the host cell. On the other hand, omicron may enter cells in both a TMPRSS2-dependent and independent manner, as it can avoid the endosome, and may also infect ACE2 cells. Omicron enters cells through endocytosis, as its spike proteins drive the fusion of viral and endosomal membranes to facilitate insertion of the viral genome into the cytoplasm. Once the cell is infected by omicron, the virions that are replicated inside are finally released to disseminate other cells. In the case of the delta variant, when the spike protein is expressed on the surface of infected cells, it may interact with the ACE2 receptors on neighboring cells and form syncytia.

cell activation with subsequent pro-coagulation signals. As fibrin deposition further compromises maternal-fetal gas exchange, further research is needed to elucidate the underlying cellular and molecular mechanisms.

ROUTES OF SARS-COV-2 INFECTION IN LUNG VS. PLACENTAL CELLS

For a cell to be permissive to SARS-CoV-2, it must express the angiotensin-converting enzyme 2 (ACE2) receptor and have protease activity—transmembrane serine protease 2 (TMPRSS2) (Matsuyama et al., 2010). After transmission, SARS-CoV-2 can replicate in the respiratory and gastrointestinal tracts, and cause disease ranging from asymptomatic to severe. The virus may then spread to other organs via the bloodstream. The syncytiotrophoblast, as the outer surface of the placental villi, is bathed in maternal blood, so placental infection may occur. In the case of COVID-19, electron microscopy has documented membrane-bound vesicles filled with virions in the syncytiotrophoblast, which in some cases is necrotic (Birkhead et al., 2021; Hosier et al., 2020).

Transcriptomic analyses have showed that only 3%–6% of lung airway epithelial cells, which are considered the primary route of SARS-CoV-2 lung infection, co-express ACE2 and TMPRSS2 (Ziegler et al., 2020).



snRNA-seq from lungs of fatal COVID-19 cases further supports the low expression of ACE2/TMPRSS2 (Melms et al., 2021). However, these findings could be explained by increased cell death in those cells expressing ACE2/TMPRSS2 and facilitating virus entry.

Other studies have assessed the expression of ACE2 and TMPRSS2 in the different cell types of the placental villi, to evaluate which are permissive to the entry of SARS-CoV-2. One showed that trophoblasts, but not the other main villous cell types, express ACE2 and TMPRSS2, and that these cells are capable of ACE2 endocytosis (Ouyang et al., 2021). ACE2 and TMPRSS2 expression in the first trimester placenta has been also been reported (Weatherbee et al., 2020). Another study using scRNA-seq analysis found that ACE2 expression was abundant throughout gestation. A subset of syncytiotrophoblast and extravillous trophoblast cells (amounting to 14% in first trimester placentas and 15% of second trimester placentas) co-expressed ACE2/TMPRSS2 (Ashary et al., 2020).

OTHER MECHANISMS POTENTIALLY CONTRIBUTING TO THE COMPLICATIONS OF COVID-19 IN PREGNANT WOMEN

A study that offered a mechanistic correlation to severe disease showed that the expression levels of placental ACE2 (but not TMPRSS2 or Furin) and the protein levels of IFITM1 and IFITM3 were higher in women with severe COVID-19 (Mourad et al., 2021). More comprehensive results from scRNA-seq in first trimester pregnant women with and without COVID-19 showed that villous trophoblast cells express low levels of ACE2 and TMPRSS2, but high levels of DDP4 (the MERS-CoV entry mediator) and CTSL, which, according to the authors (Constantino et al., 2021), could be non-canonical cell-entry mediators for SARS-CoV-2. These authors also suggest that changes in expression of the genes DAAM1 and PAICS that code for proteins predicted to interact with SARS-CoV-2 proteins during pregnancy might be of importance. DPP4 was among the scRNA-seq datasets of healthy placentas, expressed in all the cell types of the first trimester placenta and also in extravillous trophoblast in the second trimester (Ashary et al., 2020). CTSL expression was independently shown to be promoted after SARS-CoV-2 infection (Zhao et al., 2021). scRNA-seq analysis comparing healthy controls and COVID-19 cases showed that CTSL expression was more abundant than ACE2 expression in placental trophoblast cells, and this transcription was increased in decidual stromal cells and antigen-presenting cells in COVID-19 pregnancies (Lu-Culligan et al., 2021). This highlights the interaction between SARS-CoV-2 and other placental proteins, suggesting that SARS-CoV-2 may utilize multiple mediators to infect the placenta.

In addition, the detachment of syncytial knots is a source of transcriptionally active soluble fms-like tyrosine kinase-1 (sFlt-1) syncytial aggregates in the maternal circulation. Syncytiotrophoblast-derived syncytial knots, which become multinucleate syncytial aggregates that express the antiangiogenic protein sFlt-1, have previously been detected in the lungs of pregnant women, and the number of syncytial aggregates in the maternal lungs was higher in women with preeclampsia (Buurma et al., 2013). This process of accelerated syncytial knot formation, shedding, and aggregation and delivery of microparticles might contribute to the maternal vascular injury seen in COVID-19 lung pathology. However, the immune cascade potentially triggered by viral syncytia has not been clarified, either in the placenta or in other tissues.

Infection by the SARS-CoV-2 delta variant in pregnant women might induce increased pathogenicity via epigenomic regulation or other non-canonical entry routes. Future research should focus on comparison of the alveolar and placental barriers, cell-to-cell communication, and the cellular and molecular partners involved.

Key to syncytialization is the hypomethylation (Matousková et al., 2006) and expression of the syncytin genes producing syncytin-1 and 2 that are known to derive from human endogenous retroviruses (Dupressoir et al., 2012). Altered expression of syncytin-1 and 2 (Liu et al., 2018) was also correlated with placental pathophysiology, including in preeclampsia (Knerr et al., 2002; Vargas et al., 2011). Further studies are needed to elucidate the epigenome landscape changes during gestation in the SARS-CoV-2-infected placenta and lungs.

Unlike the alveolo-capillary barrier in the lungs, the syncytiotrophoblast expresses little or no caveolin-1 (Mohanty et al., 2010) or caveolin-2 or 3 (Lyden et al., 2001). The role of caveolin-1 in signal transduction could be of importance as it can induce inflammatory cascades via nuclear factor kappa-light-chain-enhancer of activated B cells (NFkB) and leukocyte activation (Celik et al., 2020), and its downregulation was recently documented in immature alveolar endothelial cells in fatal COVID-19 snRNA-analysis (Melms et al., 2021).



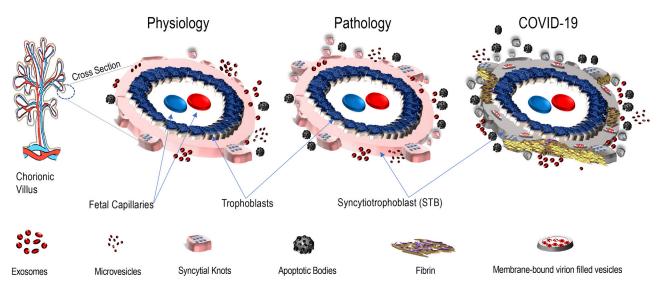


Figure 2. Cytotrophoblasts fuse with the overlying giant multinucleated syncytiotrophoblasts and form the outer layer of the placental microvilli to render the tissue impermeable and enable mother-child immune tolerance

The cytotrophoblasts may regenerate the syncytiotrophoblast if damaged, but if the process is significantly compromised, pathological conditions such as preeclampsia and intra-uterine fetal growth restriction (FGR) might ensue. Increased numbers of syncytial knots, as seen in the placenta under conditions of hypoxia, hyperoxia, or in the presence of reactive oxygen species (ROS), and in preeclampsia, were also observed with COVID-19. There may be small amounts of fibrin deposition in normal placentas, which is increased in hypertensive disorders of pregnancy. But massive intervillous fibrin deposition has also been reported as a hallmark of COVID-19 pregnancies. As systemic SARS-COV-2 infection leads to hypoxia and reduced utero-placental perfusion, it induces focal necrosis and extensive fibrin deposition. This increased amount of intravillous and perivillous fibrin might result from impaired fibrinolytic capacity of the compromised maternal endothelium or from immune cell activation with subsequent pro-coagulation signals. As fibrin deposition further compromises maternal-fetal gas exchange, further research is needed to elucidate the underlying cellular and molecular mechanisms.

Intercellular communication via gap junctions is also necessary for trophoblastic cell fusion and syncytiotrophoblast formation. The involvement of the gap junction protein connexin-43 was immunodetected at the intercellular boundaries between aggregated cells; the expression disappeared after cellular fusion (Frendo et al., 2003). There is scant information on the role of the pannexins in the development of syncytiotrophoblast, but Pannexin-1 (Panx-1) channel opening is known to accelerate viral entry, replication, cellto-cell spread, and inflammation (Malik and Eugenin, 2019). The involvement of Panx-1 in lungs has been assessed in the context of patients with COVID-19 suffering from hyperinflammation (Swayne et al., 2020) and the Panx-1 channel was shown to open in response to SARS-CoV-2 (Luu et al., 2021).

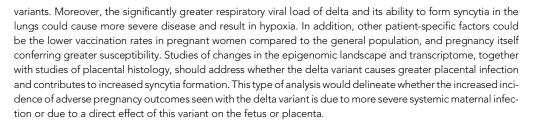
Finally, the extensive syncytial formation due to SARS-CoV-2 infection might contribute to increased viral dissemination and cause greater and more extensive placental tissue damage (Figure 2). Moreover, the role of the increased numbers of circulating placental syncytia, their molecular cargo, and their effect and entrapment in lung vessels should be delineated. This would help further evaluate whether the ability of the delta variant to spread among cells, by direct contact via syncytia without the need for extracellular virion release, could explain how the virus partly escapes existing humoral response (Al-Beltagi et al., 2021). The suggested decreased fusogenicity of the omicron variant may also translate into more favorable pregnancy outcomes following infection (Meng et al., 2022; HKUMed 2022).

Limitations of the study

The definitions of the severity of COVID-19 may vary among investigators. The data presented in this review are limited by the increasing rate of vaccination among pregnant women, the small number of published studies and preprints, and in some cases, the small sample size analyzed and the limited sequencing results in the studies of the pregnant patients.

OUTSTANDING QUESTIONS

During the delta variant wave of the COVID-19 pandemic, the rate of admission of pregnant women to hospital and to ICU was increased compared to the first wave and the alpha wave of the pandemic. This could be due to various variant-specific factors, such as the increased delta variant transmissibility compared to the previous



SEARCH STRATEGY AND SELECTION CRITERIA

Data for this Review were identified by searches of Pubmed/Medline, Current Contents, BiorXiv, and references from relevant articles and preprints, using the search terms "pregnancy", "COVID-19", "delta variant", "omicron", and "syncytia".

AUTHOR CONTRIBUTIONS

Writing original draft AS, Revisions AS, AK, POB, EH. **AS and EH would like to acknowledge** funding by the Swedish Research Council (2019-01157), Region Stockholm (20190400), the Karolinska Institutet and the Swedish Brain Foundation (FO2019-0087).

DECLARATION OF INTEREST

The authors have no interest to declare.

REFERENCES

Adhikari, E.H., SoRelle, J.A., McIntire, D.D., and Spong, C.Y. (2022a). Increasing severity of COVID-19 in pregnancy with Delta (B.1.617.2) variant surge. Am. J. Obstet. Gynecol. 226, 149–151. https://doi.org/10.1016/j.ajog.2021.09. 008.

Adhikari, E.H., MacDonald, L., SoRelle, J.A., Morse, J., Pruszynski, J., and Spong, C.Y. (2022b). COVID-19 cases and disease severity in pregnancy and neonatal positivity associated with delta (B.1.617.2) and omicron (B.1.1.529) variant predominance. JAMA 327, 1500–1502. https://doi.org/10.1001/jama.2022.4356.

Al-Beltagi, S., Goulding, L.V., Chang, D.K.E., Mellits, K.H., Hayes, C.J., Gershkovich, P., Coleman, C.M., and Chang, K.C. (2021). Emergent SARS-CoV-2 variants: comparative replication dynamics and high sensitivity to thapsigargin. Virulence 12, 2946–2956. https:// doi.org/10.1080/21505594.2021.2006960.

Allotey, J., Stallings, E., Bonet, M., Yap, M., Chatterjee, S., Kew, T., Debenham, L., Llavall, A.C., Dixit, A., Zhou, D., et al.; for PregCOV-19 Living Systematic Review Consortium (2020). Clinical manifestations, risk factors, and maternal and perinatal outcomes of coronavirus disease 2019 in pregnancy: living systematic review and meta-analysis. BMJ *370*, m3320. https://doi.org/ 10.1136/bmj.m3320.

Alsaadi, E.A.J., Neuman, B.W., and Jones, I.M. (2019). A fusion peptide in the spike protein of MERS coronavirus. Viruses 11, 825. https://doi. org/10.3390/v11090825.

Arora, P., Sidarovich, A., Krüger, N., Kempf, A., Nehlmeier, I., Graichen, L., Moldenhauer, A.S., Winkler, M.S., Schulz, S., Jäck, H.M., et al. (2021). B.1.617.2 enters and fuses lung cells with increased efficiency and evades antibodies induced by infection and vaccination. Cell Rep. **37**, 109825. https://doi.org/10.1016/j.celrep. 2021.109825.

Ashary, N., Bhide, A., Chakraborty, P., Colaco, S., Mishra, A., Chhabria, K., Jolly, M.K., and Modi, D. (2020). Single-cell RNA-seq identifies cell subsets in human placenta that highly expresses factors driving pathogenesis of SARS-CoV-2. Front. Cell Dev. Biol. *8*, 783. https://doi.org/10.3389/fcell. 2020.00783.

Bailey, A.L., Dmytrenko, O., Greenberg, L., Bredemeyer, A.L., Ma, P., Liu, J., Penna, V., Winkler, E.S., Sviben, S., Brooks, E., et al. (2021). SARS-CoV-2 infects human engineered heart tissues and models COVID-19 myocarditis. JACC Basic Transl. Sci. 6, 331–345. https://doi.org/10. 1016/j.jacbts.2021.01.002.

Bentley, E.G., Kirby, A., Sharma, P., Kipar, A., Mega, D.F., Bramwell, C., Penrice-Randal, R., Prince, T., Brown, J.C., Zhou, J., et al. (2021). SARS-CoV-2 Omicron-B.1.1.529 Variant leads to less severe disease than Pango B and Delta variants strains in a mouse model of severe COVID-19. Preprint at bioRxiv. https://doi.org/10.1101/ 2021.12.26.474085.

Birkhead, M., Glass, A.J., Allan-Gould, H., Goossens, C., and Wright, C.A. (2021). Ultrastructural evidence for vertical transmission of SARS-CoV-2. Int. J. Infect Dis. 111, 10–11. https://doi.org/10.1016/j.ijid.2021.08.020.

Birol Ilter, P., Prasad, S., Berkkan, M., Mutlu, M.A., Tekin, A.B., Celik, E., Ata, B., Turgal, M., Yildiz, S., Turkgeldi, E., et al. (2022). Clinical severity of SARS-CoV-2 infection among vaccinated and unvaccinated pregnancies during the Omicron wave. Ultrasound Obstet. Gynecol. *59*, 560–562. https://doi.org/10.1002/uog.24893.

Braga, L., Ali, H., Secco, I., Chiavacci, E., Neves, G., Goldhill, D., Penn, R., Jimenez-Guardeño, J.M., Ortega-Prieto, A.M., Bussani, R., et al. (2021). Drugs that inhibit TMEM16 proteins block SARS-CoV-2 spike-induced syncytia. Nature 594, 88–93. https://doi.org/10.1038/s41586-021-03491-6.

Buchrieser, J., Dufloo, J., Hubert, M., Monel, B., Planas, D., Rajah, M.M., Planchais, C., Porrot, F., Guivel-Benhassine, F., Van der Werf, S., et al. (2020). Syncytia formation by SARS-CoV-2infected cells. EMBO J. *39*, e106267. https://doi. org/10.15252/embj.2020106267.

Burton, G.J., and Fowden, A.L. (2015). The placenta: a multifaceted, transient organ. Philos. Trans. R. Soc. Lond. B Biol. Sci. *370*, 20140066. https://doi.org/10.1098/rstb.2014.0066.

Bussani, R., Schneider, E., Zentilin, L., Collesi, C., Ali, H., Braga, L., Volpe, M.C., Colliva, A., Zanconati, F., Berlot, G., et al. (2020). Persistence of viral RNA, pneumocyte syncytia and thrombosis are hallmarks of advanced COVID-19 pathology. EBioMedicine *61*, 103104. https://doi. org/10.1016/j.ebiom.2020.103104.

Buurma, A.J., Penning, M.E., Prins, F., Schutte, J.M., Bruijn, J.A., Wilhelmus, S., Rajakumar, A., Bloemenkamp, K.W., Karumanchi, S.A., and Baelde, H.J. (2013). Preeclampsia is associated with the presence of transcriptionally active placental fragments in the maternal lung. Hypertension *62*, 608–613. https://doi.org/10. 1161/HYPERTENSIONAHA.113.01505.

Celik, O., Saglam, A., Baysal, B., Derwig, I.E., Celik, N., Ak, M., Aslan, S.N., Ulas, M., Ersahin, A., Tayyar, A.T., et al. (2020). Factors preventing materno-fetal transmission of SARS-CoV-2. Placenta 97, 1–5. https://doi.org/10.1016/j. placenta.2020.05.012.

Constantino, F.B., Cury, S.S., Nogueira, C.R., Carvalho, R.F., and Justulin, L.A. (2021). Prediction of non-canonical routes for SARS-CoV-2 infection in human placenta cells. Front. Mol.



Biosci. 8, 614728. https://doi.org/10.3389/fmolb. 2021.614728.

DeSisto, C.L., Wallace, B., Simeone, R.M., et al. (2021). Risk for stillbirth among women with and without COVID-19 at delivery hospitalization — United States, march 2020–September 2021. MMWR Morb. Mortal Wkly Rep. 70, 1640–1645. https://doi.org/10.15585/mmwr.mm7047e1external icon.

Donati, S., Corsi, E., Maraschini, A., and Salvatore, M.A.; ItOSS-COVID-19 Working Group (2022). SARS-CoV-2 infection among hospitalised pregnant women and impact of different viral strains on COVID-19 severity in Italy: a national prospective population-based cohort study. BJOG 129, 221–231. https://doi.org/10.1111/ 1471-0528.16980.

Dupressoir, A., Lavialle, C., and Heidmann, T. (2012). From ancestral infectious retroviruses to bona fide cellular genes: role of the captured syncytins in placentation. Placenta 33, 663–671. https://doi.org/10.1016/j.placenta.2012.05.005.

Engjom, H., van den Akker, T., Aabakke, A., Ayras, O., Bloemenkamp, K., Donati, S., Cereda, D., Overtoom, E., and Knight, M. (2022). Severe COVID-19 in pregnancy is almost exclusively limited to unvaccinated women - time for policies to change. Lancet Reg. Health Eur. 13, 100313. https://doi.org/10.1016/j.lanepe.2022.100313.

Fabre, M., Calvo, P., Ruiz-Martinez, S., Peran, M., Oros, D., Medel-Martinez, A., Strunk, M., Benito Ruesca, R., Schoorlemmer, J., and Paules, C. (2021). Frequent placental SARS-CoV-2 in patients with COVID-19-associated hypertensive disorders of pregnancy. Fetal Diagn. Ther. 48, 801–811. https://doi.org/10.1159/000520179.

Fox, H. (1967). Perivillous fibrin deposition in the human placenta. Am. J. Obstet. Gynecol. *98*, 245–251. https://doi.org/10.1016/s0002-9378(16) 34594-x.

Frendo, J.L., Cronier, L., Bertin, G., Guibourdenche, J., Vidaud, M., Evain-Brion, D., and Malassine, A. (2003). Involvement of connexin 43 in human trophoblast cell fusion and differentiation. J. Cell Sci. 116, 3413–3421. https://doi.org/10.1242/jcs.00648.

Gao, L., Ren, J., Xu, L., Ke, X., Xiong, L., Tian, X., Fan, C., Yan, H., and Yuan, J. (2021). Placental pathology of the third trimester pregnant women from COVID-19. Diagn. Pathol. 16, 8. https://doi. org/10.1186/s13000-021-01067-6.

Guan, M., Johannesen, E., Tang, C.Y., Hsu, A.L., Barnes, C.L., Burnam, M., McElroy, J.A., and Wan, X.F. (2022). Intrauterine fetal demise in the third trimester of pregnancy associated with mild infection with the SARS-CoV-2 Delta variant without protection from vaccination. J. Infect Dis. 225, 748–753. jiac007. https://doi.org/10.1093/ infdis/jiac007.

Heazell, A.E., Moll, S.J., Jones, C.J., Baker, P.N., and Crocker, I.P. (2007). Formation of syncytial knots is increased by hyperoxia, hypoxia and reactive oxygen species. Placenta 28 (Suppl A), S33–S40. Erratum in: Placenta. 2007 Aug-Sep;28(8-9):973. https://doi.org/10.1016/j. placenta.2006.10.007.

HKUMed (2022). HKUMed finds omicron SARS-CoV-2 can infect faster and better than delta in human bronchus but with less severe infection in lung. https://www.med.hku.hk/en/news/press/ 20211215-omicron-sars-cov-2-infection.

Hosier, H., Farhadian, S.F., Morotti, R.A., Deshmukh, U., Lu-Culligan, A., Campbell, K.H., Yasumoto, Y., Vogels, C.B., Casanovas-Massana, A., Vijayakumar, P., et al. (2020). SARS-CoV-2 infection of the placenta. J. Clin. Invest. 130, 4947–4953. https://doi.org/10.1172/JCl139569.

Huppertz, B., and Kingdom, J.C. (2004). Apoptosis in the trophoblast-role of apoptosis in placental morphogenesis. J. Soc. Gynecol. Investig. 11, 353–362. https://doi.org/10.1016/j. jsgi.2004.06.002.

Husen, M.F., van der Meeren, L.E., Verdijk, R.M., Fraaij, P.L.A., van der Eijk, A.A., Koopmans, M.P.G., Freeman, L., Bogers, H., Trietsch, M.D., Reiss, I.K.M., et al. (2021). Unique severe COVID-19 placental signature independent of severity of clinical maternal symptoms. Viruses 13, 1670. https://doi.org/10.3390/v13081670.

Jaiswal, N., Puri, M., Agarwal, K., Singh, S., Yadav, R., Tiwary, N., Tayal, P., and Vats, B. (2021). COVID-19 as an independent risk factor for subclinical placental dysfunction. Eur. J. Obstet. Gynecol. Reprod. Biol. 259, 7-11. https://doi.org/ 10.1016/j.ejogrb.2021.01.049.

Kasehagen, L., Byers, P., Taylor, K., Kittle, T., Roberts, C., Collier, C., Rust, B., Ricaldi, J.N., Green, J., Zapata, L.B., et al. (2021). COVID-19-Associated deaths after SARS-CoV-2 infection during pregnancy - Mississippi, March 1, 2020-October 6, 2021. MMWR Morb. Mortal Wkly Rep. 70, 1646– 1648. https://doi.org/10.15585/mmwr.mm7047e2.

Knerr, I., Beinder, E., and Rascher, W. (2002). Syncytin, a novel human endogenous retroviral gene in human placenta: evidence for its dysregulation in preeclampsia and HELLP syndrome. Am. J. Obstet. Gynecol. 186, 210–213. https://doi.org/10.1067/mob.2002.119636.

Knight, M., Ramakrishnan, R., Bunch, K., Vousden, N., J Kurinczuk, J.J., Dunn, S., Norman, L., Aisling Barry, A., Harrison, E., Docherty, A., and Semple, C. (2021). Females in hospital with SARS-CoV-2 infection, the association with pregnancy and pregnancy outcomes: a UKOSS/ISARIC/CO-CIN Investigation Gov.UK. https://assets.publishing. service.gov.uk/government/uploads/system/ uploads/attachment_data/file/977287/s1171ukoss-isaric-co-cin-covid-19-young-femalespregnancy-report.pdf.

Li, W., Moore, M.J., Vasilieva, N., Sui, J., Wong, S.K., Berne, M.A., Somasundaran, M., Sullivan, J.L., Luzuriaga, K., Greenough, T.C., et al. (2003). Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature 426, 450–454. https://doi.org/10.1038/nature02145.

Lin, L., Liu, Y., Tang, X., and He, D. (2021). The disease severity and clinical outcomes of the SARS-CoV-2 variants of concern. Front. Public Health 9, 775224. https://doi.org/10.3389/fpubh. 2021.775224.

Liu, Y., Fan, X., Wang, R., Lu, X., Dang, Y.L., Wang, H., Lin, H.Y., Zhu, C., Ge, H., Cross, J.C., and Wang, H. (2018). Single-cell RNA-seq reveals the diversity of trophoblast subtypes and patterns of differentiation in the human placenta. Cell Res. 28, 819–832. https://doi.org/10.1038/s41422-018-0066-v. Lu-Culligan, A., Chavan, A.R., Vijayakumar, P., Irshaid, L., Courchaine, E.M., Milano, K.M., Tang, Z., Pope, S.D., Song, E., Vogels, C.B.F., Lu-Culligan, W.J., Campbell, K.H., Casanovas-Massana, A., Bermejo, S., Toothaker, J.M., Lee, H.J., Liu, F., Schulz, W., Fournier, J., Muenker, M.C., Moore, A.J., Yale IMPACT Team, Konnikova, L., Neugebauer, K.M., Ring, A., Grubaugh, N.D., Ko, A.I., Morotti, R., Guller, S., Kliman, H.J., Iwasaki, A., and Farhadian, S.F. (2021). Maternal respiratory SARS-CoV-2 infection in pregnancy is associated with a robust inflammatory response at the maternal-fetal interface. Preprint at medRxiv 2, 591–610.e10. https:// doi.org/10.1016/j.medj.2021.04.016.

Luo, C.H., Morris, C.P., Sachithanandham, J., Amadi, A., Gaston, D., Li, M., Swanson, N.J., Schwartz, M., Klein, E.Y., Pekosz, A., and Mostafa, H.H. (2021). Infection with the SARS-CoV-2 delta variant is associated with higher infectious virus loads compared to the alpha variant in both unvaccinated and vaccinated individuals. Preprint at medRxiv. Update in: Clin Infect Dis. 2021 Dec 18: PMID: 34462756; PMCID: PMC8404894. https://doi.org/10.1101/2021.08.15.21262077.

Luu, R., Valdebenito, S., Scemes, E., Cibelli, A., Spray, D.C., Rovegno, M., Tichauer, J., Cottignies-Calamarte, A., Rosenberg, A., Capron, C., et al. (2021). Pannexin-1 channel opening is critical for COVID-19 pathogenesis. iScience 24, 103478. https://doi.org/10.1016/j. isci.2021.103478.

Lyden, T.W., Robinson, J.M., Tridandapani, S., Teillaud, J.L., Garber, S.A., Osborne, J.M., Frey, J., Budde, P., and Anderson, C.L. (2001). The Fc receptor for IgG expressed in the villus endothelium of human placenta is Fc gamma RIIb2. J. Immunol. 166, 3882–3889. https://doi. org/10.4049/jimmunol.166.6.3882.

Malik, S., and Eugenin, E.A. (2019). Role of connexin and pannexin containing channels in HIV infection and NeuroAIDS. Neurosci. Lett. 695, 86–90. https://doi.org/10.1016/j. neulet.2017.09.005.

Mamun, M.M.A., and Khan, M.R. (2021). COVID-19 delta variant-of-concern: a real concern for pregnant women with gestational diabetes mellitus. Front. Endocrinol. (Lausanne) 12, 778911. https://doi.org/10.3389/fendo.2021. 778911.

Mao, Q., Chu, S., Shapiro, S., Young, L., Russo, M., and De Paepe, M.E. (2022). Placental SARS-CoV-2 distribution correlates with level of tissue oxygenation in COVID-19-associated necrotizing histiocytic intervillositis/perivillous fibrin deposition. Placenta 117, 187–193. https://doi.org/10. 1016/j.placenta.2021.12.002.

Matousková, M., Blazková, J., Pajer, P., Pavlícek, A., and Hejnar, J. (2006). CpG methylation suppresses transcriptional activity of human syncytin-1 in non-placental tissues. Exp. Cell Res. 312, 1011–1020. https://doi.org/10.1016/j.yexcr. 2005.12.010.

Matsuyama, S., Nao, N., Shirato, K., Kawase, M., Saito, S., Takayama, I., Nagata, N., Sekizuka, T., Katoh, H., Kato, F., et al. (2020). Enhanced isolation of SARS-CoV-2 by TMPRSS2-expressing cells. Proc. Natl. Acad. Sci. U S A *117*, 7001–7003. https://doi.org/10.1073/pnas.2002589117.

iScience Review



Matsuyama, S., Nagata, N., Shirato, K., Kawase, M., Takeda, M., and Taguchi, F. (2010). Efficient activation of the severe acute respiratory syndrome coronavirus spike protein by the transmembrane protease TMPRSS2. J. Virol. *84*, 12658–12664. https:// doi.org/10.1128/JVI.01542-10.

Mehta, P., McAuley, D.F., Brown, M., Sanchez, E., Tattersall, R.S., and Manson, J.J.; HLH Across Speciality Collaboration, UK (2020). COVID-19: consider cytokine storm syndromes and immunosuppression. Lancet *395*, 1033–1034. https://doi.org/10.1016/S0140-6736(20)30628-0.

Melms, J.C., Biermann, J., Huang, H., Wang, Y., Nair, A., Tagore, S., Katsyv, I., Rendeiro, A.F., Amin, A.D., Schapiro, D., et al. (2021 Jul). A molecular single-cell lung atlas of lethal COVID-19. Nature 595, 114–119. Erratum in: Nature. 2021 Oct;598(7882):E2. https://doi.org/10.1038/ s41586-021-03569-1.

Meng, B., Abdullahi, A., Ferreira, I.A.T.M., Goonawardane, N., Saito, A., Kimura, I., Yamasoba, D., Gerber, P.P., Fatihi, S., Rathore, S., et al. (2022). Altered TMPRSS2 usage by SARS-CoV-2 omicron impacts infectivity and fusogenicity. Nature 603, 706–714. https://doi.org/10. 1038/s41586-022-04474-x.

Mohanty, S., Anderson, C.L., and Robinson, J.M. (2010). The expression of caveolin-1 and the distribution of caveolae in the murine placenta and yolk sac: parallels to the human placenta. Placenta 31, 144–150. https://doi.org/10.1016/j. placenta.2009.11.007.

Mourad, M., Jacob, T., Sadovsky, E., Bejerano, S., Simone, G.S., Bagalkot, T.R., Zucker, J., Yin, M.T., Chang, J.Y., Liu, L., et al. (2021). Placental response to maternal SARS-CoV-2 infection. Sci. Rep. 11, 14390. https://doi.org/10.1038/s41598-021-93931-0.

Ouyang, Y., Bagalkot, T., Fitzgerald, W., Sadovsky, E., Chu, T., Martínez-Marchal, A., Brieňo-Enríquez, M., Su, E.J., Margolis, L., Sorkin, A., and Sadovsky, Y. (2021). Term human placental trophoblasts express SARS-CoV-2 entry factors ACE2, TMPRSS2, and Furin. mSphere 6. e00250-21. https://doi.org/10.1128/mSphere. 00250-21.

Overtoom, E., Rosman, A., Zwart, J., et al. (2021). SARS-CoV-2 en zwangerschap in Nederland: registratie tijdens een pandemie door NethOSS. Nederlands Tijdschrift voor Obstetrie & Gynaecologie 134, 406–409.

Peacock, T.P., Brown, J.C., Zhou, J., Thakur, N., Kugathasan, R., Sukhova, K., Kforou, M., Bailey, D., and Barclay, W.S. (2022). The SARS-CoV-2 variant, Omicron, shows rapid replication in human primary nasal epithelial cultures and efficiently uses the endosomal route of entry. Preprint at bioRxiv. https://www.biorxiv.org/ content/10.1101/2021.12.31.474653v1.

Pötgens, A.J., Schmitz, U., Bose, P., Versmold, A., Kaufmann, P., and Frank, H.G. (2002). Mechanisms of syncytial fusion: a review. Placenta 23 (Suppl A), S107–S113. https://doi.org/10. 1053/plac.2002.0772.

Rangchaikul, P., and Venketaraman, V. (2021). SARS-CoV-2 and the immune response in pregnancy with delta variant considerations. Infect Dis. Rep. 13, 993–1008. https://doi.org/10. 3390/idr13040091.

Rajah, M.M., Bernier, A., Buchrieser, J., and Schwartz, O. (2021). The mechanism and consequences of SARS-CoV-2 spike-mediated fusion and syncytia formation. J. Mol. Biol. 434, 167280. https://doi.org/10.1016/j.jmb.2021. 167280.

Redline, R.W., and Patterson, P. (1995). Preeclampsia is associated with an excess of proliferative immature intermediate trophoblast. Hum. Pathol. 26, 594–600. https://doi.org/10. 1016/0046-8177(95)90162-0.

Robbins, J.R., and Bakardjiev, A.I. (2012). Pathogens and the placental fortress. Curr. Opin. Microbiol. 15, 36–43. https://doi.org/10.1016/j. mib.2011.11.006.

Sanders, D.W., Jumper, C.C., Ackerman, P.J., Bracha, D., Donlic, A., Kim, H., Kenney, D., Castello-Serrano, I., Suzuki, S., Tamura, T., et al. (2021). SARS-CoV-2 requires cholesterol for viral entry and pathological syncytia formation. Elife 10, e65962. https://doi.org/10.7554/eLife.65962.

Sankar, K.D., Bhanu, P.S., Kiran, S., Ramakrishna, B.A., and Shanthi, V. (2012). Vasculosyncytial membrane in relation to syncytial knots complicates the placenta in preeclampsia: a histomorphometrical study. Anat. Cell Biol. 45, 86–91. https://doi.org/10.5115/acb.2012.45.2.86.

Shchegolev, A.I., Kulikova, G.V., Lyapin, V.M., Shmakov, R.G., and Sukhikh, G.T. (2021). The number of syncytial knots and VEGF expression in placental villi in parturient woman with COVID-19 depends on the disease severity. Bull Exp. Biol. Med. 171, 399–403. https://doi.org/10.1007/ s10517-021-05236-x.

Shen, C.J., Fu, Y.C., Lin, Y.P., Shen, C.F., Sun, D.J., Chen, H.Y., and Cheng, C.M. (2022). Evaluation of Transplacental Antibody Transfer in SARS-CoV-2-Immunized Pregnant Women. Vaccines (Basel) 10, 101. https://doi.org/10.3390/ vaccines10010101.

Shook, L.L., Brigida, S., Regan, J., Flynn, J.P., Mohammadi, A., Etemad, B., Siegel, M.R., Clapp, M.A., Li, J.Z., Roberts, D.J., and Edlow, A.G. (2022). SARS-CoV-2 placentitis associated with B.1.617.2 (Delta) variant and fetal distress or demise. J. Infect Dis. 225, 754–758. jiac008. https://doi.org/10.1093/infdis/jiac008.

Schwartz, D.A., Baldewijns, M., Benachi, A., Bugatti, M., Collins, R.R.J., De Luca, D., Facchetti, F., Linn, R.L., Marcelis, L., Morotti, D., et al. (2021). Chronic histiocytic intervillositis with trophoblast necrosis is a risk factor Associated with placental infection from coronavirus disease 2019 (COVID-19) and intrauterine maternal-fetal severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) transmission in live-born and stillborn infants. Arch. Pathol. Lab Med. 145, 517–528. https://doi. org/10.5858/arpa.2020-0771-SA.

Schwartz, D.A., and Morotti, D. (2020). Placental pathology of COVID-19 with and without fetal and neonatal infection: trophoblast necrosis and chronic histiocytic intervillositis as risk factors for transplacental transmission of SARS-CoV-2. Viruses 12, 1308. https://doi.org/10.3390/ vi2111308.

Seasely, A.R., Blanchard, C.T., Arora, N., Battarbee, A.N., Casey, B.M., Dionne-Odom, J., Leal, S.M., et al. (2021). MPH, on behalf of the CWRH COVID-19 working group maternal and perinatal outcomes associated with the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) delta (B.1.617.2) variant. Obstet. Gynecol. 138, 842–844. https://doi.org/10.1097/AOG. 0000000000004607.

Gychka, S.G., Kuchyn, I.L., Savchuk, T.V., Nikolaienko, S.I., Zhezhera, V.M., Chermak, I.I., and Suzuki, Y.J. (2021). Placental vascular remodeling in pregnant women with COVID-19. Preprint at medRxiv. https://doi.org/10.1101/ 2021.07.01.21259860.

Singh, N., Buckley, T., and Shertz, W. (2021). Placental pathology in COVID-19: case series in a community hospital setting. Cureus 13, e12522. PMID: 3564526; PMCID: PMC7863052. https:// doi.org/10.7759/cureus.12522.

Swayne, L.A., Johnstone, S.R., Ng, C.S., Sanchez-Arias, J.C., Good, M.E., Penuela, S., Lohman, A.W., Wolpe, A.G., Laubach, V.E., Koval, M., and Isakson, B.E. (2020). Consideration of Pannexin 1 channels in COVID-19 pathology and treatment. Am. J. Physiol. Lung Cell Mol Physiol. *319*, L121– L125. https://doi.org/10.1152/ajplung.00146. 2020.

Suzuki, R., Yamasoba, D., Kimura, I., Wang, L., Kishimoto, M., Ito, J., Morioka, Y., Nao, N., Nasser, H., Uriu, K., Kosugi, Y., Tsuda, M., Orba, Y., Sasaki, M., Shimizu, R., Kawabata, R., Yoshimatsu, K., Asakura, H., Nagashima, M., Sadamasu, K., Yoshimura, K., Genotype to phenotype Japan (G2P-Japan) consortium, Sawa, H., Ikeda, T., Irie, T., Matsuno, K., Tanaka, S., Fukuhara, T., and Sato, K. (2022). Attenuated fusogenicity and pathogenicity of SARS-CoV-2 omicron variant. Nature *603*, 700–705. https://doi. ora/10.1038/s41586-022-04462-1.

UKOSS MBRRACE Infographic v13, 2021, UKOSS Infographic MBRRACE-UK_Rapid_COVID_19_ DEC_2021_-_Infographic_v13.Pdf (ox.ac.uk). https://www.npeu.ox.ac.uk/assets/downloads/ npeu-news/MBRRACE-UK_Rapid_COVID_ 19_DEC_2021_-_Infographic_v13.pdf.

Vargas, A., Toufaily, C., LeBellego, F., Rassart, É., Lafond, J., and Barbeau, B. (2011). Reduced expression of both syncytin 1 and syncytin 2 correlates with severity of preeclampsia. Reprod. Sci. 18, 1085–1091. https://doi.org/10.1177/ 1933719111404608.

Villar, J., Ariff, S., Gunier, R.B., Thiruvengadam, R., Rauch, S., Kholin, A., Roggero, P., Prefumo, F., do Vale, M.S., Cardona-Perez, J.A., et al. (2021). Maternal and neonatal morbidity and mortality among pregnant women with and without COVID-19 infection: the INTERCOVID multinational cohort study. JAMA Pediatr. *175*, 817–826. Erratum in: JAMA Pediatr. 2022 Jan 1;176(1):104. https://doi.org/10.1001/jamapediatrics.2021. 1050.

Vousden, N., Ramakrishnan, R., Bunch, K., Morris, E., Simpson, N., Gale, C., O'Brien, P., Quigley, M., Brocklehurst, P., Kurinczuk, J.J., and Knight, M. (2021). Impact of SARS-CoV-2 variant on the severity of maternal infection and perinatal outcomes: data from the UK Obstetric Surveillance System national cohort. Preprint at medRxiv. https://doi.org/10.1101/2021.07.22. 21261000.







Vousden, N., Ramakrishnan, R., Bunch, K., Morris, E., Simpson, N.A.B., Gale, C., O'Brien, P., Quigley, M., Brocklehurst, P., Kurinczuk, J.J., and Knight, M. (2022). Severity of maternal infection and perinatal outcomes during periods of SARS-CoV-2 wildtype, alpha, and delta variant dominance in the UK: prospective cohort study. BMJ Med. 1, e000053. https://doi.org/10.1136/ bmjmed-2021-000053.

Weatherbee, B.A., Glover, D.M., and Zernicka-Goetz, M. (2020). Expression of SARS-CoV-2 receptor ACE2 and the protease TMPRSS2 suggests susceptibility of the human embryo in the first trimester. Open Biol. 10, 200162. https://doi. org/10.1098/rsob.200162.

Willett, B.J., Grove, J., MacLean, O.A., Wilkie, C., Logan, N., De Lorenzo, G., Furnon, W., Scott, S., Manali, M., Szemiel, A., et al. (2022). The hypertransmissible SARS-CoV-2 omicron variant exhibits significant antigenic change, vaccine escape and a switch in cell entry mechanism. Preprint at medRxiv. https://www.medrxiv.org/ content/10.1101/2022.01.03.21268111v1.

Xia, S., Liu, M., Wang, C., Xu, W., Lan, Q., Feng, S., Qi, F., Bao, L., Du, L., Liu, S., et al. (2020). Inhibition of SARS-CoV-2 (previously 2019-nCoV) infection by a highly potent pan-coronavirus fusion inhibitor targeting its spike protein that harbors a high capacity to mediate membrane fusion. Cell Res. 30, 343–355. https://doi.org/10. 1038/s41422-020-0305-x.

Zaigham, M., and Andersson, O. (2020). Maternal and perinatal outcomes with COVID-19: a systematic review of 108 pregnancies. Acta Obstet. Gynecol. Scand. *99*, 823–829. Zhao, M.M., Yang, W.L., Yang, F.Y., Zhang, L., Huang, W.J., Hou, W., Fan, C.F., Jin, R.H., Feng, Y.M., Wang, Y.C., and Yang, J.K. (2021). Cathepsin L plays a key role in SARS-CoV-2 infection in humans and humanized mice and is a promising target for new drug development. Signal Transduct. Target Ther. *6*, 134. https://doi. org/10.1038/s41392-021-00558-8.

Ziegler, C.G.K., Allon, S.J., Nyquist, S.K., Mbano, I.M., Miao, V.N., Tzouanas, C.N., Cao, Y., Yousif, A.S., Bals, J., Hauser, B.M., et al.; HCA Lung Biological Network (2020). Electronic address: lung-network@humancellatlas.org; HCA lung biological network. SARS-CoV-2 receptor ACE2 is an interferon-stimulated gene in human airway epithelial cells and is detected in specific cell subsets across tissues. Cell 181, 1016–1035. https://doi.org/10.1016/j.cell.2020.04.035.