

Supplementary Material

Supplementary Methods.....	2
Supplementary Table 1: <i>SCN</i> Functionally Characterized Variants.....	5
Supplementary Table 2: Corresponding variants, phenotypes, and function across different sodium channels	51
Supplementary Table 3: Detailed <i>SCNI-11A</i> Analysis.....	55
Supplementary Figure 1: Study selection	58
Supplementary Figure 2: Voltage sensing regions (S4) structure zoom across D1-D4.....	59

Supplementary Methods

Functional classification of mutations:

For each mutation described we assessed the overall effect with respect to channel function in terms of net current flow during voltage steps. Channel function is not necessarily the same as overall effect on *cellular* function as elegantly shown recently in Liu et al., 2019,¹ where a gain in function for the channel, led to a paradoxical loss of function through depolarising block at the cellular level. As the vast majority of channels are characterised in non-excitabile mammalian cell lines in voltage clamp only, we have used this baseline to allow comparisons between published studies. Assessing the net effect where a mutation can have multiple effects on channels, we have looked at different parameters in the following approach:

1. Peak currents: we first checked whether a mutation reduced or increased the peak current substantially. Where data allow (e.g. for FHM3 mutations in Cestele et al., 2013)² we have also asked whether these effects on peak current are dependent on expression system in a way that means they may not be physiologically relevant in the endogenous cells. If there are data suggesting the effects on peak current are dependent on system, we have noted this as ‘mixed’ in the first instance.
2. Voltage dependence of inactivation: as the channels must be able to activate in order to pass current, we have asked whether there were shifts in the voltage dependence of inactivation that suggest a substantial portion of channels would be stuck in inactivated states in resting cells at ~-70 or -80 mV. In a very few cases we have suggested that were a large shift in voltage dependence of inactivation would effectively make channels non-functional, we have indicated this is likely an overall LoF effect, even if the peak current is increased and voltage dependence of activation (see #3) was also shifted (we gave inactivation precedence as the channels would be unable to open in resting cells if inactivated at resting potentials).
3. Voltage dependence of activation: given functional channels (i.e. peak current is not lost) that are not stuck in inactivated states at rest (i.e. voltage dependence of inactivation not shifted so all channels are inactive at -80 mV), we asked how much stimulation is required to open channels – that is to say, is the voltage dependence of activation shifted significantly? Here a shift to more

negative voltage dependences of activation would be a gain of function, but again – only if there are sufficient currents, and a significant population of non-inactivated channels.

4. Persistent currents: These can be increased, sometimes even when peak currents are reduced. Increases in persistent currents are included as gain of function, unless peak currents or voltage dependence of inactivation are shifted so much that channels are unlikely to be open at all, in which case, given the profound effects of sodium leaks on cellular activity we have still called cases with large increases in persistent currents ‘mixed’.

5. Gating pore currents: Few groups using neuronal channels have been able to interrogate whether in the cases of S4 arginines these include gating pore leak, we have not rigorously separated gating pore leaks (but see R853Q in SCN2A for one example where these currents have been seen in neuronal channels; Mason et al., 2019)³. Where gating pore leaks are described, these are a gain of function (and one that is highly likely to be pathogenic in any cell). Further technically challenging work on neuronal channels will be needed to confirm the conservation of these gating pore currents across channel subtypes.

6. There are many additional features that can come into play, and where the above 5 criteria do not give an overriding answer, we have looked at these. For example, many groups also look at recovery from inactivation, but in our survey, this was rarely changed enough to be the dominating feature of a channel’s functional change. However, where there were large impacts on recovery from inactivation and these were in contrast to other effects on the channel, we have given the results as mixed.

In practice this functional hierarchy, albeit simplistic and incomplete, typically agrees with the author assessments of the channels, with rare exceptions. We emphasise that this functional assessment is from the perspective of the channel, not the cell or the organism (indeed as these are mutations associated with diseases, they would all be loss of function from the organism’s perspective).

Additional studies, including dynamic clamp, expression in excitable cells and interrogating current clamp properties, modelling, and the golden standard of knock-in studies in mice, all will bring additional insights to the consequences of the mutations on the cells, networks and behaviours, but for the purposes of comparing the effects of mutations across different channels we have relied on the reductionist, most commonly used experimental approach. It remains to be

seen whether functional effects that alter, for example channel trafficking, are conserved in different types of sodium channels expressed in different cellular backgrounds. In addition, some effects, for example where the different threshold of activation of SCN9A has specific effects on cellular activity (in this case in DRG neurons, Dib-Hajj et al., 2012)⁴ which are highly unlikely to be conserved in (for example) SCN4A in muscle cells. In these cases, the reductionist effect on voltage dependence of activation may be conserved, but the cellular effect divergent.

References

1. Liu Y, Schubert J, Sonnenberg L, et al. Neuronal mechanisms of mutations in SCN8A causing epilepsy or intellectual disability. *Brain*. 2019;142(2):376-390.
2. Cestèle S, Schiavon E, Rusconi R, Franceschetti S, Mantegazza M. Nonfunctional NaV1.1 familial hemiplegic migraine mutant transformed into gain of function by partial rescue of folding defects. *Proc Natl Acad Sci USA*. 2013;110(43):17546-17551.
3. Mason ER, Wu F, Patel RR, Xiao Y, Cannon SC, Cummins TR. Resurgent and Gating Pore Currents Induced by De Novo SCN2A Epilepsy Mutations. *eNeuro*. 2019;6(5)
4. Dib-Hajj SD, Yang Y, Black JA, Waxman SG. The NaV1.7 sodium channel: from molecule to man. *Nature Reviews Neuroscience*. 2013;14(1):49-62.

Supplementary Table 1: SCN Functionally Characterized Variants

No.	Gene	Variant	Overall effect	Reference for function	Primary disease	Reference for phenotype	SCN1A Equivalent	gnomAD frequency	Conservation
1-128 Cytoplasmic segment									
1	SCN5A	G9V	STW	Glazer (2020) ¹	LQT3	Millat (2006) ² Gutter (2013) ³	G10V	-	NC
2	SCN9A	Q10R	GoF	Han (2009) ⁴	IEM	Han (2009) ⁴	D12R	1.29e-4	NC
3	SCN2A	D12N	LoF	Ben-Shalom (2017) ⁵	ASD	Ben-Shalom (2017) ⁵	D12N	-	NC
4	SCN5A	R18W	GoF	Gutter (2013) ³	LQT3	Tester (2005) ⁶	R19W	2.57e-4	NC
5	SCN5A	R27H	LoF	Gutter (2013) ³	BrS	Priori (2002) ⁷	R28H	2.36e-4	NC
6	SCN5A	E30G	STW	Kapplinger (2015) ⁸	LQT3	Kapplinger (2015) ⁸	E31G	-	NC
7	SCN5A	R43Q	STW	Lin (2008) ⁹	LQT3	Lin (2008) ⁹	K41Q	4.85e-5	NC
8	SCN1A	E78D	LoF	Kluckova (2020) ¹⁰	EPI	Mancardi (2006) ¹¹	E78D	-	-
9	SCN5A	D84N	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	D81N	-	-
10	SCN2A	D82G	LoF	Ben-Shalom (2017) ⁵	ASD	Ben-Shalom (2017) ⁵	D81G	-	-
11	SCN5A	F93S	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	F90S	-	-
12	SCN10A	V94G	LoF	Jabbari (2015) ¹³	AF	Jabbari (2015) ¹³	V92G	-	-
13	SCN4A	R104H	LoF	Zaharieva (2016) ¹⁴	CMS	Zaharieva (2016) ¹⁴	R101H	4.01e-6	-
14	SCN5A	R104Q	LoF	Gutter (2013) ³	BrS	Levy-Nissenbaum (2001) ¹⁵	R101Q	-	-
15	SCN5A	R104W	LoF	Clatot (2012) ¹⁶	BrS	Clatot (2012) ¹⁶	R101W	4.01e-6	-
16	SCN5A	N109K	STW	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	S106K	4.01e-6	NC
17	SCN5A	R121W	LoF	Clatot (2012) ¹⁶ Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Holst (2010) ¹⁷	R118W	-	-
18	SCN5A	A124D	LoF	Moreau (2012) ¹⁸	BrS	Moreau (2012) ¹⁸	A121D	-	NC
19	SCN5A	V125L	GoF	Gutter (2013) ³	LQT3	Kapplinger (2009) ¹⁹	I122L	2.17e-4	NC

20	SCN5A	K126E	LoF	Gutter (2013) ³	BrS	Gutter (2013) ³	K123E	-	NC
129-147 S1 of D1									
21	SCN5A	L136P	LoF	Glazer (2020) ¹	BrS	Yokokawa (2007) ²⁰ Yamagata (2017) ²¹	L133P	-	NC
22	SCN8A	M139I	GoF	Zaman (2019) ²²	EPI	Zaman (2019) ²²	M135I	-	NC
23	SCN4A	I141V	GoF	Petitprez (2008) ²³	PMC	Petitprez (2008) ²³	I138V	-	NC
24	SCN9A	I136V	GoF	Cheng (2008) ²⁴	IEM	Lee (2007) ²⁵		-	NC
25	SCN1A	M145T	LoF	Mantegazza (2005) ²⁶	EPI	Mantegazza (2005) ²⁶	M145T	-	-
148-154 Extracellular									
155-175 S2 of D1									
26	SCN5A	E161K	LoF	Smits (2005a) ²⁷	BrS	Smits (2005a) ²⁷	E158K	4.15e-6	-
27	SCN10A	Y158D	GoF	Savio-Galimberti (2014) ²⁸	AF	Savio-Galimberti (2014) ²⁸	Y159D	2.58e-4	NC
28	SCN5A	K175N	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	K172N	-	-
29	SCN5A	A178G	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	A175G	-	-
176-189 Cytoplasmic									
30	SCN1A	G177A	LoF	Nissenkorn (2019) ²⁹	EPI	Nissenkorn (2019) ²⁹	G177A	-	-
31	SCN1A	G177E	LoF	Ohmori (2006) ³⁰	EPI	Ohmori (2006) ³⁰	G177E	-	-
32	SCN5A	T187I	LoF	Makiyama (2005) ³¹	BrS	Makiyama (2005) ³¹	T184I	-	NC
33	SCN2A	R188W	Mixed	Sugawara (2001) ³²	EPI	Sugawara (2001) ³²	R187W	1.99e-5	-
190-207 S3 of D1									
34	SCN4A	M203K	LoF	Zaharieva (2016) ¹⁴	CMS	Zaharieva (2016) ¹⁴	F200K	7.99e-6	NC
35	SCN2A	V208E	GoF	Lauxmann (2018) ³³	EPI	Lauxmann (2018) ³³ Lemke (2012) ³⁴	V207E	-	NC
208-213 Extracellular									

36	SCN5A	S216L	Mixed	Marangoni (2011) ³⁵ Wang (2007) ³⁶	LQT3; BrS	Marangoni (2011) ³⁵ Wang (2007) ³⁶	S213L	8.09e-5	NC
37	SCN9A	S211P	GoF	Estacion (2010) ³⁷	IEM	Estacion (2010) ³⁷	S213P	-	NC
214-230 S4 of D1									
38	SCN11A	R222H	GoF	Han (2017) ³⁸	PPN	Han (2017) ³⁸ Okuda (2016) ³⁹	R216H	-	-
39	SCN5A	T220I	STW	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	T217I	7.14e-4	NC
40	SCN9A	F216S	GoF	Choi (2006) ⁴⁰ Sheets (2007) ⁴¹	IEM	Drenth (2005) ⁴² Kim (2013) ⁴³	F218S	-	-
41	SCN8A	R223G	LoF	De Kovel (2014) ⁴⁴	EPI	De Kovel (2014) ⁴⁴	R219G	-	-
42	SCN5A	V223L	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	V220L	-	-
43	SCN4A	R225W	LoF	Zaharieva (2016) ¹⁴	CMS	Zaharieva (2016) ¹⁴	R222W	-	-
44	SCN5A	R225W	LoF	Bezzina (2003) ⁴⁵	LQT3; BrS	Kapplinger (2009) ¹⁹ Kapplinger (2010) ¹²		-	-
45	SCN5A	R225P	GoF	Beckermann (2014) ⁴⁶	LQT3	Beckermann (2014) ⁴⁶	R222P	-	-
46	SCN5A	A226V	LoF	Tan (2016) ⁴⁷	BrS	Tan (2016) ⁴⁷	A223V	-	-
47	SCN1A	T226M	Mixed	Berecki (2019) ⁴⁸	EPI	Sadleir (2017) ⁴⁹	T226M	-	NC
48	SCN1A	I227S	LoF	Ohmori (2006) ³⁰	EPI	Nabbout (2003) ⁵⁰	I227S	-	NC
49	SCN5A	V232I	STW	Barajaz-Martinez (2008) ⁵¹	BrS	Barajaz-Martinez (2008) ⁵¹ Kapplinger (2010) ¹² [compound missense with L1308F]	V229I	-	NC
50	SCN9A	I228M	GoF	Estacion (2011) ⁵²	SFN	Estacion (2011) ⁵² Faber (2012) ⁵³	I230M	8.58-e4	NC
231-249 Cytoplasmic									
51	SCN2A	T236S	GoF	Thompson (2020) ⁵⁴	EPI	Nakamura (2013) ⁵⁵	T235S	3.98e-6	NC
52	SCN9A	I234T	GoF	Ahn (2010) ⁵⁶	IEM	Ahn (2010) ⁵⁶	I236T	-	-
53	SCN4A	S246L	STW	Tsujino (2003) ⁵⁷	CMS	Tsujino (2003) ⁵⁷	S243L	-	NC
54	SCN9A	S241T	GoF	Lampert (2006) ⁵⁸	IEM	Lampert (2006) ⁵⁸ Michiels (2005) ⁵⁹	S243T	-	-
55	SCN10A	S242T	GoF	Han (2018) ⁶⁰	PPN	Han (2018) ⁶⁰		1.91-e4	-

56	SCN3A	L247P	LoF	Lamar (2017) ⁶¹	EPI	Lamar (2017) ⁶¹	L247P	-	-
57	SCN1A	D249E	LoF	Kluckova (2020) ¹⁰	EPI	Kluckova (2020) ¹⁰	D249E	-	NC
250-269 S5 of D1									
58	SCN1A	S259R	LoF	Nissenkorn (2019) ²⁹	EPI	Nissenkorn (2019) ²⁹	S259R	-	-
59	SCN2A	A263V	GoF	Liao (2010a) ⁶²	EPI	Liao (2010a) ⁶² Schwarz (2016) ⁶³	A262V	-	-
60	SCN1A	L263V	GoF	Kahlig (2008) ⁶⁴	FHM	Kahlig (2008) ⁶⁴	L263V	-	-
61	SCN8A	G269R	LoF	Wengert (2019) ⁶⁵	NDD without epilepsy	Wengert (2019) ⁶⁵	G265R	-	-
62	SCN4A	Q270K	Mixed	Carle (2009) ⁶⁶	PMC	Carle (2009) ⁶⁶	Q267K	-	-
63	SCN5A	Q270K	Mixed	Calloe (2011) ⁶⁷	LQT3	Calloe (2011) ⁶⁷		-	-
270-367 Extracellular									
64	SCN5A	L276Q	LoF	Glazer (2020) ¹	BrS	Sommariva (2013) ⁶⁸ Yamagata (2017) ²¹	L273Q	-	-
65	SCN5A	R282H	LoF	Polezing (2006) ⁶⁹	BrS	Priori (2002) ⁷ Itoh (2005) ⁷⁰	Q279H	1.60e-5	NC
66	SCN5A	R282C	LoF	Glazer (2020) ¹	BrS	Andorin (2016) ⁷¹	Q279C	-	NC
67	SCN1A	T297I	LoF	Binini (2017) ⁷²	EPI	Binini (2017) ⁷²	T297I	-	NC
68	SCN5A	L325R	LoF	Keller (2005) ⁷³	BrS	Keller (2005) ⁷³	L335R	-	NC
69	SCN5A	C335R	LoF	Glazer (2020) ¹	BrS	Van Malderen (2017) ⁷⁴	C345R	-	-
70	SCN5A	P336L	LoF	Cordeiro (2006) ⁷⁵	BrS	Cordeiro (2006) ⁷⁵	P346L	-	NC
71	SCN3A	K354Q	GoF	Estacion (2010) ⁷⁶	EPI	Holland (2008) ⁷⁷	K353Q	-	NC
72	SCN5A	E346D	Mixed	Glazer (2020) ¹	BrS	Probst (2006) ⁷⁸	R356D	-	NC
73	SCN4A	P382T	LoF	Zaharieva (2016) ¹⁴	CMS	Zaharieva (2016) ¹⁴	P358T	-	-
74	SCN5A	D349N	LoF	Glazer (2020) ¹	BrS	Savastano (2014) ⁷⁹	N359N	1.43e-5	NC
75	SCN5A	T353I	LoF	Pfahnl (2007) ⁸⁰ Zhang (2015) ⁸¹ Glazer (2020) ¹	BrS	Pfahnl (2007) ⁸⁰	T363I	-	-

76	SCN5A	D356N	LoF	Makiyama (2005) ³¹	BrS	Makiyama (2005) ³¹	D366N	4.02e-6	-
368-392 Pore-forming									
77	SCN5A	R367C	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Amin (2011) ⁸²	R377C	1.07e-5	-
78	SCN2A	R379H	LoF	Ben-Shalom (2017) ⁵	ASD	Ben-Shalom (2017) ⁵	R377H	-	-
79	SCN5A	R367H	LoF	Hong (2004) ⁸³	BrS	Hong (2004) ⁸³		-	-
80	SCN5A	R367L	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	R377L	-	-
81	SCN5A	M369K	LoF	Glazer (2020) ¹	BrS	Probst (2006) ⁷⁸ Probst (2007) ⁸⁴ Andorin (2016) ⁷¹	M379K	-	-
82	SCN5A	W374G	LoF	Nakajima (2021) ⁸⁵	BrS	Kapplinger (2010) ¹² Nakajima (2021) ⁸⁵	W384G	-	-
83	SCN5A	R376H	LoF	Rossenbacker (2004) ⁸⁶ Frustaci (2005) ⁸⁷ Peters (2016) ⁸⁸	BrS	Rossenbacker (2004) ⁸⁶ Frustaci (2005) ⁸⁷	N386H	8.08e-6	NC
84	SCN8A	N374K	GoF	Zaman (2019) ²²	EPI	Zaman (2019) ²²	N386K	-	NC
393-399 Extracellular									
85	SCN1A	R393H	LoF	Ohmori (2006) ³⁰	EPI	Claes (2003) ⁸⁹	R393H	-	-
86	SCN5A	G386R	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	G396R	-	-
400-420 S6 of D1									
87	SCN11A	I381T	GoF	Huang (2014) ⁹⁰	SFN	Huang (2014) ⁹⁰	V404T	7.96e-6	NC
88	SCN5A	V396L	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	V406L	-	NC
89	SCN10A	L388M	LoF	Gando (2020) ⁹¹	SUD	Gando (2020) ⁹¹	L414M	-	-
90	SCN4A	N440K	GoF	Lossin (2012) ⁹²	PMC; PAM	Lossin (2012) ⁹²	N416K	-	-
91	SCN5A	N406K	Mixed	Hu (2018) ⁹³ Kato (2014) ⁹⁴	LQT3	Hu (2018) ⁹³ Kato (2014) ⁹⁴		-	-
92	SCN9A	N395K	GoF	Sheets (2007) ⁴¹	IEM	Drenth (2005) ⁴²		-	-
93	SCN5A	N406S	LoF	Itoh (2007) ⁹⁵	BrS	Itoh (2007) ⁹⁵	N416S	-	-
421-768 Cytoplasmic									

94	SCN2A	V423L	GoF	Wolff (2017) ⁹⁶	EPI	Wolff (2017) ⁹⁶	V421L	-	-
95	SCN4A	V445M	GoF	Wang (1999) ⁹⁷ Huang (2020) ⁹⁸	PMC	Liu (2015) ⁹⁹ Huang (2020) ⁹⁸	V421M	-	-
96	SCN5A	V411M	GoF	Horne (2011) ¹⁰⁰ Zhou (2015) ¹⁰¹	LQT3	Horne (2011) ¹⁰⁰ Zhou (2015) ¹⁰¹		-	-
97	SCN9A	V400M	GoF	Fischer (2009) ¹⁰²	IEM	Fischer (2009) ¹⁰²		-	-
98	SCN1A	Y426N	LoF	Ohmori (2006) ³⁰	EPI	Nabbout (2003) ⁵⁰	Y426N	-	-
99	SCN5A	R458C	GoF	Winkel (2015) ¹⁰³	LQT3	Winkel (2015) ¹⁰³	D481C	1.45e-4	NC
100	SCN5A	A551T	LoF	Chiang (2009) ¹⁰⁴ Juang (2014) ¹⁰⁵	BrS	Chiang (2009) ¹⁰⁴ Juang (2014) ¹⁰⁵	F598T	-	NC
101	SCN5A	H558R	STW	Ye (2003) ¹⁰⁶ Tester (2010) ¹⁰⁷ Veltmann (2016) ¹⁰⁸	LQT3; BrS	Ye (2003) ¹⁰⁶ Veltmann (2016) ¹⁰⁸ Juang (2014) ¹⁰⁵	R605R	0.22	NC
102	SCN5A	L567Q	LoF	Wan (2001) ¹⁰⁹	BrS	Priori (2000) ¹¹⁰	H614Q	-	NC
103	SCN5A	R569W	GoF	Kapplinger (2015) ⁸	LQT3	Kapplinger (2015) ⁸	E616W	8.03e-6	NC
104	SCN5A	A572D	STW	Tester (2010) ¹⁰⁷	LQT3	Tester (2010) ¹⁰⁷	N619D	5.18e-3	NC
105	SCN10A	L554P	GoF	Faber (2012) ¹¹¹	PPN	Faber (2012) ¹¹¹	R630P	8.60e-5	NC
106	SCN5A	N592K	LoF	Juang (2014) ¹⁰⁵	BrS	Juang (2014) ¹⁰⁵	H642K	3.23e-5	NC
107	SCN9A	D623N	GoF	Ahn (2013) ¹¹²	SFN	Faber (2012) ⁵³	D646N	-	-
108	SCN5A	L619F	GoF	Wehrens (2003) ¹¹³	LQT3	Wehrens (2003) ¹¹³	L668F	4.03e-5	NC
109	SCN5A	R620H	STW	Calloe (2013) ¹¹⁴ Glazer (2020) ¹	BrS	Calloe (2013) ¹¹⁴	P669H	3.14e-5	NC
110	SCN5A	R689H	LoF	Hong (2012) ¹¹⁵	BrS	Hong (2012) ¹¹⁵	K740H	9.25e-5	NC
111	SCN10A	M650K	Mixed	Kist (2016) ¹¹⁶	IEM	Kist (2016) ¹¹⁶	Y753K	5.45e-4	NC
769-787 S1 of D2									
112	SCN9A	I739V	GoF	Han (2012) ¹¹⁷	SFN	Faber (2012) ⁵³ Han (2012) ¹¹⁷	I774V	2.47e-3	-
113	SCN2A	T773I	GoF	Lauxmann (2018) ³³	EPI	Lauxmann (2018) ³³	T782I	-	-
114	SCN8A	T767I	GoF	Estacion (2014) ¹¹⁸ Pan (2020) ¹¹⁹	EPI	Estacion (2014) ¹¹⁸		-	-

115	SCN5A	M734V	LoF	Glazer (2020) ¹	BrS	Le Scouarnec (2015) ¹²⁰	M785V	-	NC
116	SCN5A	A735E	LoF	Glazer (2020) ¹	BrS	Priori (2002) ⁷ Nakajima (2011) ¹²¹	A786E	-	-
117	SCN5A	A735T	LoF	Glazer (2020) ¹	BrS	García-Castro (2010) ¹²²	A786T	-	-
118	SCN5A	A735V	LoF	De la Roche (2019) ¹²³	BrS	De la Roche (2019) ¹²³	A786V	4.01e-6	-
788-798 Extracellular									
119	SCN1A	E788K	LoF	Kluckova (2020) ¹⁰	EPI	Kluckova (2020) ¹⁰	E788K	-	-
120	SCN1A	Y790C	LoF	Bechi (2015) ¹²⁴	EPI	Annesi (2003) ¹²⁵	Y790C	-	NC
121	SCN5A	E746K	LoF	Glazer (2020) ¹	BrS	Peters (2008) ¹²⁶	N797K	2.14e-5	NC
799-818 S2 of D2									
122	SCN5A	G752R	LoF	Glazer (2020) ¹	BrS	Smits (2002) ¹²⁷ Probst (2006) ⁷⁸ Probst (2009) ¹²⁸ Hoogendijk (2010) ¹²⁹	G803R	4.03e-6	-
123	SCN1A	T808S	Mixed	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹ [compound missense with N1011I]	T808S	-	-
819-832 Cytoplasmic									
124	SCN5A	D772N	Mixed	Glazer (2020) ¹	LQT3; BrS	Kapplinger (2009) ¹⁹ Kapplinger (2010) ¹²	D823N	2.01e-5	-
125	SCN5A	P773S	STW	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	P824S	-	NC
833-852 S3 of D2									
126	SCN5A	D785N	LoF	Glazer (2020) ¹	BrS	Sayeed (2014) ¹³²	D836N	-	-
127	SCN8A	G822R	LoF	Wengert (2019) ⁶⁵	NDD without epilepsy	Wengert (2019) ⁶⁵	G837R	-	NC
853-854 Extracellular									
855-872 S4 of D2									
128	SCN9A	L823R	Mixed	Lampert (2009) ¹³³	IEM	Lampert (2009) ¹³³	L858R	-	-
129	SCN10A	R756W	LoF	Gando (2020) ⁹¹	SUD	Gando (2020) ⁹¹	R859W	5.68e-5	-
130	SCN1A	R859H	LoF	Volkers (2011) ¹³⁴	EPI	Volkers (2011) ¹³⁴	R859H	7.99e-6	-

131	SCN4A	R669H	LoF	Kuzmenkin (2002) ¹³⁵	HypoPP	Bulman (1999) ¹³⁶		8.05e-6	-
132	SCN1A	R859C	LoF	Bechi (2015) ¹²⁴	EPI	Depienne (2009) ¹³⁷	R859C	-	-
133	SCN5A	R808C	LoF	Glazer (2020) ¹	BrS	Kotta (2010) ¹³⁸		8.07e-6	-
134	SCN9A	F826Y	GoF	Wu (2017) ¹³⁹	IEM	Wu (2017) ¹³⁹	F861Y	-	-
135	SCN4A	R672G	LoF	Jurkatt-Rott (2000) ¹⁴⁰ Kuzmenkin (2002) ¹³⁵	HypoPP	Jurkatt-Rott (2000) ¹⁴⁰	R862G	-	-
136	SCN4A	R672H	LoF	Jurkatt-Rott (2000) ¹⁴⁰ Kuzmenkin (2002) ¹³⁵	HypoPP	Jurkatt-Rott (2000) ¹⁴⁰	R862H	1.21e-5	-
137	SCN5A	R811H	LoF	Calloe (2013) ¹¹⁴	BrS	Calloe (2013) ¹¹⁴		1.22e-5	-
138	SCN2A	R853Q	Mixed	Berecki (2018) ¹⁴¹ Mason (2019) ¹⁴²	EPI	Nakamura (2013) ⁵⁵ Epi (2013) ¹⁴³ Samanta (2015) ¹⁴⁴ Kobayashi (2016) ¹⁴⁵ Li (2016) ¹⁴⁶ Wolff (2017) ⁹⁶ Berecki (2018) ¹⁴¹	R862Q	-	-
139	SCN3A	L855P	GoF	Zaman (2020) ¹⁴⁷	Fetal Akinesia	Zaman (2020) ¹⁴⁷	L863P	-	NC
140	SCN5A	L812Q	LoF	Wang (2015) ¹⁴⁸	BrS	Wang (2015) ¹⁴⁸	L863Q	-	-
141	SCN1A	R865G	GoF	Volkers (2011) ¹³⁴	EPI	Volkers (2011) ¹³⁴	R865G	-	-
142	SCN4A	R675Q	Mixed	Wu (2014) ¹⁴⁹	NormoPP	Wu (2014) ¹⁴⁹	R865Q	8.19e-6	-
143	SCN5A	R814Q	Mixed	Glazer (2020) ¹	LQT3; BrS	Frigo (2007) ¹⁵⁰ Sommariva (2013) ⁶⁸ Itoh (2016) ¹⁵¹ Yamagata (2017) ²¹		2.51e-5	-
144	SCN5A	K817E	LoF	Kinoshita (2016) ¹⁵²	BrS	Kinoshita (2016) ¹⁵²	K868E	-	-
873-888 Cytoplasmic									
145	SCN1A	T875M	LoF	Lossin (2002) ¹⁵³	EPI	Escayg (2000) ¹⁵⁴	T875M	-	-
146	SCN3A	I875T	GoF	Zaman (2018) ¹⁵⁵ Zaman (2020) ¹⁴⁷	EPI/PMG	Zaman (2018) ¹⁵⁵ Miyatake (2018) ¹⁵⁶ Zaman (2020) ¹⁴⁷	I883T	-	-
147	SCN9A	I848T	GoF	Cummins (2004) ¹⁵⁷ Namer (2015) ¹⁵⁸ Thiele (2011) ¹⁵⁹	IEM	Yang (2004) ¹⁶⁰ Drenth (2005) ⁴² Namer (2015) ¹⁵⁸		-	-
148	SCN11A	G699R	GoF	Han (2015) ¹⁶¹	SFN	Han (2015) ¹⁶¹	G888R	1.63e-4	-
889-907 S5 of D2									

149	SCN5A	L839P	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	L890P	-	-
150	SCN9A	G856D	GoF	Hoeijmakers (2012) ¹⁶²	IEM; SFN	Hoeijmakers (2012) ¹⁶²	G891D	-	-
151	SCN9A	G856R	GoF	Tanaka (2017) ¹⁶³	IEM	Tanaka (2017) ¹⁶³	G891R	-	-
152	SCN9A	L858F	GoF	Han (2006) ¹⁶⁴ Han (2007) ¹⁶⁵ Cregg (2014) ¹⁶⁶	IEM	Han (2006) ¹⁶⁴ Drenth (2005) ⁴²	L893F	-	-
153	SCN9A	L858H	GoF	Cummins (2004) ¹⁵⁷ Estacion (2010) ¹⁶⁷ Thiele (2011) ¹⁵⁹ Vasylyev (2014) ¹⁶⁸	IEM	Yang (2004) ¹⁶⁰	L893H	-	-
154	SCN4A	T704M	GoF	Bendahhou (1999) ¹⁶⁹	HyperPP; PMC	Huang (2019) ¹⁷⁰	T894M	-	-
155	SCN9A	A863P	GoF	Harty (2006) ¹⁷¹	IEM	Harty (2006) ¹⁷¹	A898P	-	NC
156	SCN1A	F902C	LoF	Rhodes (2004) ¹⁷²	EPI	Ohmori (2002) ¹⁷³ - reported as F891C	F902C	-	-
157	SCN5A	F851L	LoF	Glazer (2020) ¹	BrS	Priori (2002) ⁷	F902L	7.07e-6	-
158	SCN9A	V872G	GoF	Choi (2009) ¹⁷⁴	IEM	Choi (2009) ¹⁷⁴	V907G	-	-
908-936 Extracellular									
159	SCN2A	G899S	LoF	Wolff (2017) ⁹⁶	EPI	Wolff (2017) ⁹⁶	G908S	-	-
160	SCN1A	M909K	LoF	Kluckova (2020) ¹⁰	EPI	Kluckova (2020) ¹⁰	M909K	-	NC
161	SCN9A	Q875E	GoF	Stadler (2015) ¹⁷⁵	IEM	Skeik 2012 ¹⁷⁶	Q910E	-	-
162	SCN10A	R814H	GoF	Savio-Galimberti (2014) ²⁸	AF	Savio-Galimberti (2014) ²⁸	K917H	3.08e-4	NC
163	SCN2A	K908E	GoF	Lauxmann (2018) ³³	EPI	Wolff (2017) ⁹⁶ Lauxmann (2018) ³³	K917E	-	NC
164	SCN9A	R896Q	LoF	Cox (2010) ¹⁷⁷	CIP	Cox (2010) ¹⁷⁷	R931Q	1.06e-5	NC
165	SCN5A	W879R	LoF	Glazer (2020) ¹	BrS	Glazer (2020) ¹	W932R	-	-
937-957 Pore-forming									
166	SCN1A	H939Q	LoF	Ohmori (2006) ³⁰	EPI	Ohmori (2006) ³⁰	H939Q	-	-
167	SCN5A	I890T	LoF	Tarradas (2013) ¹⁷⁸	BrS	Tarradas (2013) ¹⁷⁸	I943T	-	NC
168	SCN5A	F892I	LoF	Glazer (2020) ¹	BrS	Savastano (2014) ¹⁷⁹	F945L	-	-

169	SCN1A	R946C	LoF	Volkers (2011) ¹³⁴	EPI	Volkers (2011) ¹³⁴	R946C	-	-
170	SCN2A	R937C	LoF	Ben-Shalom (2017) ⁵ Begemann (2019) ¹⁸⁰	ASD	Ben-Shalom (2017) ⁵ Begemann (2019) ¹⁸⁰ Rauch (2012) ¹⁸¹		-	-
171	SCN1A	R946H	LoF	Liao (2010b) ¹⁸² Volkers (2011) ¹³⁴	EPI	Liao (2010b) ¹⁸² Volkers (2011) ¹³⁴	R946H	-	-
172	SCN2A	R937H	LoF	Ben-Shalom (2017) ⁵	ASD	Ben-Shalom (2017) ⁵		-	-
173	SCN5A	E901K	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	E954K	-	-
174	SCN1A	M956T	LoF	Bechi (2015) ¹²⁴	EPI	Bechi (2015) ¹²⁴	M956T	-	-
958-970 Extracellular									
175	SCN1A	C959R	LoF	Ohmori (2006) ³⁰	EPI	Ohmori (2006) ³⁰	C959R	-	-
176	SCN5A	S910L	LoF	Pambrun (2014) ¹⁸³	BrS	Pambrun (2014) ¹⁸³	A963L	3.99e-6	NC
177	SCN10A	L867F	LoF	Gando (2020) ⁹¹	SUD	Gando (2020) ⁹¹	L969F	2.85e-5	NC
971-991 S6 of D2									
178	SCN1A	G979R	LoF	Sugawara (2003) ¹⁸⁴ Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹	G979R	-	-
179	SCN8A	G964R	LoF	Wagnon (2017) ¹⁸⁵	NDD without epilepsy	Wagnon (2017) ¹⁸⁵		-	-
180	SCN5A	N927S	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	N980S	-	NC
181	SCN5A	L928P	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	L981P	-	-
182	SCN1A	V983A	LoF	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹	V983A	-	-
183	SCN1A	N985I	LoF	Sugawara (2003) ¹⁸⁴	EPI	Fujiwara (2003) ¹³¹	N985I	-	-
184	SCN1A	L986F	LoF	Lossin (2003) ¹⁸⁶ Thompson (2012) ¹⁸⁷	EPI	Claes (2001) ¹⁸⁸	L986F	-	-
185	SCN4A	L796V	GoF	Elia (2020) ¹⁸⁹	PMC	Elia (2020) ¹⁸⁹	L986V	-	-
992-1219 Cytoplasmic									
186	SCN5A	S941N	GoF	Ruan (2007) ¹⁹⁰	LQT3	Schwarz (2000) ¹⁹¹	S994N	-	-
187	SCN11A	N816K	GoF	Huang (2019) ¹⁹²	FEP	Huang (2019) ¹⁹²	A997K	3.99e-6	NC

188	SCN2A	E999K	GoF	Miao (2020) ¹⁹³ Thompson (2020) ⁵⁴	EPI	Nakamura (2013) ⁵⁵ Miao (2020) ¹⁹³	E1008K	-	NC
189	SCN1A	N1011I	LoF	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹ [compound missense with T808S]	N1011I	-	NC
190	SCN5A	R965C	LoF	Hsueh (2009) ¹⁹⁴	BrS	Hsueh (2009) ¹⁹⁴	R1018C	6.49e-5	-
191	SCN5A	P1014S	STW	Glazer (2020) ¹	BrS	Glazer (2020) ¹	H1065S	-	NC
192	SCN5A	R1023H	LoF	Frustaci (2005) ⁸⁷	BrS	Frustaci (2005) ⁸⁷	L1073H	2.50-e4	NC
193	SCN10A	P1102S	GoF	Gando (2020) ⁹¹	SUD	Gando (2020) ⁹¹	P1167S	4.02e-6	NC
194	SCN5A	S1103Y	GoF	Splawski (2002) ¹⁹⁵	LQT3	Splawski (2002) ¹⁹⁵ - reported as S1102Y Plant (2006) ¹⁹⁶	-	7.69e-3	NC
195	SCN1A	T1174S	Mixed	Cestele (2013) ¹⁹⁷	EPI	Cestele (2013) ¹⁹⁷	T1174S	1.71e-3	NC
196	SCN9A	W1150R	GoF	Estacion (2009) ¹⁹⁸	IEM	Drenth (2005) ⁴² Estacion (2009) ¹⁹⁸	Q1187R	0.88	NC
197	SCN5A	P1177L	GoF	Winkel (2012) ¹⁹⁹	LQT3	Winkel (2012) ¹⁹⁹	K1190L	-	NC
198	SCN1A	W1204R	Mixed	Lossin (2002) ¹⁵³ Bechi (2015) ¹²⁴	EPI	Escayg (2001) ²⁰⁰ Marini (2007) ²⁰¹	W1204R	-	NC
199	SCN5A	R1193Q	LoF	Wang (2004) ²⁰² Huang (2006) ²⁰³ Abdelsayed (2015) ²⁰⁴ Peters (2016) ⁸⁸ Abe (2018) ²⁰⁵ Li (2019) ²⁰⁶	LQT3; BrS	Takahata (2003) ²⁰⁷ Wang (2004) ²⁰² Huang (2006) ²⁰³ Li (2019) ²⁰⁶	N1206Q	5.18-e3	NC
1220-1237 S1 of D3									
200	SCN2A	E1211K	Mixed	Ogiwara (2009) ²⁰⁸	EPI	Ogiwara (2009) ²⁰⁸	E1221K	-	-
201	SCN5A	S1218I	LoF	Calloe (2013) ¹¹⁴	BrS	Calloe (2013) ¹¹⁴	S1231I	-	-
1238-1250 Extracellular									
202	SCN5A	E1225K	LoF	Glazer (2020) ¹	LQT3; BrS	Schulze-Bahr (2003) ²⁰⁹ Tester (2005) ⁶ Crotti (2012) ²¹⁰ Sommariva (2013) ⁶⁸ Andorin (2016) ⁷¹ Yamagata (2017) ²¹ Van Malderen (2017) ⁷⁴	E1238K	4.01e-6	-
203	SCN5A	R1232W	LoF	Baroudi (2002) ²¹¹ Makita (2008) ²¹²	BrS	Chen (1998) ²¹³ Baroudi (2002) ²¹¹	R1245W	-	NC
1251-1269 S2 of D3									

204	SCN4A	D1069N	LoF/Mixed	Zaharieva (2016) ¹⁴	CMS	Zaharieva (2016) ¹⁴	D1256N	1.20e-5	-
205	SCN5A	D1243N	LoF/Mixed	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²		1.45e-4	-
206	SCN1A	M1267I	LoF	Nissenkorn (2019) ²⁹	EPI	Nissenkorn (2019) ²⁹	M1267I	-	-
1270-1283 Cytoplasmic									-
207	SCN1A	A1273V	Mixed	Peters (2016) ²¹⁴	EPI	Peters (2016) ²¹⁴	A1273V	-	-
208	SCN5A	G1262S	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	G1275S	2.83e-5	-
1284-1302 S3 of D3									
209	SCN3A	V1280I	STW	Zaman (2020) ¹⁴⁷	EPI	Zaman (2020) ¹⁴⁷	V1292I	1.2e-5	NC
210	SCN5A	V1281F	Mixed	Glazer (2020) ¹	BrS	Hermida (2013) ²¹⁵	V1294F	-	-
1303-1310 Extracellular									
211	SCN5A	A1294G	LoF	Zaytseva (2019) ²¹⁶	BrS	Zaytseva (2019) ²¹⁶	S1307G	2.86e-5	NC
212	SCN5A	E1295K	GoF	Abriel (2001) ²¹⁷	LQT3	Abriel (2001) ²¹⁷	E1308K	4.02e-6	NC
1311-1329 S4 of D3									
213	SCN9A	R1279P	GoF	Huang (2014) ²¹⁸	PPN	Huang (2014) ²¹⁸	R1316P	-	-
214	SCN5A	T1304M	GoF	Wang (2007) ³⁶	LQT3	Wang (2007) ³⁶ Kapplinger (2009) ¹⁹ Olesen (2012) ²¹⁹	T1317M	1.65e-4	-
215	SCN5A	L1308F	STW	Barajaz-Martinez (2008) ⁵¹	BrS	Barajaz-Martinez (2008) ⁵¹ Kapplinger (2010) ¹² [compound missense with V232I]	L1321F	4.71e-4	NC
216	SCN2A	R1312T	LoF	Lossin (2012) ²²⁰	EPI	Shi (2009) ²²¹	R1322T	-	-
217	SCN11A	L1158P	GoF	Huang (2014) ⁹⁰	SFN	Huang (2014) ⁹⁰	L1327P	4.71e-4	-
218	SCN2A	R1319Q	LoF	Misra (2008) ²²²	EPI	Berkovic (2004) ²²³ Wolff (2017) ⁹⁶	R1329Q	-	NC
1330-1346 Cytoplasmic									
219	SCN5A	G1319V	LoF	Casini (2007) ²²⁴	BrS	Casini (2007) ²²⁴	G1332V	4.08e-5	-
220	SCN9A	V1298F	GoF	Jarecki (2008) ²²⁵ Cheng (2010) ²²⁶ Estacion (2010) ¹⁶⁷	PEPD	Fertleman (2006) ²²⁷	V1335F	-	-

221	SCN9A	V1299F	GoF	Jarecki (2008) ²²⁵ Thiele (2011) ¹⁵⁹	PEPD	Fertleman (2006) ²²⁷	V1336F	-	-
222	SCN5A	N1325S	GoF	Wang (1996) ²²⁸ Tian (2004) ²²⁹ Glazer (2020) ¹ Li (2020) ²³⁰	LQT3	Wang (1995) ²³¹	N1338S	-	NC
223	SCN2A	L1330F	LoF	Misra (2008) ²²²	EPI	Heron (2002) ²³²	L1340F	-	-
224	SCN5A	V1328M	GoF	Turker (2016) ²³³	BrS	Turker (2016) ²³³	L1341M	-	NC
225	SCN5A	A1330P	GoF	Wedekind (2001) ²³⁴ Berecki (2006) ²³⁵	LQT3	Wedekind (2001) ²³⁴	A1343P	-	-
226	SCN4A	A1156T	GoF	Palmio (2017) ²³⁶	HyperPP; PMC	McClatchey (1992) ²³⁷	A1343T	5.33e-5	-
227	SCN5A	A1330T	GoF	Smits (2005b) ²³⁸	LQT3	Smits (2005b) ²³⁸		-	-
228	SCN3A	P1333L	GoF	Zaman (2018) ¹⁵⁵ Zaman (2020) ¹⁴⁷	EPI	Zaman (2018) ¹⁵⁵	P1345L	-	-
229	SCN4A	P1158L	GoF	Desaphy (2016) ²³⁹	PAM	Desaphy (2016) ²³⁹		-	-
230	SCN5A	P1332L	GoF	Ruan (2007) ¹⁹⁰	LQT3	Ruan (2007) ¹⁹⁰ Schulze-Bahr (2004) ²⁴⁰		-	-
231	SCN9A	P1308L	GoF	Cheng (2010) ²²⁶	IEM	Cheng (2010) ²²⁶		-	-
232	SCN4A	P1158S	Mixed	Sugiara (2003) ²⁴¹ Webb (2008) ²⁴²	HypoPP	Sugiara (2003) ²⁴¹	P1345S	-	-
233	SCN5A	S1333Y	GoF	Huang (2009) ²⁴³	LQT3	Huang (2009) ²⁴³	S1346Y	-	NC
234	SCN2A	S1336Y	GoF	Thompson (2020) ⁵⁴	EPI	Nakamura (2013) ⁵⁵		-	NC
1347-1366 S5 of D3									
235	SCN8A	I1327V	GoF	Barker (2016) ²⁴⁴	EPI	Vaher (2014) ²⁴⁵ Singh (2015) ²⁴⁶	I1347V	-	-
236	SCN2A	N1339D	GoF	Miao (2020) ¹⁹³	EPI	Miao (2020) ¹⁹³	N1349D	-	-
237	SCN8A	L1331V	GoF	Patel (2016) ²⁴⁷	EPI	Carvill (2013) ²⁴⁸ - reported as L1290V	L1351V	-	-
238	SCN2A	L1342P	Mixed	Begemann (2019) ¹⁸⁰	EPI	Hackenberg (2014) ²⁴⁹ Matalon (2014) ²⁵⁰ Dimassi (2016) ²⁵¹ Wolff (2017) ⁹⁶ Begemann (2019) ¹⁸⁰	L1352P	-	-
239	SCN9A	V1316A	GoF	Wu (2013) ²⁵² Estacion (2013) ²⁵³	IEM	Huang (2016) ²⁵⁴ Estacion (2013) ²⁵³	V1353A	-	-
240	SCN11A	V1184A	GoF	Leipold (2015) ²⁵⁵	PPN	Leipold (2015) ²⁵⁵		-	-

241	SCN5A	V1340I	LoF	Samani (2009) ²⁵⁶	BrS	Samani (2009) ²⁵⁶	V1353I	4.60e-5	-
242	SCN1A	V1353L	LoF	Lossin (2003) ¹⁸⁶	EPI	Wallace (2001) ²⁵⁷	V1353L	-	-
243	SCN5A	F1344S	LoF	Keller (2006) ²⁵⁸	BrS	Keller (2006) ²⁵⁸	F1357S	-	-
244	SCN5A	W1345C	LoF	Glazer (2020) ¹	BrS	Lee (2010) ²⁵⁹	W1358C	-	-
245	SCN5A	L1346P	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	L1359P	-	-
246	SCN1A	V1366I	LoF	Bechi (2015) ¹²⁴	EPI	Osaka (2007) ²⁶⁰	V1366I	-	-
247	SCN5A	V1353M	STW	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	V1366M	2.78e-5	NC
1367-1418 Extracellular									
248	SCN10A	A1304T	GoF	Faber (2012) ¹¹¹	PPN	Faber (2012) ¹¹¹	A1370T	4.60e-5	NC
249	SCN8A	T1360N	LoF	Wengert (2019) ⁶⁵	NDD without epilepsy	Wengert (2019) ⁶⁵	T1380N	-	-
250	SCN5A	V1378M	LoF	Moreau (2012) ¹⁸	BrS	Moreau (2012) ¹⁸	V1390M	3.99e-6	NC
251	SCN5A	N1380K	LoF	Glazer (2020) ¹	BrS	Rudnik-Schöneborn (2011) ²⁶¹	N1392K	-	-
252	SCN5A	S1382I	LoF	Glazer (2020) ¹	BrS	Probst (2009) ¹²⁸	T1394I	-	NC
253	SCN4A	C1209F	LoF	Zaharieva (2016) ¹⁴	CMS	Zaharieva (2016) ¹⁴	C1396F	-	-
254	SCN2A	C1386R	LoF	Ben-Shalom (2017) ⁵	ASD	Ben-Shalom (2017) ⁵	C1396R	-	-
255	SCN5A	V1405L	LoF	Glazer (2020) ¹	BrS	Amin (2011) ⁸²	V1418L	-	-
256	SCN5A	V1405M	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Zumhagen (2016) ²⁶² Yamagata (2017) ²¹	V1418M	-	-
1419-1440 Pore-forming									
257	SCN5A	G1406E	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	G1419E	-	NC
258	SCN5A	G1406R	LoF	Tan (2006) ²⁶³	BrS	Kyndt (2001) ²⁶⁴	G1419R	-	NC
259	SCN1A	G1421W	LoF	Kim (2018) ²⁶⁵	EPI	Kim (2018) ²⁶⁵	G1421W	-	NC
260	SCN2A	T1420M	LoF	Ben-Shalom (2017) ⁵	ASD	Ben-Shalom (2017) ⁵	T1430M	-	-

261	SCN5A	G1420R	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	G1433R	-	-
262	SCN5A	G1420V	LoF	Glazer (2020) ¹	BrS	Hermida (2010) ²⁶⁶	G1433V	-	-
1441-1457 Extracellular									
263	SCN5A	A1428S	LoF	Zhu (2015) ²⁶⁷	BrS	Zhu (2015) ²⁶⁷	A1441S	3.19e-5	-
264	SCN5A	A1428V	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Van Malderen (2017) ⁷⁴	A1441V	-	-
265	SCN5A	D1430N	LoF	Maury (2013) ²⁶⁸	BrS	Maury (2013) ²⁶⁸	D1443N	-	-
266	SCN5A	R1432G	LoF	Deschenes (2000) ²⁶⁹ Glazer (2020) ¹	BrS	Deschenes (2000) ²⁶⁹	R1445G	-	NC
1458-1479 S6 of D3									
267	SCN5A	Y1449C	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Hothi (2015) ²⁷⁰ Van Malderen (2017) ⁷⁴	Y1462C	-	-
268	SCN5A	T1461S	LoF	Glazer (2020) ¹	BrS	Sacilotto (2017) ²⁷¹	T1474S	-	-
1480-1542 Cytoplasmic									
269	SCN4A	V1293I	GoF	Farinato (2019) ²⁷²	PMC	Koch (1995) ²⁷³	V1481I	-	-
270	SCN2A	I1473M	GoF	Ogiwara (2009) ²⁰⁸	EPI	Ogiwara (2009) ²⁰⁸	I1483M	-	-
271	SCN3A	I1468R	Mixed	Zaman (2020) ¹⁴⁷	EPI	Zaman (2020) ¹⁴⁷	I1483R	-	-
272	SCN4A	N1297K	GoF	Farinato (2019) ²⁷²	PMC	Farinato (2019) ²⁷² Gay (2008) ²⁷⁴	N1485K	-	-
273	SCN4A	N1297S	GoF	Maggi (2017) ²⁷⁵ Farinato (2019) ²⁷²	PMC	Maggi (2017) ²⁷⁵	N1485S	-	-
274	SCN4A	F1298C	GoF	Farinato (2019) ²⁷²	PAM	Farinato (2019) ²⁷²	F1486C	-	-
275	SCN5A	F1473C	GoF	Bankston (2007) ²⁷⁶	LQT3	Bankston (2007) ²⁷⁶		-	-
276	SCN5A	F1473S	GoF	Cai (2016) ²⁷⁷	LQT3	Ruan (2010) ²⁷⁸	F1486S	-	-
277	SCN9A	F1449V	GoF	Dib-Hajj (2005) ²⁷⁹ Gurkiewicz (2011) ²⁸⁰	IEM	Dib-Hajj (2005) ²⁷⁹	F1486V	-	-
278	SCN5A	Q1475P	GoF	Gando (2020b) ²⁸¹	LQT3	Tan (2017) ²⁸²	Q1488P	-	-
279	SCN1A	Q1489H	GoF	Barbieri (2019) ²⁸³	FHM	Vahedi (2009) ²⁸⁴	Q1489H	-	-

280	SCN1A	Q1489K	GoF	Kahlig (2008) ⁶⁴ Cestèle (2008) ²⁸⁵	FHM	Kahlig (2008) ⁶⁴ Cestèle (2008) ²⁸⁵	Q1489K	-	-
281	SCN5A	Q1476R	GoF	Moreau (2013) ²⁸⁶	LQT3	Moreau (2013) ²⁸⁶	Q1489R	-	-
282	SCN4A	G1306E	GoF	Farinato (2019) ²⁷²	PAM	Lerche (1993) ²⁸⁷	G1494E	-	-
283	SCN8A	G1475R	GoF	Liu (2019) ²⁸⁸ Zaman (2019) ²²	EPI	Parrini (2017) ²⁸⁹ Wang (2017) ²⁹⁰ Xiao (2018) ²⁹¹ Gardella (2018) ²⁹² Liu (2019) ²⁸⁸ Zaman (2019) ²²	G1494R	-	-
284	SCN1A	I1498M	LoF	Barbieri (2019) ²⁸³	FHM	Weller (2014) ²⁹³	I1498M	-	-
285	SCN4A	I1310N	GoF	Farinato (2019) ²⁷²	PMC	Farinato (2019) ²⁷²	I1498N	-	-
286	SCN9A	I1461T	GoF	Fertleman (2006) ²²⁷	PEPD	Fertleman (2006) ²²⁷	I1498T	-	-
287	SCN5A	F1486L	GoF	Wang (2007) ³⁶	LQT3	Wang (2007) ³⁶	F1499L	-	-
288	SCN1A	F1499L	GoF	Barbieri (2019) ²⁸³	FHM	Vahedi (2009) ²⁸⁴		-	-
289	SCN1A	M1500V	GoF	Barbieri (2019) ²⁸³	FHM	Domitrz (2016) ²⁹⁴	M1500V	-	-
290	SCN4A	T1313A	GoF	Bouhours (2004) ²⁹⁵	PMC	Bouhours (2004) ²⁹⁵	T1501A	-	-
291	SCN4A	T1313M	GoF	Farinato (2019) ²⁷²	PMC	Farinato (2019) ²⁷²	T1501M	4.00e-6	-
292	SCN9A	T1464I	GoF	Fertleman (2006) ²²⁷ Thiele (2011) ¹⁵⁹	PEPD	Fertleman (2006) ²²⁷	T1501I	-	-
293	SCN3A	T1486I	GoF	Zaman (2020) ¹⁴⁷	EPI/PMG	Zaman (2020) ¹⁴⁷		-	-
294	SCN8A	A1491V	GoF	Zaman (2019) ²²	EPI	Zaman (2019) ²²	A1510V	-	-
295	SCN5A	P1506S	LoF	Saber (2015) ²⁹⁶	BrS	Saber (2015) ²⁹⁶	P1519S	-	-
296	SCN5A	R1512W	LoF	Deschenes (2000) ²⁶⁹ Zheng (2016) ²⁹⁷	BrS	Deschenes (2000) ²⁶⁹ Smits (2002) ¹²⁷ Zheng (2016) ²⁹⁷	R1525W	5.57e-5	-
297	SCN1A	R1525Q	LoF	Binini (2017) ⁷²	EPI	Binini (2017) ⁷²	R1525Q	3.99e-6	-
1543-1560 S1 of D4									
298	SCN5A	N1541D	LoF	Dharmawan (2019) ²⁹⁸	BrS	Dharmawan (2019) ²⁹⁸	N1554D	-	-
299	SCN4A	N1366S	GoF	Ke (2017) ²⁹⁹	PMC	Ke (2017) ²⁹⁹	N1554S	-	-

1561-1571 Extracellular									
1572-1590 S2 of D4									
300	SCN2A	L1563V	Mixed	Misra (2008) ²²² Berecki (2018) ¹⁴¹ Begemann (2019) ¹⁸⁰	EPI	Lewis (1996) ³⁰⁰ Heron (2002) ²³²	L1573V	-	-
301	SCN9A	W1538R	GoF	Cregg (2013) ³⁰¹	IEM	Cregg (2013) ³⁰¹	R1575R	2.02e-3	NC
302	SCN1A	R1575C	GoF	Ohmori (2008) ³⁰²	EPI	Ohmori (2008) ³⁰²	R1575C	7.18e-5	NC
303	SCN2A	I1571T	GoF	Miao (2020) ¹⁹³	EPI	Miao (2020) ¹⁹³	I1581T	-	NC
304	SCN10A	V1518I	LoF	Gando (2020) ⁹¹	SUD	Gando (2020) ⁹¹	L1583I	7.78e-5	NC
305	SCN1A	E1587K	LoF	Kluckova (2020) ¹⁰	EPI	Kluckova (2020) ¹⁰	E1587K	-	-
306	SCN5A	E1574K	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²		-	-
1591-1602 Cytoplasmic									
307	SCN1A	R1596C	LoF	Kluckova (2020) ¹⁰	EPI	Harkin (2007) ³⁰³ Depienne (2009) ¹³⁷ Kim (2014) ³⁰⁴	R1596C	-	-
308	SCN5A	R1583C	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²		8.03e-6	-
309	SCN2A	Y1589C	GoF	Lauxmann (2013) ³⁰⁵	EPI	Lauxmann (2013) ³⁰⁵	Y1599C	-	-
1603-1620 S3 of D4									
310	SCN5A	I1593M	STW	Kapplinger (2015) ⁸	LQT3	Kapplinger (2015) ⁸	I1606M	4.02e-6	NC
311	SCN2A	F1597L	GoF	Wolff (2017) ⁹⁶	EPI	Wolff (2017) ⁹⁶	F1607L	-	-
312	SCN5A	D1595N	LoF	Wang (2002) ³⁰⁶	PCCD; BrS	Wang (2002) ³⁰⁶	D1608N	-	-
313	SCN1A	V1611F	GoF	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹	V1611F	4.03e-6	-
1621-1633 Extracellular									
314	SCN5A	S1609L	GoF	Winkel (2015) ¹⁰³	LQT3	Winkel (2015) ¹⁰³	A1622L	-	NC
315	SCN1A	L1624P	GoF	Fan (2016) ³⁰⁷	FHM	Fan (2016) ³⁰⁷	L1624P	-	NC
316	SCN4A	V1442E	LoF	Tsujino (2003) ⁵⁷	CMS	Tsujino (2003) ⁵⁷	V1630E	-	NC

317	SCN1A	P1632S	LoF	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹	P1632S	-	-
318	SCN2A	P1622S	LoF	Wolff (2017) ⁹⁶	EPI	Wolff (2017) ⁹⁶		-	-
319	SCN5A	T1620K	Mixed	Surber (2008) ³⁰⁸	LQT3	Surber (2008) ³⁰⁸	T1633K	-	-
320	SCN2A	T1623N	GoF	Thompson (2020) ⁵⁴	EPI	Nakamura (2013) ⁵⁵	T1633N	-	-
321	SCN5A	T1620M	LoF	Baroudi (2002) ²¹¹ Wang (2000) ³⁰⁹ Makita (2008) ²¹²	BrS	Chen (1998) ²¹³ Baroudi (2002) ²¹¹	T1633M	3.99e-6	-
1634-1650 S4 of D4									
322	SCN3A	R1621G	GoF	Zaman (2020) ¹⁴⁷	EPI/PMG	Zaman (2020) ¹⁴⁷	R1636G	-	-
323	SCN3A	R1621Q	GoF	Zaman (2020) ¹⁴⁷	EPI/PMG	Zaman (2020) ¹⁴⁷	R1636Q	3.98e-6	-
324	SCN5A	R1623Q	GoF	Kambouris (1998) ³¹⁰ Tsurugi (2009) ³¹¹ Li (2020) ²³⁰	LQT3	Kambouris (2000) ³¹² Miura (2003) ³¹³		-	-
325	SCN8A	R1617Q	GoF	Wagnon (2015) ³¹⁴	EPI	Ohba (2014) ³¹⁵ Kong (2015) ³¹⁶ Larsen (2015) ³¹⁷		-	-
326	SCN4A	R1451C	LoF	Poulin (2018) ³¹⁸	HypoPP	Poulin (2018) ³¹⁸	R1639C	1.21e-5	-
327	SCN5A	R1626P	GoF	Ruan (2007) ¹⁹⁰	LQT3	Ruan (2007) ¹⁹⁰	R1639P	-	-
328	SCN4A	R1451L	LoF	Poulin (2018) ³¹⁸	PMC	Poulin (2018) ³¹⁸	R1639L	4.04e-6	-
329	SCN8A	R1620L	LoF	Liu (2019) ²⁸⁸	ASD	Liu (2019) ²⁸⁸		-	-
330	SCN8A	A1622D	GoF	Liu (2019) ²⁸⁸	ASD	Liu (2019) ²⁸⁸	A1641D	-	-
331	SCN5A	R1629Q	LoF	Zeng (2013) ³¹⁹	BrS	Zeng (2013) ³¹⁹	R1642Q	1.19e-5	-
332	SCN4A	R1454W	LoF	Habbout (2016) ³²⁰	CMS	Habbout (2016) ³²⁰	R1642W	1.61e-5	-
333	SCN4A	I1455T	Mixed	Bednarz (2016) ³²¹	PMC	Bednarz (2016) ³²¹	I1643T	1.20e-5	-
334	SCN5A	G1631D	GoF	Wang (2008) ³²²	LQT3	Wang (2008) ³²²	G1644D	-	-
335	SCN9A	G1607R	GoF	Choi (2011) ³²³	PEPD	Choi (2011) ³²³	G1644R	-	-
336	SCN5A	R1632C	LoF	Nakajima (2015) ³²⁴ Dharmawan (2019) ²⁹⁸	BrS	Nakajima (2015) ³²⁴ García-Molina (2016) ³²⁵	R1645C	3.98e-6	-
337	SCN4A	R1457H	LoF	Arnold (2015) ³²⁶	CMS	Arnold (2015) ³²⁶	R1645H	4.01e-6	-

338	SCN5A	R1632H	Mixed	Benson (2003) ³²⁷ Glazer (2020) ¹	BrS	Robyns (2014) ³²⁸		7.96e-6	-
339	SCN1A	R1648C	Mixed	Rhodes (2004) ¹⁷² Thompson (2012) ¹⁸⁷	EPI	Ohmori (2002) ¹⁷³ – reported as R1638C	R1648C	-	-
340	SCN1A	R1648H	Mixed	Lossin (2002) ¹⁵³ Vanoye (2006) ³²⁹ Kahlig (2010) ³³⁰	EPI	Escayg (2000) ¹⁵⁴	R1648H	-	-
341	SCN4A	R1460Q	Mixed	Elia (2019) ³³¹	CMS	Elia (2019) ³³¹	R1648Q	8.01e-6	-
342	SCN4A	R1460W	Mixed	Elia (2019) ³³¹	CMS	Elia (2019) ³³¹	R1648W	2.13e-5	-
343	SCN9A	L1612P	GoF	Suter (2015) ³³²	PEPD	Suter (2015) ³³²	L1649P	-	-
344	SCN1A	L1649Q	Mixed	Kahlig (2008) ⁶⁴ Cestèle (2013) ³³³	FHM	Kahlig (2008) ⁶⁴ Cestèle (2013) ³³³	L1649Q	-	-
1651-1669 Cytoplasmic									
345	SCN10A	R1588Q	LoF	Jabbari (2015) ¹³	AF	Jabbari (2015) ¹³	K1651Q	1.19e-5	NC
346	SCN5A	G1642E	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	G1655E	-	-
347	SCN1A	I1656M	LoF	Lossin (2003) ¹⁸⁶ Liu (2013) ³³⁴	EPI	Wallace (2001) ²⁵⁷	I1656M	-	-
348	SCN1A	R1657C	LoF	Lossin (2003) ¹⁸⁶	EPI	Lossin (2003) ¹⁸⁶	R1657C	-	-
349	SCN5A	R1644C	LoF	Frustaci (2005) ⁸⁷	BrS	Frustaci (2005) ⁸⁷		3.98e-6	-
350	SCN8A	R1638C	LoF	Wengert (2019) ⁶⁵	NDD without epilepsy	Wengert (2019) ⁶⁵		-	-
351	SCN5A	R1644H	GoF	Wang (1996) ²²⁸ Nieto-Marin (2019) ³³⁵	LQT3	Nieto-Marin (2019) ³³⁵	R1657H	-	-
352	SCN3A	F1646C	GoF	Zaman (2020) ¹⁴⁷	EPI/PMG	Zaman (2020) ¹⁴⁷	F1661C	-	-
353	SCN1A	F1661L	GoF	Barbieri (2019) ²⁸³	FHM	Weller (2014) ²⁹³	F1661L	-	-
354	SCN1A	F1661S	Mixed	Rhodes (2004) ¹⁷² Thompson (2012) ¹⁸⁷	EPI	Claes (2003) ⁸⁹	F1661S	-	-
355	SCN4A	F1473S	GoF	Fleischhauer (1998) ³³⁶	PMC	Fleischhauer (1998) ³³⁶		-	-
356	SCN1A	M1664K	LoF	Bechi (2015) ¹²⁴	EPI	Depienne (2010) ³³⁷	M1664K	-	-
357	SCN9A	M1627K	GoF	Fertleman (2006) ²²⁷ Dib-Hajj (2008) ³³⁸ Thiele (2011) ¹⁵⁹	PEPD	Fertleman (2006) ²²⁷		-	-
358	SCN5A	M1652R	GoF	Ruan (2007) ¹⁹⁰ Li (2020) ²³⁰	LQT3	Ruan (2007) ¹⁹⁰	M1665R	-	-

359	SCN2A	P1658S	LoF	Miao (2020) ¹⁹³	EPI	Miao (2020) ¹⁹³	P1668S		-
360	SCN5A	A1656D	GoF	Kim (2019) ³³⁹	LQT3	Kim (2019) ³³⁹	A1669D	-	-
361	SCN9A	A1632E	GoF	Estacion (2008) ³⁴⁰ Rühlmann (2020) ³⁴¹	IEM; PEPD	Estacion (2008) ³⁴⁰	A1669E	-	-
362	SCN9A	A1632G	GoF	Yang (2016) ³⁴²	IEM	Yang (2016) ³⁴²	A1669G	-	-
363	SCN9A	A1632T	GoF	Eberhardt (2014) ³⁴³	IEM	Eberhardt (2014) ³⁴³	A1669T	-	NC
1670-1687 S5 of D4									
364	SCN1A	L1670W	Mixed	Bertelli (2018) ³⁴⁴ Dhifallah (2018) ³⁴⁵	FHM	Dhifallah (2018) ³⁴⁵	L1670W	-	-
365	SCN5A	I1660V	LoF	Cordeiro (2006) ⁷⁵	BrS	Cordeiro (2006) ⁷⁵	I1673V	3.18e-5	-
366	SCN1A	G1674R	LoF	Rhodes (2004) ¹⁷² Thompson (2012) ¹⁸⁷	EPI	Ohmori (2002) ¹⁷³ - reported as G1664R	G1674R	-	-
367	SCN5A	G1661R	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Van Malderen (2017) ⁷⁴		-	-
368	SCN5A	V1667I	GoF	Nakajima (2020) ³⁴⁶	LQT3	Nakajima (2020) ³⁴⁶	V1680I	3.98e-6	NC
369	SCN4A	I1495F	Mixed	Bendahhou (1999) ¹⁶⁹	HyperPP	Bendahhou (1999) ¹⁶⁹	I1683F	-	-
370	SCN3A	Y1669C	LoF	Zaman (2020) ¹⁴⁷	ASD	Zaman (2020) ¹⁴⁷	Y1684C	-	NC
371	SCN1A	A1685D	LoF	Sugiura (2012) ³⁴⁷	EPI	Fujiwara (2003) ¹³¹	A1685D	-	NC
372	SCN1A	A1685V	LoF	Lossin (2003) ¹⁸⁶ Sugiura (2012) ³⁴⁷	EPI	Sugawara (2001) ³⁴⁸	A1685V	-	NC
373	SCN5A	S1672Y	LoF	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹² Andorin (2016) ⁷¹	A1685Y	-	NC
1688-1709 Extracellular									
374	SCN5A	A1680T	STW	Glazer (2020) ¹	BrS	Kapplinger (2010) ¹²	A1693T	4.6e-5	NC
375	SCN10A	D1639N	LoF	Kaluza (2018) ³⁴⁹	SFN	Dabby (2016) ³⁵⁰	D1702N	8.75e-5	NC
376	SCN5A	D1690N	LoF	Zeng (2016) ³⁵¹ Nunez (2013) ³⁵²	BrS	Zeng (2016) ³⁵¹ Nunez (2013) ³⁵²	D1703N	3.98e-6	-
377	SCN1A	T1709I	LoF	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹	T1709I	-	-
1710-1732 Pore-forming									

378	SCN5A	T1709M	LoF	Glazer (2020) ¹	LQT3; BrS	Kapplinger (2010) ¹² Lakshmanadoss (2016) ³⁵³	T1722M	3.98e-6	-
379	SCN5A	G1712C	LoF	Chen (2016) ³⁵⁴	BrS	Kapplinger (2015) ⁸ Chen (2016) ³⁵⁴	G1725C	-	-
380	SCN10A	G1662S	GoF	Han (2014) ³⁵⁵	SFN	Han (2014) ³⁵⁵	G1725S	1.36e-3	-
381	SCN5A	D1714G	LoF	Amin (2005) ³⁵⁶	BrS	Amin (2005) ³⁵⁶	D1727G	-	-
1733-1762 Extracellular									
382	SCN5A	N1722D	LoF	Glazer (2020) ¹	BrS	Probst (2009) ¹²⁸	N1735D	-	NC
383	SCN5A	P1730H	LoF	Glazer (2020) ¹	BrS	Van Malderen (2017) ⁷⁴	P1743H	-	NC
384	SCN1A	G1749E	LoF	Rhodes (2004) ¹⁷²	EPI	Claes (2003) ⁸⁹	G1749E	-	NC
385	SCN5A	G1740R	LoF	Baroudi (2004) ³⁵⁷	BrS	Baroudi (2004) ³⁵⁷ Kapplinger (2010) ¹²	G1754R	-	-
386	SCN5A	G1743R	LoF	Valdivia (2004) ³⁵⁸	BrS	Takahata (2003) ²⁰⁷ Valdivia (2004) ³⁵⁸	G1757R	-	NC
387	SCN5A	G1748D	LoF	Nunez (2013) ³⁵²	BrS	Nunez (2013) ³⁵²	G1762D	-	NC
1763-1785 S6 of D4									
388	SCN1A	F1765L	LoF	Liao (2010b) ¹⁸²	EPI	Liao (2010b) ¹⁸²	F1765L	-	NC
389	SCN10A	I1706V	GoF	Huang (2013) ³⁵⁹	SFN	Huang (2013) ³⁵⁹	I1770V	-	-
390	SCN1A	F1774S	GoF	Bertelli (2018) ³⁴⁴	FHM	Chastan (2016) ³⁶⁰	F1774S	-	-
391	SCN5A	I1762A	GoF	Chang (2004) ³⁶¹	LQT3	Chang (2004) ³⁶¹	V1776A	-	NC
392	SCN5A	V1763M	GoF	Chang (2004) ³⁶¹	LQT3	Chang (2004) ³⁶¹	V1777M	-	NC
393	SCN8A	V1758A	LoF	Zaman (2019) ²²	EPI	Zaman (2019) ²²	V1778A	-	-
394	SCN5A	V1764M	GoF	Chang (2004) ³⁶¹	LQT3	Chang (2004) ³⁶¹	V1778M	-	-
395	SCN3A	M1765I	GoF	Zaman (2020) ¹⁴⁷	EPI/PMG	Zaman (2020) ¹⁴⁷	M1780I	-	-
396	SCN8A	M1760I	GoF	Liu (2019) ²⁸⁸	EPI	Liu (2019) ²⁸⁸		-	-
397	SCN5A	M1766L	Mixed	Valdivia (2002) ³⁶² Ye (2003) ¹⁰⁶	LQT3	Valdivia (2002) ³⁶² Ye (2003) ¹⁰⁶	M1780L	-	-

398	SCN5A	Y1767C	GoF	Huang (2006) ²⁰³ Huang (2011) ³⁶³	LQT3	Huang (2011) ³⁶³	Y1781C	-	-
399	SCN5A	I1768V	GoF	Rivolta (2002) ³⁶⁴ Clancy (2003) ³⁶⁵	LQT3	Rivolta (2002) ³⁶⁴	I1782V	-	-
400	SCN9A	A1746G	GoF	Cregg (2013) ³⁰¹	IEM	Cregg (2013) ³⁰¹	A1783G	-	-
401	SCN2A	A1773T	LoF	Miao (2020) ¹⁹³	EPI	Miao (2020) ¹⁹³	A1783T	-	-
402	SCN3A	V1769A	GoF	Zaman (2018) ¹⁵⁵ Zaman (2020) ¹⁴⁷	EPI	Zaman (2018) ¹⁵⁵	V1784A	-	NC
1786-2009 Cytoplasmic									
403	SCN5A	N1774D	GoF	Kato (2014) ⁹⁴	LQT3	Kato (2014) ⁹⁴	N1788D	-	-
404	SCN8A	N1768D	GoF	Veeramah (2012) ³⁶⁶ Patel (2016) ²⁴⁷ Baker (2018) ³⁶⁷	EPI	Veeramah (2012) ³⁶⁶		-	-
405	SCN5A	N1774H	Mixed	Neubauer (2019) ³⁶⁸	LQT3	Neubauer (2019) ³⁶⁸	N1788H	-	-
406	SCN5A	E1784K	Mixed	Deschenes (2000) ²⁶⁹ Abdelsayed (2015) ²⁰⁴ Peters (2016) ⁸⁸ Veltmann (2016) ¹⁰⁸ Abdelsayed (2017) ³⁶⁹ Abdelsayed (2018) ³⁷⁰ Glazer (2020) ¹	LQT3; BrS	Deschenes (2000) ²⁶⁹ Takahashi (2014) ³⁷¹ Veltmann (2016) ¹⁰⁸	E1798K	-	NC
407	SCN5A	S1787N	GoF	Hu (2015) ³⁷²	LQT3	Splawski (2000) ³⁷³	S1801N	8.29e-4	NC
408	SCN5A	D1790G	Mixed	An (1998) ³⁷⁴ Wehrens (2000) ³⁷⁵ Baroudi (2000) ³⁷⁶	LQT3; BrS	Benhorin (1998) ³⁷⁷ Blich (2015) ³⁷⁸	D1804G	-	-
409	SCN1A	F1808L	Mixed	Rhodes (2005) ¹³⁰	EPI	Fujiwara (2003) ¹³¹	F1808L	-	-
410	SCN5A	Y1795C	GoF	Rivolta (2001) ³⁷⁹ Berecki (2006) ²³⁵ Fredj (2006) ³⁸⁰	LQT3	Rivolta (2001) ³⁷⁹ Benito (2008) ³⁸¹ Kapplinger (2015) ⁸	Y1809C	-	-
411	SCN5A	Y1795H	LoF	Rivolta (2001) ³⁷⁹	BrS	Rivolta (2001) ³⁷⁹	Y1809H	-	-
412	SCN2A	E1803G	GoF	Begemann (2019) ¹⁸⁰	EPI	Papuc (2019) ³⁸² Begemann (2019) ¹⁸⁰	E1813G	-	-
413	SCN4A	Q1633E	GoF	Kubota (2009) ³⁸³	PAM	Kubota (2009) ³⁸³	Q1821E	-	-
414	SCN1A	F1831S	LoF	Sugawara (2003) ¹⁸⁴	EPI	Fujiwara (2003) ¹³¹	F1831S	-	-
415	SCN5A	L1825P	LoF	Liu (2005) ³⁸⁴	LQT3	Makita (2002) ³⁸⁵	L1839P	-	-
416	SCN5A	Q1832E	LoF	Gando (2017) ³⁸⁶	BrS	Gando (2017) ³⁸⁶	K1846E	9.97e-5	NC

417	SCN1A	M1852T	Mixed	Rusconi (2007) ³⁸⁷	EPI	Annesi (2003) ¹²⁵	M1852T	-	NC
418	SCN5A	C1850S	LoF	Petitprez (2008) ³⁸⁸	BrS	Petitprez (2008) ³⁸⁸	C1864S	-	-
419	SCN1A	D1866Y	GoF	Spampanato (2004) ³⁸⁹	EPI	Spampanato (2004) ³⁸⁹	D1866Y	-	-
420	SCN2A	M1879T	GoF	Adney (2020) ³⁹⁰	EPI	Adney (2020) ³⁹⁰	M1889T	-	-
421	SCN8A	R1872L	GoF	Wagon (2015) ³¹⁴ Zaman (2019) ²²	EPI	Wagon (2015) ³¹⁴ Zaman (2019) ²²	R1892L	-	NC
422	SCN2A	R1882G	GoF	Schwarz (2016) ⁶³	EPI	Schwarz (2016) ⁶³	R1892G	-	NC
423	SCN2A	R1882Q	GoF	Wolff (2017) ⁹⁶ Berecki (2018) ¹⁴¹ Mason (2019) ¹⁴²	EPI	Howell (2015) ³⁹¹ Trump (2016) ³⁹² Wolff (2017) ⁹⁶ Berecki (2018) ¹⁴¹	R1892Q	-	NC
424	SCN8A	R1872Q	GoF	Wagon (2015) ³¹⁴ Aktin (2018) ³⁹³	EPI	Wagon (2015) ³¹⁴ Atkin (2018) ³⁹³		4.02e-6	NC
425	SCN8A	R1872W	GoF	Liu (2019) ²⁸⁸ Zaman (2019) ²²	EPI	Gardella (2016) ³⁹⁴ Zaman (2019) ²²	R1892W	-	NC
426	SCN1A	T1909I	Mixed	Ohmori (2006) ³⁰	EPI	Ohmori (2002) ³⁰	T1909I	-	-
427	SCN5A	R1897W	LoF	Olesen (2012) ²¹⁹	LQT3	Kapplinger (2009) ¹⁹	K1911W	-	NC
428	SCN5A	R1898C	LoF	Glazer (2020) ¹	BrS	Selga (2015) ³⁹⁵ Zhang (2016) ³⁹⁶	R1912C	3.56e-5	NC
429	SCN1A	Q1923R	LoF	Nissenkorn (2019) ²⁹	EPI	Shi (2012) ⁴⁰ Nissenkorn (2019) ²⁹	Q1923R	-	-
430	SCN5A	Q1909R	Mixed	Winkel (2015) ¹⁰³ Abdelsayed (2017) ³⁶⁹	LQT3	Winkel (2015) ¹⁰³ Kapplinger (2015) ⁸		-	-
431	SCN1A	R1927G	LoF	Rusconi (2009) ³⁹⁷	EPI	Rusconi (2009) ³⁹⁷	R1927G	-	-
432	SCN1A	T1934I	LoF	Kluckova (2020) ¹⁰	EPI	Kluckova (2020) ¹⁰	T1934I	3.19e-5	NC
433	SCN10A	A1886V	GoF	Savio-Galimberti (2014) ²⁸	AF	Savio-Galimberti (2014) ²⁸	G1950V	1.20e-3	NC
434	SCN5A	I1968S	LoF	Frustaci (2005) ⁸⁷	BrS	Frustaci (2005) ⁸⁷	M1977S	1.64e-5	NC
435	SCN5A	Y1977N	Mixed	Casini (2019) ³⁹⁸	LQT3	Casini (2019) ³⁹⁸	Y1986N	-	NC
436	SCN5A	F2004L	LoF	Bebarova (2008) ³⁹⁹	LQT3; BrS	Bebarova (2008) ³⁹⁹ Arnestad (2007) ⁴⁰⁰	-	1.02e-5	NC
437	SCN5A	P2006A	GoF	Shinlapawittayatorn (2011) ⁴⁰¹	LQT3	Shinlapawittayatorn(2011) ⁴⁰¹	-	1.11e-3	NC

Legend:

gnomAD frequencies (marked in grey)

NC = not conserved (marked in grey) = corresponding position of variants do not share the same amino acid

STW = Similar to Wildtype function (variant marked in grey)

Phenotypical features: AF = atrial fibrillation, ASD = autism spectrum disorder, BrS = Brugada syndrome, CAP = cold aggravated pain, CMS = congenital myasthenic syndrome, DEE = developmental and epileptic encephalopathy, DS = Dravet syndrome, ECG = electrocardiogram, echo = echocardiogram, EPI = epilepsy, FHM3 = familial hemiplegic migraine type 3, GEFS+ = genetic epilepsy with febrile seizures plus, Hyper-PP = hyperkalaemic periodic paralysis, Hypo-PP = hypokalaemic periodic paralysis, IEM = inherited erythromelalgia, LQT3 = long QT3 syndrome, NDD = neurodevelopmental disorder, PAM = potassium-aggravated myotonia, PDN = painful diabetic neuropathy, PEPD = paroxysmal extreme pain disorder, PMC = paramyotonia congenita, PMG = polymicrogyria, PPN = painful peripheral neuropathy, SCB = sodium channel blocker, SCD = sudden cardiac death, SIDS = sudden infant death syndrome, SSS = sick sinus syndrome, SUD = sudden unexplained death, Sz = seizure, TdP = torsade de pointes, VT = ventricular tachycardia

References

1. Glazer AM, Wada Y, Li B, et al. High-Throughput Reclassification of SCN5A Variants. *Am J Hum Genet* 2020.
2. Millat G, Chevalier P, Restier-Miron L, et al. Spectrum of pathogenic mutations and associated polymorphisms in a cohort of 44 unrelated patients with long QT syndrome. *Clinical Genetics* 2006;70:214-27.
3. Gutter C, Benndorf K, Zimmer T. Characterization of N-terminally mutated cardiac Na(+) channels associated with long QT syndrome 3 and Brugada syndrome. *Front Physiol* 2013;4:153.
4. Han C, Dib-Hajj SD, Lin Z, et al. Early- and late-onset inherited erythromelalgia: genotype-phenotype correlation. *Brain* 2009;132:1711-22.
5. Ben-Shalom R, Keeshen CM, Berrios KN, An JY, Sanders SJ, Bender KJ. Opposing Effects on NaV1.2 Function Underlie Differences Between SCN2A Variants Observed in Individuals With Autism Spectrum Disorder or Infantile Seizures. *Biol Psychiatry* 2017;82:224-32.
6. Tester DJ, Will ML, Haglund CM, Ackerman MJ. Compendium of cardiac channel mutations in 541 consecutive unrelated patients referred for long QT syndrome genetic testing. *Heart Rhythm* 2005;2:507-17.
7. Priori SG, Napolitano C, Gasparini M, et al. Natural history of Brugada syndrome: insights for risk stratification and management. *Circulation* 2002;105:1342-7.
8. Kapplinger JD, Giudicessi JR, Ye D, et al. Enhanced Classification of Brugada Syndrome-Associated and Long-QT Syndrome-Associated Genetic Variants in the SCN5A-Encoded Na(v)1.5 Cardiac Sodium Channel. *Circ Cardiovasc Genet* 2015;8:582-95.
9. Lin MT, Wu MH, Chang CC, et al. In utero onset of long QT syndrome with atrioventricular block and spontaneous or lidocaine-induced ventricular tachycardia: compound effects of hERG pore region mutation and SCN5A N-terminus variant. *Heart Rhythm* 2008;5:1567-74.
10. Kluckova D, Kolnikova M, Lacinova L, et al. A Study among the Genotype, Functional Alternations, and Phenotype of 9 SCN1A Mutations in Epilepsy Patients. *Sci Rep* 2020;10:10288.
11. Mancardi MM, Striano P, Gennaro E, et al. Familial occurrence of febrile seizures and epilepsy in severe myoclonic epilepsy of infancy (SMEI) patients with SCN1A mutations. *Epilepsia* 2006;47:1629-35.
12. Kapplinger JD, Tester DJ, Alders M, et al. An international compendium of mutations in the SCN5A-encoded cardiac sodium channel in patients referred for Brugada syndrome genetic testing. *Heart Rhythm* 2010;7:33-46.
13. Jabbari J, Olesen MS, Yuan L, et al. Common and Rare Variants in *SCN10A* Modulate the Risk of Atrial Fibrillation. *Circulation: Cardiovascular Genetics* 2015;8:64-73.
14. Zaharieva IT, Thor MG, Oates EC, et al. Loss-of-function mutations in SCN4A cause severe foetal hypokinesia or 'classical' congenital myopathy. *Brain* 2016;139:674-91.
15. Levy-Nissenbaum E, Eldar M, Wang Q, et al. Genetic analysis of Brugada syndrome in Israel: two novel mutations and possible genetic heterogeneity. *Genet Test* 2001;5:331-4.
16. Clatot J, Ziyadeh-Isleem A, Maugenre S, et al. Dominant-negative effect of SCN5A N-terminal mutations through the interaction of Na(v)1.5 α -subunits. *Cardiovasc Res* 2012;96:53-63.
17. Holst AG, Liang B, Jespersen T, et al. Sick sinus syndrome, progressive cardiac conduction disease, atrial flutter and ventricular tachycardia caused by a novel SCN5A mutation. *Cardiology* 2010;115:311-6.
18. Moreau A, Keller DI, Huang H, et al. Mexiletine differentially restores the trafficking defects caused by two brugada syndrome mutations. *Front Pharmacol* 2012;3:62.
19. Kapplinger JD, Tester DJ, Salisbury BA, et al. Spectrum and prevalence of mutations from the first 2,500 consecutive unrelated patients referred for the FAMILION long QT syndrome genetic test. *Heart rhythm* 2009;6:1297-303.
20. Yokokawa M, Noda T, Okamura H, et al. Comparison of Long-Term Follow-Up of Electrocardiographic Features in Brugada Syndrome Between the SCN5A-Positive Proband and the SCN5A-Negative Proband. *The American Journal of Cardiology* 2007;100:649-55.
21. Yamagata K, Horie M, Aiba T, et al. Genotype-Phenotype Correlation of SCN5A Mutation for the Clinical and Electrocardiographic Characteristics of Proband With Brugada Syndrome: A Japanese Multicenter Registry. *Circulation* 2017;135:2255-70.

22. Zaman T, Abou Tayoun A, Goldberg EM. A single-center SCN8A-related epilepsy cohort: clinical, genetic, and physiologic characterization. *Ann Clin Transl Neurol* 2019;6:1445-55.
23. Petitprez S, Tiab L, Chen L, et al. A novel dominant mutation of the Nav1.4 alpha-subunit domain I leading to sodium channel myotonia. *Neurology* 2008;71:1669-75.
24. Cheng X, Dib-Hajj SD, Tyrrell L, Waxman SG. Mutation I136V alters electrophysiological properties of the Na(v)1.7 channel in a family with onset of erythromelalgia in the second decade. *Mol Pain* 2008;4:1.
25. Lee MJ, Yu HS, Hsieh ST, Stephenson DA, Lu CJ, Yang CC. Characterization of a familial case with primary erythromelalgia from Taiwan. *J Neurol* 2007;254:210-4.
26. Mantegazza M, Gambardella A, Rusconi R, et al. Identification of an Nav1.1 sodium channel (SCN1A) loss-of-function mutation associated with familial simple febrile seizures. *Proc Natl Acad Sci USA* 2005;102:18177-82.
27. Smits JP, Koopmann TT, Wilders R, et al. A mutation in the human cardiac sodium channel (E161K) contributes to sick sinus syndrome, conduction disease and Brugada syndrome in two families. *J Mol Cell Cardiol* 2005;38:969-81.
28. Savio-Galimberti E, Weeke P, Muhammad R, et al. SCN10A/Nav1.8 modulation of peak and late sodium currents in patients with early onset atrial fibrillation. *Cardiovasc Res* 2014;104:355-63.
29. Nissenkorn A, Almog Y, Adler I, et al. In vivo, in vitro and in silico correlations of four de novo SCN1A missense mutations. *PLoS One* 2019;14:e0211901.
30. Ohmori I, Kahlig KM, Rhodes TH, Wang DW, George AL, Jr. Nonfunctional SCN1A is common in severe myoclonic epilepsy of infancy. *Epilepsia* 2006;47:1636-42.
31. Makiyama T, Akao M, Tsuji K, et al. High risk for bradyarrhythmic complications in patients with Brugada syndrome caused by SCN5A gene mutations. *J Am Coll Cardiol* 2005;46:2100-6.
32. Sugawara T, Tsurubuchi Y, Agarwala KL, et al. A missense mutation of the Na⁺ channel alpha II subunit gene Na(v)1.2 in a patient with febrile and afebrile seizures causes channel dysfunction. *Proc Natl Acad Sci U S A* 2001;98:6384-9.
33. Lauxmann S, Verbeek NE, Liu Y, et al. Relationship of electrophysiological dysfunction and clinical severity in SCN2A-related epilepsies. *Hum Mutat* 2018;39:1942-56.
34. Lemke JR, Riesch E, Scheurenbrand T, et al. Targeted next generation sequencing as a diagnostic tool in epileptic disorders. *Epilepsia* 2012;53:1387-98.
35. Marangoni S, Di Resta C, Rocchetti M, et al. A Brugada syndrome mutation (p.S216L) and its modulation by p.H558R polymorphism: standard and dynamic characterization. *Cardiovasc Res* 2011;91:606-16.
36. Wang DW, Desai RR, Crotti L, et al. Cardiac sodium channel dysfunction in sudden infant death syndrome. *Circulation* 2007;115:368-76.
37. Estacion M, Choi JS, Eastman EM, et al. Can robots patch-clamp as well as humans? Characterization of a novel sodium channel mutation. *J Physiol* 2010;588:1915-27.
38. Han C, Yang Y, Te Morsche RH, et al. Familial gain-of-function Nav1.9 mutation in a painful channelopathy. *J Neurol Neurosurg Psychiatry* 2017;88:233-40.
39. Okuda H, Noguchi A, Kobayashi H, et al. Infantile Pain Episodes Associated with Novel Nav1.9 Mutations in Familial Episodic Pain Syndrome in Japanese Families. *PLoS One* 2016;11:e0154827.
40. Choi JS, Dib-Hajj SD, Waxman SG. Inherited erythromelalgia: limb pain from an S4 charge-neutral Na channelopathy. *Neurology* 2006;67:1563-7.
41. Sheets PL, Jackson JO, 2nd, Waxman SG, Dib-Hajj SD, Cummins TR. A Nav1.7 channel mutation associated with hereditary erythromelalgia contributes to neuronal hyperexcitability and displays reduced lidocaine sensitivity. *J Physiol* 2007;581:1019-31.
42. Drenth JP, te Morsche RH, Guillet G, Taieb A, Kirby RL, Jansen JB. SCN9A mutations define primary erythromelalgia as a neuropathic disorder of voltage gated sodium channels. *J Invest Dermatol* 2005;124:1333-8.
43. Kim MK, Yuk JW, Kim HS, Park KJ, Kim DS. Autonomic dysfunction in SCN9A-associated primary erythromelalgia. *Clin Auton Res* 2013;23:105-7.
44. de Kovel CG, Meisler MH, Brilstra EH, et al. Characterization of a de novo SCN8A mutation in a patient with epileptic encephalopathy. *Epilepsy Res* 2014;108:1511-8.

45. Bezzina CR, Rook MB, Groenewegen WA, et al. Compound heterozygosity for mutations (W156X and R225W) in SCN5A associated with severe cardiac conduction disturbances and degenerative changes in the conduction system. *Circ Res* 2003;92:159-68.
46. Beckermann TM, McLeod K, Murday V, Potet F, George AL, Jr. Novel SCN5A mutation in amiodarone-responsive multifocal ventricular ectopy-associated cardiomyopathy. *Heart Rhythm* 2014;11:1446-53.
47. Tan BY, Yong RY, Barajas-Martinez H, et al. A Brugada syndrome proband with compound heterozygote SCN5A mutations identified from a Chinese family in Singapore. *Europace* 2016;18:897-904.
48. Berecki GB, A.: Terhag, J.: Maljevic, S.: Gazina, E. V.: Hill, S. L.: Petrou, S. SCN1A gain of function in early infantile encephalopathy. *Ann Neurol* 2019;85:514-25.
49. Sadleir LG, Mountier EI, Gill D, et al. Not all SCN1A epileptic encephalopathies are Dravet syndrome: Early profound Thr226Met phenotype. *Neurology* 2017;89:1035-42.
50. Nabbout R, Gennaro E, Dalla Bernardina B, et al. Spectrum of SCN1A mutations in severe myoclonic epilepsy of infancy. *Neurology* 2003;60:1961-7.
51. Barajas-Martínez HM, Hu D, Cordeiro JM, et al. Lidocaine-induced Brugada syndrome phenotype linked to a novel double mutation in the cardiac sodium channel. *Circ Res* 2008;103:396-404.
52. Estacion M, Han C, Choi JS, et al. Intra- and interfamily phenotypic diversity in pain syndromes associated with a gain-of-function variant of Nav1.7. *Mol Pain* 2011;7:92.
53. Faber CG, Hoeijmakers JG, Ahn HS, et al. Gain of function Nav1.7 mutations in idiopathic small fiber neuropathy. *Ann Neurol* 2012;71:26-39.
54. Thompson CH, Ben-Shalom R, Bender KJ, George AL. Alternative splicing potentiates dysfunction of early-onset epileptic encephalopathy SCN2A variants. *J Gen Physiol* 2020;152.
55. Nakamura K, Kato M, Osaka H, et al. Clinical spectrum of SCN2A mutations expanding to Ohtahara syndrome. *Neurology* 2013;81:992-8.
56. Ahn HS, Dib-Hajj SD, Cox JJ, et al. A new Nav1.7 sodium channel mutation I234T in a child with severe pain. *Eur J Pain* 2010;14:944-50.
57. Tsujino A, Maertens C, Ohno K, et al. Myasthenic syndrome caused by mutation of the SCN4A sodium channel. *Proc Natl Acad Sci U S A* 2003;100:7377-82.
58. Lampert A, Dib-Hajj SD, Tyrrell L, Waxman SG. Size matters: Erythromelalgia mutation S241T in Nav1.7 alters channel gating. *J Biol Chem* 2006;281:36029-35.
59. Michiels JJ, te Morsche RH, Jansen JB, Drenth JP. Autosomal dominant erythromelalgia associated with a novel mutation in the voltage-gated sodium channel alpha subunit Nav1.7. *Arch Neurol* 2005;62:1587-90.
60. Han C, Themistocleous AC, Estacion M, et al. The Novel Activity of Carbamazepine as an Activation Modulator Extends from Nav1.7 Mutations to the Nav1.8-S242T Mutant Channel from a Patient with Painful Diabetic Neuropathy. *Mol Pharmacol* 2018;94:1256-69.
61. Lamar T, Vanoye CG, Calhoun J, et al. SCN3A deficiency associated with increased seizure susceptibility. *Neurobiol Dis* 2017;102:38-48.
62. Liao Y, Anttonen AK, Liukkonen E, et al. SCN2A mutation associated with neonatal epilepsy, late-onset episodic ataxia, myoclonus, and pain. *Neurology* 2010;75:1454-8.
63. Schwarz N, Hahn A, Bast T, et al. Mutations in the sodium channel gene SCN2A cause neonatal epilepsy with late-onset episodic ataxia. *J Neurol* 2016;263:334-43.
64. Kahlig KM, Rhodes TH, Pusch M, et al. Divergent sodium channel defects in familial hemiplegic migraine. *Proc Natl Acad Sci USA* 2008;105:9799-804.
65. Wengert ER, Tronhjems CE, Wagnon JL, et al. Biallelic inherited SCN8A variants, a rare cause of SCN8A-related developmental and epileptic encephalopathy. *Epilepsia* 2019;60:2277-85.
66. Carle T, Fournier E, Sternberg D, Fontaine B, Tabti N. Cold-induced disruption of Na⁺ channel slow inactivation underlies paralysis in highly thermosensitive paramyotonia. *J Physiol* 2009;587:1705-14.
67. Calloe K, Schmitt N, Grubb S, et al. Multiple arrhythmic syndromes in a newborn, owing to a novel mutation in SCN5A. *Can J Physiol Pharmacol* 2011;89:723-36.

68. Sommariva E, Pappone C, Martinelli Boneschi F, et al. Genetics can contribute to the prognosis of Brugada syndrome: a pilot model for risk stratification. *Eur J Hum Genet* 2013;21:911-7.
69. Poelzing S, Forleo C, Samodell M, et al. SCN5A polymorphism restores trafficking of a Brugada syndrome mutation on a separate gene. *Circulation* 2006;114:368-76.
70. Itoh H, Shimizu M, Mabuchi H, Imoto K. Clinical and electrophysiological characteristics of Brugada syndrome caused by a missense mutation in the S5-pore site of SCN5A. *J Cardiovasc Electrophysiol* 2005;16:378-83.
71. Andorin A, Behr ER, Denjoy I, et al. Impact of clinical and genetic findings on the management of young patients with Brugada syndrome. *Heart Rhythm* 2016;13:1274-82.
72. Binini N, Sancini G, Villa C, et al. Identification of two mutations in cis in the SCN1A gene in a family showing genetic epilepsy with febrile seizures plus (GEFS+) and idiopathic generalized epilepsy (IGE). *Brain Res* 2017;1677:26-32.
73. Keller DI, Rougier JS, Kucera JP, et al. Brugada syndrome and fever: genetic and molecular characterization of patients carrying SCN5A mutations. *Cardiovasc Res* 2005;67:510-9.
74. Van Malderen SCH, Daneels D, Kerkhove D, et al. Prolonged Right Ventricular Ejection Delay in Brugada Syndrome Depends on the Type of SCN5A Variant - Electromechanical Coupling Through Tissue Velocity Imaging as a Bridge Between Genotyping and Phenotyping. *Circ J* 2017;82:53-61.
75. Cordeiro JM, Barajas-Martinez H, Hong K, et al. Compound heterozygous mutations P336L and I1660V in the human cardiac sodium channel associated with the Brugada syndrome. *Circulation* 2006;114:2026-33.
76. Estacion M, Gasser A, Dib-Hajj SD, Waxman SG. A sodium channel mutation linked to epilepsy increases ramp and persistent current of Nav1.3 and induces hyperexcitability in hippocampal neurons. *Exp Neurol* 2010;224:362-8.
77. Holland KD, Kearney JA, Glauser TA, et al. Mutation of sodium channel SCN3A in a patient with cryptogenic pediatric partial epilepsy. *Neuroscience Letters* 2008;433:65-70.
78. Probst V, Allouis M, Sacher F, et al. Progressive cardiac conduction defect is the prevailing phenotype in carriers of a Brugada syndrome SCN5A mutation. *J Cardiovasc Electrophysiol* 2006;17:270-5.
79. Savastano S, Rordorf R, Vicentini A, et al. A comprehensive electrocardiographic, molecular, and echocardiographic study of Brugada syndrome: Validation of the 2013 diagnostic criteria. *Heart Rhythm* 2014;11:1176-83.
80. Pfahnl AE, Viswanathan PC, Weiss R, et al. A sodium channel pore mutation causing Brugada syndrome. *Heart Rhythm* 2007;4:46-53.
81. Zhang J, Chen Y, Yang J, et al. Electrophysiological and trafficking defects of the SCN5A T353I mutation in Brugada syndrome are rescued by alpha-allocryptopine. *Eur J Pharmacol* 2015;746:333-43.
82. Amin AS, Boink GJ, Atrafi F, et al. Facilitatory and inhibitory effects of SCN5A mutations on atrial fibrillation in Brugada syndrome. *Europace* 2011;13:968-75.
83. Hong K, Berruezo-Sanchez A, Pongvarin N, et al. Phenotypic characterization of a large European family with Brugada syndrome displaying a sudden unexpected death syndrome mutation in SCN5A. *J Cardiovasc Electrophysiol* 2004;15:64-9.
84. Probst V, Denjoy I, Meregalli PG, et al. Clinical aspects and prognosis of Brugada syndrome in children. *Circulation* 2007;115:2042-8.
85. Nakajima T, Dharmawan T, Kawabata-Iwakawa R, et al. Reduced current density, partially rescued by mexiletine, and depolarizing shift in activation of SCN5A W374G channels as a cause of severe form of Brugada syndrome. *Ann Noninvasive Electrocardiol* 2021:e12828.
86. Rossenbacker T, Carroll SJ, Liu H, et al. Novel pore mutation in SCN5A manifests as a spectrum of phenotypes ranging from atrial flutter, conduction disease, and Brugada syndrome to sudden cardiac death. *Heart Rhythm* 2004;1:610-5.
87. Frustaci A, Priori SG, Pieroni M, et al. Cardiac histological substrate in patients with clinical phenotype of Brugada syndrome. *Circulation* 2005;112:3680-7.
88. Peters CH, Abdelsayed M, Ruben PC. Triggers for arrhythmogenesis in the Brugada and long QT 3 syndromes. *Prog Biophys Mol Biol* 2016;120:77-88.
89. Claes L, Ceulemans B, Audenaert D, et al. De novo SCN1A mutations are a major cause of severe myoclonic epilepsy of infancy. *Hum Mutat* 2003;21:615-21.

90. Huang J, Han C, Estacion M, et al. Gain-of-function mutations in sodium channel Na(v)1.9 in painful neuropathy. *Brain* 2014;137:1627-42.
91. Gando I, Williams N, Fishman GI, Sampson BA, Tang Y, Coetzee WA. Functional characterization of SCN10A variants in several cases of sudden unexplained death. *Forensic Sci Int* 2019;301:289-98.
92. Lossin C, Nam TS, Shahangian S, et al. Altered fast and slow inactivation of the N440K Nav1.4 mutant in a periodic paralysis syndrome. *Neurology* 2012;79:1033-40.
93. Hu RM, Tester DJ, Li R, et al. Mexiletine rescues a mixed biophysical phenotype of the cardiac sodium channel arising from the SCN5A mutation, N406K, found in LQT3 patients. *Channels (Austin)* 2018;12:176-86.
94. Kato K, Makiyama T, Wu J, et al. Cardiac channelopathies associated with infantile fatal ventricular arrhythmias: from the cradle to the bench. *J Cardiovasc Electrophysiol* 2014;25:66-73.
95. Itoh H, Tsuji K, Sakaguchi T, et al. A paradoxical effect of lidocaine for the N406S mutation of SCN5A associated with Brugada syndrome. *Int J Cardiol* 2007;121:239-48.
96. Wolff M, Johannesen KM, Hedrich UBS, et al. Genetic and phenotypic heterogeneity suggest therapeutic implications in SCN2A-related disorders. *Brain* 2017;140:1316-36.
97. Wang DW, VanDeCarr D, Ruben PC, George AL, Jr., Bennett PB. Functional consequences of a domain 1/S6 segment sodium channel mutation associated with painful congenital myotonia. *FEBS Lett* 1999;448:231-4.
98. Huang CW, Lai HJ, Lin PC, Lee MJ. Changes of Resurgent Na(+) Currents in the Nav1.4 Channel Resulting from an SCN4A Mutation Contributing to Sodium Channel Myotonia. *Int J Mol Sci* 2020;21.
99. Liu XL, Huang XJ, Luan XH, et al. Mutations of SCN4A gene cause different diseases: 2 case reports and literature review. *Channels (Austin)* 2015;9:82-7.
100. Horne AJ, Eldstrom J, Sanatani S, Fedida D. A novel mechanism for LQT3 with 2:1 block: a pore-lining mutation in Nav1.5 significantly affects voltage-dependence of activation. *Heart Rhythm* 2011;8:770-7.
101. Zhou H, Li Z, Ali Raza G, et al. [High incidence of sudden cardiac death in one family with type-3 long QT syndrome: molecular genetics and electrophysiology mechanism analysis]. *Zhonghua Xin Xue Guan Bing Za Zhi* 2015;43:1046-50.
102. Fischer TZ, Gilmore ES, Estacion M, et al. A novel Nav1.7 mutation producing carbamazepine-responsive erythromelalgia. *Ann Neurol* 2009;65:733-41.
103. Winkel BG, Yuan L, Olesen MS, et al. The role of the sodium current complex in a nonreferred nationwide cohort of sudden infant death syndrome. *Heart Rhythm* 2015;12:1241-9.
104. Chiang KC, Lai LP, Shieh RC. Characterization of a novel Nav1.5 channel mutation, A551T, associated with Brugada syndrome. *J Biomed Sci* 2009;16:76.
105. Juang JM, Lu TP, Lai LC, et al. Utilizing multiple in silico analyses to identify putative causal SCN5A variants in Brugada syndrome. *Sci Rep* 2014;4:3850.
106. Ye B, Valdivia CR, Ackerman MJ, Makielski JC. A common human SCN5A polymorphism modifies expression of an arrhythmia causing mutation. *Physiol Genomics* 2003;12:187-93.
107. Tester DJ, Valdivia C, Harris-Kerr C, et al. Epidemiologic, molecular, and functional evidence suggest A572D-SCN5A should not be considered an independent LQT3-susceptibility mutation. *Heart Rhythm* 2010;7:912-9.
108. Veltmann C, Barajas-Martinez H, Wolpert C, et al. Further Insights in the Most Common SCN5A Mutation Causing Overlapping Phenotype of Long QT Syndrome, Brugada Syndrome, and Conduction Defect. *J Am Heart Assoc* 2016;5.

109. Wan X, Chen S, Sadeghpour A, Wang Q, Kirsch GE. Accelerated inactivation in a mutant Na(+) channel associated with idiopathic ventricular fibrillation. *Am J Physiol Heart Circ Physiol* 2001;280:H354-60.
110. Priori SG, Napolitano C, Giordano U, Collisani G, Memmi M. Brugada syndrome and sudden cardiac death in children. *Lancet* 2000;355:808-9.
111. Faber CG, Lauria G, Merkies IS, et al. Gain-of-function Nav1.8 mutations in painful neuropathy. *Proc Natl Acad Sci U S A* 2012;109:19444-9.
112. Ahn HS, Vasylyev DV, Estacion M, et al. Differential effect of D623N variant and wild-type Na(v)1.7 sodium channels on resting potential and interspike membrane potential of dorsal root ganglion neurons. *Brain Res* 2013;1529:165-77.
113. Wehrens XH, Rossenbacker T, Jongbloed RJ, et al. A novel mutation L619F in the cardiac Na⁺ channel SCN5A associated with long-QT syndrome (LQT3): a role for the I-II linker in inactivation gating. *Hum Mutat* 2003;21:552.
114. Calloe K, Refaat MM, Grubb S, et al. Characterization and mechanisms of action of novel Nav1.5 channel mutations associated with Brugada syndrome. *Circ Arrhythm Electrophysiol* 2013;6:177-84.
115. Hong K, Hu J, Yu J, Brugada R. Concomitant Brugada-like and short QT electrocardiogram linked to SCN5A mutation. *Eur J Hum Genet* 2012;20:1189-92.
116. Kist AM, Sagafos D, Rush AM, et al. SCN10A Mutation in a Patient with Erythromelalgia Enhances C-Fiber Activity Dependent Slowing. *PLoS One* 2016;11:e0161789.
117. Han C, Hoeijmakers JG, Ahn HS, et al. Nav1.7-related small fiber neuropathy: impaired slow-inactivation and DRG neuron hyperexcitability. *Neurology* 2012;78:1635-43.
118. Estacion M, O'Brien JE, Conravey A, et al. A novel de novo mutation of SCN8A (Nav1.6) with enhanced channel activation in a child with epileptic encephalopathy. *Neurobiol Dis* 2014;69:117-23.
119. Pan Y, Cummins TR. Distinct functional alterations in SCN8A epilepsy mutant channels. *J Physiol* 2020;598:381-401.
120. Le Scouarnec S, Karakachoff M, Gourraud JB, et al. Testing the burden of rare variation in arrhythmia-susceptibility genes provides new insights into molecular diagnosis for Brugada syndrome. *Hum Mol Genet* 2015;24:2757-63.
121. Nakajima T, Kaneko Y, Saito A, et al. Identification of six novel SCN5A mutations in Japanese patients with Brugada syndrome. *Int Heart J* 2011;52:27-31.
122. García-Castro M, García C, Reguero JR, et al. The spectrum of SCN5A gene mutations in Spanish Brugada syndrome patients. *Rev Esp Cardiol* 2010;63:856-9.
123. de la Roche J, Angsutararux P, Kempf H, et al. Comparing human iPSC-cardiomyocytes versus HEK293T cells unveils disease-causing effects of Brugada mutation A735V of Na(V)1.5 sodium channels. *Sci Rep* 2019;9:11173.
124. Bechi G, Rusconi R, Cestele S, Striano P, Franceschetti S, Mantegazza M. Rescuable folding defective Nav1.1 (SCN1A) mutants in epilepsy: properties, occurrence, and novel rescuing strategy with peptides targeted to the endoplasmic reticulum. *Neurobiol Dis* 2015;75:100-14.
125. Annesi G, Gambardella A, Carrideo S, et al. Two novel SCN1A missense mutations in generalized epilepsy with febrile seizures plus. *Epilepsia* 2003;44:1257-8.
126. Peters S. Arrhythmogenic right ventricular dysplasia-cardiomyopathy and provokable coved-type ST-segment elevation in right precordial leads: clues from long-term follow-up. *Europace* 2008;10:816-20.

127. Smits JP, Eckardt L, Probst V, et al. Genotype-phenotype relationship in Brugada syndrome: electrocardiographic features differentiate SCN5A-related patients from non-SCN5A-related patients. *J Am Coll Cardiol* 2002;40:350-6.
128. Probst V, Wilde AA, Barc J, et al. SCN5A mutations and the role of genetic background in the pathophysiology of Brugada syndrome. *Circ Cardiovasc Genet* 2009;2:552-7.
129. Hoogendijk MG, Potse M, Linnenbank AC, et al. Mechanism of right precordial ST-segment elevation in structural heart disease: excitation failure by current-to-load mismatch. *Heart Rhythm* 2010;7:238-48.
130. Rhodes TH, Vanoye CG, Ohmori I, Ogiwara I, Yamakawa K, George AL, Jr. Sodium channel dysfunction in intractable childhood epilepsy with generalized tonic-clonic seizures. *J Physiol* 2005;569:433-45.
131. Fujiwara T, Sugawara T, Mazaki-Miyazaki E, et al. Mutations of sodium channel alpha subunit type 1 (SCN1A) in intractable childhood epilepsies with frequent generalized tonic-clonic seizures. *Brain* 2003;126:531-46.
132. Sayeed MZ, Salam MA, Haque MZ, Islam AK. Brugada syndrome with a novel missense mutation in SCN5A gene: a case report from Bangladesh. *Indian Heart J* 2014;66:104-7.
133. Lampert A, Dib-Hajj SD, Eastman EM, et al. Erythromelalgia mutation L823R shifts activation and inactivation of threshold sodium channel Nav1.7 to hyperpolarized potentials. *Biochemical and Biophysical Research Communications* 2009;390:319-24.
134. Volkens L, Kahlig KM, Verbeek NE, et al. Nav 1.1 dysfunction in genetic epilepsy with febrile seizures-plus or Dravet syndrome. *Eur J Neurosci* 2011;34:1268-75.
135. Kuzmenkin A, Muncan V, Jurkat-Rott K, et al. Enhanced inactivation and pH sensitivity of Na(+) channel mutations causing hypokalaemic periodic paralysis type II. *Brain* 2002;125:835-43.
136. Bulman DE, Scoggan KA, van Oene MD, et al. A novel sodium channel mutation in a family with hypokalemic periodic paralysis. *Neurology* 1999;53:1932-6.
137. Depienne C, Trouillard O, Saint-Martin C, et al. Spectrum of SCN1A gene mutations associated with Dravet syndrome: analysis of 333 patients. *J Med Genet* 2009;46:183-91.
138. Kotta CM, Anastasakis A, Gatzoulis K, Manolis AS, Stefanadis C. Novel sodium channel SCN5A mutations in Brugada syndrome patients from Greece. *Int J Cardiol* 2010;145:45-8.
139. Wu B, Zhang Y, Tang H, et al. A Novel SCN9A Mutation (F826Y) in Primary Erythromelalgia Alters the Excitability of Nav1.7. *Curr Mol Med* 2017;17:450-7.
140. Jurkat-Rott K, Mitrovic N, Hang C, et al. Voltage-sensor sodium channel mutations cause hypokalemic periodic paralysis type 2 by enhanced inactivation and reduced current. *Proc Natl Acad Sci U S A* 2000;97:9549-54.
141. Berecki G, Howell KB, Deerasooriya YH, et al. Dynamic action potential clamp predicts functional separation in mild familial and severe de novo forms of SCN2A epilepsy. *Proc Natl Acad Sci U S A* 2018;115:E5516-e25.
142. Mason ER, Wu F, Patel RR, Xiao Y, Cannon SC, Cummins TR. Resurgent and Gating Pore Currents Induced by De Novo SCN2A Epilepsy Mutations. *eNeuro* 2019;6.
143. Epi K, Allen AS, Berkovic SF, et al. De novo mutations in epileptic encephalopathies. *Nature* 2013;501:217-21.
144. Samanta D, Ramakrishnaiah R. De novo R853Q mutation of SCN2A gene and West syndrome. *Acta Neurol Belg* 2015;115:773-6.

145. Kobayashi Y, Tohyama J, Kato M, et al. High prevalence of genetic alterations in early-onset epileptic encephalopathies associated with infantile movement disorders. *Brain Dev* 2016;38:285-92.
146. Li J, Cai T, Jiang Y, et al. Genes with de novo mutations are shared by four neuropsychiatric disorders discovered from NPdenovo database. *Mol Psychiatry* 2016;21:298.
147. Zaman T, Helbig KL, Clatot J, et al. SCN3A-related neurodevelopmental disorder: A spectrum of epilepsy and brain malformation. *Ann Neurol* 2020;88:348-62.
148. Wang L, Meng X, Yuchi Z, et al. De Novo Mutation in the SCN5A Gene Associated with Brugada Syndrome. *Cell Physiol Biochem* 2015;36:2250-62.
149. Wu L, Zhang B, Kang Y, Wu W. Enhanced slow inactivation of the human skeletal muscle sodium channel causing normokalemic periodic paralysis. *Cell Mol Neurobiol* 2014;34:707-14.
150. Frigo G, Rampazzo A, Baucé B, et al. Homozygous SCN5A mutation in Brugada syndrome with monomorphic ventricular tachycardia and structural heart abnormalities. *Europace* 2007;9:391-7.
151. Itoh H, Berthet M, Fressart V, et al. Asymmetry of parental origin in long QT syndrome: preferential maternal transmission of KCNQ1 variants linked to channel dysfunction. *Eur J Hum Genet* 2016;24:1160-6.
152. Kinoshita K, Takahashi H, Hata Y, et al. SCN5A(K817E), a novel Brugada syndrome-associated mutation that alters the activation gating of Nav1.5 channel. *Heart Rhythm* 2016;13:1113-20.
153. Lossin C, Wang DW, Rhodes TH, Vanoye CG, George AL, Jr. Molecular basis of an inherited epilepsy. *Neuron* 2002;34:877-84.
154. Escayg A, MacDonald BT, Meisler MH, et al. Mutations of SCN1A, encoding a neuronal sodium channel, in two families with GEFS+2. *Nat Genet* 2000;24:343-5.
155. Zaman T, Helbig I, Bozovic IB, et al. Mutations in SCN3A cause early infantile epileptic encephalopathy. *Ann Neurol* 2018;83:703-17.
156. Miyatake S, Kato M, Sawaishi Y, et al. Recurrent SCN3A p.Ile875Thr variant in patients with polymicrogyria. *Ann Neurol* 2018;84:159-61.
157. Cummins TR, Dib-Hajj SD, Waxman SG. Electrophysiological properties of mutant Nav1.7 sodium channels in a painful inherited neuropathy. *J Neurosci* 2004;24:8232-6.
158. Namer B, Orstavik K, Schmidt R, et al. Specific changes in conduction velocity recovery cycles of single nociceptors in a patient with erythromelalgia with the I848T gain-of-function mutation of Nav1.7. *Pain* 2015;156:1637-46.
159. Theille JW, Jarecki BW, Piekarczyk AD, Cummins TR. Nav1.7 mutations associated with paroxysmal extreme pain disorder, but not erythromelalgia, enhance Navbeta4 peptide-mediated resurgent sodium currents. *J Physiol* 2011;589:597-608.
160. Yang Y, Wang Y, Li S, et al. Mutations in SCN9A, encoding a sodium channel alpha subunit, in patients with primary erythromelalgia. *J Med Genet* 2004;41:171-4.
161. Han C, Yang Y, de Greef BT, et al. The Domain II S4-S5 Linker in Nav1.9: A Missense Mutation Enhances Activation, Impairs Fast Inactivation, and Produces Human Painful Neuropathy. *Neuromolecular Med* 2015;17:158-69.
162. Hoeijmakers JG, Han C, Merkies IS, et al. Small nerve fibres, small hands and small feet: a new syndrome of pain, dysautonomia and acromesomelia in a kindred with a novel Nav1.7 mutation. *Brain* 2012;135:345-58.

163. Tanaka BS, Nguyen PT, Zhou EY, et al. Gain-of-function mutation of a voltage-gated sodium channel Na(V)1.7 associated with peripheral pain and impaired limb development. *J Biol Chem* 2017;292:9262-72.
164. Han C, Rush AM, Dib-Hajj SD, et al. Sporadic onset of erythromelgia: a gain-of-function mutation in Nav1.7. *Ann Neurol* 2006;59:553-8.
165. Han C, Lampert A, Rush AM, et al. Temperature dependence of erythromelgia mutation L858F in sodium channel Nav1.7. *Mol Pain* 2007;3:3.
166. Cregg R, Cox JJ, Bennett DL, Wood JN, Werdehausen R. Mexiletine as a treatment for primary erythromelgia: normalization of biophysical properties of mutant L858F Nav 1.7 sodium channels. *Br J Pharmacol* 2014;171:4455-63.
167. Estacion M, Waxman SG, Dib-Hajj SD. Effects of ranolazine on wild-type and mutant hNav1.7 channels and on DRG neuron excitability. *Mol Pain* 2010;6:35.
168. Vasylyev DV, Han C, Zhao P, Dib-Hajj S, Waxman SG. Dynamic-clamp analysis of wild-type human Nav1.7 and erythromelgia mutant channel L858H. *J Neurophysiol* 2014;111:1429-43.
169. Bendahhou S, Cummins TR, Tawil R, Waxman SG, Ptáček LJ. Activation and inactivation of the voltage-gated sodium channel: role of segment S5 revealed by a novel hyperkalaemic periodic paralysis mutation. *J Neurosci* 1999;19:4762-71.
170. Huang S, Zhang W, Chang X, Guo J. Overlap of periodic paralysis and paramyotonia congenita caused by SCN4A gene mutations two family reports and literature review. *Channels (Austin)* 2019;13:110-9.
171. Harty TP, Dib-Hajj SD, Tyrrell L, et al. Na(V)1.7 mutant A863P in erythromelgia: effects of altered activation and steady-state inactivation on excitability of nociceptive dorsal root ganglion neurons. *J Neurosci* 2006;26:12566-75.
172. Rhodes TH, Lossin C, Vanoye CG, Wang DW, George AL, Jr. Noninactivating voltage-gated sodium channels in severe myoclonic epilepsy of infancy. *Proc Natl Acad Sci U S A* 2004;101:11147-52.
173. Ohmori I, Ouchida M, Ohtsuka Y, Oka E, Shimizu K. Significant correlation of the SCN1A mutations and severe myoclonic epilepsy in infancy. *Biochem Biophys Res Commun* 2002;295:17-23.
174. Choi JS, Zhang L, Dib-Hajj SD, et al. Mexiletine-responsive erythromelgia due to a new Na(v)1.7 mutation showing use-dependent current fall-off. *Exp Neurol* 2009;216:383-9.
175. Stadler T, O'Reilly AO, Lampert A. Erythromelgia mutation Q875E Stabilizes the activated state of sodium channel Nav1.7. *J Biol Chem* 2015;290:6316-25.
176. Skeik N, Rooke TW, Davis MD, et al. Severe case and literature review of primary erythromelgia: novel SCN9A gene mutation. *Vasc Med* 2012;17:44-9.
177. Cox JJ, Sheynin J, Shorer Z, et al. Congenital insensitivity to pain: novel SCN9A missense and in-frame deletion mutations. *Hum Mutat* 2010;31:E1670-86.
178. Tarradas A, Selga E, Beltran-Alvarez P, et al. A novel missense mutation, I890T, in the pore region of cardiac sodium channel causes Brugada syndrome. *PLoS One* 2013;8:e53220.
179. Savastano S, Rordorf R, Vicentini A, et al. A comprehensive electrocardiographic, molecular, and echocardiographic study of Brugada syndrome: validation of the 2013 diagnostic criteria. *Heart Rhythm* 2014;11:1176-83.
180. Begemann A, Acuna MA, Zweier M, et al. Further corroboration of distinct functional features in SCN2A variants causing intellectual disability or epileptic phenotypes. *Mol Med* 2019;25:6.

181. Rauch A, Wieczorek D, Graf E, et al. Range of genetic mutations associated with severe non-syndromic sporadic intellectual disability: an exome sequencing study. *Lancet* 2012;380:1674-82.
182. Liao WP, Shi YW, Long YS, et al. Partial epilepsy with antecedent febrile seizures and seizure aggravation by antiepileptic drugs: associated with loss of function of Na(v) 1.1. *Epilepsia* 2010;51:1669-78.
183. Pambrun T, Mercier A, Chatelier A, et al. Myotonic dystrophy type 1 mimics and exacerbates Brugada phenotype induced by Nav1.5 sodium channel loss-of-function mutation. *Heart Rhythm* 2014;11:1393-400.
184. Sugawara T, Tsurubuchi Y, Fujiwara T, et al. Nav1.1 channels with mutations of severe myoclonic epilepsy in infancy display attenuated currents. *Epilepsy Res* 2003;54:201-7.
185. Wagnon JL, Barker BS, Ottolini M, et al. Loss-of-function variants of SCN8A in intellectual disability without seizures. *Neurol Genet* 2017;3:e170.
186. Lossin C, Rhodes TH, Desai RR, et al. Epilepsy-associated dysfunction in the voltage-gated neuronal sodium channel SCN1A. *J Neurosci* 2003;23:11289-95.
187. Thompson CH, Porter JC, Kahlig KM, Daniels MA, George AL, Jr. Nontruncating SCN1A mutations associated with severe myoclonic epilepsy of infancy impair cell surface expression. *J Biol Chem* 2012;287:42001-8.
188. Claes L, Del-Favero J, Ceulemans B, Lagae L, Van Broeckhoven C, De Jonghe P. De novo mutations in the sodium-channel gene SCN1A cause severe myoclonic epilepsy of infancy. *Am J Hum Genet* 2001;68:1327-32.
189. Elia N, Nault T, McMillan HJ, Graham GE, Huang L, Cannon SC. Myotonic Myopathy With Secondary Joint and Skeletal Anomalies From the c.2386C>G, p.L769V Mutation in SCN4A. *Front Neurol* 2020;11:77.
190. Ruan Y, Liu N, Bloise R, Napolitano C, Priori SG. Gating properties of SCN5A mutations and the response to mexiletine in long-QT syndrome type 3 patients. *Circulation* 2007;116:1137-44.
191. Schwartz PJ, Priori SG, Dumaine R, et al. A Molecular Link between the Sudden Infant Death Syndrome and the Long-QT Syndrome. *New England Journal of Medicine* 2000;343:262-7.
192. Huang J, Estacion M, Zhao P, et al. A Novel Gain-of-Function Nav1.9 Mutation in a Child With Episodic Pain. *Front Neurosci* 2019;13:918.
193. Miao P, Tang S, Ye J, et al. Electrophysiological features: The next precise step for SCN2A developmental epileptic encephalopathy. *Mol Genet Genomic Med* 2020:e1250.
194. Hsueh CH, Chen WP, Lin JL, et al. Distinct functional defect of three novel Brugada syndrome related cardiac sodium channel mutations. *J Biomed Sci* 2009;16:23.
195. Splawski I, Timothy KW, Tateyama M, et al. Variant of SCN5A sodium channel implicated in risk of cardiac arrhythmia. *Science* 2002;297:1333-6.
196. Plant LD, Bowers PN, Liu Q, et al. A common cardiac sodium channel variant associated with sudden infant death in African Americans, SCN5A S1103Y. *J Clin Invest* 2006;116:430-5.
197. Cestele S, Labate A, Rusconi R, et al. Divergent effects of the T1174S SCN1A mutation associated with seizures and hemiplegic migraine. *Epilepsia* 2013;54:927-35.
198. Estacion M, Harty TP, Choi JS, Tyrrell L, Dib-Hajj SD, Waxman SG. A sodium channel gene SCN9A polymorphism that increases nociceptor excitability. *Ann Neurol* 2009;66:862-6.

199. Winkel BG, Larsen MK, Berge KE, et al. The prevalence of mutations in KCNQ1, KCNH2, and SCN5A in an unselected national cohort of young sudden unexplained death cases. *J Cardiovasc Electrophysiol* 2012;23:1092-8.
200. Escayg A, Heils A, MacDonald BT, Haug K, Sander T, Meisler MH. A Novel SCN1A Mutation Associated with Generalized Epilepsy with Febrile Seizures Plus—and Prevalence of Variants in Patients with Epilepsy. *The American Journal of Human Genetics* 2001;68:866-73.
201. Marini C, Mei D, Temudo T, et al. Idiopathic Epilepsies with Seizures Precipitated by Fever and SCN1A Abnormalities. *Epilepsia* 2007;48:1678-85.
202. Wang Q, Chen S, Chen Q, et al. The common SCN5A mutation R1193Q causes LQTS-type electrophysiological alterations of the cardiac sodium channel. *J Med Genet* 2004;41:e66.
203. Huang H, Zhao J, Barrane FZ, Champagne J, Chahine M. Nav1.5/R1193Q polymorphism is associated with both long QT and Brugada syndromes. *Can J Cardiol* 2006;22:309-13.
204. Abdelsayed M, Peters CH, Ruben PC. Differential thermosensitivity in mixed syndrome cardiac sodium channel mutants. *J Physiol* 2015;593:4201-23.
205. Abe M, Kinoshita K, Matsuoka K, et al. Lack of modulatory effect of the SCN5A R1193Q polymorphism on cardiac fast Na⁺ current at body temperature. *PLoS One* 2018;13:e0207437.
206. Li L, Ruan Y, Liu N, et al. "Pill-in-the-Pocket" Treatment of Propafenone Unmasks ECG Brugada Pattern in an Atrial Fibrillation Patient With a Common SCN5A R1193Q Polymorphism. *Front Physiol* 2019;10:353.
207. Takahata T, Yasui-Furukori N, Sasaki S, et al. Nucleotide changes in the translated region of SCN5A from Japanese patients with Brugada syndrome and control subjects. *Life Sci* 2003;72:2391-9.
208. Ogiwara I, Ito K, Sawaishi Y, et al. De novo mutations of voltage-gated sodium channel alphaII gene SCN2A in intractable epilepsies. *Neurology* 2009;73:1046-53.
209. Schulze-Bahr E, Eckardt L, Breithardt G, et al. Sodium channel gene (SCN5A) mutations in 44 index patients with Brugada syndrome: different incidences in familial and sporadic disease. *Hum Mutat* 2003;21:651-2.
210. Crotti L, Marcou CA, Tester DJ, et al. Spectrum and prevalence of mutations involving BrS1- through BrS12-susceptibility genes in a cohort of unrelated patients referred for Brugada syndrome genetic testing: implications for genetic testing. *J Am Coll Cardiol* 2012;60:1410-8.
211. Baroudi G, Acharfi S, Larouche C, Chahine M. Expression and intracellular localization of an SCN5A double mutant R1232W/T1620M implicated in Brugada syndrome. *Circ Res* 2002;90:E11-6.
212. Makita N, Mochizuki N, Tsutsui H. Absence of a trafficking defect in R1232W/T1620M, a double SCN5A mutant responsible for Brugada syndrome. *Circ J* 2008;72:1018-9.
213. Chen Q, Kirsch GE, Zhang D, et al. Genetic basis and molecular mechanism for idiopathic ventricular fibrillation. *Nature* 1998;392:293-6.
214. Peters C, Rosch RE, Hughes E, Ruben PC. Temperature-dependent changes in neuronal dynamics in a patient with an SCN1A mutation and hyperthermia induced seizures. *Sci Rep* 2016;6:31879.
215. Hermida JS, Arnalsteen-Dassonville E, Kubala M, et al. Dual phenotypic transmission in Brugada syndrome. *Arch Cardiovasc Dis* 2013;106:366-72.
216. Zaytseva AK, Karpushev AV, Kiselev AM, et al. Characterization of a novel SCN5A genetic variant A1294G associated with mixed clinical phenotype. *Biochem Biophys Res Commun* 2019;516:777-83.

217. Abriel H, Cabo C, Wehrens XH, et al. Novel arrhythmogenic mechanism revealed by a long-QT syndrome mutation in the cardiac Na⁺ channel. *Circ Res* 2001;88:740-5.
218. Huang J, Yang Y, Dib-Hajj SD, et al. Depolarized inactivation overcomes impaired activation to produce DRG neuron hyperexcitability in a Nav1.7 mutation in a patient with distal limb pain. *J Neurosci* 2014;34:12328-40.
219. Olesen MS, Yuan L, Liang B, et al. High prevalence of long QT syndrome-associated SCN5A variants in patients with early-onset lone atrial fibrillation. *Circ Cardiovasc Genet* 2012;5:450-9.
220. Lossin C, Shi X, Rogawski MA, Hirose S. Compromised function in the Na(v)1.2 Dravet syndrome mutation R1312T. *Neurobiol Dis* 2012;47:378-84.
221. Shi X, Yasumoto S, Nakagawa E, Fukasawa T, Uchiya S, Hirose S. Missense mutation of the sodium channel gene SCN2A causes Dravet syndrome. *Brain Dev* 2009;31:758-62.
222. Misra SN, Kahlig KM, George AL, Jr. Impaired Nav1.2 function and reduced cell surface expression in benign familial neonatal-infantile seizures. *Epilepsia* 2008;49:1535-45.
223. Berkovic SF, Heron SE, Giordano L, et al. Benign familial neonatal-infantile seizures: characterization of a new sodium channelopathy. *Ann Neurol* 2004;55:550-7.
224. Casini S, Tan HL, Bhuiyan ZA, et al. Characterization of a novel SCN5A mutation associated with Brugada syndrome reveals involvement of DIIIS4-S5 linker in slow inactivation. *Cardiovasc Res* 2007;76:418-29.
225. Jarecki BW, Sheets PL, Jackson JO, 2nd, Cummins TR. Paroxysmal extreme pain disorder mutations within the D3/S4-S5 linker of Nav1.7 cause moderate destabilization of fast inactivation. *J Physiol* 2008;586:4137-53.
226. Cheng X, Dib-Hajj SD, Tyrrell L, Wright DA, Fischer TZ, Waxman SG. Mutations at opposite ends of the DIII/S4-S5 linker of sodium channel Na V 1.7 produce distinct pain disorders. *Mol Pain* 2010;6:24.
227. Fertleman CR, Baker MD, Parker KA, et al. SCN9A mutations in paroxysmal extreme pain disorder: allelic variants underlie distinct channel defects and phenotypes. *Neuron* 2006;52:767-74.
228. Wang DW, Yazawa K, George AL, Jr., Bennett PB. Characterization of human cardiac Na⁺ channel mutations in the congenital long QT syndrome. *Proc Natl Acad Sci U S A* 1996;93:13200-5.
229. Tian XL, Yong SL, Wan X, et al. Mechanisms by which SCN5A mutation N1325S causes cardiac arrhythmias and sudden death in vivo. *Cardiovasc Res* 2004;61:256-67.
230. Li G, Woltz RL, Wang CY, et al. Gating Properties of Mutant Sodium Channels and Responses to Sodium Current Inhibitors Predict Mexiletine-Sensitive Mutations of Long QT Syndrome 3. *Front Pharmacol* 2020;11:1182.
231. Wang Q, Shen J, Li Z, et al. Cardiac sodium channel mutations in patients with long QT syndrome, an inherited cardiac arrhythmia. *Hum Mol Genet* 1995;4:1603-7.
232. Heron SE, Crossland KM, Andermann E, et al. Sodium-channel defects in benign familial neonatal-infantile seizures. *Lancet* 2002;360:851-2.
233. Turker I, Makiyama T, Vatta M, et al. A Novel SCN5A Mutation Associated with Drug Induced Brugada Type ECG. *PLoS One* 2016;11:e0161872.
234. Wedekind H, Smits JP, Schulze-Bahr E, et al. De novo mutation in the SCN5A gene associated with early onset of sudden infant death. *Circulation* 2001;104:1158-64.

235. Berecki G, Zegers JG, Bhuiyan ZA, Verkerk AO, Wilders R, van Ginneken AC. Long-QT syndrome-related sodium channel mutations probed by the dynamic action potential clamp technique. *J Physiol* 2006;570:237-50.
236. Palmio J, Sandell S, Hanna MG, Männikkö R, Penttilä S, Udd B. Predominantly myalgic phenotype caused by the c.3466G>A p.A1156T mutation in SCN4A gene. *Neurology* 2017;88:1520-7.
237. McClatchey AI, McKenna-Yasek D, Cros D, et al. Novel mutations in families with unusual and variable disorders of the skeletal muscle sodium channel. *Nat Genet* 1992;2:148-52.
238. Smits JP, Veldkamp MW, Bezzina CR, et al. Substitution of a conserved alanine in the domain IIIIS4-S5 linker of the cardiac sodium channel causes long QT syndrome. *Cardiovasc Res* 2005;67:459-66.
239. Desaphy JF, Carbonara R, D'Amico A, et al. Translational approach to address therapy in myotonia permanens due to a new SCN4A mutation. *Neurology* 2016;86:2100-8.
240. Schulze-Bahr E, Fenge H, Etzrodt D, et al. Long QT syndrome and life threatening arrhythmia in a newborn: molecular diagnosis and treatment response. *Heart* 2004;90:13-6.
241. Sugiura Y, Makita N, Li L, et al. Cold induces shifts of voltage dependence in mutant SCN4A, causing hypokalemic periodic paralysis. *Neurology* 2003;61:914-8.
242. Webb J, Cannon SC. Cold-induced defects of sodium channel gating in atypical periodic paralysis plus myotonia. *Neurology* 2008;70:755-61.
243. Huang H, Millat G, Rodriguez-Lafrasse C, et al. Biophysical characterization of a new SCN5A mutation S1333Y in a SIDS infant linked to long QT syndrome. *FEBS Lett* 2009;583:890-6.
244. Barker BS, Ottolini M, Wagnon JL, Hollander RM, Meisler MH, Patel MK. The SCN8A encephalopathy mutation p.Ile1327Val displays elevated sensitivity to the anticonvulsant phenytoin. *Epilepsia* 2016;57:1458-66.
245. Vaher U, Noukas M, Nikopentis T, et al. De novo SCN8A mutation identified by whole-exome sequencing in a boy with neonatal epileptic encephalopathy, multiple congenital anomalies, and movement disorders. *J Child Neurol* 2014;29:NP202-6.
246. Singh R, Jayapal S, Goyal S, Jungbluth H, Lascelles K. Early-onset movement disorder and epileptic encephalopathy due to de novo dominant SCN8A mutation. *Seizure* 2015;26:69-71.
247. Patel RR, Barbosa C, Brustovetsky T, Brustovetsky N, Cummins TR. Aberrant epilepsy-associated mutant Nav1.6 sodium channel activity can be targeted with cannabidiol. *Brain* 2016;139:2164-81.
248. Carvill GL, Heavin SB, Yendle SC, et al. Targeted resequencing in epileptic encephalopathies identifies de novo mutations in CHD2 and SYNGAP1. *Nat Genet* 2013;45:825-30.
249. Hackenberg A, Baumer A, Sticht H, et al. Infantile epileptic encephalopathy, transient choreoathetotic movements, and hypersomnia due to a De Novo missense mutation in the SCN2A gene. *Neuropediatrics* 2014;45:261-4.
250. Matalon D, Goldberg E, Medne L, Marsh ED. Confirming an expanded spectrum of SCN2A mutations: a case series. *Epileptic Disord* 2014;16:13-8.
251. Dimassi S, Labalme A, Ville D, et al. Whole-exome sequencing improves the diagnosis yield in sporadic infantile spasm syndrome. *Clin Genet* 2016;89:198-204.
252. Wu MT, Huang PY, Yen CT, Chen CC, Lee MJ. A novel SCN9A mutation responsible for primary erythromelalgia and is resistant to the treatment of sodium channel blockers. *PLoS One* 2013;8:e55212.

253. Estacion M, Yang Y, Dib-Hajj SD, et al. A new Nav1.7 mutation in an erythromelalgia patient. *Biochem Biophys Res Commun* 2013;432:99-104.
254. Huang CW, Lai HJ, Huang PY, Lee MJ, Kuo CC. The Biophysical Basis Underlying Gating Changes in the p.V1316A Mutant Nav1.7 Channel and the Molecular Pathogenesis of Inherited Erythromelalgia. *PLoS Biol* 2016;14:e1002561.
255. Leipold E, Hanson-Kahn A, Frick M, et al. Cold-aggravated pain in humans caused by a hyperactive Nav1.9 channel mutant. *Nat Commun* 2015;6:10049.
256. Samani K, Wu G, Ai T, et al. A novel SCN5A mutation V1340I in Brugada syndrome augmenting arrhythmias during febrile illness. *Heart Rhythm* 2009;6:1318-26.
257. Wallace RH, Scheffer IE, Barnett S, et al. Neuronal sodium-channel alpha1-subunit mutations in generalized epilepsy with febrile seizures plus. *Am J Hum Genet* 2001;68:859-65.
258. Keller DI, Huang H, Zhao J, et al. A novel SCN5A mutation, F1344S, identified in a patient with Brugada syndrome and fever-induced ventricular fibrillation. *Cardiovasc Res* 2006;70:521-9.
259. Lee YS, Baek JS, Kim SY, et al. Childhood brugada syndrome in two korean families. *Korean Circ J* 2010;40:143-7.
260. Osaka H, Ogiwara I, Mazaki E, et al. Patients with a sodium channel alpha 1 gene mutation show wide phenotypic variation. *Epilepsy Res* 2007;75:46-51.
261. Rudnik-Schöneborn S, Schaupp M, Lindner A, et al. Brugada-like cardiac disease in myotonic dystrophy type 2: report of two unrelated patients. *Eur J Neurol* 2011;18:191-4.
262. Zumhagen S, Zeidler EM, Stallmeyer B, Ernsting M, Eckardt L, Schulze-Bahr E. Tpeak-Tend interval and Tpeak-Tend/QT ratio in patients with Brugada syndrome. *Europace* 2016;18:1866-72.
263. Tan BH, Valdivia CR, Song C, Makielski JC. Partial expression defect for the SCN5A missense mutation G1406R depends on splice variant background Q1077 and rescue by mexiletine. *Am J Physiol Heart Circ Physiol* 2006;291:H1822-8.
264. Kyndt F, Probst V, Potet F, et al. Novel SCN5A mutation leading either to isolated cardiac conduction defect or Brugada syndrome in a large French family. *Circulation* 2001;104:3081-6.
265. Kim HW, Quan Z, Kim YB, et al. Differential effects on sodium current impairments by distinct SCN1A mutations in GABAergic neurons derived from Dravet syndrome patients. *Brain Dev* 2018;40:287-98.
266. Hermida JS, Dassonville E, Six I, et al. Prospective evaluation of the familial prevalence of the brugada syndrome. *Am J Cardiol* 2010;106:1758-62.
267. Zhu JF, Du LL, Tian Y, et al. Novel heterozygous mutation c.4282G>T in the SCN5A gene in a family with Brugada syndrome. *Exp Ther Med* 2015;9:1639-45.
268. Maury P, Moreau A, Hidden-Lucet F, et al. Novel SCN5A mutations in two families with "Brugada-like" ST elevation in the inferior leads and conduction disturbances. *J Interv Card Electrophysiol* 2013;37:131-40.
269. Deschenes I, Baroudi G, Berthet M, et al. Electrophysiological characterization of SCN5A mutations causing long QT (E1784K) and Brugada (R1512W and R1432G) syndromes. *Cardiovasc Res* 2000;46:55-65.
270. Hothi SS, Ara F, Timperley J. p.Y1449C SCN5A mutation associated with overlap disorder comprising conduction disease, Brugada syndrome, and atrial flutter. *J Cardiovasc Electrophysiol* 2015;26:93-7.
271. Sacilotto L, Epifanio HB, Darrieux FC, et al. Compound Heterozygous SCN5A Mutations in a Toddler - Are they Associated with a More Severe Phenotype? *Arq Bras Cardiol* 2017;108:70-3.

272. Farinato A, Altamura C, Imbrici P, et al. Pharmacogenetics of myotonic hNav1.4 sodium channel variants situated near the fast inactivation gate. *Pharmacol Res* 2019;141:224-35.
273. Koch MC, Baumbach K, George AL, Ricker K. Paramyotonia congenita without paralysis on exposure to cold: a novel mutation in the SCN4A gene (Val1293Ile). *Neuroreport* 1995;6:2001-4.
274. Gay S, Dupuis D, Faivre L, et al. Severe neonatal non-dystrophic myotonia secondary to a novel mutation of the voltage-gated sodium channel (SCN4A) gene. *Am J Med Genet A* 2008;146a:380-3.
275. Maggi L, Ravaglia S, Farinato A, et al. Coexistence of CLCN1 and SCN4A mutations in one family suffering from myotonia. *Neurogenetics* 2017;18:219-25.
276. Bankston JR, Yue M, Chung W, et al. A novel and lethal de novo LQT-3 mutation in a newborn with distinct molecular pharmacology and therapeutic response. *PLoS One* 2007;2:e1258.
277. Cai ZQ, Li WP, Chen X, et al. [The reverse effects of allitridum on sodium current decrease caused by SCN5A-F1473S mutation]. *Yao Xue Xue Bao* 2016;51:1852-7.
278. Ruan Y, Denegri M, Liu N, et al. Trafficking Defects and Gating Abnormalities of a Novel *SCN5A* Mutation Question Gene-Specific Therapy in Long QT Syndrome Type 3. *Circulation Research* 2010;106:1374-83.
279. Dib-Hajj SD, Rush AM, Cummins TR, et al. Gain-of-function mutation in Nav1.7 in familial erythromelalgia induces bursting of sensory neurons. *Brain* 2005;128:1847-54.
280. Gurkiewicz M, Korngreen A, Waxman SG, Lampert A. Kinetic modeling of Nav1.7 provides insight into erythromelalgia-associated F1449V mutation. *J Neurophysiol* 2011;105:1546-57.
281. Gando I, Campana C, Tan RB, Cecchin F, Sobie EA, Coetzee WA. A distinct molecular mechanism by which phenytoin rescues a novel long QT 3 variant. *J Mol Cell Cardiol* 2020;144:1-11.
282. Tan RB, Chakravarti S, Busovsky-McNeal M, Walsh A, Cecchin F. Complexity of ranolazine and phenytoin use in an infant with long QT syndrome type 3. *HeartRhythm Case Rep* 2017;3:104-8.
283. Barbieri R, Bertelli S, Pusch M, Gavazzo P. Late sodium current blocker GS967 inhibits persistent currents induced by familial hemiplegic migraine type 3 mutations of the SCN1A gene. *J Headache Pain* 2019;20:107.
284. Vahedi K, Depienne C, Le Fort D, et al. Elicited repetitive daily blindness. A new phenotype associated with hemiplegic migraine and *SCN1A* mutations 2009;72:1178-83.
285. Cestele S, Scalmani P, Rusconi R, Terragni B, Franceschetti S, Mantegazza M. Self-limited hyperexcitability: functional effect of a familial hemiplegic migraine mutation of the Nav1.1 (SCN1A) Na⁺ channel. *J Neurosci* 2008;28:7273-83.
286. Moreau A, Krahn AD, Gosselin-Badaroudine P, et al. Sodium overload due to a persistent current that attenuates the arrhythmogenic potential of a novel LQT3 mutation. *Front Pharmacol* 2013;4:126.
287. Lerche H, Heine R, Pika U, et al. Human sodium channel myotonia: slowed channel inactivation due to substitutions for a glycine within the III-IV linker. *J Physiol* 1993;470:13-22.
288. Liu Y, Schubert J, Sonnenberg L, et al. Neuronal mechanisms of mutations in SCN8A causing epilepsy or intellectual disability. *Brain* 2019;142:376-90.
289. Parrini E, Marini C, Mei D, et al. Diagnostic Targeted Resequencing in 349 Patients with Drug-Resistant Pediatric Epilepsies Identifies Causative Mutations in 30 Different Genes. *Hum Mutat* 2017;38:216-25.

290. Wang J, Gao H, Bao X, et al. SCN8A mutations in Chinese patients with early onset epileptic encephalopathy and benign infantile seizures. *BMC Med Genet* 2017;18:104.
291. Xiao Y, Xiong J, Mao D, et al. Early-onset epileptic encephalopathy with de novo SCN8A mutation. *Epilepsy Res* 2018;139:9-13.
292. Gardella E, Marini C, Trivisano M, et al. The phenotype of SCN8A developmental and epileptic encephalopathy. *Neurology* 2018;91:e1112-e24.
293. Weller CM, Pelzer N, de Vries B, et al. Two novel SCN1A mutations identified in families with familial hemiplegic migraine. *Cephalalgia* 2014;34:1062-9.
294. Domitrz I, Kosiorek M, Żekanowski C, Kamińska A. Genetic studies of Polish migraine patients: screening for causative mutations in four migraine-associated genes. *Hum Genomics* 2016;10:3.
295. Bouhours M, Sternberg D, Davoine CS, et al. Functional characterization and cold sensitivity of T1313A, a new mutation of the skeletal muscle sodium channel causing paramyotonia congenita in humans. *J Physiol* 2004;554:635-47.
296. Saber S, Amarouch MY, Fazelifar AF, et al. Complex genetic background in a large family with Brugada syndrome. *Physiol Rep* 2015;3.
297. Zheng J, Zhou F, Su T, et al. The biophysical characterization of the first SCN5A mutation R1512W identified in Chinese sudden unexplained nocturnal death syndrome. *Medicine (Baltimore)* 2016;95:e3836.
298. Dharmawan T, Nakajima T, Iizuka T, et al. Enhanced closed-state inactivation of mutant cardiac sodium channels (SCN5A N1541D and R1632C) through different mechanisms. *J Mol Cell Cardiol* 2019;130:88-95.
299. Ke Q, Ye J, Tang S, et al. N1366S mutation of human skeletal muscle sodium channel causes paramyotonia congenita. *J Physiol* 2017;595:6837-50.
300. Lewis TB, Shevell MI, Andermann E, Ryan SG, Leach RJ. Evidence of a third locus for benign familial convulsions. *J Child Neurol* 1996;11:211-4.
301. Cregg R, Laguda B, Werdehausen R, et al. Novel mutations mapping to the fourth sodium channel domain of Nav1.7 result in variable clinical manifestations of primary erythromelalgia. *Neuromolecular Med* 2013;15:265-78.
302. Ohmori I, Ouchida M, Kobayashi K, et al. Rasmussen encephalitis associated with SCN 1 A mutation. *Epilepsia* 2008;49:521-6.
303. Harkin LA, McMahon JM, Iona X, et al. The spectrum of SCN1A-related infantile epileptic encephalopathies. *Brain* 2007;130:843-52.
304. Kim YO, Bellows S, McMahon JM, et al. Atypical multifocal Dravet syndrome lacks generalized seizures and may show later cognitive decline. *Dev Med Child Neurol* 2014;56:85-90.
305. Lauxmann S, Boutry-Kryza N, Rivier C, et al. An SCN2A mutation in a family with infantile seizures from Madagascar reveals an increased subthreshold Na⁽⁺⁾ current. *Epilepsia* 2013;54:e117-21.
306. Wang DW, Viswanathan PC, Balsler JR, George AL, Jr., Benson DW. Clinical, genetic, and biophysical characterization of SCN5A mutations associated with atrioventricular conduction block. *Circulation* 2002;105:341-6.
307. Fan C, Wolking S, Lehmann-Horn F, et al. Early-onset familial hemiplegic migraine due to a novel SCN1A mutation. *Cephalalgia* 2016;36:1238-47.

308. Surber R, Hensellek S, Prochnau D, et al. Combination of cardiac conduction disease and long QT syndrome caused by mutation T1620K in the cardiac sodium channel. *Cardiovasc Res* 2008;77:740-8.
309. Wang DW, Makita N, Kitabatake A, Balsler JR, George AL, Jr. Enhanced Na(+) channel intermediate inactivation in Brugada syndrome. *Circ Res* 2000;87:E37-43.
310. Kambouris NG, Nuss HB, Johns DC, Tomaselli GF, Marban E, Balsler JR. Phenotypic characterization of a novel long-QT syndrome mutation (R1623Q) in the cardiac sodium channel. *Circulation* 1998;97:640-4.
311. Tsurugi T, Nagatomo T, Abe H, et al. Differential modulation of late sodium current by protein kinase A in R1623Q mutant of LQT3. *Life Sci* 2009;84:380-7.
312. Kambouris NG, Nuss HB, Johns DC, Marban E, Tomaselli GF, Balsler JR. A revised view of cardiac sodium channel "blockade" in the long-QT syndrome. *J Clin Invest* 2000;105:1133-40.
313. Miura M, Yamagishi H, Morikawa Y, Matsuoka R. Congenital long QT syndrome and 2:1 atrioventricular block with a mutation of the SCN5A gene. *Pediatr Cardiol* 2003;24:70-2.
314. Wagnon JL, Barker BS, Hounshell JA, et al. Pathogenic mechanism of recurrent mutations of SCN8A in epileptic encephalopathy. *Ann Clin Transl Neurol* 2016;3:114-23.
315. Ohba C, Kato M, Takahashi S, et al. Early onset epileptic encephalopathy caused by de novo SCN8A mutations. *Epilepsia* 2014;55:994-1000.
316. Kong W, Zhang Y, Gao Y, et al. SCN8A mutations in Chinese children with early onset epilepsy and intellectual disability. *Epilepsia* 2015;56:431-8.
317. Larsen J, Carvill GL, Gardella E, et al. The phenotypic spectrum of SCN8A encephalopathy. *Neurology* 2015;84:480-9.
318. Poulin H, Gosselin-Badaroudine P, Vicart S, et al. Substitutions of the S4DIV R2 residue (R1451) in Na(V)1.4 lead to complex forms of paramyotonia congenita and periodic paralyses. *Sci Rep* 2018;8:2041.
319. Zeng Z, Zhou J, Hou Y, et al. Electrophysiological characteristics of a SCN5A voltage sensors mutation R1629Q associated with Brugada syndrome. *PLoS One* 2013;8:e78382.
320. Habbout K, Poulin H, Rivier F, et al. A recessive Nav1.4 mutation underlies congenital myasthenic syndrome with periodic paralysis. *Neurology* 2016;86:161-9.
321. Bednarz M, Stunnenberg BC, Kusters B, et al. A novel Ile1455Thr variant in the skeletal muscle sodium channel alpha-subunit in a patient with a severe adult-onset proximal myopathy with electrical myotonia and a patient with mild paramyotonia phenotype. *Neuromuscul Disord* 2017;27:175-82.
322. Wang DW, Crotti L, Shimizu W, et al. Malignant perinatal variant of long-QT syndrome caused by a profoundly dysfunctional cardiac sodium channel. *Circ Arrhythm Electrophysiol* 2008;1:370-8.
323. Choi JS, Boralevi F, Brissaud O, et al. Paroxysmal extreme pain disorder: a molecular lesion of peripheral neurons. *Nat Rev Neurol* 2011;7:51-5.
324. Nakajima T, Kaneko Y, Saito A, Ota M, Iijima T, Kurabayashi M. Enhanced fast-inactivated state stability of cardiac sodium channels by a novel voltage sensor SCN5A mutation, R1632C, as a cause of atypical Brugada syndrome. *Heart Rhythm* 2015;12:2296-304.
325. García-Molina E, Sabater-Molina M, Muñoz C, Ruiz-Espejo F, Gimeno JR. An R1632C variant in the SCN5A gene causing Brugada syndrome. *Mol Med Rep* 2016;13:4677-80.

326. Arnold WD, Feldman DH, Ramirez S, et al. Defective fast inactivation recovery of Nav 1.4 in congenital myasthenic syndrome. *Ann Neurol* 2015;77:840-50.
327. Benson DW, Wang DW, Dymment M, et al. Congenital sick sinus syndrome caused by recessive mutations in the cardiac sodium channel gene (SCN5A). *J Clin Invest* 2003;112:1019-28.
328. Robyns T, Nuyens D, Van Casteren L, et al. Reduced Penetrance and Variable Expression of SCN5A Mutations and the Importance of Co-inherited Genetic Variants: Case Report and Review of the Literature. *Indian Pacing Electrophysiol J* 2014;14:133-49.
329. Vanoye CG, Lossin C, Rhodes TH, George AL, Jr. Single-channel properties of human Nav1.1 and mechanism of channel dysfunction in SCN1A-associated epilepsy. *J Gen Physiol* 2006;127:1-14.
330. Kahlig KM, Lepist I, Leung K, Rajamani S, George AL. Ranolazine selectively blocks persistent current evoked by epilepsy-associated Nav1.1 mutations. *Br J Pharmacol* 2010;161:1414-26.
331. Elia N, Palmio J, Castañeda MS, et al. Myasthenic congenital myopathy from recessive mutations at a single residue in Na(V)1.4. *Neurology* 2019;92:e1405-e15.
332. Suter MR, Bhuiyan ZA, Laedermann CJ, et al. p.L1612P, a novel voltage-gated sodium channel Nav1.7 mutation inducing a cold sensitive paroxysmal extreme pain disorder. *Anesthesiology* 2015;122:414-23.
333. Cestèle S, Schiavon E, Rusconi R, Franceschetti S, Mantegazza M. Nonfunctional Nav1.1 familial hemiplegic migraine mutant transformed into gain of function by partial rescue of folding defects. *Proc Natl Acad Sci USA* 2013;110:17546-51.
334. Liu SZ, P. Altered PKA modulation in the Na(v)1.1 epilepsy variant I1656M. *J Neurophysiol* 2013;110:2090-8.
335. Nieto-Marin P, Jimenez-Jaimez J, Tinaquero D, et al. Digenic Heterozygosity in SCN5A and CACNA1C Explains the Variable Expressivity of the Long QT Phenotype in a Spanish Family. *Rev Esp Cardiol (Engl Ed)* 2019;72:324-32.
336. Fleischhauer R, Mitrovic N, Deymeer F, Lehmann-Horn F, Lerche H. Effects of temperature and mexiletine on the F1473S Na⁺ channel mutation causing paramyotonia congenita. *Pflugers Arch* 1998;436:757-65.
337. Depienne C, Trouillard O, Gourfinkel-An I, et al. Mechanisms for variable expressivity of inherited SCN1A mutations causing Dravet syndrome. *J Med Genet* 2010;47:404-10.
338. Dib-Hajj SD, Estacion M, Jarecki BW, et al. Paroxysmal extreme pain disorder M1627K mutation in human Nav1.7 renders DRG neurons hyperexcitable. *Mol Pain* 2008;4:37.
339. Kim HJ, Kim BG, Park JE, et al. Characterization of a novel LQT3 variant with a selective efficacy of mexiletine treatment. *Sci Rep* 2019;9:12997.
340. Estacion M, Dib-Hajj SD, Benke PJ, et al. Nav1.7 gain-of-function mutations as a continuum: A1632E displays physiological changes associated with erythromelalgia and paroxysmal extreme pain disorder mutations and produces symptoms of both disorders. *J Neurosci* 2008;28:11079-88.
341. Rühlmann AH, Körner J, Hausmann R, et al. Uncoupling sodium channel dimers restores the phenotype of a pain-linked Na(v) 1.7 channel mutation. *Br J Pharmacol* 2020;177:4481-96.
342. Yang Y, Huang J, Mis MA, et al. Nav1.7-A1632G Mutation from a Family with Inherited Erythromelalgia: Enhanced Firing of Dorsal Root Ganglia Neurons Evoked by Thermal Stimuli. *J Neurosci* 2016;36:7511-22.
343. Eberhardt M, Nakajima J, Klinger AB, et al. Inherited pain: sodium channel Nav1.7 A1632T mutation causes erythromelalgia due to a shift of fast inactivation. *J Biol Chem* 2014;289:1971-80.

344. Bertelli S, Barbieri R, Pusch M, Gavazzo P. Gain of function of sporadic/familial hemiplegic migraine-causing SCN1A mutations: Use of an optimized cDNA. *Cephalalgia* 2019;39:477-88.
345. Dhifallah S, Lancaster E, Merrill S, Leroudier N, Mantegazza M, Cestele S. Gain of Function for the SCN1A/hNav1.1-L1670W Mutation Responsible for Familial Hemiplegic Migraine. *Front Mol Neurosci* 2018;11:232.
346. Nakajima T, Dharmawan T, Kawabata-Iwakawa R, et al. Biophysical defects of an SCN5A V1667I mutation associated with epinephrine-induced marked QT prolongation. *J Cardiovasc Electrophysiol* 2020.
347. Sugiura Y, Ogiwara I, Hoshi A, Yamakawa K, Ugawa Y. Different degrees of loss of function between GEFS+ and SMEI Nav 1.1 missense mutants at the same residue induced by rescuable folding defects. *Epilepsia* 2012;53:e111-4.
348. Sugawara T, Mazaki-Miyazaki E, Ito M, et al. Nav1.1 mutations cause febrile seizures associated with afebrile partial seizures. *Neurology* 2001;57:703-5.
349. Kaluza L, Meents JE, Hampl M, et al. Loss-of-function of Nav1.8/D1639N linked to human pain can be rescued by lidocaine. *Pflugers Arch* 2018;470:1787-801.
350. Dabby R, Sadeh M, Broitman Y, Yosovich K, Dickman R, Leshinsky-Silver E. Painful small fiber neuropathy with gastroparesis: A new phenotype with a novel mutation in the SCN10A gene. *J Clin Neurosci* 2016;26:84-8.
351. Zeng Z, Xie Q, Huang Y, Zhao Y, Li W, Huang Z. p.D1690N sodium voltage-gated channel alpha subunit 5 mutation reduced sodium current density and is associated with Brugada syndrome. *Mol Med Rep* 2016;13:5216-22.
352. Nunez L, Barana A, Amoros I, et al. p.D1690N Nav1.5 rescues p.G1748D mutation gating defects in a compound heterozygous Brugada syndrome patient. *Heart Rhythm* 2013;10:264-72.
353. Lakshmanadoss U, Mertens A, Gallagher M, Kutinsky I, Williamson B. Sudden cardiac arrest due to a single sodium channel mutation producing a mixed phenotype of Brugada and Long QT3 syndromes. *Indian Pacing Electrophysiol J* 2016;16:66-9.
354. Chen YY, Liu SR, Xie LZ, et al. [Functional analysis of a novel SCN5A mutation G1712C identified in Brugada syndrome]. *Nan Fang Yi Ke Da Xue Xue Bao* 2016;37:256-60.
355. Han C, Vasylyev D, Macala LJ, et al. The G1662S Nav1.8 mutation in small fibre neuropathy: impaired inactivation underlying DRG neuron hyperexcitability. *J Neurol Neurosurg Psychiatry* 2014;85:499-505.
356. Amin AS, Verkerk AO, Bhuiyan ZA, Wilde AA, Tan HL. Novel Brugada syndrome-causing mutation in ion-conducting pore of cardiac Na⁺ channel does not affect ion selectivity properties. *Acta Physiol Scand* 2005;185:291-301.
357. Baroudi G, Napolitano C, Priori SG, Del Bufalo A, Chahine M. Loss of function associated with novel mutations of the SCN5A gene in patients with Brugada syndrome. *Can J Cardiol* 2004;20:425-30.
358. Valdivia CR, Tester DJ, Rok BA, et al. A trafficking defective, Brugada syndrome-causing SCN5A mutation rescued by drugs. *Cardiovasc Res* 2004;62:53-62.
359. Huang J, Yang Y, Zhao P, et al. Small-fiber neuropathy Nav1.8 mutation shifts activation to hyperpolarized potentials and increases excitability of dorsal root ganglion neurons. *J Neurosci* 2013;33:14087-97.
360. Chastan N, Lebas A, Legoff F, Parain D, Guyant-Marechal L. Clinical and electroencephalographic abnormalities during the full duration of a sporadic hemiplegic migraine attack. *Neurophysiol Clin* 2016;46:307-11.
361. Chang CC, Acharfi S, Wu MH, et al. A novel SCN5A mutation manifests as a malignant form of long QT syndrome with perinatal onset of tachycardia/bradycardia. *Cardiovasc Res* 2004;64:268-78.

362. Valdivia CR, Ackerman MJ, Tester DJ, et al. A novel SCN5A arrhythmia mutation, M1766L, with expression defect rescued by mexiletine. *Cardiovasc Res* 2002;55:279-89.
363. Huang H, Priori SG, Napolitano C, O'Leary ME, Chahine M. Y1767C, a novel SCN5A mutation, induces a persistent Na⁺ current and potentiates ranolazine inhibition of Nav1.5 channels. *Am J Physiol Heart Circ Physiol* 2011;300:H288-99.
364. Rivolta I, Clancy CE, Tateyama M, Liu H, Priori SG, Kass RS. A novel SCN5A mutation associated with long QT-3: altered inactivation kinetics and channel dysfunction. *Physiol Genomics* 2002;10:191-7.
365. Clancy CE, Tateyama M, Liu H, Wehrens XH, Kass RS. Non-equilibrium gating in cardiac Na⁺ channels: an original mechanism of arrhythmia. *Circulation* 2003;107:2233-7.
366. Veeramah KR, O'Brien JE, Meisler MH, et al. De novo pathogenic SCN8A mutation identified by whole-genome sequencing of a family quartet affected by infantile epileptic encephalopathy and SUDEP. *Am J Hum Genet* 2012;90:502-10.
367. Baker EM, Thompson CH, Hawkins NA, et al. The novel sodium channel modulator GS-458967 (GS967) is an effective treatment in a mouse model of SCN8A encephalopathy. *Epilepsia* 2018;59:1166-76.
368. Neubauer J, Wang Z, Rougier JS, et al. Functional characterization of a novel SCN5A variant associated with long QT syndrome and sudden cardiac death. *Int J Legal Med* 2019;133:1733-42.
369. Abdelsayed M, Baruteau AE, Gibbs K, et al. Differential calcium sensitivity in Nav 1.5 mixed syndrome mutants. *J Physiol* 2017;595:6165-86.
370. Abdelsayed M, Ruprai M, Ruben PC. The efficacy of Ranolazine on E1784K is altered by temperature and calcium. *Sci Rep* 2018;8:3643.
371. Takahashi K, Shimizu W, Miyake A, Nabeshima T, Nakayashiro M, Ganaha H. High prevalence of the SCN5A E1784K mutation in school children with long QT syndrome living on the Okinawa islands. *Circulation Journal* 2014:CJ-13-1516.
372. Hu RM, Tan BH, Tester DJ, et al. Arrhythmogenic Biophysical Phenotype for SCN5A Mutation S1787N Depends upon Splice Variant Background and Intracellular Acidosis. *PLoS One* 2015;10:e0124921.
373. Splawski I, Shen J, Timothy KW, et al. Spectrum of mutations in long-QT syndrome genes. KVLQT1, HERG, SCN5A, KCNE1, and KCNE2. *Circulation* 2000;102:1178-85.
374. An RH, Wang XL, Kerem B, et al. Novel LQT-3 mutation affects Na⁺ channel activity through interactions between alpha- and beta1-subunits. *Circ Res* 1998;83:141-6.
375. Wehrens XH, Abriel H, Cabo C, Benhorin J, Kass RS. Arrhythmogenic mechanism of an LQT-3 mutation of the human heart Na(+) channel alpha-subunit: A computational analysis. *Circulation* 2000;102:584-90.
376. Baroudi G, Chahine M. Biophysical phenotypes of SCN5A mutations causing long QT and Brugada syndromes. *FEBS Lett* 2000;487:224-8.
377. Benhorin J, Goldmit M, MacCluer JW, et al. Identification of a new SCN5A mutation, D1840G, associated with the long QT syndrome. *Mutations in brief no. 153*. Online. *Hum Mutat* 1998;12:72.
378. Blich M, Efrati E, Marai I, Suleiman M, Gepstein L, Boulous M. Novel Clinical Manifestation of the Known SCN5A D1790G Mutation. *Cardiology* 2015;132:228-32.
379. Rivolta I, Abriel H, Tateyama M, et al. Inherited Brugada and long QT-3 syndrome mutations of a single residue of the cardiac sodium channel confer distinct channel and clinical phenotypes. *J Biol Chem* 2001;276:30623-30.

380. Fredj S, Sampson KJ, Liu H, Kass RS. Molecular basis of ranolazine block of LQT-3 mutant sodium channels: evidence for site of action. *Br J Pharmacol* 2006;148:16-24.
381. Benito B, Brugada R, Perich RM, et al. A mutation in the sodium channel is responsible for the association of long QT syndrome and familial atrial fibrillation. *Heart Rhythm* 2008;5:1434-40.
382. Papuc SM, Abela L, Steindl K, et al. The role of recessive inheritance in early-onset epileptic encephalopathies: a combined whole-exome sequencing and copy number study. *Eur J Hum Genet* 2019;27:408-21.
383. Kubota T, Kinoshita M, Sasaki R, et al. New mutation of the Na channel in the severe form of potassium-aggravated myotonia. *Muscle Nerve* 2009;39:666-73.
384. Liu K, Yang T, Viswanathan PC, Roden DM. New mechanism contributing to drug-induced arrhythmia: rescue of a misprocessed LQT3 mutant. *Circulation* 2005;112:3239-46.
385. Makita N, Horie M, Nakamura T, et al. Drug-induced long-QT syndrome associated with a subclinical SCN5A mutation. *Circulation* 2002;106:1269-74.
386. Gando I, Morganstein J, Jana K, McDonald TV, Tang Y, Coetzee WA. Infant sudden death: Mutations responsible for impaired Nav1.5 channel trafficking and function. *Pacing Clin Electrophysiol* 2017;40:703-12.
387. Rusconi R, Scalmani P, Cassulini RR, et al. Modulatory proteins can rescue a trafficking defective epileptogenic Nav1.1 Na⁺ channel mutant. *J Neurosci* 2007;27:11037-46.
388. Petitprez S, Jespersen T, Pruvot E, et al. Analyses of a novel SCN5A mutation (C1850S): conduction vs. repolarization disorder hypotheses in the Brugada syndrome. *Cardiovasc Res* 2008;78:494-504.
389. Spampinato J, Kearney JA, de Haan G, et al. A novel epilepsy mutation in the sodium channel SCN1A identifies a cytoplasmic domain for beta subunit interaction. *J Neurosci* 2004;24:10022-34.
390. Adney SK, Millichap JJ, DeKeyser JM, Abramova T, Thompson CH, George AL, Jr. Functional and pharmacological evaluation of a novel SCN2A variant linked to early-onset epilepsy. *Ann Clin Transl Neurol* 2020;7:1488-501.
391. Howell KB, McMahan JM, Carvill GL, et al. SCN2A encephalopathy: A major cause of epilepsy of infancy with migrating focal seizures. *Neurology* 2015;85:958-66.
392. Trump N, McTague A, Brittain H, et al. Improving diagnosis and broadening the phenotypes in early-onset seizure and severe developmental delay disorders through gene panel analysis. *J Med Genet* 2016;53:310-7.
393. Atkin TA, Maher CM, Gerlach AC, et al. A comprehensive approach to identifying repurposed drugs to treat SCN8A epilepsy. *Epilepsia* 2018;59:802-13.
394. Gardella E, Becker F, Moller RS, et al. Benign infantile seizures and paroxysmal dyskinesia caused by an SCN8A mutation. *Ann Neurol* 2016;79:428-36.
395. Selga E, Campuzano O, Pinsach-Abuin ML, et al. Comprehensive Genetic Characterization of a Spanish Brugada Syndrome Cohort. *PLoS One* 2015;10:e0132888.
396. Zhang L, Tester DJ, Lang D, et al. Does Sudden Unexplained Nocturnal Death Syndrome Remain the Autopsy-Negative Disorder: A Gross, Microscopic, and Molecular Autopsy Investigation in Southern China. *Mayo Clin Proc* 2016;91:1503-14.
397. Rusconi R, Combi R, Cestele S, et al. A rescuable folding defective Nav1.1 (SCN1A) sodium channel mutant causes GEFS+: common mechanism in Nav1.1 related epilepsies? *Hum Mutat* 2009;30:E747-60.

398. Casini S, Albesa M, Wang Z, et al. Functional Consequences of the SCN5A-p.Y1977N Mutation within the PY Ubiquitylation Motif: Discrepancy between HEK293 Cells and Transgenic Mice. *Int J Mol Sci* 2019;20.
399. Bebarova M, O'Hara T, Geelen JL, et al. Subepicardial phase 0 block and discontinuous transmural conduction underlie right precordial ST-segment elevation by a SCN5A loss-of-function mutation. *Am J Physiol Heart Circ Physiol* 2008;295:H48-58.
400. Arnestad M, Crotti L, Rognum TO, et al. Prevalence of long-QT syndrome gene variants in sudden infant death syndrome. *Circulation* 2007;115:361-7.
401. Shinlapawittayatorn K, Du XX, Liu H, Ficker E, Kaufman ES, Deschênes I. A common SCN5A polymorphism modulates the biophysical defects of SCN5A mutations. *Heart Rhythm* 2011;8:455-62.

Supplementary Table 2: Corresponding variants, phenotypes, and function across different sodium channels

Pair	SCN1A Position	Gene/Variant	Function	Phenotype	Reference*	Corresponding Gene/Variant	Function	Phenotype	Reference*
1	I138V	SCN4A ; I141V DI S1	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	Sodium channel myotonia	Petitprez (2008) ²³	SCN9A ; I136V DI S1	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	IEM	Cheng (2008) ²⁴
2	R222W	SCN4A ; R225W DI S4	LoF; WCC: Y, ↓CD, →V _{1/2} Act., no change V _{1/2} FI	Congenital Myopathy	Zaharieva (2016) ¹⁴	SCN5A ; R225W DI S4	LoF; WCC: Y, ↓↓CD, →V _{1/2} Act., →V _{1/2} FI	Severe conduction disease	Bezzina (2003) ⁴⁵
3	S243T	SCN9A ; S241T DI S4-5	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	IEM	Lampert (2006) ⁵⁸	SCN10A ; S242T DI S4-5	GoF; WCC: Y, ←V _{1/2} Act., ←V _{1/2} FI	PPN, PDN; carbamazepin e responder	Han (2018) ⁶⁰
4	Q267K	SCN4A ; Q270K DI S5	GoF/Mixed; WCC: Y, →V _{1/2} Act., →V _{1/2} FI	PMC	Carle (2009) ⁶⁶	SCN5A ; Q270K DI S5	GoF/Mixed; WCC: Y, ↓CD, ↑I _{NaP} , →V _{1/2} Act., →V _{1/2} FI	LQT3/BrS overlap syndrome; ECG: fetal tachycardia/fibrillation TdP, LQT	Calloe (2011) ⁶⁷
5	R377H	SCN2A ; R379H DI S5-6	LoF; WCC: None	ASD	Ben-Shalom (2017) ⁵	SCN5A ; R367H DI S5-6	LoF; WCC: None	BrS; SCD; ECG: ST elevation	Hong (2004) ⁸³
6	N416K	SCN4A ; N440K DI S6	GoF; WCC: Y, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	PMC	Lossin (2012) ⁹²	SCN9A ; N395K DI S6	GoF; WCC: Y, ←V _{1/2} Act., no change V _{1/2} FI	IEM	Sheets (2007) ⁴¹
						SCN5A ; N406K DI S6	GoF/Mixed; WCC: Y, ↓CD, ↑I _{NaP} , no change V _{1/2} Act., no change V _{1/2} FI,	LQT3; ECG: TdP, LQT, polymorphic VT, mexiletine responder	Hu (2018) ⁹³ Kato (2014) ⁹⁴
7	V421M	SCN4A ; V445M DI S6	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., ←V _{1/2} FI	PMC	Wang (1999) ⁹⁷ Huang (2020) ⁹⁸	SCN5A ; V411M DI S6	GoF; WCC: Y, ↑CD, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	LQT3; ECG: neonatal LQT with 2:1 block	Horne (2011) ¹⁰⁰ Zhou (2015) ¹⁰¹
						SCN9A ; V400M DI S6	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., →V _{1/2} FI	IEM; carbamazepin e responder	Fischer (2009) ¹⁰²
8	T782I	SCN2A ; T773I D2 S1	GoF; WCC: Y, no change CD, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	DEE, Sz onset 1 day	Lauxmann (2018) ³³	SCN8A ; T767I D2 S1	GoF; WCC: Y, ↓CD, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	DEE, Sz onset 2 weeks	Pan (2020) ¹¹⁸ Estacion (2014) ¹¹⁹
9	R859H	SCN1A ; R859H D2 S4	LoF/mixed; WCC: Y, ↑I _{NaP} , no change CD, ←V _{1/2} Act., ←V _{1/2} FI	GEFS+	Volkers (2011) ¹³⁴	SCN4A ; R669H D2 S4	LoF/mixed; WCC: Y, ↓CD, no change V _{1/2} Act., ←V _{1/2} FI	HypoPP	Kuzmenkin (2002) ¹³⁵
10	R859C	SCN1A ; R859C D2 S4	LoF; WCC: Y, ↓CD, no change V _{1/2} Act., no change V _{1/2} FI	EPI	Bechi (2015) ¹²⁴	SCN5A ; R808C D2 S4	LoF; WCC: Y, ↓CD, no change V _{1/2} Act., ←V _{1/2} FI	BrS	Glazer (2020) ¹

11	R862H	SCN4A ; R672H D2 S4	LoF; WCC: Y, ↓CD; →V _{1/2} Act., ←V _{1/2} FI	Hypo-PP	Jurkatt-Rott (2000) ¹⁴⁰ Kuzmenkin (2002) ¹³⁵	SCN5A ; R811H D2 S4	LoF; WCC: Y, ↓CD; no change V _{1/2} Act., ←V _{1/2} FI	BrS; family history of sudden death,	Calloe (2013) ¹¹⁴
12	R865Q	SCN4A ; R675Q D2 S4	Mixed; WCC: Y, no change CD, no change V _{1/2} Act., no change V _{1/2} FI, ←V _{1/2} SI	Potassium sensitive normoPP	Wu (2014) ¹⁴⁹	SCN5A ; R814Q D2 S4	Mixed; WCC: Y, no change CD, no change V _{1/2} Act., no change V _{1/2} FI	LQT3; BrS	Glazer (2020) ¹
13	I883T	SCN3A ; I875T D2 S4-5	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., →V _{1/2} FI	EPI/PMG	Zaman (2018) ¹⁵⁵	SCN9A ; I848T D2 S4-5	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	IEM	Cummins (2004) ¹⁵⁷ Namer (2015) ¹⁵⁸ Theile (2011) ¹⁵⁹
14	R946C	SCN1A ; R946C; D2 S5-6	LoF; WCC: None	DS	Volkers (2011) ¹³⁴	SCN2A ; R937C; D2 S5-6	LoF; WCC: None	ASD	Begemann (2019) ¹⁸⁰
15	R946H	SCN1A ; R946H; D2 S5-6	LoF; WCC: None	DS	Liao (2010) ¹⁸² Volkers (2011) ¹³⁴	SCN2A ; R937H; D2 S5-6	LoF; WCC: None	ASD	Ben-Shalom (2017) ⁵
16	G979R	SCN1A ; G979R; D2 S6	LoF; WCC: None	DS	Sugawara (2003) ¹⁸⁴ Rhodes (2005) ¹³⁰	SCN8A ; G964R; D2 S6	LoF; WCC: None	NDD without EPI	Wagnon (2017) ¹⁸⁵
17	D1256N	SCN4A ; D1069N D3 S2	LoF/Mixed; WCC: Y, no change CD, →V _{1/2} Act., →V _{1/2} FI	Congenital myopathy	Zaharieva (2016) ¹⁴	SCN5A ; D1243N D3 S2	LoF/Mixed; WCC: Y, no change CD, →V _{1/2} Act., →V _{1/2} FI	BrS	Glazer (2020) ¹
18	A1343T	SCN4A ; A1156T D3 S4-5	GoF; WCC: Y, no change V _{1/2} Act., →V _{1/2} FI	PMC with prominent myalgia	Palmio (2017) ²³⁶	SCN5A ; A1330T D3 S4-5	GoF; WCC: Y, no change V _{1/2} Act., →V _{1/2} FI	LQT3; SCD	Smits (2005) ²³⁸
19	PI345L	SCN3A ; PI333L D3 S4-5	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	EPI	Zaman (2018) ¹⁵⁵	SCN4A ; PI158L D3 S4-5	GoF; WCC: Y, no change V _{1/2} Act., →V _{1/2} FI	Sodium channel myotonia	Desaphy (2016) ²³⁹
						SCN5A ; PI332L D3 S4-5	GoF/mixed; WCC: Y, ←V _{1/2} Act., ←V _{1/2} FI	LQT3, ECG: TdP, LQT, mexiletine responder	Ruan (2007) ¹⁹⁰
						SCN9A ; PI308L D3 S4-5	GoF; WCC: Y, ↓CD, ←V _{1/2} Act., no change V _{1/2} FI	IEM	Cheng (2010) ²²⁶
20	SI346Y	SCN2A ; SI336Y D3 S4-5	GoF/Mixed (Na _v 1.2N); WCC: Y, ↓CD, ←V _{1/2} Act., →V _{1/2} FI	DEE	Thompson (2020) ⁵⁴	SCN5A ; SI333Y D3 S4-5	GoF; WCC: Y, no change CD, ←V _{1/2} Act., →V _{1/2} FI	SIDS; LQT3	Huang (2009) ²⁴³
21	VI353A	SCN9A ; VI316A D3 S4-5	GoF; WCC: Y, ←V _{1/2} Act., →V _{1/2} FI	IEM	Wu (2013) ²⁵² , Estacion (2013) ²⁵³	SCN11A ; VI184A D3 S5	GoF; WCC: Y, ↑CD, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	PPN, CAP	Leipold (2015) ²⁵⁵
22	FI486C	SCN4A ; FI298C D3-D4 linker	GoF/Mixed; WCC: Y, →V _{1/2} Act., →V _{1/2} FI	Sodium channel myotonia	Farinato (2019) ²⁷²	SCN5A ; FI473C D3-D4 linker	GoF; WCC: Y, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	LQT3; ECG: TdP, LQT with 2:1 block, mexiletine responder	Bankston (2007) ²⁷⁶

23	FI499L	SCN1A ; FI499L D3-D4 linker	GoF; WCC: Y, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	FHM	Barbieri (2019) ²⁸³	SCN5A ; FI486L D3-D4 linker	GoF; WCC: Y, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	LQT3	Wang (2007) ³⁶
24	TI501I	SCN3A ; TI486I D3-D4 linker	GoF; WCC: Y, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	EPI/PMG	Zaman (2020) ¹⁴⁷	SCN9A ; TI464I D3-D4 linker	GoF; WCC: Y, ↑I _{NaP} , →V _{1/2} Act., →V _{1/2} FI	PEPD, responsive to SCB	Fertleman (2006) ²²⁷ Theile (2011) ¹⁵⁹
25	EI587K	SCN1A ; EI587K D4 S2	LoF; WCC: None	EPI	Kluckova (2020) ¹⁰	SCN5A ; EI574K D4 S2	LoF; WCC: Y, ↓↓CD, →V _{1/2} Act.	BrS	Glazer (2020) ¹
26	RI596C	SCN1A ; RI596C D4 S2-3	LoF; WCC: None	EPI	Kluckova (2020) ¹⁰	SCN5A ; RI583C D4 S2-3	LoF; WCC: Y, ↓CD, no change I _{NaP} , no change V _{1/2} Act., no change V _{1/2} FI	BrS	Glazer (2020) ¹
27	PI632S	SCN1A ; PI632S; D4 S3-4	LoF; WCC: Y, ←V _{1/2} Act., ←V _{1/2} FI	DS	Rhodes (2005) ¹³⁰	SCN2A ; PI622S; D4 S3-4	LoF; WCC: Y, ←V _{1/2} FI	ASD and Sz onset 21 months	Wolff (2017) ⁹⁶
28	RI636Q	SCN3A ; RI621Q D4 S4	GoF; WCC: Y, no change CD, ↑I _{NaP} , ←V _{1/2} Act., no change V _{1/2} FI	EPI/PMG	Zaman (2020) ¹⁴⁷	SCN8A ; RI617Q D4 S4	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act., →V _{1/2} FI	DEE, Sz onset 3 months	Wagnon (2015) ³¹⁴
						SCN5A ; RI623Q D4 S4	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2} Act.	LQT3; ECG: TdP, LQT, mexiletine responder	Kambouris (1998) ³¹⁰ Tsurugi (2009) ³¹¹
29	RI639L	SCN4A ; RI451L D4 S4	LoF; WCC: Y, ↓CD, no change V _{1/2} Act., ←V _{1/2} FI	Complex phenotype including myotonia and paralysis (both potassium sensitive and hypopp)	Poulin (2018) ³¹⁸	SCN8A ; RI620L D4 S4	LoF; WCC: Y, ↓↓CD, no change V _{1/2} Act., ←V _{1/2} FI	ASD	Liu (2019) ²⁸⁸
30	RI645H	SCN4A ; RI457H D4 S4	LoF; WCC: Y, no change V _{1/2} Act., ←V _{1/2} FI	CMS (in patient homozygous for RI457H variant)	Arnold (2015) ³²⁶	SCN5A ; RI632H D4 S4	LoF; WCC: Y, no change CD, no change V _{1/2} Act., ←V _{1/2} FI	SSS; ECG: bradycardia, absent atrial depolarizatio ns, prolonged QRS, 1° heart block	Benson (2003) ³²⁷
31	RI657C	SCN1A ; RI657C; D4 S4-5	LoF; WCC: Y, ↓CD, →V _{1/2} Act., no change V _{1/2} FI	GEFS+	Lossin (2003) ¹⁸⁶	SCN5A ; RI644C D4 S4-5	LoF; WCC: Y, →V _{1/2} Act., no change V _{1/2} FI	BrS; ECG: ST elevation, echo: CM changes	Frustaci (2005) ⁸⁷
						SCN8A ; RI638C; D4 S4-5	LoF; WCC: Y, →V _{1/2} Act., no change V _{1/2} FI	NDD without epilepsy	Wengert (2019) ⁶⁵
32	FI661S	SCN1A ; FI661S D4 S4-5	Mixed; 50% reduction in trafficking WCC: Y, ↓CD, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	DS	Rhodes (2004) ¹⁷² Thompson (2012) ¹⁸⁷	SCN4A ; FI473S D4 S4-5	GoF; WCC: Y, CD not reported, ↑I _{NaP} , no change V _{1/2} Act., →V _{1/2} FI	PMC	Fleischhauer (1998) ³³⁶
33	MI664K	SCN1A ; MI664K D4 S4-5	LoF; 90% reduction in peak current and trafficking not allowing for detailed SCN1A biophysics	GEFS+/DS	Bechi (2015) ¹²⁴	SCN9A ; MI627K D4 S4-5	GoF; WCC: Y, no change CD, no change V _{1/2} Act., →V _{1/2} FI	PEPD	Fertleman (2006) ²²⁷ Dib-Hajj (2008) ³³⁸ Theile (2011) ¹⁵⁹

34	G1674R	SCN1A ; G1674R D4 S5	LoF; WCC: None ↓↓CD	EPI	Rhodes (2004) ¹⁷² Thompson (2012) ¹⁸⁷	SCN5A ; G1661R D4 S5	LoF; WCC: Y (barely) ↓↓CD	BrS	Glazer (2020) ¹
35	M1780I	SCN3A ; M1765I D4 S6	GoF; WCC: Y, ↑I _{NaP} , ←V _{1/2 Act.} , no change V _{1/2 FI}	EPI/PMG	Zaman (2020) ¹⁴⁷	SCN8A ; M1760I D4 S6	GoF; WCC: Y, ←V _{1/2 Act.} , no change V _{1/2 FI}	EPI	Liu (2019) ²⁸⁸
36	N1788D	SCN5A ; N1774D C-Term	GoF; WCC: Y, ↑CD, ↑I _{NaP} , ←V _{1/2 Act.} , no change V _{1/2 FI}	LQT3; ECG: TdP, LQT with 2:1 block, mexiletine responder	Kato (2014) ⁹⁴	SCN8A ; N1768D C-Term	GoF; WCC: Y, ↑I _{NaP} , no change V _{1/2 Act.} , →V _{1/2 FI}	DEE, Sz onset 6 months	Veeramah (2012) ³⁶⁶ Patel (2016) ²⁴⁷ Baker (2018) ³⁶⁷
37	R1892Q	SCN2A ; R1882Q; C-Term	GoF; WCC: Y, ↑CD, ↑I _{NaP} , ←V _{1/2 Act.} , →V _{1/2 FI}	DEE, Sz onset 1 day	Berecki (2018) ¹⁴¹ Mason (2019) ¹⁴² Wolff (2017) ⁹⁶	SCN8A ; R1872Q; C-Term	GoF; WCC: Y, ↑CD, ←V _{1/2 Act.} , →V _{1/2 FI}	DEE, Sz onset 4 months	Wagnon (2015) ³¹⁴ Aktin (2018) ³⁹³
38	Q1923R	SCN1A ; Q1923R C-Term	LoF; WCC: None	DS	Nissenkorn (2019) ²⁹	SCN5A ; Q1909R C-Term	Mixed; ←V _{1/2 Act.} , no change V _{1/2 FI} decrease in peak current by 50%.	SIDS (not a known cardiac patient)	Winkel (2015) ¹⁰³ Abdelsayed (2017) ³⁶⁹

Corresponding variant = identical variant among different SCN at the same position/location in the SCN protein (the corresponding sequence numbers are not identical as the amino acid sequence between SCN variants differs slightly).

Rows marked in grey denote variant pairs with divergent functional properties.

*References relate to those detailed in Supplementary Table 1.

Phenotypical features: ASD = autism spectrum disorder, BrS = Brugada syndrome, CAP = cold aggravated pain, CM changes = cardiomyopathic changes, CMS = congenital myasthenic syndrome, DEE = developmental and epileptic encephalopathy, DS = Dravet syndrome, ECG = electrocardiogram, echo = echocardiogram, EPI = epilepsy, FHM3 = familial hemiplegic migraine type 3, GEFS+ = genetic epilepsy with febrile seizures plus, Hyper-PP = hyperkalaemic periodic paralysis, Hypo-PP = hypokalaemic periodic paralysis, IEM = inherited erythromelalgia, LQT3 = long QT3 syndrome, Na_v1.2N = neonatal proteoform, NDD = neurodevelopmental disorder, PAM = potassium-aggravated myotonia, PDN = painful diabetic neuropathy, PEPD = paroxysmal extreme pain disorder, PMC = paramyotonia congenita, PMG = polymicrogyria, PPN = painful peripheral neuropathy, SCB = sodium channel blocker, SCD = sudden cardiac death, SIDS = sudden infant death syndrome, SSS = sick sinus syndrome, SUD = sudden unexplained death, Sz = seizure, TdP = torsade de pointes, VT = ventricular tachycardia

Electrophysiological key features: Arrows (→) are used for electrophysiological parameters. The direction of the arrows indicates hyperpolarizing (←) or depolarizing shifts (→), as well as an increase (↑) or decrease (↓) of parameters, (↓↓ = >50% decrease)

Electrophysiological abbreviations: GoF: gain-of-function, LoF: loss-of-function, WCC: whole cell current (Y = measurable, N = not measurable), Act: activation, CD: current density, FI: fast inactivation, SI: slow inactivation, I_{NaP}: persistent sodium current, V_{1/2 Act.}: half-activation of steady-state activation curve, V_{1/2 FI}: half-inactivation of steady-state fast inactivation curve.

Supplementary Table 3: Detailed *SCN1-11A* Analysis

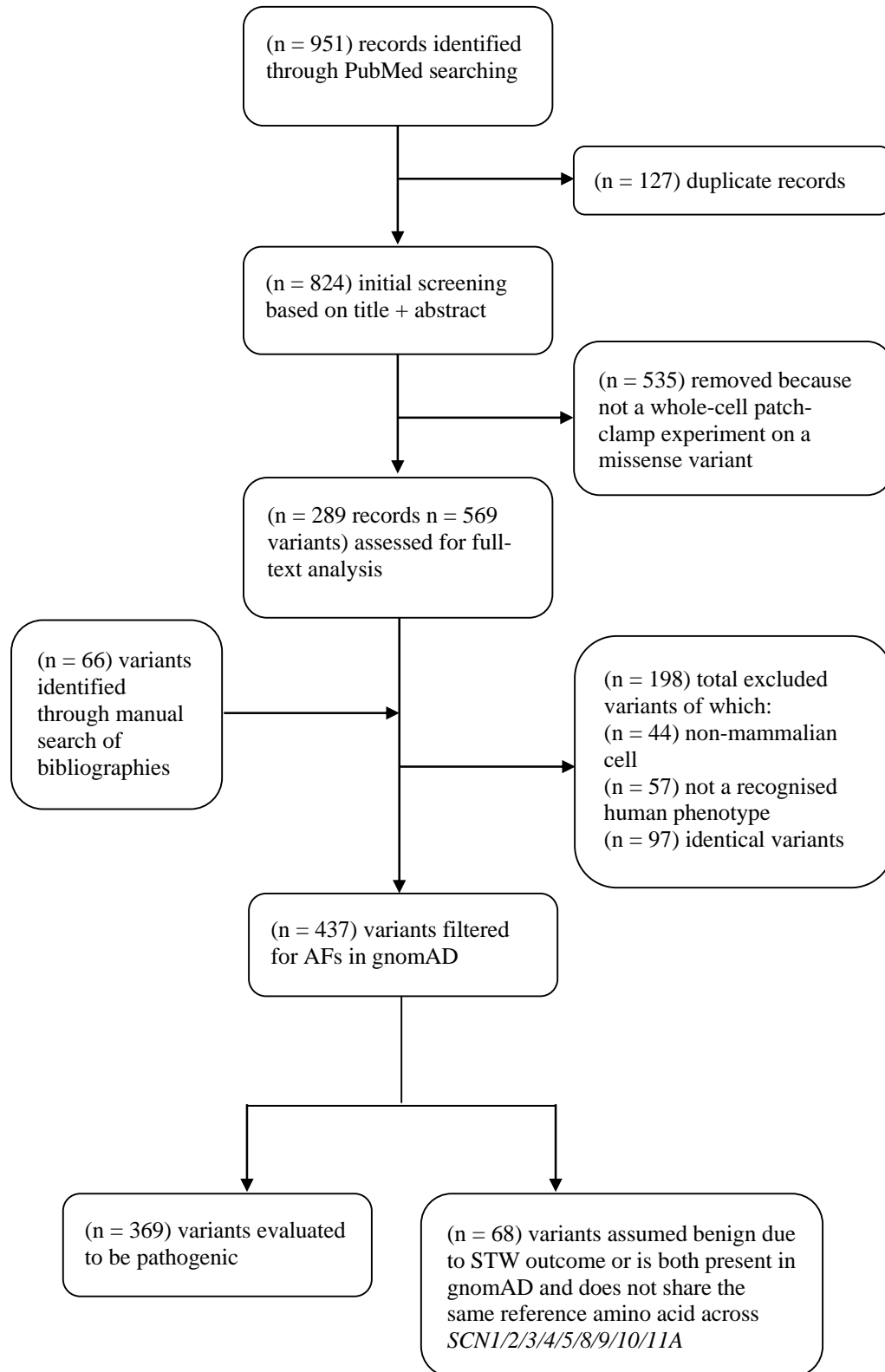
SCN1A					
Disorder	No.	Functional Effects	Distribution	Clinical Context	
EPI	60 (85%)	<ul style="list-style-type: none"> GoF – (3/60, 5%) LoF – (47/60, 78%) Mixed – (10/60, 17%) 	<i>SCN1A</i> variants clustered across pore-loop regions (S5, S5-6 & S6) and 88% of variants showed LoF (23/26)	Variants displaying LoF were more frequently associated with DS, whereas variants showing GoF were FHM3-associated (p<0.001)	
FHM3	11 (15%)	<ul style="list-style-type: none"> GoF – (8/11, 73%) LoF – (1/11, 9%) Mixed – (2/11, 18%) 	55% (6/11) of FHM3-associated variants occurred in sites implicated in channel inactivation, including the DIII-IV linker, DIVS4-5 and DIVS6, predominantly showing GoF effects. Of these, 83% were GoF (5/6). While most FHM3-associated variants are associated with GoF effects, the variant located in DIVS4 was clearly mixed. The other mixed variant was located in DIVS5		
SCN2A					
Disorder	No.	Functional Effect	Distribution	Clinical Context	
EPI	29 (81%)	<ul style="list-style-type: none"> GoF – (17/29, 59%) LoF – (7/29, 24%) Mixed – (5/29, 17%) 	<i>SCN2A</i> epilepsy-associated variants were evenly distributed across homologous domains but very few were found in pore-forming regions	Variants displaying LoF effects were frequently associated with ASD, while GoF variants were epilepsy-associated (p = 0.001)	
ASD	7 (19%)	All variants showed LoF	71% of ASD-associated variants clustered in pore-loop regions (5/7), all displaying LoF		
SCN3A					
Disorder	No.	Functional Effect	Distribution	Clinical Context	
EPI	5 (38%)	<ul style="list-style-type: none"> GoF – 3/5 (60%) LoF – 1/5 (20%) Mixed – 1/5 (20%) 	Variants associated with an epilepsy or mixed phenotype were evenly distributed across the protein and only two were observed in pore-forming regions	Epilepsy and PMG variants appear to be mainly GoF	
EPI/PMG	6 (46%)	All variants showed GoF			
ASD	1 (8%)	LoF			DIVS5
Fetal Akinesia	1 (8%)	GoF			DIIS4

SCN4A				
Disorder	No.	Functional Effect	Distribution	Clinical Context
PMC	14 (36%)	<ul style="list-style-type: none"> GoF – (11/14, 86%) LoF – (1/14, 7%) Mixed – (2/14, 7%) 	43% occurred in the DIII-IV linker (6/14) and the remainder across the protein	<p>GoF variants were frequently PAM/PMC-associated, whereas LoF variants were CMS-associated (p<0.001). Overall, 32% of variants occurred in inactivation sites (12/38), 92% of which showed GoF (11/12). 29% of variants were found in S4 sites (11/38), 70% of which showed LoF (8/11). 18% occurred in pore-forming regions (7/38) and the remainder across the protein.</p>
CMS	10 (26%)	<ul style="list-style-type: none"> LoF – (8/10, 80%) Mixed – (2/10, 20%) 	Two variants occurred in pore-loops and two in voltage-sensing regions.	
PAM	4 (11%)	All variants showed GoF	Three variants were found in inactivation sites and one in the C-terminus	
HypoPP	5 (13%)	Four variants displayed LoF and one variant showed mixed function	All four variants displaying LoF were found in S4 sites while the LoF variant was found in DIIIS4-5	
HyperPP	1 (3%)	Mixed	DIVS5	
NormoPP	1 (3%)	Mixed	DIIS4	
Mixed Phenotype	3 (8%)	All variants showed GoF	Variants occurred in DIS6, DIIS5 and DIIIS4-5	
SCN5A				
Disorder	No.	Functional Effect	Distribution	Clinical Context
BrS	100 (69%)	96% of variants showed LoF (96/100), three were mixed effect and one displayed GoF	52% of BrS-associated variants occurred in pore-loop regions (S5, S5-6 & S6) (52/100), while only 6% were found at sites of channel inactivation (6/100)	<p>Variants displaying LoF effects were more frequently BrS-associated, while GoF variants were LQT3-associated, (p<0.001)</p>
LQT3	38 (26%)	<ul style="list-style-type: none"> GoF – (30/38, 79%) LoF – (2/38, 5%) Mixed – (6/38, 16%) 	47% of LQT3 variants clustered in sites of inactivation (18/38), showing predominantly GoF (94%, 17/18)	
Mixed	8 (5%)	<ul style="list-style-type: none"> LoF – 4/8 (50%) Mixed – 4/8 (50%) 	Two variants occurred in the C-terminus, one in DIS4, one in DIIS4 and the rest across the protein	

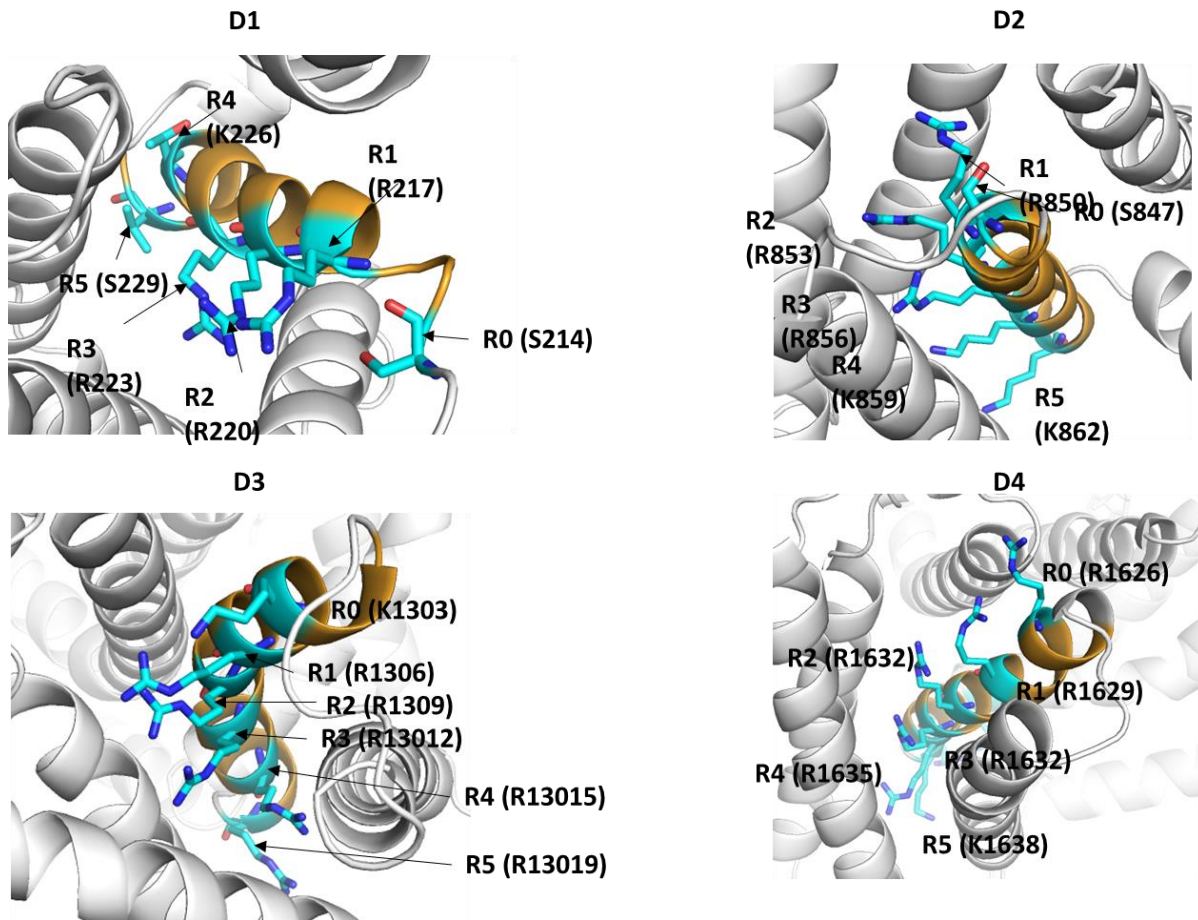
SCN8A				
Disorder	No.	Functional Effect	Distribution	Clinical Context
EPI	14 (67%)	<ul style="list-style-type: none"> GoF – (12/14, 86%) LoF – (2/14, 14%) 	29% occurred in S4 regions (4/14), 14% in the C-terminus (2/14), 14% in inactivation sites (2/14) and the remainder across the protein. None occurred in cytoplasmic regions.	Variants displaying LoF effects were frequently NDD/ASD-associated, while GoF variants were epilepsy-associated, (p=0.003).
NDD	5 (23%)	All variants showed LoF	Three were found in pore-forming regions, one in DIVS4-5 and one in DIIS3	
ASD	2 (9%)	One variant showed GoF while the other LoF	Both variants were located in DIVS4	
SCN9A				
Disorder	No.	Functional Effect	Distribution	Clinical Context
PPN	34	<ul style="list-style-type: none"> GoF – 33/34 (97%) Mixed – 1/34 (3%) 	Of 33 GoF variants, 33% were found in inactivation sites (11/33), 18% in DIIS5 (6/33), 15% in S4 regions (5/33) and the remainder across the protein. The mixed effect variant was located in DIIS4.	The majority of variants appear to be GoF
SCN10A				
Disorder	No.	Functional Effect	Distribution	Clinical Context
AF	1	LoF	The LoF variant was observed in the N-terminus	GoF variants appear to be associated with PPN
SUD	2	Both variants showed LoF	Variants were observed in DIS6 and DIIS4	
PPN	3	All variants showed GoF	Variants were found at DIS4-S5, DIVS5-6 and DIVS6	
SCN11A				
Disorder	No.	Functional Effect	Distribution	Clinical Context
PPN	4	All variants showed GoF	PPN-associated variants were found in DIS4, DIIS4-5, DIIS4 and DIIS5	The majority of variants appear to be GoF.

Phenotypical features: FHM3 = familial hemiplegic migraine type 3, EPI = epilepsy, ASD = autism spectrum disorder, NDD = neurodevelopmental disorder, PMG = polymicrogyria, Hypo-PP = hypokalaemic periodic paralysis, Hyper-PP = hyperkalaemic periodic paralysis, PMC = paramyotonia congenita, CMS = congenital myasthenic syndrome, PAM = potassium-aggravated myotonia, BrS = Brugada syndrome, LQT3 = long QT3 syndrome, SCD = sudden cardiac death, PPN = peripheral painful neuropathy (including, PEPD = paroxysmal extreme pain disorder, IEM = inherited erythromelalgia and SFN = small fibre neuropathy), AF = atrial fibrillation, SUD = sudden unexpected death.

Supplementary Figure 1: Study Selection



Supplementary Figure 2: Voltage sensing regions (S4) structure zoom across D1-D4



Voltage sensing regions (S4) structure zoom across D1-D4. Close-up view of S4 voltage sensing regions across all four domains (D1-D4) illustrating conserved Arginines R0-R4.