**Heterologous effects of infant bacille Calmette-Guérin vaccination: potential mechanisms of immunity**

***Authors’ names and affiliations***

Egle Butkeviciute[[1]](#footnote-1), Christine E Jones[[2]](#footnote-2), [[3]](#footnote-3), Steven G Smith1, [[4]](#footnote-4)\*

***Abstract***

The current anti-tuberculosis vaccine, bacille Calmette-Guérin (BCG), was derived in the 1920s, yet the mechanisms of BCG-induced protective immunity and the variability of protective efficacy among populations are still not fully understood. BCG challenges the concept of vaccine specificity, as there is evidence that BCG may protect immunised infants from pathogens other than *Mycobacterium tuberculosis* – resulting in heterologous or non-specific protection. This review summarises the up-to-date evidence for this phenomenon, potential immunological mechanisms and implications for improved childhood vaccine design. BCG induces functional changes in infant innate and adaptive immune compartments, encouraging their collaboration in the first year of life. Understanding biological mechanisms beyond heterologous BCG effects is crucial to improve infant protection from infectious diseases.

***Keywords***

BCG, childhood immunisation, infant immunity, trained immunity, innate memory, heterologous vaccine effects, monocytes, natural killer cells, T-cells, humoral responses

***Introduction***

The bacille Calmette-Guérin (BCG) is a live attenuated strain of *Mycobacterium bovis* and is the only currently licensed vaccine against tuberculosis (TB). It is routinely administered to infants at or shortly after birth in regions where TB is endemic. BCG vaccination confers consistent efficacy against disseminated forms of TB in childhood, such as TB meningitis and miliary TB, however, its protective efficacy against adult-type pulmonary TB varies [1, 2]. Factors that have been implicated include BCG strain, route of administration, geographical location, exposure to environmental mycobacteria and helminth infection [3]. There is increasing evidence, especially in regions affected by a high infectious disease burden, that apart from protecting against TB, BCG may reduce infant mortality from unrelated infections. Here we review the evidence for this phenomenon, discuss potential mechanisms and outline the possible implications for future vaccine candidates.

***All-cause infant mortality reduction***

Observational studies reported that BCG, alone or in combination with other vaccines, might decrease the all-cause mortality risk up to 30-50% for up to 2 years of age in West Africa [4-6], extending to up to 5 years of age in Uganda [7]. More specifically, it was shown that immunising low-birth-weight infants with BCG at birth could significantly improve their survival for the first month of life because of decreased infection risk [8]. Similar findings were reported in India, where mortality rates were lower in BCG-vaccinated infants for up to 6 months of age, compared to the unvaccinated infant group [9]. Studies in Malawi and Guinea-Bissau also found a trend for reduced mortality among infants vaccinated with BCG [10, 11]. Although the extent of BCG-dependent non-specific reduction in infant mortality is difficult to evaluate, with the efficacy estimates reaching 6-72% in clinical trials or 2-95% in observational studies [12, 13], this evidence suggests that BCG may exert a beneficial, heterologous influence on infant survival, reducing mortality unrelated to TB.

***Impact on acquisition of infectious diseases***

BCG may also reduce the acquisition of non-mycobacterial infections. In Uganda, BCG-vaccinated HIV-positive adults had lower risk of intestinal nematode infection than unvaccinated individuals [14]. BCG vaccination also decreased risk of heterologous infections in infants. In Guinea-Bissau, BCG-immunised infants had lower rates of neonatal sepsis and respiratory infection [8]. Similarly, hospitalisation rate due to non-tubercular respiratory infections and sepsis in Spain was lower among the BCG-vaccinated children [15]. A recent analysis of infant immunisation with BCG in 33 countries suggested BCG vaccination may reduce acute lower respiratory infection incidence by 17-37% [16]. In contrast, a randomised trial in Denmark found no association between neonatal BCG vaccination and infection incidence [17]. The reasons for the discrepancies are not clear, although it has been suggested that benefits of infant BCG immunisation in low-income settings may be partially accounted by lowered undiagnosed mycobacterial infection rate [18]. Such infections would be less likely in a setting with low infectious disease burden (e.g. Denmark). Together, these studies imply that non-specific BCG effects may be particularly beneficial in countries with high infectious disease load, reducing both the all-cause mortality and the disease incidence.

***Factors potentially contributing to the heterologous effects of BCG vaccination***

*BCG timing and interaction with other vaccines*

Some studies suggested that diphtheria-tetanus-pertussis (DTP) vaccine might affect the impact of BCG on childhood mortality [4, 9], implicating that other vaccines may modulate the non-specific effects of BCG immunisation (**Table 1**). In contrast, a study in Burkina Faso found that risk of mortality before two years of age was reduced to a similar extent in infants vaccinated with BCG-only or both BCG and DTP [5]. Vaccination timing and sequence were suggested as potentially important to the non-specific effects of vaccines, as studies in Senegal and Philippines found that immunising infants with DTP at or following BCG administration was associated with enhanced survival [6, 19-21]. Proposals were made that BCG following DTP might reduce all-cause infant mortality even further [22-24]. The WHO Strategic Advisory Group of Experts (SAGE) addressed the controversy of non-specific BCG and DTP interactions in 2014 and concluded that the evidence for such effects was insufficient [13].

*Infant age and time post-BCG vaccination*

The extent to which heterologous effects of BCG vaccination are apparent may depend on age of the infant. BCG-dependent reduction in overall infant mortality may be the most evident in the first few months of life, before the non-specific infant protection becomes influenced by the administration of subsequent vaccines [4, 8]; however, in other studies heterologous effects are still apparent up to 24 months of age [5, 6]. Some studies reported BCG-associated reduction in overall mortality of children aged up to 5 years [7, 25] or decrease in hospitalization rates due to non-mycobacterial infections in children up to 14 years of age [15]. This may be a consequence of improved early childhood survival as BCG-related reduction in hospitalisation due to sepsis was the most significant in children aged 1 to 4 years, diminishing in older children [15]. This implies that the heterologous BCG effects manifest soon after immunisation, but wane over time and may be the most apparent for neonates vaccinated at birth. Another possibility is that as infants grow older, they become exposed to pathogens more frequently, with the resulting development of the classical immunity against infectious diseases eventually overcoming the heterologous beneficial effects of BCG.

*Sex-differential effects of BCG vaccination*

Some studies indicate that BCG-dependent non-specific infant mortality reduction may be influenced by male or female sex, females possibly benefitting from heterologous BCG effects more than males [9, 26, 27]. A review of female-male twin pair datasets from Guinea-Bissau and Senegal found that the survival benefit of BCG vaccination in females was variable, possibly due to low death rates observed among the vaccinated twin pairs [28]. No BCG-associated survival benefit in female infants was observed over the first 8 months of life in an Indian infant cohort [29], although this could be attributable to excess background female neonatal mortality in this region. No sex-related differences in heterologous BCG protection were observed in Burkina Faso [5]. SAGE addressed this issue in 2014; however, no evidence for differences in BCG-immunised female or male heterologous mortality reduction was found [13]. Of interest, some studies indicated that BCG-vaccinated females may produce higher levels of inflammatory cytokines in response to non-mycobacterial stimuli than males at 4 weeks [30] or 1 and 12 weeks [31] post vaccination. Therefore, immunological mechanisms of heterologous effects of BCG vaccination may be sex-dependent; however, their contribution to heterologous infant mortality is not clear.

***Mechanisms implicated in heterologous BCG-vaccinated infant protection***

*BCG-inducible trained innate immunity*

A tempting candidate to explain the heterologous effects of BCG is the phenomenon of innate immune response training [32-34]. Originally identified in natural killer (NK) cells, innate memory or training enables the innate cells to respond more rapidly and strongly to antigens unrelated to the original stimulus and was shown to be BCG-inducible in monocytes. Pre-exposure of murine macrophages to BCG was demonstrated to increase their ability to cope with *Candida albicans* (*C. albicans*) infection both *in vitro* and *in vivo* [35]. In humans, monocytes of BCG-vaccinated adults had increased expression of surface markers of activation (TLR4, CD11b, and CD14) and produced more IL-1β, IL-6, IFNγ and TNFα in response to *Staphylococcus aureus* or *C. albicans* for up to three months post-vaccination compared to monocytes isolated before vaccination from the same adults [36-38] (**Figure 1**). Interestingly, while surface receptor expression on monocytes from BCG-vaccinated adults was upregulated for up to a year, IL-1β and TNFα production upon non-mycobacterial antigen stimulation diminished by this time [39]. This suggests that the most potent effects of heterologous BCG-trained immunity manifest over the first few months post-BCG vaccination. Apart from inducing functional, lasting monocyte changes, BCG enhanced the vaccinated adult NK cell IL-1β and IL-6 production in response to *C. albicans* and *S. aureus* for up to 3 months, also, improving T- and B-cell deficient mice, infected with *C. albicans,* survival [36, 40]. This is consistent with the observations that BCG-associated reduction in all-cause infant mortality is the most significant during the first few months of life [4, 5, 8], implying that BCG-trained monocyte and NK cell immunity may contribute to broad infant protection from infectious diseases when they are the most susceptible.

*Metabolic changes induced by innate immune training*

Functional monocyte changes induced by BCG training have been associated with a metabolic shift from oxidative phosphorylation to aerobic glycolysis (**Figure 1**). First demonstrated in monocyte *in vitro* training with β-glucan (fungal cell wall component), this shift is characterised by increased glucose consumption, lactate production and improved mammalian target of rapamycin (mTOR) phosphorylation [41]. Similar changes were observed in BCG-dependent monocyte training [42]. Peripheral blood mononuclear cells (PBMCs) isolated from BCG-vaccinated adults at 2 weeks and 3 months post-immunisation were shown to secrete more lactate and express higher levels of glycolysis pathway components hexokinase-2 (HK2) and platelet phosphofructokinase (PFKP) upon heterologous stimulation than cells obtained prior to the vaccination [42]. This correlates with the previous findings that monocytes obtained from BCG-immunised donors at these time points produce higher cytokine levels than monocytes isolated before the immunisation [36-38]. Of note, inhibition of mTOR and glycolysis pathways diminished *ex vivo* BCG-trained human monocyte production of lactate, TNF and IL-6 upon LPS challenge, supporting the role for glycolysis in the innate immune training [42]. Importantly, polymorphisms of HK2 and PFKP were associated with the ability of monocytes to be trained and produce cytokines in response to LPS [42]. This implies BCG-inducible training may be ineffective in some individuals with metabolic component polymorphisms. Other pathways may be involved in innate immune training. Monocytes trained *in vitro* with BCG or oxidised low-density lipoprotein (oxLDL) were shown to increase reactive oxygen species production upon stimulation with zymosan, a yeast-derived ligand of TLR2 [43]. BCG enhanced IL-6 and TNFα production in histone 3 lysine 4 trimethylation (H3K4me3) dependent manner, this effect also demonstrated for oxLDL [36, 37]. Interestingly, oxLDL stimulated monocyte scavenger receptor and CD36 expression and differentiation to foam cells [44]. Mycobacteria can interfere with the host's lipid metabolism and drive foam cell formation [45], suggesting that BCG may also exploit lipid metabolism to induce monocyte training.

*Epigenetic regulation of innate immune training*

Epigenetic mechanisms, largely, histone modifications, regulate monocyte training (**Figure 1, Table 2**). For example, enhanced surface activation marker and inflammatory cytokine expression upon non-mycobacterial stimulation of monocytes from BCG-vaccinated adults was associated with intracellular nucleotide sensor NOD2 (nucleotide-binding oligomerisation domain-containing protein 2) dependent H3K4 trimethylation of promoters of genes encoding these monocyte markers and cytokines [36, 37]. In addition, active promoters of β-glucan-trained monocytes contained higher levels of permissive histone modifications, such as H3K4me3 and histone 3 lysine 27 acetylation (H3K27ac) than promoters in untrained monocytes [41]. The accumulation of these epigenetic markers of promoter activation at the glycolysis and mTOR pathway component genes implied cellular metabolism in innate immune training [41]. BCG-inducible monocyte training enriched the activating H3K4me3 modification at mTOR, glycolytic enzyme, TNF and IL-6 gene promoters [42]. However, the regulatory patterns of training-related histone modifications seem to be complex as mTOR or glutamine pathway inhibition cancelled H3K4me3 accumulation at the cytokine promoters [42]. Importantly, not only the permissive, but also inhibitory histone modifications, such as histone 3 lysine 9 trimethylation (H3K9me3) regulate glycolysis and mTOR pathway component or inflammatory cytokine expression in BCG-trained cells [42, 46]. BCG training was shown to suppress H3K9me3 mark while inhibition of glutamine or mTOR pathways enhanced the accumulation of this mark at the inflammatory cytokine promoters [42]. This suggests that enzymes managing histone modification patterns (methyltransferases, demethylases) may respond to intracellular metabolite changes, coordinating cytokine or other gene expression accordingly.

*Infant BCG immunisation and innate immune training*

The evidence on whether BCG induces trained innate immunity in infants and if it contributes to their protection from non-mycobacterial pathogens is somewhat controversial. Although adult BCG vaccination or *in vitro* training models suggest that BCG primes monocytes to increase surface activation markers and type 1 cytokine production in response to heterologous antigen stimulation [36-38], infant immune responses to BCG seem more difficult to define. Differently from adults, no differences in monocyte surface activation marker expression were observed upon whole blood stimulation with heterologous stimuli in the BCG-immunised infant group versus unvaccinated controls [47]. However, Pam3CSK4 (TLR1/2 agonist) stimulation upregulated NK cell activation marker CD69 in the vaccinated infant samples, implying that NK cells may mediate heterologous BCG effects in infants, similarly to the NK cells of the BCG-vaccinated adults [40]. Likewise, in agreement with the adult studies, whole blood samples from BCG-immunised low-birth-weight infants produced more TNFα, IFNγ, IL-1β and IL-6 upon Pam3CSK4 stimulation compared to the unvaccinated infants [30] (**Table 3**). Yet, different cytokine profile was identified in BCG-vaccinated UK infant whole blood cultures, with higher levels of EGF, IL-6, PDGF-AB/BB in response to Pam3CSK4, *C. albicans* and *S. aureus* challenge compared to the control group [47]. Previous studies explored more narrow cytokine profiles [30, 36, 38], so it is not clear if discrepancies reflect differences in the adult and the infant immune systems or diverse study design.

Other studies did not confirm the association between infant BCG vaccination status and heterologous immune responses. In contrast to previous findings, no TNFα production changes at 1 and 12 weeks post immunisation were found in BCG-vaccinated Gambian infants upon their PBMC stimulation with heterologous microorganisms [31] (**Table 3**). In addition, no significant changes in cytokine responses to non-specific stimuli were observed at 3 and 13 months post-randomisation in whole blood samples obtained from the BCG-vaccinated infants compared to the controls in Denmark [48]. The reasons for the discrepancies among the findings from different studies are not clear, although potentially low immunogenicity of BCG used in some studies was suggested as a possible cause [31, 48][31, 53]. In Uganda, maternal BCG scar was associated with stronger inflammatory responses in infants upon whole blood culture stimulation with TLR agonists [49], suggesting that maternal BCG status could affect infant responses. Differences in the vaccination schedules, study design or infant populations may also contribute to diverse outcomes in such studies.

*BCG and other innate immune responses*

Other mechanisms may contribute to the heterologous BCG effects. BCG-dependent immune training was shown to elevate levels of IL-6, TNFα and IL-1β in BCG-vaccinated adults and infants 2 weeks to several months post vaccination in response to heterologous stimuli [30, 36-38, 47]. These cytokines can mediate the acute phase responses, suggesting that BCG-primed immune system might exploit plasma iron regulation upon encounter with infectious microorganisms. However, a study of Gambian neonates found no association between the vaccination status and plasma iron, haemoglobin, hepcidin, ferritin or IL-6 levels in the unvaccinated controls and neonates vaccinated with OPV, HBV and BCG at birth or given BCG at 5 days of age [50]. The authors argued that early responses were measured (24-48 h and 72-96 h post immunisation), potentially missing out BCG-dependent non-specific effects and that the observed neonate plasma levels of IL-6, hepcidin and ferritin were elevated irrespective of immunisation status as a consequence of the birth process, potentially masking the non-specific effects of BCG [50]. Further studies exploring a possible relationship between the acute phase responses in infants and non-specific effects of BCG would be of interest.

*BCG-enhanced heterologous T-cell responses*

BCG may steer the immune system towards Th1-type pro-inflammatory cytokine (TNFα or IFNγ) production, activating monocytes and alveolar macrophages, so mediating classical anti-mycobacterial effects. However, this effect may extend beyond mycobacterial specificity. In mice, BCG immunisation enhanced protection from vaccinia virus via increased CD4+ T-cell IFNγ production [51]. Studies on human infant responses to BCG show similar effects (**Table 3**). BCG-Denmark improved IFNγ and IL-10 responses to tetanus toxoid at 12 months of age in a Ugandan infant cohort [52]. In Philippines, infants, given BCG at birth, had higher frequencies of tetanus toxoid specific PBMCs producing IFNγ and CD4+ memory T-cells secreting IFNγ and TNFα upon PHA stimulation [53]. In Guinea-Bissau, BCG-vaccinated infants produced more IFNγ than unvaccinated controls upon whole blood stimulation with PMA [30]. PBMCs from Gambian infants vaccinated with BCG at birth produced higher levels of IFNγ, IL-5 and IL-13 in response to hepatitis B surface antigen (HBsAg), and their lymphocytes were more proliferative compared to the cells from control infants [54]. Increased IFNγ-producing CD8+ T-cell frequency upon *C. albicans* stimulation at 1 week post BCG immunisation was observed in another Gambian infant cohort, although this effect subsided by 12 weeks post-vaccination [31]. This study also reported reduced IL-10 production in response to LPS and increased IFNγ/IL-10 ratio upon *S. pneumoniae* stimulation in BCG-vaccinated females at 12 weeks post immunisation [31]. Together, these studies suggest that BCG vaccine may enhance maturation of Th1 cells with diverse specificities, improving responses to a broad range of microbial or childhood vaccine antigens. As infant immune responses shift from Th17-like towards Th1-type in the first year of life [55], intensifying this process through BCG vaccination may contribute to heterologous infant protection from infectious diseases (**Figure 2**). However, this effect may be limited as no difference in HBsAg-specific IFNγ producing PBMC frequencies was found in BCG-vaccinated and control infants in Philippines [53], suggesting that BCG did not affect responses to hepatitis B vaccine in this population.

BCG may modulate heterologous responses in other T-cell populations – *C. albicans* and *S. aureus* boosted IL-17 and IL-22 production at 2 weeks and 1 year post-immunisation in BCG-vaccinated adults [39]. Whole blood samples from BCG-immunised infants produced less IL-13 and IL-17 upon *C. albicans* stimulation at 4 months post-immunisation than samples from unvaccinated controls [47]. Increased fraction of IL-2-producing, proliferating CD8+ *Bordetella pertussis­*-specific T-cells was found in BCG-vaccinated HIV-exposed uninfected South African infants compared to the control group [56]. This suggests that BCG may also regulate Th17, Th22 or cytotoxic T-lymphocyte subsets. It is not clear how BCG might exert this effect, but, persisting in an infant, it may prolong activation of the innate system and provide continuous cytokine signals for T-cell activation. Dendritic cells, innate lymphocytes and conventional T-cells can make IL-22, while IL-1β or IL-6 can promote its secretion, mediating immune responses to respiratory pathogens and fungal infections [57], implying that trained immunity and heterologous T-cell responses may complement one another mediating BCG-dependent heterologous infant protection from infections. Importantly, trained innate immunity wanes over time: PBMCs from BCG-vaccinated adults produce less TNFα and IL-1β in response to *C. albicans* and LPS at 1 year post-immunisation [39]. However, BCG-vaccinated infant protection from all-cause mortality extends for several years [4-7] or until adolescence [15]. Although improved neonatal or infancy survival can contribute to the long-term survival rates, the adaptive immune responses may take over heterologous infant protection from childhood infections once the trained immunity effect diminishes (**Figure 2**).

# *Potential mechanisms beyond heterologous protection from infectious diseases and cancer*

As well as reducing all-cause infant mortality, BCG may decrease the development of some cancers. A case-cohort study in Denmark suggested that BCG may reduce a risk of lymphoma [58]. Applied as a therapy against bladder cancer, BCG reduced patient mortality, tumour progression and recurrence for up to 10 years, however, this effect tended to decrease over time [59]. Infant protection from non-mycobacterial infections may share some mechanisms with BCG-dependent anti-tumour effects, with trained immunity implicated in BCG immunotherapy against bladder cancer [38]. Polymorphisms of autophagy gene *ATG2B* limited the ability of BCG-trainedmonocytes to improve IL-1β, IL-6 and TNFα production upon heterologous stimulation *in vitro* and *in vivo* and correlated with increased tumour progression and recurrence in bladder cancer patients treated with intravesical BCG [38]. Cytokines promoted by innate training, e.g. IL-1β, IL-6 and TNFα were suggested to mediate the anti-cancer effects of BCG [59], implicating overlap between BCG-mediated non-specific protection from infectious diseases and cancer. Increased frequencies of T-helper cells and IL-2, IL-12, TNFα, IFNγ and IL-10 production may also mediate the anti-cancer effects of BCG [58, 59], suggesting the involvement of heterologous T-cell responses, similarly to the observations on heterologous infant protection from infectious diseases (**Table 3**).

*Influence of BCG on humoral responses to non-mycobacterial stimuli*

Few studies explored the impact of BCG vaccination on humoral responses to heterologous antigens, however, in adults, BCG was shown to boost antibody titres against influenza vaccine [60]. Infants given BCG at birth also had higher antibody levels to HBsAg and to polio antigens than infants whose BCG vaccination was delayed [54] (**Table 4**).Elevated serum antibody concentrations to pneumococcal antigens were found in BCG-immunised Australian infants compared to the control group, although, contrary to the previous findings, lower anti-HBsAg IgG levels were detected in the BCG-vaccinated group [54, 61]. This study also observed a trend for increased concentrations of IgG against *Haemophilus influenza* and tetanus toxoid antigens [61]. Although these studies suggest BCG may have non-specific effects on antibody production to other childhood vaccines, other findings maintain the controversy over the influence of BCG on heterologous antibody titres or function. No differences in levels of antibodies to Expanded Program for Immunization (EPI) vaccine antigens were found at 12 weeks post-immunisation in BCG-vaccinated Gambian infants compared to the controls [31]. Similarly, no differences in titres of antibodies against *H. influenza*, pertussis, tetanus and hepatitis B antigens were found in South African infants immunised at birth compared to the group in which BCG immunisation was delayed [62]. A recent trial in Denmark found no association between the BCG vaccination status and antibody titres against other childhood vaccine antigens at 13 months of age [63]. Possibly, timing of BCG vaccination during early immune system development phases may influence its non-specific effects on antibody responses, as BCG may have contributed to elevated antibody titres against *H. influenza,* pertussis and several pneumococcal antigens in infants vaccinated at 2 to 7 days post-birth [63]; however, the extent of this effect is not clear. Variation in BCG strains, EPI vaccines or immunisation schedules applied in individual studies may also influence infant humoral responses. Reducing their impact in future studies may be necessary to establish whether BCG influences antibody responses to other EPI vaccines or childhood infections.

***Conclusion and future perspectives***

A large body of evidence suggests that BCG vaccination provides protection from diseases other than TB and that it may modulate the immune responses to other childhood vaccines. Several important implications for BCG and other vaccines that may exert similar beneficial heterologous effects arise from these findings.

Firstly, the immunological mechanisms beyond heterologous infant protection from infectious diseases are not understood. Although BCG or other live vaccines, such as measles-mumps-rubella (MMR) vaccine may broadly enhance monocyte activation status and function or proinflammatory, Th1-polarising responses [64], or modulate antibody responses [54, 61], the data on immune mechanisms beyond heterologous infant protection from infections is inconsistent.

To overcome this, future immunological studies or randomised trials exploring the heterologous effects of BCG and their immunological mechanisms in infants may need to reduce variation in vaccination schedules or observation timings. Although this is difficult to conduct in real-life settings, it would allow for more comparability between the studies and their outcomes. In addition, despite the controversies, future studies need to address the issue of potential vaccine interactions to ensure optimal infant protection from infectious diseases.

Further work needs to address how BCG or other childhood immunisations regulate T- and B-cell subsets of diverse antigen specificities and which memory or effector cell fractions they maintain or promote to proliferate. In parallel, the role of the innate immune responses in mediating the non-specific effects of BCG, MMR or other vaccines needs to be studied more extensively. It would also be interesting to test if other cell types (e.g. dendritic cells) could be involved in heterologous infant protection from all-cause mortality or infections. Together, this knowledge may be exploited for improving anti-TB and other childhood vaccine design.

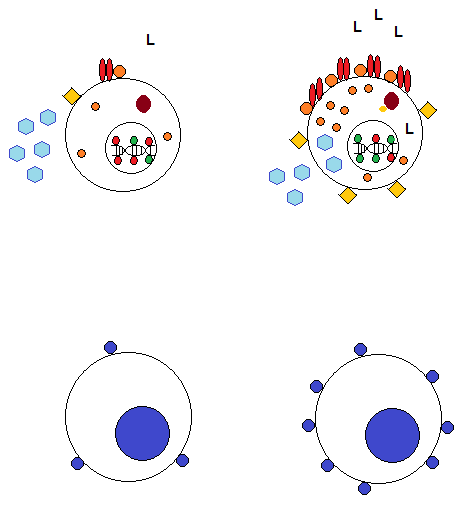
|  |
| --- |
| **Executive summary** |
| ***All-cause infant mortality reduction*** |
| * BCG reduces all-cause infant mortality, unaccounted by reduction in mycobacterial infections alone. |
| ***Impact on acquisition of infectious diseases*** |
| * BCG may decrease the risk of non-mycobacterial sepsis and respiratory infections among the vaccinated infants. |
| ***Factors potentially contributing to the heterologous BCG effect manifestation*** |
| * Heterologous effects of BCG may be modified by interactions with other vaccines, such as DTP; however, the evidence for this is controversial. * BCG can significantly decrease all-cause infant mortality and infectious disease acquisition in young infants (up to 24 months of age); however, this effect can extend throughout childhood and adolescence. * The sex of an infant may influence heterologous BCG protection from infectious diseases, with some studies reporting that female infants may benefit more from BCG-immunisation; however, the evidence for such an effect is mixed. |
| ***Mechanisms implicated in heterologous BCG-vaccinated infant protection*** |
| * BCG may enhance maturation of infant Th1 responses and monocyte and NK cell ability to cope with a broad spectrum of pathogens for prolonged time periods. * It may do so via innate immune training, a process characterised by metabolic, cytokine production and surface marker changes in monocytes and NK cells. * During innate immune training with BCG, trained cells undergo a metabolic shift from oxidative phosphorylation to glycolysis. * In parallel, changes in epigenetic regulation occur in BCG-trained cells, with accumulation of gene expression activating histone modifications accumulating at the promoters of genes encoding IL-1β IL-6 and TNFα, glycolytic pathway components and surface receptors. * The evidence for the presence of immunological mechanisms associated with trained immunity in BCG-vaccinated infants is mixed, with some studies reporting enhanced NK cell activation, elevated TNFα, IL-1β, IL-6, IFNγ, EGF or PDGF-AB/BB production upon heterologous stimulation; however, other studies found no association between BCG immunisation and heightened innate immune responses. * Improved heterologous Th1-like responses, with increased TNFα and IFNγ production in response to EPI vaccine antigens were reported in multiple sites, including Uganda, Guinea-Bissau, The Gambia or Philippines. * As BCG-vaccinated individual IL-1β and TNFα production in response to innate immunity stimuli subsides by 1 year post-immunisation, heterologous BCG-dependent T-cell activation can contribute to or maintain non-specific BCG effects once trained immunity benefits wane. * BCG may modulate infant humoral responses to other immunisations, elevating or decreasing antibody levels to such vaccines as hepatitis B or pneumococcal conjugate vaccines; however, further studies are needed to determine the extent of this effect. |
| ***Conclusion and future perspectives*** |
| * Multiple epidemiological and immunological studies confirm that BCG exerts broad, beneficial effects in the vaccinated individuals protecting them from diseases other than TB. * Trained immunity, enhanced heterologous T-cell responses and, possibly, modulated antibody responses to other vaccines may mediate the non-specific effects of BCG. * Further work is needed to address the role of these factors on BCG and other childhood vaccine dependent heterologous effects and define the underlying mechanisms. |

***Financial disclosure / Acknowledgements***

*The authors have no personal, academic or financial associations or financial involvement in an organisation or an entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. No writing assistance has been used in the creation of the manuscript.*

*Steve – may wish to add your funding here too.*

*C.E.J. has received funding from the IMmunising PRegnant women and INfants neTwork (IMPRINT), funded by the GCRF Networks in Vaccines Research and Development, which was co-funded by the MRC and BBSRC; the National Vaccine Program Office (NVPO) and Bill & Melinda Gates Foundation, Grant OPP1119788, Global Alignment of Immunization Safety Assessment in pregnancy (GAIA). CEJ is an investigator for clinical trials performed on behalf of the University of Southampton and University Hospital Southampton NHS Trust, UK, sponsored by vaccine manufacturers, including Novavax, GSK and Janssen. She has received no personal funding for these activities.*

****

**Monocytes**

**NK cells**

heterologous

microorganism

Glycolysis

Phosphorylated mTOR

+ BCG

ATP

mTOR

CD69

Lactate

+ BCG

↑IL-1β

↑EGF

↑PDGF-AB/BB

↑IL-6

↑TNFα

↑IFNγ

Oxidative phosphorylation

Glucose

H3K9me3

H3K4me3

CD11b

CD14

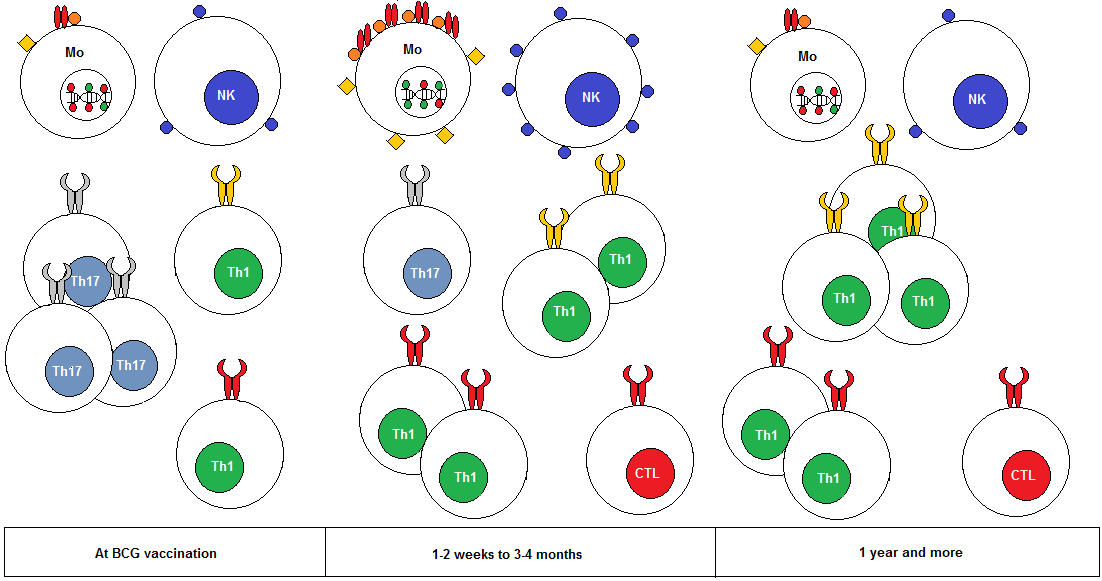
TLR4

****

↑IL-6

↑IL-1β

**Figure 1.** *BCG training-induced phenotype changes in monocytes and NK cells.* BCG training of human monocytes (top) or NK cells (bottom) *in vitro* or by vaccination increases their surface marker expression and cytokine production in response to heterologous antigen stimulation [36, 47]. In monocytes, these changes are regulated by metabolic shift from oxidative phosphorylation to glycolysis and histone modifications [42], with increased frequency of permissive H3K4me3 and reduced presence of inhibitory H3K9me3 at the promoters of cytokine, receptor and metabolic pathway component encoding genes [36-38, 42, 46]. The left side of the diagram depicts model innate immune cells prior to the BCG training and the right side – post training. Enhanced cytokine production post-training is indicated by arrows. Heterologous microorganism – secondary, non-mycobacterial infectious agent, ATP – adenosine triphosphate, EGF – epidermal growth factor, IFNγ – interferon gamma, IL-1β – interleukin 1 beta, IL-6 – interleukin 6, mTOR – mammalian target of rapamycin, PDGF-AB/BB – platelet derived growth factor AB/BB, TLR4 – Toll-like receptor 4, TNFα – tumour necrosis factor alpha. H3K4me3 – trimethylation of lysine at position 4 on histone 3, H3K9me3 – trimethylation of lysine at position 9 on histone 3; + BCG – *in vitro* or *in vivo* cell training with BCG.

****

H3K9me3

T-cell receptor

H3K4me3

CD69

CD11b

CD14

TLR4

*Time post BCG vaccination*

*Infant age*

**Figure 2.** *A model of* *cell populations mediating BCG-vaccinated infant heterologous responses.* The diagram shows the innate and adaptive immune cells implicated in non-specific infant protection and the likely timings for their involvement with respect to BCG vaccination and infant age. At of or immediately after BCG-vaccination, monocytes and NK cells of young infants are “untrained”, characterised by low surface receptor expression or cytokine production (not depicted, for simplicity). Once these cells become “trained” by BCG, they increase surface receptor expression and inflammatory cytokine production and may cope with childhood infections more readily [30, 36, 47]. This effect diminishes over time, subsiding by 1 year post vaccination [39]. BCG, however, induces mycobacteria-specific Th1 or CTL responses (yellow T-cell receptors) and may encourage the expansion of heterologous Th1 or CTL cells (red T-cell receptors), skewing the infant immune system away from Th17 responses (depicted by increasing proportion of Th1 cells and disappearance of Th17 cells) [30, 47]. BCG-supported heterologous T-cell responses may enhance trained innate immune responses from several weeks post-immunisation and provide heterologous protection from childhood infections once trained innate immunity fades. The impact of BCG on heterologous B-cell responses is not yet clear, the current evidence being contradictive (not depicted). Mo – monocyte, NK – natural killer cell, Th1 – T-helper cell 1, Th17 – T-helper cell 17, CTL – cytotoxic T-cell. The role of other cells in trained immunity or heterologous adaptive responses is not well characterised yet and is therefore not presented.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Table 1. Non-specific infant mortality reduction by BCG and interaction with DTP** | | | | | | |
| **Study** | **Study Type** | **Vaccine schedule[[5]](#footnote-5)♠** | **BCG-vaccinated vs unvaccinated infants** | **DTP-vaccinated vs unvaccinated infants** | **BCG vs BCG & DTP** | **Observed age group** |
| **Guinea-Bissau**  **(1990-1996)** [4] | Cohort | BCG & OPV at birth;  DTP at 6, 10 & 14 weeks;  MV at 9 months | Mortality rate (%) | | | Up to 5 years of age |
| At 6-12 months of age  3.9% among BCG-vaccinated  4.9% among BCG not vaccinated | At 7.5-12 months of age  4.8% among DTP-vaccinated  4.0% among DTP not vaccinated | At 7.5-9 months of age  3.9% among BCG & DTP;  2.5% among BCG only  At 10-12 months of age  5.6% among BCG & DTP; 4.1% among BCG only |
| **Burkina Faso**  **(1985-1993)** [5] | Cohort | BCG at birth;  DTP at 6, 10 & 14 weeks*[[6]](#footnote-6)#* | Mortality before two years of age risk ratio (95% CI)[[7]](#footnote-7)##: | | | Up to 2 years of age |
| **0.37 (0.29-0.48)** | **0.23 (0.12-0.43)** | BCG & DTP vs unvaccinated  **0.34 (0.29-0.40)** |
| **Senegal**  **(1989-1997)**  [6] | Cohort | DTP-IPV at 2, 4 & 6 months[[8]](#footnote-8)¶;  BCG administered with the first DTP-IPV;  MV at 9-10 months | Not analysed | | BCG & DTP vs unvaccinated  Mortality before two years of age ratio (95% CI):  Cohort 1¶: **0.70 (0.50-0.97)**  Cohort 2¶: **0.59 (0.46-0.74**) | Up to 2 years of age |
| **Senegal**  **(1996-1999)**  [21] | Cohort | **Recommended schedule:**  BCG at birth;  DTP & OPV at 6, 10 & 14 weeks;  MV at 9 months  **BCG first:**  BCG vaccinated, DTP1 not yet received;  DTP1, DTP2 or DTP3 following BCG  **BCG & DTP first:**  DTP2 or DTP3 following BCG & DTP simultaneously   **DTP first:**  BCG following DTP1, DTP2 or DTP3 | Mortality rate ratio (95% CI): | | | Up to 24 months of age |
| **0.98 (0.50-1.90)**– BCG-vaccinated, DTP1 not yet received | **1.33 (0.60-2.98)** – DTP1, no BCG  **1.41 (0.59-3.37)** – DTP2, no BCG  **0.63 (0.16-2.50)** – DTP3, no BCG | **0.98 (0.50-1.90)** – BCG-vaccinated, DTP1 not yet received  **0.96 (0.64-1.44)** – BCG first  **0.69 (0.53-0.89)** – BCG & DTP first  **1.10 (0.64-1.88)** – DTP first |
| **Guinea-Bissau**  **(2002-2006)**  [23] | Randomised trial | BCG & OPV at birth;  DTP & OPV at 6, 10 & 14 weeks;  MV at 9 months;  DTP & OPV booster at 18 months;  BCG revaccination at 19 months | BCG revaccination vs no revaccination hazard ratio (95% CI):  **1.20 (0.77-1.89)** - the whole study period | BCG revaccination vs no revaccination hazard ratio (95% CI):  **0.36 (0.13-0.99)** – DTP booster given prior to the trial  **1.78 (1.04-3.74)** – no DTP booster prior to the trial | | Up to 5 years of age |
| **India**  **(1998-2002)**  [9] | Cohort | BCG at 0-12 months;  OPV at birth, 6, 10 & 14;  DTP at 6, 10 & 14 weeks | Hazard ratio (95% CI)[[9]](#footnote-9)◊: | |  | Up to 6 months of age |
| **0.62 (0.45-0.84)** – for BCG-vaccinated vs no BCG  **0.44 (0.29-0.66)** – BCG only, no DTP  **0.72 (0.43-1.2)** – BCG & DTP | **0.70 (0.45-1.08)** - DTP prior to BCG  **0.44 (0.24-0.79)** – DTP only |
| **Philippines**  **(1988-1991)**  [19] | Cohort | BCG at 0-11 weeks;  DTP & polio vaccine at 6, 10 & 14 weeks[[10]](#footnote-10)□;  MV at 9 months | BCG-vaccinated infants only. | | Hazard ratio (95% CI)[[11]](#footnote-11)□□:  **0.18 (0.04-0.81)** – females, no DTP  **0.27 (0.14-0.54)** – DTP-vaccinated females  **0.32 (0.16-0.62)** – DTP-vaccinated males | Up to 30 months of age |
| **India**  **(1987-1989)**  [24] | Cohort | **Recommended schedule:**  BCG at birth;  DTP & OPV at 2, 3 & 4 months;  MV at 9 months;  DTP & OPV booster at 18 months  **Altered sequence of BCG and DTP vaccination:**  BCG & DTP simultaneously  BCG following DTP | Mortality rate to 1 year of age (%): | | Mortality rate ratio (95% CI) in the first 9-12 months prior to receiving MV[[12]](#footnote-12)●:  **0.11 (0.01–0.91)** – BCG & DTP simultaneously and the last in the sequence vs DTP following BCG  **0.14 (0.02–1.05)** - BCG & DTP simultaneously and the last in the sequence vs DTP only  **0.13 (0.02–0.94)** – BCG & DTP simultaneously and the last in the sequence vs DTP as the last vaccine in the sequence  **0.27 (0.09–0.83)** – BCG alone or BCG & DTP simultaneously and the last in the sequence vs DTP as the last vaccine in the sequence | Up to 5 years of age |
| 2.2% in BCG only group  3.6% in the unvaccinated group | 2.4% in DTP only group  3.6% in the unvaccinated group |
| CI – confidence interval, BCG – Bacille Calmette-Guérin, DTP1, 2 or 3 – 1st, 2nd or 3rd dose of diphtheria-tetanus-pertussis vaccine, IPV – inactivated polio vaccine, MV – measles vaccine, OPV – oral polio vaccine | | | | | | |

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **Table 2. Impact of BCG training on histone modification patterns at the promoters of the immune and metabolic genes** | | | | | | | |
| **Gene** | **Product** | **Function** | **Histone modification** | **Impact on gene expression** | **Cell type** | **Model** | **Reference** |
| ***TNFα***  ***IL-6*** | **Tumour necrosis factor α**  **Interleukin 6** | Immune responses | ↑H3K4me3 | Permissive | Monocytes | *In vivo* / BCG vaccination | [36] |
| *In vitro* /γBCG training | [37] |
| *In vitro* / BCG training | [38] |
| ↑H3K4me3  ↓H3K9me3 | Permissive  Inhibitory | Monocytes | *In vitro* / BCG training | [42] |
| ↓H3K9me3 | Inhibitory | Monocytes / macrophages | *In vitro* / BCG training | [46] |
| ***mTOR***  ***HK2***  ***PFKP*** | **Mammalian target of rapamycin**  **Hexokinase 2**  **Platelet phosphofructokinase** | Glycolysis | ↑H3K4me3  ↓H3K9me3 | Permissive  Inhibitory | Monocytes | *In vitro* / BCG training | [42] |
| ***GLS***  ***GLUD[[13]](#footnote-13)#*** | **Glutaminase**  **Glutamate dehydrogenase** | Glutaminolysis | ↑H3K4me3  ↓H3K9me3 | Permissive  Inhibitory | Monocytes | *In vitro* / BCG training | [42] |
| H3K4me3 – trimethylation of histone 3 at lysine 4, H3K9me3 – trimethylation of histone 3 at lysine 9, γBCG – γ-irradiated Bacille Calmette Guérin | | | | | | | |

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Table 3. Heterologous effects of BCG on cytokine production, cell surface marker expression and proliferation** | | | | | | | |  |
| **Study** | **Vaccine schedule[[14]](#footnote-14)♠** | **Assay** | **Age at observation** | **Secondary stimulus** | **BCG vs Control** | | | |
| **Cytokine production** | | **Surface marker expression** | **Proliferation** |
| **The Gambia**[54] | BCG-Pasteur at birth  OR  BCG at 2 months  HBV at birth, 2 & 4 months;  OPV at birth, 1, 2 & 3 months;  DTP at 2, 3 & 4 months  Control – BCG at 4.5 months | PBMCs | At birth, 2 & 4 months | HBsAg | ↑IFNγ, IL-5 and IL-13 at 2 and 4.5 months of age in infants given BCG at birth  ↑IL-5 and IL-13 at 4.5 months of age in infants given BCG at 2 months | |  | ↑Lymphocyte proliferation at 2 and 4.5 months in infants given BCG at birth  ↑Lymphocyte proliferation at 4.5 months in infants given BCG at 2 months |
| TT | ↑IL-5 at 4.5 months of age in infants given BCG at 2 months  ↑IL-13 at 4.5 months of age in infants given BCG at birth or 2 months | |  | No lymphocyte proliferation changes |
| **Uganda**[52] | BCG-Bulgaria, BCG-Denmark or BCG-Russia at birth;  OPV at birth, 6, 10 & 14 weeks;  DTP, Hib and HBV  MV at 9 months | Whole blood | 12 months | TT | ↑IFNγ in infants given BCG-Denmark  ↑IL-10 in infants given BCG-Bulgaria or BCG-Denmark | |  |  |
| PHA | ↑IFNγ, IL-10, IL-13 in infants given BCG-Denmark | |  |  |
| **South Africa**[56] | BCG-Denmark at birth  Control – BCG at 8 weeks | Whole blood | 8 & 14 weeks | SEB | n.s. | |  | ↑CD4+ T-cell proliferation at 14 weeks |
| BP | ↑IL-2+CD8+ T-cells at 8 weeks  ↑IL-13+ CD4+ and CD8+T-cells at 14 weeks |  | |  |
| **Guinea-Bissau** [30] | OPV at birth;  BCG-Denmark at birth;  Penta at 6 weeks  Control - BCG at 6 weeks | Whole blood | 4 weeks | Pam3CSK4 | ↑IL-1β, IL-6, TNFα, IFNγ | |  |  |
| PMA & ionomycin | ↑IL-6, IFNγ | |  |  |
| **The Gambia**[31] | OPV & HBV at birth;  BCG-Russia at 6 weeks;  Penta, PCV-13 & OPV at 8, 12 & 16 weeks;  Control – BCG at 18 weeks | PBMC | 6, 7 & 18 weeks[[15]](#footnote-15)\* | LPS | ↓IL-10 in females at 18 weeks\* | |  |  |
| PMA & ionomycin | ↓IFNγ in females at 18 weeks\* | |  |  |
| *C. albicans* | ↑IFNγ+CD8+ T-cells in males and females at 7 weeks\* |  | |  |
| *S. pneumoniae* | ↑IFNγ/IL-10 ratio in females at 18 weeks\* | |  |  |
| **United Kingdom**[47] | BCG-Denmark at 6 weeks  Control – no BCG | Whole blood | 4 months post vaccination | LPS | ↑IL-8  ↓GM-CSF, GRO | |  |  |
| Pam3CSK4 | ↑EGF, IL-6, PDGF-AB/BB, MCP-3, IL-7, IL-10, IL-12p40, sCD40L, Eotaxin, MIP-1α | | ↑CD69 on NK cells |  |
| *C. albicans* | ↑EGF, IL-6, PDGF-AB/BB, MCP-3  ↓IL-2, IL-13, IL-17, IP-10 | |  |  |
| *S. aureus* | ↑EGF, IL-6, PDGF-AB/BB | |  |  |
| *E. coli* | ↑EGF  ↓GM-CSF, GRO | |  |  |
| **Denmark**[48] | BCG-Denmark at 0-7 days;  DiTeKiPol/Act-Hib[[16]](#footnote-16)# & Prevenar 13 at 3, 5 & 12 months  Control – no BCG | Whole blood | 4 days, 3 & 13 months post randomisation to BCG or control groups | *C. albicans* | ↑TNFα/IL-10 at 13 months | |  |  |
| **Philippines**[53] | BCG at 0-2 weeks  OR  BCG after the first DTP & OPV dose | PBMCs | 2-3 months | TT | ↑IFNγ+ PBMCs in infants vaccinated at 0-2 weeks | |  |  |
| Polio 1-3 | ↑IFNγ+ PBMC trend in infants vaccinated at 0-2 weeks | |  |  |
| PMA & ionomycin | ↑IFNγ+TNFα+CD45RO+CD4+ T-cells in infants vaccinated at 0-2 weeks |  | |  |
| BP – whole cell *Bordetella pertussis*,CL075 – TLR7/8 agonist, HBsAg – hepatitis B surface antigen, HBV – hepatitis B vaccine, Pam3CSK4 – S)-(2,3-bis(palmitoyloxy)-(2RS) -propyl)- *N*-palmitoyl-(R)-Cys-(S)-Ser(S)-Lys4-OH trihydrochloride, PCV-13, Prevenar 13 – 13-valent pneumococcal conjugate vaccine, Penta – diphtheria-pertussis-tetanus-*Haemophilus influenzae b* –HBV, PHA – phytohaemagglutinin,PMA – phorbol myristate acetate, Polio1-3 – poliovirus types 1-3 antigens, PPD – purified protein derivative; SEB – staphylococcal enterotoxin B, TT – tetanus toxoid, n.s. – not significant | | | | | | | | |

|  |  |  |  |
| --- | --- | --- | --- |
| **Table 4. Influence of BCG on antibody responses to EPI vaccines** | | | |
| **Study** | **Vaccine schedule[[17]](#footnote-17)♠** | **Age at observation** | **BCG vs Control** |
| **The Gambia** [54] | BCG-Pasteur at birth  OR  BCG at 2 months  HBV at birth, 2 & 4 months;  OPV at birth, 1, 2 & 3 months;  DTP at 2, 3 & 4 months  Control – BCG at 4.5 months | At birth, 2 & 4 months | **↑**αHBs at 2 and 4.5 months of age in infants vaccinated with BCG at birth  ↑αPV1 at 4.5 months of age in infants vaccinated at 2 months of age |
| **Australia**[61] | BCG-Denmark, BCG-Japan or BCG-Russia at birth;  HBV at birth;  PCV-7, Infanrixhexa[[18]](#footnote-18)\* & Rota Teq[[19]](#footnote-19)\*\* at 2, 4 & 6 months;  Control – no BCG | 4 weeks after the last immunisation | ↑αPn against serotypes 9v & 18c  Trend for ↑αPn against serotype 6b  ↓αHBs |
| **South Africa**[62] | BCG-Denmark & OPV at birth;  TETRActHib, HBV & OPV at 6, 10 & 14 weeks;  MV at 9 months  Control – BCG at 14 weeks | 14, 24 and 52 weeks | No differences in levels of αHib*,* αPT, αTT and αHBs antibodies |
| **The Gambia**[31] | OPV & HBV at birth;  BCG-Russia at 6 weeks of age;  Penta[[20]](#footnote-20)▫, PCV-13 & OPV at 8, 12 & 16 weeks  Control – BCG at 18 weeks | 6, 7 & 18 weeks | No differences in levels of αPV1, αPV2, αHBs, αDP, αPT and αTT antibodies |
| **Denmark**[63] | BCG-Denmark at 0-7 days;  DiTeKiPol/Act-Hib[[21]](#footnote-21)# & Prevenar 13 at 3, 5 & 12 months  Control – no BCG | 13 months | No differences in levels of IgG against αPT, αDP, αTT, αHib or αPn against serotypes 4, 6b, 9v, 14, 18c, 19f, 23f |
| Antibodies: αDP – anti-diphtheria, αHBs – anti-HBsAg, αHib – anti-*H. influenzae,* αPV1 – anti-poliovirus type 1, αPn – anti-pneumococcal, αPT – anti-pertussis, αTT – anti-tetanus; DTP – diphtheria-tetanus-pertussis vaccine, HBV – hepatitis B vaccine, MV – measles vaccine, OPV – oral polio vaccine, PCV-7 – 7-valent pneumococcal vaccine | | | |

**References:**

1. Trunz BB, Fine PEM, Dye C. Effect of BCG vaccination on childhood tuberculous meningitis and miliary tuberculosis worldwide: a meta-analysis and assessment of cost-effectiveness*.* *Lancet.* 367(9517), 1173-1180 (2006).

2. Colditz GA, Brewer TF, Berkey CS *et al*. Efficacy of BCG vaccine in the prevention of tuberculosis. Meta-analysis of the published literature*.* *JAMA.* 271(9), 698-702 (1994).

3. Mangtani P, Abubakar I, Ariti C *et al*. Protection by BCG Vaccine Against Tuberculosis: A Systematic Review of Randomized Controlled Trials*.* *Clin. Infect. Dis.* 58(4), 470-480 (2014).

4. Kristensen I, Aaby P, Jensen H. Routine vaccinations and child survival: follow up study in Guinea-Bissau, West Africa*.* *BMJ.* 321(7274), 1435-1438 (2000).

5. Vaugelade J, Pinchinat S, Guiella G, Elguero E, Simondon F. Non-specific effects of vaccination on child survival: prospective cohort study in Burkina Faso*.* *BMJ.* 329 (2004).

6. Elguero E, Simondon KB, Vaugelade J, Marra A, Simondon F. Non-specific effects of vaccination on child survival? A prospective study in Senegal*.* *Trop. Med. Int. Health.* 10(10), 956-960 (2005).

7. Nankabirwa V, Tumwine JK, Mugaba PM, Tylleskär T, Sommerfelt H. Child survival and BCG vaccination: a community based prospective cohort study in Uganda*.* *BMC Public Health.* 15(1), 175 (2015).

8. Aaby P, Roth A, Ravn H *et al*. Randomized trial of BCG vaccination at birth to low-birth-weight children: beneficial nonspecific effects in the neonatal period? *J. Infect. Dis.* 204(2), 245-252 (2011).

9. Moulton LH, Rahmathullah L, Halsey NA, Thulasiraj RD, Katz J, Tielsch JM. Evaluation of non-specific effects of infant immunizations on early infant mortality in a southern Indian population*.* *Trop. Med. Int. Health.* 10(10), 947-955 (2005).

10. Aaby P, Vessari H, Nielsen J *et al*. Sex Differential Effects of Routine Immunizations and Childhood Survival in Rural Malawi*.* *Pediatr. Infect. Dis. J.* 25(8), 721-727 (2006).

11. Biering-Sørensen S, Aaby P, Napirna BM *et al*. Small Randomized Trial Among Low–birth-weight Children Receiving Bacillus Calmette-Guérin Vaccination at First Health Center Contact*.* *Pediatr. Infect. Dis. J.* 31(3), 306-308 (2012).

12. Higgins JP, Soares-Weiser K, Lopez-Lopez JA *et al*. Association of BCG, DTP, and measles containing vaccines with childhood mortality: systematic review*.* *BMJ.* 355(i5170), 1-13 (2016).

13. Higgins JP, Soares-Weiser K, Reingold A. Systematic review of the non-specific effects of BCG, DTP and measles containing vaccines*.* *WHO SAGE Report.*  1-34 (2014).

14. Elliott AM, Nakiyingi J, Quigley MA, French N, Gilks CF, Whitworth JA. Inverse association between BCG immunisation and intestinal nematode infestation among HIV-1-positive individuals in Uganda*.* *Lancet.* 354(9183), 1000-1001 (1999).

15. De Castro MJ, Pardo-Seco J, Martinón-Torres F. Nonspecific (Heterologous) Protection of Neonatal BCG Vaccination Against Hospitalization Due to Respiratory Infection and Sepsis*.* *Clin. Infect. Dis.* 60(11), 1611-1619 (2015).

16. Hollm-Delgado M-G, Stuart EA, Black RE. Acute Lower Respiratory Infection Among Bacille Calmette-Guérin (BCG)–Vaccinated Children*.* *Pediatrics.* 133(1), e73-e81 (2014).

17. Kjaergaard J, Birk NM, Nissen TN *et al*. Nonspecific effect of BCG vaccination at birth on early childhood infections: a randomized, clinical multicenter trial*.* *Pediatr. Res.* 80(5), 681-685 (2016).

18. Stensballe LG. Does BCG have non-specific effects on childhood mortality? *BMJ.* 356(j700), (2017).

19. Chan GJ, Moulton LH, Becker S, Muñoz A, Black RE. Non-specific effects of diphtheria–tetanus–pertussis vaccination on child mortality in Cebu, The Philippines*.* *Int. J. Epidemiol.* 36(5), 1022-1029 (2007).

20. Aaby P, Benn CS, Nielsen J, Ravn H. Sex-differential non-specific effects of BCG and DTP in Cebu, The Philippines*.* *Int. J. Epidemiol.* 38(1), 320-323 (2009).

21. Aaby P, Nielsen J, Benn CS, Trape J-F. Sex-differential and non-specific effects of routine vaccinations in a rural area with low vaccination coverage: an observational study from Senegal*.* *Trans. R. Soc. Trop. Med. Hyg.* 109(1), 77-84 (2015).

22. Aaby P, Benn C, Nielsen J, Lisse IM, Rodrigues A, Ravn H. Testing the hypothesis that diphtheria–tetanus–pertussis vaccine has negative non-specific and sex-differential effects on child survival in high-mortality countries*.* *BMJ Open.* 2(3), 1-24 (2012).

23. Roth AE, Stabell Benn C, Ravn H *et al*. Effect of revaccination with BCG in early childhood on mortality: randomised trial in Guinea-Bissau*.* *BMJ.* 340 1-11 (2010).

24. Hirve S, Bavdekar A, Juvekar S, Benn CS, Nielsen J, Aaby P. Non-specific and sex-differential effects of vaccinations on child survival in rural western India*.* *Vaccine.* 30(50), 7300-7308 (2012).

25. Roth A, Gustafson P, Nhaga A *et al*. BCG vaccination scar associated with better childhood survival in Guinea-Bissau*.* *Int. J. Epidemiol.* 34(3), 540-547 (2005).

26. Roth A, Sodemann M, Jensen H *et al*. Tuberculin Reaction, BCG Scar, and Lower Female Mortality*.* *Epidemiology.* 17(5), 562-568 (2006).

27. Aaby P, Jensen H, Walraven G. Age-specific changes in the female–male mortality ratio related to the pattern of vaccinations: An observational study from rural Gambia*.* *Vaccine.* 24(22), 4701-4708 (2006).

28. Aaby P, Jensen H, Rodrigues A *et al*. Divergent female–male mortality ratios associated with different routine vaccinations among female–male twin pairs*.* *Int. J. Epidemiol.* 33(2), 367-373 (2004).

29. Krishnan A, Srivastava R, Dwivedi P, Ng N, Byass P, Pandav CS. Non-specific sex-differential effect of DTP vaccination may partially explain the excess girl child mortality in Ballabgarh, India*.* *Trop. Med. Int. Health.* 18(11), 1329-1337 (2013).

30. Jensen KJ, Larsen N, Biering-Sørensen S *et al*. Heterologous Immunological Effects of Early BCG Vaccination in Low-Birth-Weight Infants in Guinea-Bissau: A Randomized-controlled Trial*.* *J. Infect. Dis.* 211(6), 956-967 (2015).

31. Darboe F, Adetifa JU, Reynolds J *et al*. Minimal Sex-Differential Modulation of Reactivity to Pathogens and Toll-Like Receptor Ligands following Infant Bacillus Calmette–Guérin Russia Vaccination*.* *Front. Immunol.* 8(1092), 1-13 (2017).

32. Shann F. Nonspecific Effects of Vaccines and the Reduction of Mortality in Children*.* *Clin. Ther.* 35(2), 109-114 (2013).

33. Sankoh O, Welaga P, Debpuur C *et al*. The non-specific effects of vaccines and other childhood interventions: the contribution of INDEPTH Health and Demographic Surveillance Systems*.* *Int. J. Epidemiol.* 43(3), 645-653 (2014).

34. Levy O, Levy O. Ready to benefit from training: heterologous effects of early life immunization*.* *Trans. R. Soc. Trop. Med. Hyg.* 109(1), 3-4 (2015).

35. Van't Wout JW, Poell R, Van Furth R. The Role of BCG/PPD-Activated Macrophages in Resistance against Systemic Candidiasis in Mice*.* *Scand. J. Immunol.* 36(5), 713-720 (1992).

36. Kleinnijenhuis J, Quintin J, Preijers F *et al*. Bacille Calmette-Guerin induces NOD2-dependent nonspecific protection from reinfection via epigenetic reprogramming of monocytes*.* *PNAS.* 109(43), 17537–17542 (2012).

37. Arts RJW, Blok BA, Aaby P *et al*. Long-term in vitro and in vivo effects of γ-irradiated BCG on innate and adaptive immunity*.* *J. Leukoc. Biol.* 98(6), 995-1001 (2015).

38. Buffen K, Oosting M, Quintin J *et al*. Autophagy Controls BCG-Induced Trained Immunity and the Response to Intravesical BCG Therapy for Bladder Cancer*.* *PLOS Pathog.* 10(10), 1-10 (2014).

39. Kleinnijenhuis J, Quintin J, Preijers F *et al*. Long-Lasting Effects of BCG Vaccination on Both Heterologous Th1/Th17 Responses and Innate Trained Immunity*.* *J. Innate Immun.* 6(2), 152-158 (2014).

40. Kleinnijenhuis J, Quintin J, Preijers F *et al*. BCG-induced trained immunity in NK cells: role for non-specific protection to infection*.* *Clin. Immunol.* 155(2), 213-219 (2014).

41. Cheng S-C, Quintin J, Cramer RA *et al*. mTOR- and HIF-1α–mediated aerobic glycolysis as metabolic basis for trained immunity*.* *Science.* 345(6204), (2014).

42. Arts RJW, Carvalho A, La Rocca C *et al*. Immunometabolic Pathways in BCG-Induced Trained Immunity*.* *Cell Rep.* 17(10), 2562-2571 (2016).

43. Bekkering S, Blok BA, Joosten LA, Riksen NP, Van Crevel R, Netea MG. In Vitro Experimental Model of Trained Innate Immunity in Human Primary Monocytes*.* *Clin. Vaccine Immunol.* 23(12), 926-933 (2016).

44. Bekkering S, Quintin J, Joosten LA, Van Der Meer JW, Netea MG, Riksen NP. Oxidized low-density lipoprotein induces long-term proinflammatory cytokine production and foam cell formation via epigenetic reprogramming of monocytes*.* *Arterioscler. Thromb. Vasc. Biol.* 34(8), 1731-1738 (2014).

45. Lovewell RR, Sassetti CM, Vanderven BC. Chewing the fat: lipid metabolism and homeostasis during *M. tuberculosis* infection*.* *Curr. Opin. Microbiol.* 29 30-36 (2016).

46. Arts RJW, Blok BA, Van Crevel R *et al*. Vitamin A induces inhibitory histone methylation modifications and down-regulates trained immunity in human monocytes*.* *J. Leukoc. Biol.* 98(1), 129-136 (2015).

47. Smith SG, Kleinnijenhuis J, Netea MG, Dockrell HM. Whole Blood Profiling of Bacillus Calmette–Guérin-Induced Trained Innate Immunity in Infants Identifies Epidermal Growth Factor, IL-6, Platelet-Derived Growth Factor-AB/BB, and Natural Killer Cell Activation*.* *Front. Immunol.* 8 1-11 (2017).

48. Nissen TN, Birk NM, Blok BA *et al*. Bacillus Calmette-Guérin vaccination at birth and in vitro cytokine responses to non-specific stimulation. A randomized clinical trial*.* *Eur. J. Clin. Microbiol. Infect. Dis.* 37(1), 29-41 (2017).

49. Mawa PA, Webb EL, Filali-Mouhim A *et al*. Maternal BCG scar is associated with increased infant proinflammatory immune responses*.* *Vaccine.* 35(2), 273-282 (2017).

50. Prentice S, Jallow MW, Prentice AM. The effect of BCG on iron metabolism in the early neonatal period: A controlled trial in Gambian neonates*.* *Vaccine.* 33(26), 2963-2967 (2015).

51. Mathurin KS, Martens GW, Kornfeld H, Welsh RM. CD4 T-Cell-Mediated Heterologous Immunity between Mycobacteria and Poxviruses*.* *J. Virol.* 83(8), 3528-3539 (2009).

52. Anderson EJ, Webb EL, Mawa PA *et al*. The influence of BCG vaccine strain on mycobacteria-specific and non-specific immune responses in a prospective cohort of infants in Uganda*.* *Vaccine.* 30(12), 2083-2089 (2012).

53. Libraty DH, Zhang L, Woda M *et al*. Neonatal BCG vaccination is associated with enhanced T-helper 1 immune responses to heterologous infant vaccines*.* *Trials Vaccinol.* 3 1-5 (2014).

54. Ota MOC, Vekemans J, Schlegel-Haueter SE *et al*. Influence of Mycobacterium bovis Bacillus Calmette-Guérin on Antibody and Cytokine Responses to Human Neonatal Vaccination*.* *J. Immunol.* 168(2), 919-925 (2002).

55. Kollmann TR, Levy O, Montgomery RR, Goriely S. Innate immune function by Toll-like receptors: distinct responses in newborns and the elderly*.* *Immunity.* 37(5), 771-783 (2012).

56. Blakney AK, Tchakoute CT, Hesseling AC *et al*. Delayed BCG vaccination results in minimal alterations in T cell immunogenicity of acellular pertussis and tetanus immunizations in HIV-exposed infants*.* *Vaccine.* 33(38), 4782-4789 (2015).

57. Mcaleer JP, Kolls JK. Directing traffic: IL-17 and IL-22 coordinate pulmonary immune defense*.* *Immunol. Rev.* 260(1), 129-144 (2014).

58. Villumsen M, Sørup S, Jess T *et al*. Risk of lymphoma and leukaemia after bacille Calmette-Guérin and smallpox vaccination: A Danish case-cohort study*.* *Vaccine.* 27(49), 6950-6958 (2009).

59. Gandhi NM, Morales A, Lamm DL. Bacillus Calmette-Guérin immunotherapy for genitourinary cancer*.* *BJU Internat.* 112(3), 288-297 (2013).

60. Leentjens J, Kox M, Stokman R *et al*. BCG Vaccination Enhances the Immunogenicity of Subsequent Influenza Vaccination in Healthy Volunteers: A Randomized, Placebo-Controlled Pilot Study*.* *J. Infect. Dis.* 212(12), 1930-1938 (2015).

61. Ritz N, Mui M, Balloch A, Curtis N. Non-specific effect of Bacille Calmette-Guérin vaccine on the immune response to routine immunisations*.* *Vaccine.* 31(30), 3098-3103 (2013).

62. Hesseling AC, Blakney AK, Jones CE *et al*. Delayed BCG immunization does not alter antibody responses to EPI vaccines in HIV-exposed and -unexposed South African infants*.* *Vaccine* 34(32), 3702-3709 (2016).

63. Nissen TN, Birk NM, Smits G *et al*. Bacille Calmette-Guérin (BCG) vaccination at birth and antibody responses to childhood vaccines. A randomised clinical trial*.* *Vaccine.* 35(16), 2084-2091 (2017).

64. Nicoli F, Appay V. Immunological considerations regarding parental concerns on pediatric immunizations*.* *Vaccine.* 35(23), 3012-3019 (2017).

1. Department of Immunology and Infection, Faculty of Infectious and Tropical Diseases, London School of Hygiene & Tropical Medicine, Keppel Street, London, WC1E 7HT, United Kingdom [↑](#footnote-ref-1)
2. Faculty of Medicine and Institute for Life Sciences, University of Southampton and University Hospital Southampton NHS Foundation Trust, Tremona Road, Southampton, SO16 6YD, United Kingdom [↑](#footnote-ref-2)
3. Paediatric Infectious Diseases Research Group, St George’s, University of London, Cranmer Terrace, London, SW17 0RE, United Kingdom [↑](#footnote-ref-3)
4. \* Author for correspondence: [Steven.Smith@lshtm.ac.uk](mailto:Steven.Smith@lshtm.ac.uk) [↑](#footnote-ref-4)
5. ♠ Vaccination timings correspond to infant age at time of vaccination. [↑](#footnote-ref-5)
6. # Infants were considered unvaccinated until the age of immunisation with a specified vaccine. [↑](#footnote-ref-6)
7. *##* Adjusted for the area, dispensary in a village, use of health services, diarrhoea in the first year of life and birth season. [↑](#footnote-ref-7)
8. ¶ Cohort 1 vaccinated as indicated in the *Vaccination Schedule* section. Cohort 2 received OPV instead of IPV. [↑](#footnote-ref-8)
9. ◊ Assumed hazard ratio for unvaccinated infants equals 1. [↑](#footnote-ref-9)
10. □ Type of polio vaccine (OPV vs IPV) was not specified. [↑](#footnote-ref-10)
11. □□ Assumed hazard ratio for infant males not vaccinated with DTP equals 1. The cited hazard ratio rates exclude two deaths of infants with an unknown DTP vaccination status. [↑](#footnote-ref-11)
12. ● Mortality rate ratio adjusted for most recent weight and controlled for age. [↑](#footnote-ref-12)
13. # H3K4me3 pattern change not significant upon BCG training. [↑](#footnote-ref-13)
14. ♠ Vaccination timings correspond to infant age at time of vaccination. [↑](#footnote-ref-14)
15. \* These timings correspond to the timing before the BCG vaccination, 1 week and 12 weeks post BCG vaccination, respectively. [↑](#footnote-ref-15)
16. # Due to DiTeKiPol/Act-Hib availability issues, 26 BCG-vaccinated and 48 control infants received Infanrixhexa.

    DiTeKiPol/Act-Hib contains diphtheria toxoid, tetanus toxoid, polio virus types 1-3, *H. influenzae* type b polysaccharide.

    Infanrixhexa also contains hepatitis B surface antigen and lower content of pertussis toxoid and aluminium (*Nissen et al., Vaccine, 2017*). [↑](#footnote-ref-16)
17. ♠ Vaccination timings correspond to infant age at time of vaccination. [↑](#footnote-ref-17)
18. \* Diphtheria-tetanus-pertussis-hepatitis B-inactivated polio virus-*Haemophilus influenzae* type b vaccine [↑](#footnote-ref-18)
19. \*\* Oral pentavalent rotavirus vaccine [↑](#footnote-ref-19)
20. ▫ Penta – diphtheria-pertussis-tetanus-*Haemophilus influenzae b* –HBV [↑](#footnote-ref-20)
21. # Due to DiTeKiPol/Act-Hib availability issues, 44 BCG-vaccinated and 51 control infants received Infanrixhexa. [↑](#footnote-ref-21)