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Bifunctional CD4-DC-SIGN Fusion Proteins Demonstrate Enhanced Avidity to gp120 and Inhibit HIV-1 Infection and Dissemination

Tao Du[1,2], Kai Hu[1,2], Jun Yang[1†], Jing Jin[3], Chang Li[1,2], Daniel Stieh[4], George E. Griffin[4], Robin J. Shattock[5], Qinxue Hu[1,4*]

Running title: CLDs inhibit HIV infection and dissemination

[1]State Key Laboratory of Virology, Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan 430071, China

[2]Graduate University, Chinese Academy of Sciences, Beijing 100049, China

[3]Department of Obstetrics and Gynecology, Hubei Hospital of Traditional Chinese Medicine, Wuhan 430061, China

[4]Center for Infection and Immunity, St George's University of London, London, SW17 0RE, UK

[5]Section of Infectious Diseases, Faculty of Medicine, Imperial College London, St. Mary's Campus, London W2 1PG, UK

*Corresponding author. Mailing address: State Key Laboratory of Virology, Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan 430071, China.

Phone: +86 27 8719 9992. Fax: +86 27 8719 9992. Email: qhu@wh.iov.cn

[†]Current address: Zhengzhou Tobacco Research Institute, Zhengzhou 450001, China.

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ABSTRACT

Early stages of mucosal infection are potential targets for HIV-1 prevention. CD4 is the primary receptor in HIV-1 infection while DC-SIGN likely plays an important role in HIV-1 dissemination in particular during sexual transmission. To test the hypothesis that an inhibitor simultaneously targeting both CD4 and DC-SIGN binding sites on gp120 may provide a potent anti-HIV strategy, we designed constructs by fusing the extracellular CD4 and DC-SIGN domains together with varied arrangements of the lengths of CD4, DC-SIGN and the linker. We expressed, purified and characterized a series of soluble CD4-linker-DC-SIGN (CLD) fusion proteins. Several CLDs, composed of a longer linker and an extra neck domain of DC-SIGN, had enhanced affinity for gp120 as evidenced by molecular interaction analysis. Furthermore, such CLDs exhibited significantly enhanced neutralization activity against both laboratory-adapted and primary HIV-1 isolates. Moreover, CLDs efficiently inhibited HIV-1 infection in trans via a DC-SIGN expressing cell line and primary human dendritic cells. This was further strengthened by the results from the human cervical explant model, showing that CLDs potently prevented both localized and disseminated infections. This is the first time that sDC-SIGN-based bifunctional proteins have demonstrated anti-HIV potency. Our study provides proof-of-concept that targeting both CD4 and DC-SIGN binding sites on gp120 represents a novel antiviral strategy. Given that DC-SIGN binding to gp120 increases exposure of the CD4 binding site and that the soluble forms of CD4 and DC-SIGN occur in vivo, further improvement of CLDs may render their potential to be used as prophylaxis or therapeutics.

INTRODUCTION

The majority of HIV-1 infections are acquired by mucosal exposure, with sexual transmission as the leading mode of HIV-1 infection worldwide. The distribution of dendritic cells (DCs) in cervicovagina and colorectum may allow them to be one of the first cell types to contact HIV-1 (21, 34, 57). DCs may capture HIV-1 by attachment receptors in particular DC-SIGN and transfer the virus to permissive $CD4^+$ T cells, resulting in trans-infection (11, 19, 37, 55). In addition, DC-SIGN binding to HIV-1 increases local concentration of virus on DC surface and can enhance cis-infection via the low levels of CD4 and CCR5 on DCs (38). Both HIV-1-captured and -infected DCs can efficiently release virus particles to $CD4^+$ T cells at the points of cell contact termed virological synapses (41). Evidences from colorectal explant study indicate that DC-SIGN accounts for 90% of HIV-1 binding on mucosal mononuclear cells (23). Our previous study using a cervicovaginal model demonstrates that simultaneous blockade of CD4 and mannose-binding C-type lectin receptors (MCLRs) including DC-SIGN is required to inhibit HIV-1 uptake and dissemination by migratory cells (28). Given their critical roles likely played in HIV-1 transmission, CD4 and DC-SIGN are important targets for the development of topical microbicides.

HIV-1 entry and transmission involves complex interaction between viral envelope glycoprotein (Env) and receptors on host cells. The binding of gp120 to CD4 is virtually universal among HIV-1 isolates. Soluble CD4 (sCD4), which acts as a receptor decoy to prevent the engagement of HIV-1 Env with cell surface CD4, represents a promising competitive viral attachment inhibitor. However, despite its efficient neutralization activity against laboratory-adapted HIV-1 strains, sCD4 showed poor antiviral activity against primary HIV-1 isolates and very large doses of

sCD4 were required to achieve modest reductions of viral loads in vivo (33). This is likely due to the relatively lower Env-binding affinity of sCD4 per se compared with that of target cell-bound CD4 (52). Although PRO-542 (CD4-IgG2), a tetrameric fusion protein between CD4 and immunoglobulin-G, is much more potent in vitro than the parental monomer, the translation of this improvement to clinical use remains uncertain (2, 51). The interaction of HIV-1 with DC-SIGN does not result in direct infection of DCs, but instead enhances cis- or/and trans-infection. Several studies have shown that antagonists against DC-SIGN inhibit DC-SIGN-mediated HIV-1 transmission (7, 42, 46), whereas the antiviral activity of sDC-SIGN seems more complex (25, 40). Although sDC-SIGN decreases the capture of HIV-1 by DC-SIGN (39), sDC-SIGN binding to HIV-1 Env can also increase the exposure of the CD4 binding site on gp120 which in turn contributes to enhancement of infection (25), compromising the development of DC-SIGN as a single agent.

We hypothesized that an inhibitor against both CD4 and DC-SIGN binding sites on gp120 might represent a better anti-HIV strategy and that a sCD4-DC-SIGN fusion protein could have potent antiviral activity. As a fusion protein, the binding of sDC-SIGN to Env may not only enhance the engagement of sCD4 to gp120, but also block the DC-SIGN binding sites on gp120 to prevent HIV-1 transmission. In the current study, we designed, expressed, purified and characterized a series of soluble CD4-linker-DC-SIGN (CLD) fusion proteins. We assessed the protein oligomeric state and gp120 binding affinity of CLDs, and tested their anti-HIV activity against several laboratory-adapted and primary isolates in cis-infection of target cells. We further examined the anti-HIV potency in trans-infection of target cells using DC-SIGN-expressing cell line and primary dendritic cells. The capability in inhibiting HIV-1

infection and dissemination was also evaluated in a human cervical explant model. Our findings demonstrate that several CLDs had significantly enhanced avidity to gp120 and much improved anti-HIV activity and could potently prevent both localized and disseminated infections of HIV-1. Our study proves a concept that targeting both CD4 and DC-SIGN binding sites on gp120 represents a novel antiviral strategy, and may have implications for the development of CD4/DC-SIGN based therapeutic and prophylactic antiretrovirals.

MATERIALS AND METHODS

Plasmids, cell lines, viruses and proteins.

env clones BaL, MWS2 and CH811 in pcDNA3.1, and viruses HIV-1_{BaL} and HIV-1_{RF} were described previously (29, 31, 32). U87-CD4.CCR5 cell line and pNL4-3.Luc.R^-E^- were obtained from NIH AIDS Research and Reference Reagent Program, Division of AIDS, NIH. 293T cell line was purchased from the American Type Culture Collection. Vector pET28a(+) was from Novagen. Raji/DC-SIGN cell line, and anti-DC-SIGN antibodies mAb507 and mAb 526 were described previously (28, 34). Anti-CD4 mAb RPA-T4 was from BD Biosciences. Mannan was from Sigma-Aldrich. Protein CN54 gp140 was described previously (32).

Design and genetic engineering of expression constructs

The CD4 and DC-SIGN DNA sequences were amplified from human CD4 gene in pcDNA3.1 (31) and human DC-SIGN gene in pcDNA3.1 (39), respectively. PCR primers used in this study were listed in Table S1. Amplified DNA fragments were subsequently cloned into pET28a(+) after corresponding enzyme digestion. All recombinant DNA clones were confirmed by sequencing.

Protein expression and purification

E.coli strain Rosetta (Novagen) was used for protein expression as described previously (39). After induction with isopropyl-β-D-thiogalactoside for 4 h, the bacteria were collected and lysed by ultrasonic treatment. The insoluble fraction in the lysates was washed twice with PBS supplemented with 1M GuHCl, and resuspended in denaturation buffer (10 mM Tris, 500mM NaCl, 6M GuHCl, 5 mM DTT, pH8.0). The denatured recombinant proteins were refolded for 24 h at 4°C in refolding buffer

(10 mM Tris, 500 mM NaCl, 3 mM $CaCl_2$, 10% glycerol, 3mM GSH, 0.3mM GSSG, pH8.0)(14, 53). After refolding, the solutions were loaded onto a pre-equilibrated nickel-charged chelating Sepharose Fast Flow column (GE healthcare). Proteins were purified according to the manufacturer's instructions.

SDS-PAGE and western blotting

Purified recombinant CLDs were resolved by 12% SDS-PAGE and transferred onto a PVDF membrane (Millipore) using the Microarray System (Bio-Rad). After being blocked in PBS plus 4% non-fat milk at 4°C overnight, the membrane was washed with TBS-T and subsequently incubated with anti-DC-SIGN mAb for 2 h at room temperature. After washes with TBS-T, the membrane was incubated with HRP-conjugated goat anti-mouse antibody for 2 h at room temperature. Luminometric detection of target proteins was achieved with SuperSignal West Dura Chemiluminescent Substrate (Pierce, Thermo Scientific) and visualized by a CCD camera imager (Alpha Innotech).

Analytical ultracentrifugation

Analytic ultracentrifugation was performed at 20°C using a Beckman Optima XL-I analytical ultracentrifuge equipped with an An-60Ti rotor. Purified recombinant proteins in dialysis buffer (10 mM Tris–HCl, pH 7.4, 200mM NaCl, 5 mM $CaCl_2$ and 0.1mM Glutathione) were loaded into 12mm path-length cells and centrifuged at 50,000 rpm for sCD4 and sDC-SIGN, and 40,000rpm for C35NDs60cc. A total of 200 absorbance scans (280nm) were recorded and data were analyzed with a C(s) distribution of the Lamm equation solutions calculated by the SEDFIT program (www.analyticalultracentrifugation.com/download.htm) (16, 47). Protein partial

specific volume values, solution density and solution viscosity were calculated with SEDNTERP (http://www.jphilo.mailway.com/download.htm).

Binding kinetics analysis

The buffer of tested protein was exchanged into phosphate buffered saline (pH7.4) with a desalt spin columns (Thermo Scientific). CN54 gp140 was biotinylated by mixing with Sulfo-NHS-LC-Biotinylation reagents in PBS for 30 min at room temperature according to the manufacturer's instructions (Pierce). The interactions between gp140 and CLDs were measured on a Forte-Bio Octet Red System (ForteBio) (1). This system monitors interference of light reflected from the surface of sensor to measure the thickness of molecules bound to the sensor surface. The biotinylized gp140 (5 μg/ml) was immobilized on streptavidin biosensors. After reaching baseline, sensors were dipped into different concentrations of tested proteins for association, and then moved into running buffer (3mM $CaCl_2$, 0.1% BSA, and 0.05% Tween20 in PBS, pH7.4) for dissociation. A buffer-only reference was subtracted from all curves. Octet Molecular Interaction System software was used for data analysis.

Binding in the presence of antibodies against CD4 or/and DC-SIGN was also performed. C35NDs60c was immobilized on anti-his biosensor. In the first association phase, sensors were incubated in running buffer (20mM Tris, 150mM NaCl, 3mM $CaCl_2$, 0.1% BSA, and 0.05% Tween20, pH8.0) supplemented with mAbs against CD4 (20μg/ml RPA-T4) or/and DC-SIGN (507 + 526; 20μg/ml each) as association buffer for 1800 seconds. Association was keep performing in the presence or absence of 5 μg/ml CN54 gp140 for another 1800 seconds. Dissociation was subsequently performed in running buffer for 1800 seconds.

Measurement of cytotoxicity

Cytotoxicity of fusion proteins was determined by MTT assay as described previously (30). TZM-bl cells were seeded in 96-well plates overnight and subsequently cultured in medium containing serially diluted proteins for 3 d. Medium was then removed and 100 µl of MTT solution (medium containing 0.5mg/ml 3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was added to each well and incubated for 4 h at 37°C. After removal of MTT solution, 100 µl of acid-isopropanol (0.04N HCl in isopropanol) was added and the optical density was read with a Modulus Microplate Luminometer (Turner BioSystems).

Infection

Stocks of pseudotyped reporter viruses were prepared by co-transfecting 293T cells with Env expression constructs and plasmid pNL4-3.Luc.R^-E^- as described previously (32). U87-CD4.CCR5 or TZM-b1 cells (1×10^4 cells/ well) were seeded in 96-well plates 1 d prior to infection. Env-pseudotyped viruses, HIV-1_{BaL} or HIV-1_{RF} were incubated with serially diluted CLDs for 1 h at 37°C and then the mixtures containing viruses and the proteins were added to the pre-seeded cells. The luciferase activity of cell lysate was determined 48 h post-infection with a Modulus Microplate Luminometer (Turner BioSystems). IC50 and IC90 values were calculated using Prism 4.1 (GraphPad).

Virus capture and transfer assay

HIV-1 was preincubated with CLDs at 37°C for 1 h in round bottom 96-well plates, while Raji/DC-SIGN cells (1.5×10^5 cells/ well) were pretreated with 1 mg/ml mannan.

Following incubation, Raji/DC-SIGN cells were added into CLD-pretreated HIV-1 while the same amount of HIV-1 was added to mannan-pretreated Raji/DC-SIGN cells. Following incubation at 37°C for another 2 h, unbound viruses were extensively washed with PBS. For HIV-1 capture assay, washed Raji/DC-SIGN cells were lysed directly and p24 was measured as described previously (27, 28). For virus transfer assay, washed Raji/DC-SIGN cells were then co-cultured with U87-CD4.CCR5 cells. 48 h later, cells were lysed and luciferase activity was determined.

Human monocyte-derived DCs (MDDCs) were generated from a highly enriched population of $CD14^+$ monocytes. Briefly, PBMCs were isolated using a Ficoll-Hypaque density gradient followed by negative selection using the Monocyte Isolation Kit according to the manufacturer's protocol (Miltenyi Biotec). To obtain immature MDDCs (iMDDCs), monocytes were cultured in the presence of IL-4 (500 U/ml; R&D Systems) and GM-CSF (800 U/ml; R&D Systems) for 7 d (3, 28). iMMDCs were used for HIV-1 capture and transfer assay as described above.

Culture and infection of human cervical tissues

Cervical tissues were obtained from women undergoing planned therapeutic hysterectomy in the absence of any cervical pathology at Hubei Hospital of Traditional Chinese Medicine or St George's Hospital London, with written consent obtained from all tissue donors according to the Local Research Ethics Committee. In brief, 3mm× 3mm explants were cultured in 200 µl of supplemented RPMI 1640 (28). Explants were pre-incubated in the presence or absence of C15D or antibody against CD4 or DC-SIGN for 1 h at 37°C. In dose-response experiments, explants were pre-incubated in the presence of serially diluted C35NDs60c, sCD4 or sDC-SIGN. After a

2-h exposure to HIV-1_{BaL} at 37°C, cervical explants were extensively washed to remove unbound viruses and inhibitors, and then cultured for 9 d at 37°C in fresh plates. For migratory cell experiments, after exposure to viruses, explants were cultured in the presence of 100 ng/ml recombinant human MIP-3β (R&D Systems) for 24 h at 37°C. Cells emigrating out of the explants were washed and co-cultured with PM1 cells (0.5×10^5 cells/well in 96-well plate) at 37°C. In all cases, supernatants were collected and stored at -80°C before subsequent measurement of p24 by ELISA (28).

RESULTS

Design and construction of plasmids expressing recombinant CD4-linker-DC-SIGN (CLD) fusion proteins

The CD4 binding site lies on gp120 neutralizing domain, while DC-SIGN recognizes mannose type glycans distributed around the outer domain (22, 45). Due to the distance between CD4 and DC-SIGN binding sites on gp120, an optimal linker is required to ensure that both components of the fusion protein can access their binding sites. Based on that Gly4Ser repeat has been broadly used in the construction of fusion proteins, in the current study, 3, 4, 5 and 7 Gly4Ser repeat linkers, and linkers of 5 and 7 Gly4Ser repeats integrated with DC-SIGN neck domain, were designed (Table S1 and S2). Since CD4 D1D2 domains affect the binding activity to gp120, while gp120-binding residues are all located within the first N-terminal domain (D1) of CD4 (13), we designed CD4-linker-DC-SIGN fusion proteins containing CD4 D1D2 domains, or the first 87aa or 106aa of CD4 D1 domain. In addition, modifications were made on C35D construct by truncating CD4 moiety (mC35D and sC35D contained the first 108aa and 87aa of CD4 D1, respectively) or adding DC-SIGN neck domain (C25ND and C35ND). A Ser to Cys mutation at aa 60, which can form a disulfide bond with gp120 (8), was introduced into C35ND, designated as C35NDs60c (Fig. 1a).

We generated nine pET28a constructs encoding the following recombinant fusion proteins: C15D, C20D, C25D, C35D, mC35D, sC35D, C25ND, C35ND and C35NDs60c. Two pET28a plasmids expressing sCD4 and sDC-SIGN, respectively, were also constructed (Table S1 and Table S2). The schematic diagrams of the constructs are illustrated in Fig. 1a and 1b.

Expression, refolding, purification and characterization of CLDs

The fusion proteins produced in *E.coli* Rosetta existed mainly as inclusion bodies. Modulating factors which impact protein expression, such as host bacterial strain, IPTG concentration and inducing temperature, had little effect on the solubility of CLDs. In order to obtain soluble proteins, protein refolding was performed by slowly adding the denatured recombinant proteins into the refolding buffer. Final concentration was kept at 30-50 μg/ml to avoid protein aggregation and precipitation. All recombinant proteins in this study contained his-tag at N terminus. The refolded proteins were purified with Ni-Resin column and target proteins were eluted with buffer containing 300 mM imidazole. Imidazole-free proteins were obtained by dialysis twice at 4°C.

An example of the purified fusion proteins, C35NDs60c, was analyzed by SDS-PAGE and western blotting (Fig. 1c). Reducing SDS-PAGE (Fig. 1c, lane 2) and western blot analysis revealed that C35NDs60c was correctly purified, with an expected band around 60 KD. C35NDs60c migrated mainly as a single band in both non-reducing and reducing SDS-PAGE (Fig.1c, lane 2, 3). CD4, a moiety of the fusion protein, has three oxidized isoforms, representing reduced protein (R) and the predominant disulfide-bonded CD4 isoforms (O^1/O^2) (8), while only O^1 is in functional state. C35NDs60c electrophoresed slightly faster in non-reducing SDS-PAGE than in reducing SDS-PAGE, indicating that it was in an O^1 state containing correct disulfide bonds.

Oligomeric state of recombinant CLDs

The function of recombinant fusion proteins can be impacted by their oligomeric state.

Analytic ultracentrifugation (AUC) was conducted to evaluate the molecular mass of the proteins. The distributions C(s) of protein sedimentation coefficients are shown in Fig. 1d. The predicted monomer molecular masses of sDC-SIGN, sCD4 and C35NDs60c were 18.4 KD, 22.1 KD and 60.5 KD, respectively. sDC-SIGN had a main peak at 2.0s, corresponding to apparent molecular mass of 22.4 KD. sCD4 showed a major peak around 1.9S, corresponding to apparent molecular mass of 21.5 KD. sCD4 also had a minor peak with molecular mass of 57.4 KD, but reducing SDS-PAGE confirmed that sCD4 had no band around 57.4 KD (data not shown). The minor peak in the C(s) distribution of sCD4 might be dimers caused by incorrectly formed disulfide bonds. C35NDs60c demonstrated a major peak at 6.3S, corresponding to apparent molecular mass of 214.8 KD and a minor peak 9.2S, corresponding to apparent molecular mass of 398.2KD. In non-reducing SDS-PAGE, C35NDs60c showed mainly as monomers (60.5 KD) (Fig. 1b, lane 3), although oligomers were detected, some of which likely formed in non-reducing conditions (8). The 214.8KD complex in AUC indicated that C35NDs60c existed mainly as tetramers. This tetramer formation might be contributed by the neck domain of DC-SIGN (49). C35NDs60c could also form higher oligomer during refolding procedure caused by incorrectly formed disulfide bonds, which explained the existence of the minor peak. Taken together, the AUC results showed that D1D2 domain of CD4 and carbohydrate-recognition domain of DC-SIGN existed mainly in monomer form while the majority of C35NDs60c were tetramers.

CLDs demonstrate enhanced binding affinity to gp120

We performed direct binding analysis on a Forte-Bio Octet Red System. Streptavidin biosensor immobilized with biotinylated CN54 gp140 was immersed in different

316 concentrations of CLDs (all protein molar concentrations were calculated based on
317 monomer). To characterize protein interactions, two protein binding models: langmuir
318 1:1 model and bivalent model were applied (Fig. S1). χ^2 was the sum of squared
319 deviations, where deviation was the difference between the actual data-point and the
320 fitted curve. Values close to zero indicated a good curve fit (5). As calculated χ^2
321 shown in Table 1, CLDs fitted preferably to the bivalent binding model, whereas the
322 control proteins sCD4 and sDC-SIGN fitted with the langmuir binding model. These
323 results demonstrated that, unlike sCD4 and sDC-SIGN, CLDs interacted with gp140
324 in a bivalent manner, indicating that both CD4 and DC-SIGN moieties of CLDs could
325 effectively reach their respective binding sites on gp140, either simultaneously or
326 sequentially. Moreover, as shown in Table 1, the affinity of CLDs with a 35aa linker
327 to gp140 were obviously higher than that of sCD4 (K_D= 1.21E-09 M) and sDC-SIGN
328 (K_D=1.43E-08 M). The difference of affinity between CLDs with 35aa linker and
329 C15D implied that the length of linkers impacted the interactions between CLDs and
330 gp120. Linkers in appropriate length benefitted the binding of CLD moieties to gp120,
331 while short linkers hindered such interactions. Linker with 35aa seemed long enough
332 to allow effective bindings of the fusion proteins to gp140. Other modifications on
333 CLDs, such as adding DC-SIGN neck domain or/and introducing Cys to Ser mutation
334 into CD4, also enhanced the affinity of CLDs to gp140. Compared with the off rate of
335 CD4 (K_{dis} = 6.25E-05 S^{-1}), CLDs with 35aa linker dissociated more slowly, indicating
336 the formation of more stable complexes with gp140.
337
338 When CD4 or DC-SIGN domain of C35NDs60c was blocked by antibodies, the
339 binding of gp140 to C35NDs60c-immobilized biosensor decreased, and a
340 combination of anti-CD4 and anti-DC-SIGN antibodies had an additive effect in

inhibiting C35NDs60c-gp140 interaction in the association phase (Fig. S2). In the dissociation phase, gp140-C35NDs60c complex appeared to dissociate more slowly than the antibody-C35NDs60c complex. The difference at the end of dissociation phase might be due to a competitive binding of gp140 to C35NDs60c by replacing anti-CD4 or/and anti-DC-SIGN antibodies (Fig. S2). The binding results together indicate that gp140 can bind to both CD4 and DC-SIGN domains of C35NDs60c, and that the avidity between C35NDs60c and gp140 is likely higher than that between C35NDs60c and the antibodies used in this study.

CLDs inhibit HIV-1 infection in cis

Because the binding of gp120 to CD4 is virtually universal among HIV-1 isolates, regardless of viral tropism, we expected that CLDs would be active against both R5 and X4 strains. We initially tested the antiviral potency of C15D against HIV-1_{BaL} (R5) and HIV-1_{RF} (X4) in TZM-bl cells. The IC50s were 60.5 nM and 77.8 nM, respectively (Table 2.), indicating that CLDs can neutralize HIV-1 regardless of viral tropism. Subsequent experiments were carried out by using BaL Env-pseudotyped HIV-1 to infect U87-CD4.CCR5 cells. All CLDs demonstrated anti-HIV activity (Table 2). Compared with sCD4 (IC50=25.3 nM), C35D showed enhanced neutralization activity (IC50=15.7 nM), while the CLDs with shorter linkers (C15D, C20D and C25D) had similar or lower antiviral activity. It seemed that linker length was a crucial factor affecting the anti-HIV activity of the CLDs, probably by interfering with CLD-gp120 interactions. The IC90 of the CLDs had similar trend as that of IC50.

CD4-truncated CLDs showed decreased neutralization activity. The IC50s of mC35D

and sC35D were over 1000nM, at least 40 fold higher than that of C35D. DC-SIGN neck domain has seven complete and one incomplete 23-aa long repeats, which forms helical stretches to aggregate as tetramers. Adding the neck domain may also lengthen the linker. The IC50s of C25ND and C35ND were enhanced to 4.9 nM and 5.3 nM, respectively. The similar IC50s of the two fusion proteins imply that a 25aa linker length is sufficient and the extra l0aa had little effect on the neutralization activity. The IC50 of C35NDs60c (IC50=3.3 nM) was 1.6 fold lower than that of C35ND and 7.7 fold lower than that of sCD4.

As C25ND, C35ND and C35NDs60c exhibited better anti-HIV activity compared to CLDs with linkers lacking DC-SIGN neck domain, neutralization potency against primary HIV-1 Envs was further investigated. Two primary HIV-1 Env clones were used, including MWS2, a clade C Env cloned from semen of a subject known to have infected women by vaginal intercourse, and CH811, a clade B Env isolated from a Chinese patient's blood sample. As shown in Table 2, C25ND and C35ND exhibited much weaker neutralization activity against MWS2 and CH811 than that of HIV-1_{BaL}. Remarkably, C35NDs60c demonstrated much stronger bioactivity than C25ND and C35ND, possessing potent neutralization activity against the two primary HIV-1 Envs, with an IC50 of 13.0 nM and 4.7 nM, respectively. None of the CLDs was cytotoxic to the tested cells (Table 2).

Previous studies by others reported that sCD4 or sDC-SIGN at suboptimal concentrations could enhance HIV-1 infection (25,48,54). We tested whether CLD had a similar effect. Our results indicated that C35NDs60c did not enhance HIV-1 infection at least in the tested concentration range (Fig. S3a).

CLDs inhibit HIV-1 capture and transfer via DC-SING-expressing cells and iMDDCs

Raji/DC-SIGN cells were used to assess the anti-HIV capability of CLDs against virus capture and trans-infection. As shown in Fig. 2a and 2b, sCD4 exhibited little effect in inhibiting HIV-1 capture by DC-SIGN but rendered the bound virus noninfectious to U87-CD4.CCR5 cells. Interestingly, all CLDs tested at the same concentration (1000 nM) demonstrated similar trends to sDC-SIGN in interfering with virus capture by Raji/DC-SIGN cells, as well as similar capacity as sCD4 in inhibiting virus trans-infection from Raji/DC-SIGN cells to U87-CD4.CCR5 cells.

To confirm the anti-HIV activity of CLDs in trans-infection, similar experiments were conducted in iMMDCs. Similar to controls sDC-SIGN and mannan, all CLDs demonstrated less potency against virus capture than that observed in Raji/DC-SIGN cells (Fig. 2c). Differences between the DC-SIGN-expressing cell line and iMMDCs were likely due to differences in receptor repertoire of iMMDCs which also express other attachment receptors in addition to DC-SIGN. Nevertheless, CLDs still possessed better efficacy than sCD4 and sDC-SIGN to suppress virus uptake by iMMDCs (Fig. 2c). Among those, C35NDs60c inhibited more than 50% of HIV-1 capture. Despite the incomplete inhibition of HIV-1 capture, all CLDs showed enhanced potency against trans-infection, almost completely blocking virus transfer from iMMDCs to U87-CD4.CCR5 cells (Fig. 2d). These results imply that HIV-1 may binds to iMDDCs via additional attachment receptors other than DC-SIGN, but CLDs can render the bound virus noninfectious.

Considering that saturated concentrations of proteins were used in the above study, we performed additional dose-response assays. In the neutralization assay, C35NDs60c was much more potent than a combination of sCD4 and sDC-SIGN (Fig. S3a). In viral capture and transfer assays, C35NDs60c also demonstrated better antiviral activities than sCD4 and sDC-SIGN in combination (Fig. S3b and S3c).

CLDs inhibit both localized mucosal infection and dissemination pathways

In the absence of a suitable animal model for HIV-1, ex vivo culture of human tissue explants has been generally accepted as an alternative to mimic in vivo physiological conditions. We conducted experiments to determine whether CLDs could inhibit HIV-1_{BaL} infection and dissemination in cervical tissues. As a conceptual study, we initially used C15D to conduct our experiment. Concentrations of all proteins were used at 30 µg/ml(~750 nM for C15D). As seen in Fig. 3a and 3b, anti-CD4 antibody blocked localized infection, but was incapable of inhibiting virus dissemination in mucosal tissues. Although antibodies against DC-SIGN might decrease the capture of virus by migrating cells within mucosa, receptors other than DC-SIGN could play a role in transmission. A combination of anti-CD4 and anti-DC-SIGN antibodies was required to block HIV-1_{BaL} infection and dissemination in cervical tissues. Of note, C15D not only efficiently inhibited localized infection but also prevented disseminated infection by migratory cells, with better efficacy than the combination of anti-CD4 and anti-DC-SIGN antibodies. In addition, we performed dose-response experiments using sCD4, sDC-SIGN and an improved CLD (C35NDs60c) against HIV-1_{BaL} infection in human cervical explants. While sDC-SIGN had little inhibitory effect against HIV-1 infection in the tested concentration range, C35NDs60c demonstrated significantly enhanced antiviral activity, with an IC50 one log lower than that of sCD4 (Fig. S4).

441 Our data together suggest that further improvement of CLDs may render their
442 potential to be used as prophylaxis or therapeutics, in particular for microbicide
443 development.
444

DISCUSSION

Effective microbicides need to protect against all potential routes of HIV-1 transmission across mucosal surfaces. Based our previous findings that HIV-1 uptake and dissemination by migratory dendritic cells (DCs) can occur through CD4 and mannose binding C-type lectin receptor DC-SIGN (28), in the current study, we designed, expressed, purified and characterized a series of bi-functional CD4-linker-DC-SIGN fusion proteins (CLDs). We demonstrate that several of the CLDs had enhanced gp120 binding affinity and much improved antiviral activity against HIV-1 infection and dissemination.

Soluble CD4 (sCD4) has poor antiviral activity against primary HIV-1 isolates (33). In order to improve the anti-HIV potency, several strategies have been employed to date to make fusion proteins, for instance, protein polymerization. Immunoglobulin G (IgG) is usually used as a frame protein for making fused oligomers. PRO-542, a tetramer CD4 wherein the Fv portions of both the heavy and light chains of human IgG2 have been replaced with the D1D2 domains of human CD4, binds to gp120 with high affinity (2). However, the increased size by fusing to antibody may diminish the neutralization potency of candidate molecule (10). The large size likely also makes IgG-anchored fusion protein less able to penetrate into tissues (35). When we designed and characterized a range of CLDs, we observed that the integration of DC-SIGN neck domain into fusion proteins greatly increased tetramer formation, and significantly enhanced the antiviral potency against both laboratory-adapted and primary isolates. This novel polymer strategy could be extended to design other fusion proteins aiming to enhance bioactivity.

Design of sCD4-based fusion proteins simultaneously targeting different binding sites on gp120 provides another alternative to improve anti-HIV potency. Most fusion proteins designed to date, including sCD4-17b, m35-sCD4 and $CD4_{HC}$-(GS7)-IgGE51, have been focusing on the CD4 binding site and CD4-binding-induced epitopes on gp120 (9, 36, 56). Different to the molecules described above, CLDs, binding to CD4-binding site and glycans on gp120, were designed to interrupt the interactions between gp120 and the entry receptor CD4 as well as the attachment receptor DC-SIGN. Such unique binding mode not only enhanced the engagement of sCD4 to gp120, but also inhibited the DC-SIGN binding sites on gp120. Although CD4 and DC-SIGN have little direct interaction, DC-SIGN binding to gp120 causes an allosterically induced exposure of the CD4 binding site and therefore facilitates a more stable binding of CD4 to gp120. Indeed, CLDs not only enhanced the neutralization activity, but also inhibited cis- and trans-infection of HIV-1 in both cellular and human cervical explant models. Given that there is limited research on sDC-SIGN-based antiretrovirals and that sDC-SIGN alone is unlikely to be used in practice, our study provides evidence that DC-SIGN fused to other proteins targeting gp120 may provide an important new strategy for enhancing anti-HIV potency. As DC-SIGN functions as an attachment receptor for a range of enveloped viruses such as Dengue, HCV, Ebola and so on (17, 26), the method of fusing DC-SIGN with other functional moieties may be extended to prevent or treat these additional types of viruses.

Several CLDs, including C35D, C25ND, C35ND and C35NDs60c, demonstrated much improved neutralizing activity against HIV-1. In contrast, C15D, C20D and C25D had similar or decreased antiviral activity compared with that of sCD4. This implies that fusion proteins expressing two components binding to different sites on

gp120 per se does not guarantee enhanced bioactivity. The amino acids involved in CD4 recognition distribute discontinuously around C2, C3 and V5 regions of gp120 (4, 24, 45), whereas DC-SIGN interacts with discontinuously distributed high-mannose oligosaccharides on gp120 (20). Short linkers likely hindered the two moieties of the fusion proteins from simultaneously interacting with gp120, while a linker with suitable length could render CLDs higher gp120-binding avidity and better anti-HIV potency. To this end we did not observe significant difference in terms of anti-HIV activity when C25ND and C35ND were tested against both laboratory-adapted and primary isolates, including one cloned from semen of a subject known to have infected women by vaginal intercourse, suggesting that, a linker of 25 amino acids integrated with an extra DC-SIGN neck domain, may be sufficient for simultaneous binding of the two moieties to gp120, at least for the isolates tested.

The signaling pathway activated by binding of HIV-1 to DC-SIGN is thought to cause immunosuppressive responses and triggers HIV-1 transmission and replication (18). In addition to masking the CD4-binding site on gp120, CLDs interrupted gp120 glycan-DC-SIGN interaction and inhibited the uptake of virus by host cells, avoiding potential triggering of the downstream signaling pathways. Several carbohydrate-binding agents, including cyanovirin-N and griffithsin, also target gp120 glycans and have potent antiviral activity (6). However, those proteins were originated from bacteria or plants. Whether they can be used prophylactically or therapeutically requires further evaluation. Given that the soluble forms of CD4 and DC-SIGN occur normally in vivo and that the Gly4Ser repeat liker is poorly immunogenic (43, 44), further improvement of CLDs may render their potential to be used as prophylaxis or therapeutics.

Beyond a novel antiretroviral proof-of-concept, CLDs designed in the current study may have additional potential. CD4 engagement to gp120 induces the exposures of immunogenic epitopes, including V3 and chemokine receptor binding sites. Immunization with CD4-gp120 complex has been shown to enhance viremia control in nonhuman primates (12, 15). In those studies, immunogens were crosslinked CD4-gp120 complexes or recombinant fusion proteins of CD4 and gp120, in which the native epitopes on gp120 might be affected. In contrast, CLDs, especially C35NDs60c, have a lower off rate than sCD4, facilitating the formation of a more stable CD4-gp120 complex, probably induced by DC-SIGN engagement. Given that DC-SIGN binding to HIV-1 gp120 increases exposure of the CD4 binding site (25), it will be interesting to determine in future studies whether CLD-bound gp120/gp140 complexes can be used as immunogenic components to elicit better neutralizing antibodies.

Despite the described favorable characteristics of CLDs, further improvements in antiviral activity are needed. As both CD4 and DC-SIGN components used to make CLDs do not contain glycans, we used a bacterial expression system to produce proteins. Due to the poor solubility, the target proteins were expressed mainly in the form of inclusion bodies. Even though purified target proteins showed high anti-HIV activity after refolding, a certain degree of incorrect formation of disulfide bonds or/and aggregation occurred, and this could affect their bioactivity. While careful refolding technology could decrease such wrong disulfide bonds and aggregation, this process would be difficult to scale up. Other expression systems, such as transgenic plants (50), may provide an alternative to produce proteins in a larger scale.

545

546 In conclusion, this is the first time that sDC-SIGN-based bifunctional proteins have
547 demonstrated anti-HIV potency. The designed and expressed CLDs are novel
548 bifunctional proteins with increased gp120 binding avidity. CLDs inhibit HIV-1
549 infection and dissemination in cell lines, primary dendritic cells and mucosal cervical
550 tissues.

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744 **Table 1. Kinetic parameters of gp140-CLD binding**

	C35NDs60c	C35ND	C35D	C15D	sCD4[a]	sDC-SIGN[b]
K_{D1} (M)	3.20E-10	1.66E-09	1.04E-09	2.62E-08	1.21E-09	1.43E-08
K_{D2} (M)	1.17E-09	2.11E-10	5.96E-10	1.95E-09		
K_{a1} (1/Ms)	9.05E+04	1.41E+04	1.41E+04	1.09E+04	5.16E+04	1.57E+04
K_{a2} (1/Ms)	1.12E+04	8.28E+04	6.99E+04	1.10E+04		
K_{dis1} (1/s)	2.90E-05	2.34E-05	1.46E-05	2.86E-04	6.25E-05	2.24E-04
K_{dis2} (1/s)	1.31E-05	1.75E-05	4.17E-05	2.14E-05		
$\chi^2_{langmuir}$	12.52	5.39	5.51	4.08	0.49	0.05
$\chi^2_{bivalent}$	0.38	0.10	0.59	0.91	0.90	0.52

745

746 K_D: Equilibrium (affinity) constant

747 K_a: Association rate constant

748 K_d: Dissociation rate constant

749 a: the N terminal 183 aa of CD4

750 b: the carbohydrate-recognition domain of DC-SIGN

751 Data are representative of three independent experiments.

752 **Table 2.** Anti-HIV activity of variant CLD forms

		sCD4[e]	C15D	C20D	C25D	C35D	C25ND	C35ND	C35NDs60c
HIV-1_{BaL}[a]	IC90[c]	148.1 ± 10.4	335.7 ± 53.2	ND	ND	ND	ND	ND	ND
	IC50[c]	27.1 ± 3.6	60.5 ± 12.9	ND	ND	ND	ND	ND	ND
HIV-1_{RF}[a]	IC90[c]	199.8 ± 7.2	384.2 ± 44.6	ND	ND	ND	ND	ND	ND
	IC50[c]	34.5 ± 2.2	77.8 ± 15.4	ND	ND	ND	ND	ND	ND
BaL[b]	IC90[c]	153.9 ± 4.3	355.2 ± 30.4	308.2 ± 30.7	126.6 ± 25.3	**79.6 ± 25.1 (1.9 ↓)**	**30.7 ± 3.6 (5.0 ↓)**	**26.7 ± 2.0 (5.8 ↓)**	**28.1 ± 1.3 (5.5 ↓)**
	IC50[c]	25.3 ± 1.8	66.9 ± 5.2	55.9 ± 7.3	26.4 ± 4.9	**15.7 ± 4.7 (1.6 ↓)**	**4.9 ± 1.0 (5.2 ↓)**	**5.3 ± 0.5 (4.8 ↓)**	**3.3 ± 0.3 (7.7 ↓)**
MSW2[b]	IC90[c]	>1000[d]	ND	ND	ND	ND	977.1 ± 77.6	990.7 ± 85.2	**65.6 ± 13.5 (>15.2↓)**
	IC50[c]	>1000[d]	ND	ND	ND	ND	**578.6 ± 53.1 (>1.7 ↓)**	**667.3 ± 98.3 (>1.5 ↓)**	**13.0 ± 3.7 (>76.9 ↓)**
CH811[b]	IC90[c]	>1000[d]	ND	ND	ND	ND	**423.5 ± 47.6 (>2.4 ↓)**	**354.7 ± 60.4 (>2.8 ↓)**	**36.0 ± 3.5 (>27.8 ↓)**
	IC50[c]	277.8 ± 69.1	ND	ND	ND	ND	**56.7 ± 12.9 (4.9 ↓)**	**42.8 ± 17.6 (6.5 ↓)**	**4.7 ± 0.6 (59.1 ↓)**
CC50[c,d]		>1600	>1600	>1600	>1600	>1600	>1600	>1600	>1600

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754 Bold data imply that CLDs had significantly increased anti-HIV-1 activity

755 IC50: 50% inhibitory concentration; IC90: 90% inhibitory concentration; CC50: 50% cytotoxicity concentration. All protein molar concentrations were calculated

756 based on monomer.

757 a; Replication competent HIV-1

b: Env-pseudotyped HIV-1

c: nM

d: highest concentration tested

e: the N terminal 183 aa of CD4

ND: not determined

Data are mean ± SD of at least three independent experiments (↓ fold decrease), with each condition performed in triplicate.

764 **Fig. 1. Schematic diagrams of CLD-expressing plasmids and biochemical**
765 **characterization of purified fusion proteins**
766 (a) Schematic representation of fusion proteins. Abbrevations: sCD4, the N terminal
767 183 aa of CD4; mC, the N terminal 106 aa of CD4; sC, the N terminal 87 aa of CD4;
768 N, DC-SIGN neck domain; sDC-SIGN, DC-SIGN carbohydrate-recognition domain
769 (CRD); 15, 20, 25, 35, the number of amino acids; s60c, a Cys to Ser mutation at aa
770 60 on CD4. (b) Schematics of CLD-expressing plasmids. All CD4 and DC-SIGN
771 moieties for CLDs were cloned into pET-28a(+) using restriction enzyme sites Nde I,
772 BamH I and Hind III. Restriction enzyme sites and linker sequences were introduced
773 into CD4 and DC-SIGN sequences by PCR. (c) SDS-PAGE and western blot analysis
774 of C35NDs60c. Purified C35NDs60c was resolved in 12% SDS-PAGE in reducing or
775 non-reducing condition, followed by detection with Coomassie Blue staining or
776 western blotting (reducing condition only). Lane1: western blotting analysis.
777 C35NDs60c was detected by mAb 507 against DC-SIGN, followed by an HRP-
778 conjugated secondery antibody; lane2 and lane3: Coomassie Blue staining of
779 C35NDs60c in reducing and non-reducing conditions, respectively; lane4: molecular
780 marker in reducing condition. (d) Analytical ultracentrifugation in sedimentation
781 velocity mode. CD4, peaks at 1.9S and 3.8S, corresponding to apparent moleuclar
782 mass 21.5KD and 57.4KD; DC-SIGN, peaks at 2.0S, corresponding to apparent
783 moleuclar mass 22.4KD; C35NDs60c, peak at 6.3S and 9.2S, corresponding to
784 apparent moleuclar mass 214.8KD and 398.2KD. One out of three independent
785 experiments is shown.
786
787 **Fig. 2. CLDs inhibit HIV-1 capture and transfer by Raji/DC-SIGN cells and**
788 **iMMDCs.** BaL Env-pseudotyed HIV-1 was pre-incubated with or without inhibitor
789 for 1 h at 37°C before the addition to Raji/DC-SIGN cells or iMMDCs. Cells pre-

790 treated with or without mannan were exposed to viruses as controls. After exposure
791 to virues for 2 h at 37°C, cells were extensively washed and either lysed for capture
792 assay or cocultured with U87-CD4.CCR5 cells for transfer assay. (a) HIV-1 captured
793 by Raji/DC-SIGN cells. Medium alone was defined as 100% and its p24
794 concentration was 1.79 ng/ml. (b) RLU of HIV-1 trans-infection from Raji/DC-SIGN
795 cells to U87-CD4.CCR5 cells. Medium alone was defined as 100%. (c) HIV-1
796 captured by iMDDCs. Medium alone was defined as 100% and its p24 concentration
797 was 2.87 ng/ml. (d) RLU of HIV-1 trans-infection from iMDDCs to U87-CD4.CCR5
798 cells. Medium alone was defined as 100%. Data shown are mean ± SD of three
799 independent experiments, with each condition performed in triplicate.
800
801 **Fig. 3. CLD inhibits localized mucosal HIV-1 infection and dissemination.** Human
802 cervical explants were pre-incubated with or withour inhibitors for 1 h at 37°C before
803 exposure to HIV-1_{BaL} for 2 h at 37°C. After incubation, explants were extensively
804 washed and cultured in the presence of 100 ng/ml of MIP-3β for 48 h. Emigrating
805 cells were collected, washed, and cocultured with PM1 cells. The explants were
806 cultured in separate wells. Data are representative of three independent experiments,
807 with each condition performed in triplicate. Data shown are p24 antigen (mean± SD)
808 released from both (a) cultured explants and (b) PM1 cocultured migratory cells at
809 day 9. p24 in the absence of inhibitor was defined as 100%, and was 1.56 and 2.74
810 ng/ml for the cervical explants and PM1 cocultured migratory cells, respectively.
811 Anti-CD4: RPA-T4. Anti-DC-SIGN: 507+ 526.