Ataxia-Pancytopenia Syndrome due to a de Novo SAMD9L Mutation

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Introduction

Ataxia pancytopenia syndrome (ATXPC; MIM 159550) is an autosomal dominant cerebellar ataxia associated with hematological abnormalities including pancytopenia and a predisposition to haematological malignancy (myelodysplasia and acute myeloid leukaemia). To date, 4 families have been described harboring gain of function *SAMD9L* mutations which underlie ATXPC.¹⁻⁴ Here we describe a de novo *SAMD9L* pathogenic variant in a patient with myelodysplasia, presenting with a subacute cerebellar syndrome.

Case Presentation

A 28-year-old woman presented to a tertiary neurologic center with a 6-week history of headache, diplopia, and ataxia, which started after cellulitis at the site of a pneumococcal and influenza vaccination. On examination, there was mild gait ataxia, slow saccades, diplopia in all directions of gaze without extraocular nerve palsy, and prominent downbeating nystagmus. Her medical history confirmed myelodysplastic syndrome, diagnosed 12 years previously. There was no relevant family history.

Initial investigations, including CSF and a broad panel of autoimmune and metabolic investigations, were unremarkable. MRI of the brain demonstrated confluent, symmetrical periventricular and deep white matter T2 hyperintensity, extending to involve the striatum, with periventricular cystic foci surrounding the lateral ventricles (figure 1A). There was marked cerebellar atrophy with both vermian and hemispheric volume loss (figure 1B). Whole body [¹⁸F]FDG-PET demonstrated diffuse cerebellar hypometabolism (figure 1C) and excluded active malignancy. Electroencephalography revealed occasional sharpened theta waves over the frontal region against a posterior background activity in the alpha range. Ophthalmology examination demonstrated no ocular abnormality with normal optical coherence tomography.

Whole genome sequencing of the proband and both parents revealed a heterozygous missense pathogenic variant of the *SAMD9L* gene (NM_152703.5:c.2956C>T, p.(Arg986Cys)) in the proband only and confirmed parentage. Sanger sequencing was carried out in the proband and both parents, confirming that the variant had arisen de novo in the patient (figure 1D). This variant is not seen in the gnomAD population database but was previously described by Tesi et al.¹ in a 3-generation family with ATXPC (F1).

Array comparative genomic hybridization (CGH) demonstrated a mosaic interstitial deletion on the long arm of chromosome 7 within the hematopoietic cell line, from bands q11.23 to q33

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sequencing of the proband and asymptomatic parents (\breve{D}).

(arr[GRCh37] 7q11.23q33 (76030573_137802149)x1[0.5], figure e-1, links.lww.com/NXG/A404). This deletion encompasses the *SAMD9L* gene and was present in 50% of cells.

Discussion

ATXPC is caused by gain of function pathogenic variants of the *SAMD9L* tumor suppressor gene. The disorder is extremely rare; of those 35 patients with *SAMD9L* mutations reported worldwide by the Human Gene Mutation Database, 6 have ATXPC, whereas the remainder are characterized by myelodysplastic syndrome or bone marrow failure.² The literature reports 4 families in addition to the case reported here.^{1,3,4}

Overactivity of *SAMD9L* results in the cerebellar atrophy and pancytopenia that engender the clinical presentation. Brain MRI demonstrates cerebellar volume loss accompanied frequently by periventricular white matter T2 hyperintensities.^{1,4,5} Histopathologic studies have shown diffuse loss of Purkinje

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cells and—to a lesser extent—cerebellar granule cells, the mechanism for which remains unclear.⁴

Germline *SAMD9L* gain of function pathogenic variants cause bone marrow suppression with pancytopenia. Acquired monosomy 7 or partial deletion of chromosome 7 may arise to circumvent the repression of hematopoiesis. These can act as a "rescue mechanism" that inactivates the gain of function mutation by nonrandom loss of the mutated *SAMD9L* allele (figure e-2, links.lww.com/NXG/A405).⁶ Consequently, spontaneous recovery from the pancytopenia may occur ("hemopoietic reversion"). However, partial or total deletions of chromosome 7 may lead to clonal expansion of a cell line (myelodysplastic syndrome), as occurred in our patient, and predispose to acute myeloid leukaemia.⁷

The age at onset of neurologic symptoms in familial cases varies widely, ranging from 4 to 62 years, followed by invariably slow progression.^{4,5} The neurologic presentation is often preceded by the hematological manifestations, as in the patient reported here.

Although the exact triggers for this patient's subacute presentation are not clear, possibilities include decompensation of the already compromised cerebellar system because of intercurrent infection or parainfectious phenomenon after recent immunization.

The mainstay of the management for *SAMD9L*-associated hematological abnormalities includes monitoring for the development of hematological malignancy and allogenic stem cell transplantation. Stem cell transplantation seems to have no effect on neurologic outcomes, and the management of neurologic abnormalities of ATXPC remains supportive.¹

With input from the wider multidisciplinary team, including neuro-ophthalmology, hematology, clinical genetics, and physiotherapy, the patient reported here continues to be independent and an avid cyclist. The patient initially found improvement in her diplopia with the use of occlusive lenses; however, these are no longer required, and the only remaining abnormality on examination is mild nystagmus.

Shortly after diagnosis, the patient became pregnant. Prenatal testing, by chorionic villus sampling at 12 weeks, confirmed the baby to be negative for the familial variant in *SAMD9L* (unaffected). The patient remained well throughout an uneventful pregnancy.

This case of de novo *SAMD9L* pathogenic variant in ATXPC highlights the value of whole genome sequencing, leading to an accurate diagnosis, appropriate surveillance and treatment, and opening the option of prenatal diagnosis.

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Appendix Authors

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Josh King- Robson, BMBS BSc (Hons) MRCP	Kingʻs College Hospital, London, United Kingdom	Drafted the manuscript and illustrative figure
Joseph Marshall, MBBS MSc	King's College Hospital, London, United Kingdom	Revision of the manuscript
Leanne Willoughby, BSc (Hons)	South East Genomic Laboratory Hub, Guy's & St Thomas' NHS Foundation Trust	Interpretation and reporting of array comparative genomic hybridization results and preparation of the figure
Frances Smith, BSc (Hons) DipRCPath	Viapath at Kings College Hospital, London, United Kingdom	Performed and interpreted Sanger sequencing and prepared sequencing images
Sahar Mansour, BMBS FRCP	Thames Regional Genetics Service, St George's University of London, London, United Kingdom	Interpretation of genetic results and revision of the manuscript
Laszlo Sztriha, MD PhD FRCP	King's College Hospital, London, United Kingdom	Performed initial clinical diagnosis, interpretation of data, and revision of the manuscript

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