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DNA target recognition domains in the Type I restriction and modification systems of *Staphylococcus aureus*.

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Abbreviations: Restriction and Modification, RM; Horizontal Gene Transfer, HGT; hsd, host specificity for DNA; Methyltransferase, MTase; N6-methyl adenine, m6A; N4-methyl cytosine, m4C; C5-methyl cytosine, 5mC; Clonal Complex, CC; Sequence Type, ST; Target Recognition Domain, TRD; Single Molecule Real Time, SMRT; Enhanced Green Fluorescent Protein, EGFP; Polymerase Chain Reaction, PCR; ATP hydrolysis, ATPase; Lysogeny Broth, LB; S-adenosyl-L-methionine, SAM.

Abstract

Staphylococcus aureus displays a clonal population structure in which horizontal gene transfer between different lineages is extremely rare. This is due, in part, to the presence of a Type I DNA restriction and modification (RM) system given the generic name of *Sau1*, which maintains different patterns of methylation on specific target sequences on the genomes of different lineages. We have determined the target sequences recognised by the *Sau1* Type I RM systems present in a wide range of the most prevalent *S. aureus* lineages and assigned the sequences recognised to particular target recognition domains within the RM enzymes. We used a range of biochemical assays on purified enzymes and single molecule real-time sequencing on genomic DNA to determine these target sequences and their patterns of methylation. Knowledge of the main target sequences for *Sau1* will facilitate the synthesis of new vectors for transformation of the most prevalent lineages of this "untransformable" bacterium.

44 Introduction

45 Type I DNA restriction-modification (RM) systems are found in about half of the sequenced
46 prokaryotic genomes (1-4). They present a formidable barrier to the invasion of the host cell by
47 foreign DNA whether by transduction, transformation or conjugation and thus exercise control over
48 horizontal gene transfer (HGT) (1,4-8). As an example of their effectiveness, less than 1 in 10^4 or 10^5
49 phage infections can successfully avoid the classical EcoKI Type I RM system of *Escherichia coli* K12.
50 In some circumstances, such as when antirestriction systems are absent (9), when there are multiple
51 target sites on the phage (10) or when RM expression is raised (11), the barrier due to this single RM
52 system can be even greater. RM systems operate by methylating defined target sequences on the
53 host genome and they maintain this methylation pattern through each round of DNA replication
54 (modification). Foreign DNA entering the cell often contains the same target sequence but in an
55 unmethylated state. These unmethylated target sequences are targeted for endonucleolytic
56 cleavage by the RM system (restriction). The Type I RM system comprises three *hsd* (host specificity
57 for DNA) genes, *hsdR*, *hsdM* and *hsdS* for restriction, modification and target sequence specificity
58 respectively. The gene products form an $R_2M_2S_1$ complex in which HsdS (or S) recognises the target
59 sequence, HsdM (or M) recognises the methylation status of the target and methylates
60 hemimethylated targets while HsdR (or R) cleaves the DNA containing unmethylated targets after a
61 complex reaction involving ATP hydrolysis and DNA translocation (12). An M_2S_1 complex can act
62 solely as a methyltransferase (MTase) (13). Type I RM enzymes almost always recognise and
63 methylate adenine nucleotides in their target sequences to form N6-methyl adenine (6mA) although
64 a few forming N4-methyl cytosine (m4C) are now known (3,14). In addition to the protection offered
65 by Type I, II and III RM systems, Type IV restriction systems can attack foreign DNA containing
66 methylated sequences not found in the host (15).

67
68 The presence of multiple RM systems in a single host can increase the barrier to HGT still further. For
69 instance, *Staphylococcus aureus* often contains two related Type I RM systems making its
70 transformation extremely inefficient and hindering the genetic analysis of this organism (16-19).
71 These genomes contain two *hsdM* and two *hsdS* and share a single *hsdR*, although some *S. aureus*
72 strains have different numbers of *hsdM* and *hsdS* (Figure 1a). The presence of only a single *hsdR* is
73 not a problem as it can interact with each *hsdMhsdS* pair. It has long been known that *S. aureus*
74 displays a clonal population structure (20) in which HGT between different clonal complexes is
75 exceedingly rare. Multi-locus sequence typing, microarray analysis and whole genome sequencing
76 divides lineages of *S. aureus* and close relatives into the clonal complexes (CC) (20-23), each of which
77 carries a different range of mobile genetic elements and antibiotic resistance genes on the genome
78 (24-27). Each CC can be further subdivided into sequence types (ST) (22). Waldron and Lindsay (16)
79 first realised that each CC of *S. aureus* contained a unique pair of Type I RM systems. A Type IV
80 restriction system, SauUSI, was also identified later and recognised as a methyl-dependent
81 restriction enzyme which would prevent the uptake of foreign DNA containing C5-methyl cytosine
82 (5mC) (28,29). Thus most genetic manipulation of *S. aureus* is confined to strain RN4220, which has a
83 defective Type I RM system due to a premature stop codon in *hsdR*. Furthermore, to avoid the Type
84 IV system, DNA needs to be prepared from an *Escherichia coli* strain, such as *E. coli* ER2796, lacking
85 the Dcm 5mC MTase (30).

86
87 The Type I RM systems in different strains of *S. aureus* were given the informal name of *Sau1* by
88 Waldron and Lindsay (16) and it is clear from not only a comparison of the sequences of genes and
89 proteins but also from the ability to use subunits from one strain to complement subunits from
90 other strains (31) that the term *Sau1* describes a classical "family" of Type I RM systems. Type I RM
91 families, Type IA to Type IE, were originally defined in *E. coli* and *Salmonella enterica* by DNA
92 hybridisation, antibody cross reactivity and subunit complementation (32,33), although now it is
93 more usual to use the high levels of sequence identity (over 90%) in HsdM and HsdR to define a
94 family *in silico*. Although the name *Sau1* for this family of Type I RM systems in *S. aureus* is an

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3 95 informal one not following the usual conventions (34), we retain it as it is established in the
4 96 literature. However, it is important to note that some strains of *S. aureus* show additional Type I RM
5 97 systems, which show limited amino acid sequence identity to the HsdR, HsdM and HsdS of *Sau1*
6 98 (Figure 1a). For instance, Monk *et al.* (35) identified an active Type I RM system, SauJKDIII, in *S.*
7 99 *aureus* JKD6159 which showed low sequence identity to members of the *Sau1* family. This is clearly
8
9 100 a member of a new and different Type I RM family whose subunits will be unable to interact with the
10 101 *Sau1* HsdM and HsdR (DTFD, JAL and MTG Holden, manuscript in preparation).
102

103 The *Sau1* Type I RM systems are so effective because they show great variability in the target
104 sequences recognised thus preventing HGT between CC but allowing HGT between strains within a
105 CC (31,35,36). This variability in target sequences is due to the modular construction of the Type I
106 RM systems (Figure 1b). The S subunit contains two target recognition domains (TRD) each of which
107 recognises one half of a bipartite target, for example the first Type I RM system in CC1, given the
108 generic name CC1-1, recognises CCAYNNNNNTTAA (adenine methylation sites are underlined)
109 (35,36). Swapping TRDs between S subunits generates new targets, for example the second Type I
110 RM enzyme in CC1, termed CC1-2, couples the first TRD of CC1-1 with a different second TRD to
111 recognise CCAYNNNNNNIGT. This swapping is easy because the DNA for S subunits contain
112 conserved sequences bounding each TRD. Most *S. aureus* strains have two copies of *hsdS*, two of
113 *hsdM* and one of *hsdR*. Thus there are often four TRDs in each CC, which define the restriction
114 barrier against HGT. Some Type I RM enzymes have half-size HsdS incorporating only a single TRD. It
115 has been shown that these products are often able to dimerise and recognise symmetric target
116 sequences (37-39). We have been able to recapitulate these results on “half-HsdS” enzymes by
117 manipulating the CC398-1 *S. aureus* system (EKM Bower and DTFD, unpublished results).
118

119 Previously we have identified the target sequences recognised by several common community-
120 associated, hospital-associated and livestock-associated) MRSA clonal complexes (31,36) and
121 recently several more have been identified (3,35,40). Monk *et al.* (35) and Jones *et al.* (40) have used
122 this information to prepare DNA methylated by the MTase M₂S₁ component enzymes to aid the
123 transformation of *S. aureus* strains that are usually resistant to transformation.
124

125 The identification of further targets recognised by the S subunits of *Sau1* Type I RM systems would in
126 principle allow more CC to be transformed for genetic analysis. In addition, further understanding of
127 the structural requirements for TRDs to recognise different specific DNA sequences is of intense
128 interest as the Type I RM systems are very widespread in bacteria and archaea (1,4) and exert a
129 considerable pressure on HGT and the evolution of prokaryotes. For instance, the use of multiple
130 TRDs being exchanged between strains has been observed in *Helicobacter* (41), *Mycoplasma* (42,43),
131 *Streptococci* (44,45) and *Bacteroides* (46).
132

133 Here we identify many further TRDs and their targets using both biochemical and PacBio single-
134 molecule real-time (SMRT) sequencing methods to define the barriers to HGT in a wide range of *S.*
135 *aureus* CC of global importance.
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137

138 **Materials and Methods**

139 **Nomenclature for expression plasmids encoding new MTases.**

140 As each Type I S subunit contains two TRDs and we propose to determine the targets recognised by
141 each TRD, we have given each TRD a single letter code, Table 1, and refer to the plasmids as
142 pSauTRD1-TRD2, e.g. pSauBI expresses an S subunit containing TRD B and TRD I and the M subunit. If
143 the TRD combination is the same as that found in a known clonal complex, then that CC is also given
144 in brackets. The MTase would be called M.SauBI in this example and the S subunit S.SauBI and is
145 from CC22. All sequences are given in the supplementary information.

146

Preparation of M.SauBI (CC22-1), M.SauCD (CC30-1), M.SauJK (CC30-2) and M.SauCL (CC45-1).

These four MTases were prepared as EGFP-His tag fusions as described in Roberts *et al.* (31). pSauBI-EGFP (CC22-1, genomic DNA from MRSA5906), pSauCD-EGFP (CC30-1, genomic DNA from MRSA252), pSauJK-EGFP (CC30-2, genomic DNA from MRSA252) and pSauCL-EGFP (CC45-1, genomic DNA from strain 70642) were all constructed by the polymerase chain reaction (PCR) with their *hsdS* fused to DNA encoding EGFP and a His-tag, with the following locus-specific oligonucleotides priming from the 3' end of the genes encoding the S subunits:

CC22-1 BI BS 5'GATCGAATTCGGATCCAATAAACATCTTTTGTA AAAACAC3'

CC30-1 CD BS 5'GATCGAATTCGGATCCTAAGAACATCTTTTGTA AAAAAGG3'

CC30-2 JK BS 5'GATCGAATTCGGATCCTATAAAAATTTTTGAAGTAATCCTTG3'

CC45-1 CL R167K BS 5'GATCGAATTCGGATCCAATAAACATCGATTTAAGTAAGGC3'

The sequence for CC45-1 introduced a single mutation R167K in the first TRD in the S subunit but since this change is found in other *S. aureus* isolates containing this TRD, the change is presumed to be completely neutral.

A new vector for MTase expression: pJF118his.

Although we had not experienced problems in examining the fusion proteins of S subunits and EGFP in biochemical work, we decided to construct a vector encoding *hsdS* with only a C-terminal His-tag. Vector pJF118his was made by PCR of the plasmid encoding the MTase CC5-1-EGFP constructed in Roberts *et al.* (31) with these two oligonucleotides:

pJFMShisTS 5'AGCTTCGAGAGGATCCCATCATCATCATCATTAAGAATTCAGCTTGGCTGTTTTGGCGG3'

and pJFMSEGFhisBS 5'GAGTGAATCCCCGGGGATCCGTCGACC3'.

The resulting PCR product was cut with BamHI and unimolecular religation gave pJF118his into which the *hsdMS* operon could be ligated as BamHI fragments and from which all subsequent MTase clones were descended.

Construction of an MTase plasmid to allow TRD swaps: pSaudeltaXmal.

A PCR-based strategy was devised to allow free pairwise assortment of desired TRDs in HsdS. Many, but not all of the HsdS subunits, including that encoded by the Type I system in CC398 (36), have a predicted proline-glycine sequence near the N-terminus. This dipeptide can be encoded by CCCGGG, which would be a target site for SmaI or XmaI. Oligonucleotides were designed which would introduce this motif in the N-terminus (a replacement with no amino acid changes) and at the C terminus (an insertion of two amino acids) of the S subunit of the CC398 system (36), by a two stage PCR fusion. Thus, primary PCR products were generated by reactions primed by: PromoterJF 5'GCTTCTGGCGTCAGGCAGCC3' with 398SmaIOligoBS 5'CCCATTCGCCTTCAAACCCGGGGAATCTCAACTCTGGCAC3' and 398SmaIOligoTS 5'GTGCCAGAGTTGAGATTCGCCGGGTTGAAGGCGAATGGG3' with 398SmaIBamHI 5'GATCGATCGGATCCCCGGGAATAAACATCTTTTGAAGTAATGAC3'.

The purified PCR products were then fused in a secondary PCR reaction primed by PromoterJF with 398SmaIBamHI. The product was then cut with BamHI, and ligated into the BamHI site of pJF118his as pSauNE-Xmal. This mutated form of the CC398-1 MTase, could assemble the complete restriction enzyme that proved to be active in endonucleolytic cleavage (36). This indicated that insertion of a proline and glycine towards the C-terminus did not affect the function of the enzyme. Subsequently, on reanalysing the DNA sequence, a single PCR mutation was discovered within the XmaI fragment. This caused a mutation A50S but this clearly did not affect the specificity or function of the S subunit in our assays. Digestion of pSauNE-Xmal with XmaI followed by intramolecular religation of the vector fragment generates pSaudeltaXmal, into which any pairwise combination of TRDS with XmaI cohesive ends may be inserted.

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3 196 **Construction of MTases M.SauNI, M.SauND, M.SauNK, M.SauNL, M.SauBE, M.SauJE and M.SauCE**
4 197 **(ST425-1) containing hybrid S subunits.**

5 198 The DNA for each TRD of these S subunits was fused to the DNA for the reciprocal TRD of S.SauNE
6 199 (CC398-1). This was achieved by creating primary PCRs with a short area of homology, which then
7 200 allowed base pairing of single strands of each PCR, in a secondary PCR. For example, S.SauBE TRD B
8 201 was generated from an appropriate plasmid template by PCR with oligonucleotides,
9 202 TRD1FOR398SmaIOligoTS 5'GTGCCAGAGTTGAGATTCCTCCGGGTTTGAAGGCGAATGGG3' paired with
10 203 TRD1nearuniversal 5'GTTCTTCTAATTCAATTTGT3'. TRD E was similarly generated by PCR from
11 204 plasmid template with oligonucleotides TRD2nearuniversal 5'ACAAATTGAATTAGAAGAAC3' and
12 205 398SmaIBamHI 5'GATCGATCGGATCCCCCGGAATAAACATCTTTTGAAGTAATGAC3'. The final insert
13 206 was then generated by PCR with the two gel-purified primary oligonucleotides and
14 207 TRD1FOR398SmaIOligoTS 5'GTGCCAGAGTTGAGATTCCTCCGGGTTTGAAGGCGAATGGG3' and
15 208 398SmaIBamHI 5'GATCGATCGGATCCCCCGGAATAAACATCTTTTGAAGTAATGAC3'. S.SauCL was the
16 209 only subunit for which we could not use the central universal oligonucleotides for PCR and required
17 210 specific substitutes: TRDLFOR/CC45-1
18 211 5'ACAAATTGAATTAGAAGAACAAAACTTGAATTACTTCAACAACAG3' and TRDC/CC45-1
19 212 5'GTTCTTCTAATTCAATTTGTGCGATCGAGTTTGCTGAAGAAG3'. Each C-terminus is unique and where
20 213 TRD2 was not TRD E, a specific oligonucleotide was employed: TRDIREV/CC22-1c-termssmal
21 214 5'GATCGATCGGATCCCCCGGAATAAACATCTTTTGTAAAAACAC3', TRDDREV/CC30-1c-termssmal
22 215 5'GATCGATCGGATCCCCCGGTAAGAACATCTTTTGTAAAAAGGATTG3', TRDKREV/CC30-2c-termssmal
23 216 5'GATCGATCGGATCCCCCGGTATAAAAATTTTTTGAAGTAATCCTTG3' and TRDLREV/CC45-1c-termssmal
24 217 5'GATCGATCGGATCCCCCGGAATAAACATCGATTTAAGTAAGGC3'. Each pure secondary PCR product
25 218 was cut with XmaI and ligated into the XmaI site of pSaudeltaXmaI.
26 219

27 220 **Construction of further MTases with further combinations of TRDs using synthetic genes.**

28 221 Additional *hdsS* sequences were obtained as synthetic genes from GeneArt (ThermoFisher Scientific)
29 222 with sequences optimised for expression in *E. coli* (Supplementary information). All the first TRDs
30 223 begin with 5'CCCGGGTTTGAAGGCGAATGGGAG3', except that for CC80-2 which begins with
31 224 5'CCCGGGTTTGAAGGCGAATATTCT3'. All the first TRDs end with
32 225 5'CAAATTGAATTAGAAGAACAGAAG3'. All the second TRDs begin with
33 226 3'CAAATTGAATTAGAAGAACAGAAG5' and have a universal reverse oligonucleotide, Trd2univrev
34 227 5'GATCGATCGGATCCCCCGG3'. These conserved sequences were used to create oligonucleotides to
35 228 prime PCR reactions. Each pure secondary PCR product was cut with XmaI and ligated into the XmaI
36 229 site of pSaudeltaXmaI. The orientation of the fragments was determined by PCR.
37 230

38 231 **Expression and purification of MTases.**

39 232 These new MTases and the R subunit of CC5 were expressed in *E. coli* BL21(DE3) and purified via
40 233 HisTrap chromatography, size exclusion chromatography, diethylaminoethyl (DEAE) anion exchange
41 234 chromatography and, if necessary, Heparin HiTrap chromatography (GE Healthcare, Uppsala,
42 235 Sweden) as described previously (31).
43 236

44 237 **Nuclease and ATPase assays.**

45 238 Purified MTases were mixed with the CC5 R subunit and used in assays for ATP hydrolysis (ATPase)
46 239 activity (coupled enzyme assay following a change in absorbance of NADH) and DNA cleavage
47 240 activity (plasmid cutting assay with analysis via agarose gel electrophoresis) as previously described
48 241 (31,36).
49 242

50 243 **Preparation of genomic DNA for SMRT sequencing.**

51 244 The expression plasmids harbouring the various MTases were used to transform a non-methylating
52 245 (*dam*⁻ *dcm*⁻) strain of *E. coli* ER2796 (30). Single colonies from the transformation plate of Lysogeny
53 246 Broth (LB) agar medium supplemented with 10 µg/ml kanamycin, 10 µg/ml tetracycline as well as
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3 247 100 µg/ml carbenicillin, which acted as a selection marker for the expression construct, were picked
4 248 and used to inoculate 5 mL of LB containing the same cocktail of antibiotics. The cultures were
5 249 incubated overnight with shaking at 37°C and 1 mL aliquots of the overnight culture were then
6 250 pelleted by centrifugation (6000 g, 6 min, 4°C). The culture medium was carefully removed and the
7 251 cell pellets stored at -20°C until required. Genomic DNA was prepared from each cell pellet using the
8 252 Wizard Genomic DNA purification kit (Promega, Madison, WI) according to the manufacturer's
9 253 instructions. The quality of the genomic DNA preparations was initially assessed by agarose gel
10 254 electrophoresis and from the shape of the absorbance profile from 240 to 340 nm. Genomic DNA
11 255 from *S. aureus* strains LGA251 (a kind gift from Mark Holmes) and NCTC13435 (a kind gift from
12 256 Angela Kearns) was prepared by using the PurElut Bacterial Genomic Kit (EdgeBio, Gaithersburg, MD
13 257 20877, USA). The DNA library for SMRT sequencing was prepared and subsequently analysed as
14 258 described in Anton *et al.* (30).
15 259

16 260 **Methylation of plasmids using M.EcoGII.**

17 261 M.EcoGII was kindly supplied by Dr. Iain Murray (New England Biolabs) and used to modify plasmids
18 262 E2, E5, E10, E11 and E12 previously described (31) and plasmid pCN36 (47). 0.45 µg DNA was
19 263 methylated using 2.0 U of M.EcoGII for 100 min at 37°C in a 50 µl volume. The reaction was in
20 264 1xNEB4 buffer (50 mM potassium acetate, 20 mM Tris acetate, 10 mM Mg acetate, 1 mM DTT (pH
21 265 7.9@25°C) supplemented with 320 µM S-adenosyl-L-methionine (SAM). As a negative control, DNA
22 266 was incubated in the same buffer without M.EcoGII. The DNA samples were then supplemented with
23 267 ATP (20 µM) and additional SAM (160 µM) and then digested with a Type I enzyme (CC5-1, CC5-2,
24 268 CC30-1, CC45-1 or the NY TRD hybrid) for 14 min at 37°C. As a control, methylated and
25 269 unmethylated DNA was digested with EcoRI.
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28 272 **Results and Discussion**

29 273 **Assigning TRDs to target sequences.**

30 274 Each TRD was given a one letter code (A to Z and a* to f*), Table 1. There were 14 TRD1 examples
31 275 and 18 TRD2 examples in our survey and these are found in 17 different CC or ST groups. Table 1 lists
32 276 the target specificity and site of methylation for each TRD in our survey. These data were obtained
33 277 by pairing TRDs and determining the complete target for each TRD pair as described in the next
34 278 section and in full in the supplementary information. Of interest are the TRD pairs B and P and U and
35 279 c*. These pairs recognise the same DNA sequence namely AGG and GAY respectively. Amino acid
36 280 sequence comparisons of B with P and U with c* are shown in Figure 2.
37 281

38 282 TRD B and TRD P are virtually identical throughout the TRD region even though TRD B is the first TRD
39 283 in the HsdS subunit and TRD P is the second TRD in the HsdS subunit, Figure 2a. While the high level
40 284 of sequence identity is expected for Type I systems in the same family, the high level of identity
41 285 between TRDs found in the first or second position in the HsdS subunit is more unusual. However,
42 286 such a situation has previously been observed in comparisons of the Type I systems in *Salmonella*
43 287 *blegdam* and *E. coli* R124 (48).
44 288

45 289 In contrast, TRDs U and c* are both examples of the second TRD in the HsdS subunit recognising 5'-
46 290 GAY-3' but the level of identity between them is much lower (~36%) (Figure 2b). This level of identity
47 291 between TRDs recognising the same target is expected if the TRDs are from different Type I RM
48 292 families so the low level of identity observed here is unusual. Despite this low level of sequence
49 293 identity, the predicted secondary structure elements are the same as expected from the early work
50 294 of Sturrock and Dryden (49). In fact, all of the TRDs in the *Sau1* family of RM systems align well when
51 295 secondary structure elements are taken into consideration (50) and they will have the same protein
52 296 fold (Supplementary information: PROMALS alignments). Therefore, it should in future be possible
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3 297 to **predict** the precise amino acid to nucleotide contacts involved in sequence recognition as was
4 298 done for the Type IIG TRDs (51,52).
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8 302 **Determination of complete target sequences recognised by pairs of TRDs.**

9 303 Tables 2, 3 and 4 show the TRD combinations investigated in this work and those investigated
10 304 previously by ourselves and others along with their combined target sequences, methylation
11 305 specificity and the methods used to determine these parameters. The full experimental data are
12 306 given in the supplementary information. Many of the TRDs were investigated in more than one
13 307 MTase and in more than one assay thus our set of data represents a self-consistent set. DNA
14 308 cleavage and ATP hydrolysis assays were performed on purified MTases mixed with purified R
15 309 subunit while SMRT data were collected from *E. coli* genomic DNA isolated after the hosts were
16 310 transformed with a plasmid expressing the MTase or directly from *S. aureus* genomic DNA. The
17 311 adenines targeted for methylation were determined easily by SMRT sequencing but for systems not
18 312 examined in this manner, it was assumed if there was a single adenine in the site recognised by the
19 313 TRD that this was the target for methylation.
20 314

21 315 Table 2 contains systems from a range of CC investigated previously as well as several examined in
22 316 this study. It is important to note that in our work those systems containing M.SauMRSII plus S.
23 317 SauMRSII, M.Sau133ORF1794P plus S.Sau133ORF1794P and M.SauMRSI plus S.SauMRSI are paired
24 318 with the HsdR (SauN315ORF189P) from the N315 strain of CC5 in DNA cleavage and ATPase assays.
25 319 Those shown in Tables 3 and 4 are studied as HsdS paired with the HsdM (M.SauSTORF499P) from
26 320 strain S0385 of CC398 and the HsdR (SauN315ORF189P) from the N315 strain of CC5 (if used in DNA
27 321 cleavage or ATPase assays). Therefore, these HsdS are not examined in the context of their natural
28 322 genome, but since they are all from the *Sau1* family of Type I RM systems and the HsdM and HsdR of
29 323 these RM systems are essentially identical in all of the strains, it is reasonable to assume that the
30 324 target specificities identified are those that would be recognised in their natural host.
31 325

32 326 Identifying the complete target recognised by a member of the *Sau1* Type I RM family when both
33 327 TRDs have unknown targets is difficult and ambiguous as either orientation may be correct. Hence,
34 328 we combined TRDs with unknown targets with TRD E or TRD N to make a protein recognising a
35 329 hybrid sequence in which one half of the target was already known (Table 3). A variety of methods
36 330 were used to determine the target associated with each hybrid including DNA cleavage and ATP
37 331 hydrolysis assays when the hybrid enzyme could be expressed and purified from *E. coli* and SMRT
38 332 sequencing when the expression and purification levels were low, for example, the SauJK enzyme
39 333 corresponding to the second Type I RM enzyme in CC30 did not express in *E. coli* despite its
40 334 expression in *S. aureus* by Monk *et al.* (35). The ambiguity in assignment of targets in CC93 in Monk
41 335 *et al.* (35) is resolved because the TRDs M and b* occur in more than one HsdS in our survey.
42 336

43 337 The DNA sequences for further pairs of TRDs found in a wide range of CC and ST groups were then
44 338 inserted after the *hdsM* of CC398-1 in our expression vector and examined to ascertain the spacer
45 339 sequence in the natural system (Table 4).
46 340

47 341 Genomic DNA from *S. aureus* strains NCTC13435 and LGA251 was prepared and examined using
48 342 SMRT sequencing as these strains contain two TRD pairs, XY and e*f* respectively, which we could
49 343 not express in *E. coli*. While SMRT signatures for the other Type I HsdS in these strains were very
50 344 clear (Supplementary information) and in agreement with our results from *E. coli* (Table 4) and those
51 345 of Monk *et al.* (35), these TRD pairs still showed no methylation activity even in their normal host.
52 346 Thus these TRDs pairs are not active.
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348 Analysis of spacer sequence length in *S. aureus* Type I RM systems.

349 It is apparent that the number of base pairs separating the adenines targeted for methylation and
350 the number of base pairs in the non-specific spacer between the sequences recognised by the TRDs
351 is not constant, with the former varying between 7 and 9 base pairs and the latter varying between 5
352 and 7 base pairs. This variation makes it very difficult to predict a Type I RM recognition sequence if
353 one knows only the targets recognised by the two TRDs as the length of the spacer in the target is
354 not recognised in any obvious manner by the TRDs. An example of this is the CC80-1 enzyme (Table
355 4) containing TRDs X and Y of known specificity. Since the enzyme did not methylate DNA *in vivo* for
356 the SMRT analysis, the spacer and hence the complete target for CC80-1 remain unknown until the
357 enzyme is purified and analysed biochemically. While it has been observed that insertions of
358 multiples of four amino acids into the alpha helical spacers separating the TRDs can increase the
359 length of the spacer in the target sequence in a predictable manner (65-67), it is clear from the
360 structure of HsdS subunits (Figure 1b) that the junction between the TRDs and the alpha helical
361 spacers in the conserved region is going to be of crucial importance for determining the fine details
362 of the length of the spacer in the target sequence as was found for some Type IIB RM enzymes
363 which contain a subunit equivalent to HsdS (68). Perhaps even single amino acid insertions or
364 deletions will serve to rotate the TRD with respect to the rest of the subunit and thereby change the
365 length of the spacer. Further progress in understanding the correlation between amino acid
366 sequence and the length of the target spacer would be greatly aided by an accurate atomic structure
367 of a Type I enzyme with DNA as the current models (12,13) lack sufficient resolution to be
368 informative on this point.

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370 Linking TRDs pairs to further clonal complexes and sequence types.

371 After determining the recognition sequences for all of the TRDs in Table 1 by creating artificial
372 hybrids (Table 3) we also found that some of these TRD combinations do actually occur in natural
373 systems as given in Table 5 (and supplementary information) (69). As sequence databases expand,
374 more and more of the possible TRD combinations based on the TRDs in Table 1 will be found. As
375 mentioned above, although the sequences recognised by the TRDs are known, the length of the non-
376 specific spacer separating them is unknown so that the complete target cannot be specified
377 accurately without experimentation.

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379 Further TRDs in *S. aureus* Type I RM systems.

380 Searching the publicly available sequences in the NCBI database with individual TRD sequences
381 revealed that some of those given in Table 1 can be found paired up with further novel TRDs. We
382 have found four new TRDs shown in Table 6 in *S. aureus* strains 21343 and KPL1845. Strain 21343
383 contains "NOVEL 1" paired with TRD K and the TRD pair NQ described in Table 6. Strain KPL1845 also
384 contains the TRD pair NQ and two further systems comprised of "NOVEL 2" paired with "NOVEL 3"
385 and "NOVEL 4" paired with TRD f*. Undoubtedly further TRDs will be found as sequencing continues.

386

387 Improving transformation of *S. aureus* by avoiding targets recognised by the *Sau1* Type I RM family.

388 A general method of preparing DNA suitable for transformation of *S. aureus* which can overcome the
389 RM barrier should be possible. Several DNA MTases belonging to Type II RM systems have been
390 found which have extremely short target recognition sites, namely Hin1523, Nma1821 and Hia5 (70)
391 and EcoGII recognising and methylating adenine in the targets 5'-A-3', 5'-AB-3' or 5'-BA-3'. The
392 methylation performed by these enzymes should protect any DNA molecule from the RM enzymes
393 described here (or indeed any RM barrier relying upon adenine methylation). Thus, DNA methylated
394 *in vitro* with these unusual MTases could be used in subsequent transformation experiments even
395 when major RM barriers are present.

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397 We used the M.EcoGII adenine MTase (a kind gift from Iain Murray, New England Biolabs) to modify
398 all adenines in several plasmids *in vitro*. The plasmids were from our collection of plasmids used to

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3 399 determine the target sequences of the *S. aureus* Type I enzymes and have been previously described
4 400 (31). These plasmids were then mixed with various purified *S. aureus* Type I restriction enzymes or,
5 401 as a control, the EcoRI restriction enzyme. After one hour of methylation by M.EcoGII, the plasmids
6 402 were completely resistant to digestion by EcoRI and by the *S. aureus* restriction enzymes (Figure 3).
7 403 Furthermore, the shuttle vector pCN36 (47) was also protected from digestion by these same
8 404 enzymes (data not shown). Subsequent experiments using the methylated pCN36 to transform *S.*
9 405 *aureus* were unfortunately entirely unsuccessful (unpublished results by JAL using strains HO5096
10 406 (CC22), JE2 (CC8) and RN4220 (CC8, *hsdR*). The reason for the failure of transformation with the
11 407 highly-methylated pCN36 when it should be resistant to all *Sau1* RM systems is not clear. This result
12 408 may imply a further unrecognised barrier to transformation of *S. aureus* or some aspect of the
13 409 physical properties of highly methylated DNA. Nevertheless, the method using MTases with very
14 410 short target recognition sequences may be of use for transformation of other bacterial species.
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17 413 **CONCLUSIONS**

18 414 In conclusion, we have determined the target recognition sequences of a considerable number of
19 415 TRDs and HsdS specificity subunits of the Type I RM systems in *S. aureus*. This was achieved using a
20 416 combination of gene synthesis, endonuclease activity, ATP hydrolysis activity and single molecule
21 417 real-time genome sequencing. The systems analysed cover a large proportion of the known
22 418 sequence types and clonal complexes of *S. aureus* and delineate more clearly the barrier to
23 419 horizontal gene transfer within the *S. aureus* population.
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25 421 The data obtained here will allow the construction of new *E. coli* strains for preparing methylated
26 422 shuttle vectors (35) and MTase reagents for *in vitro* methylation of DNA (40) to assist transformation
27 423 of further *S. aureus* strains. However, these approaches are time consuming and it is worth noting
28 424 that the common shuttle vector used for transformation of *S. aureus*, pCN36 (47), contains a target
29 425 site for almost every TRD pair investigated in this paper. This means that pCN36 is inevitably a poor
30 426 vector for transformation of *S. aureus*. The construction of new shuttle vectors completely lacking
31 427 *Sau1* targets via DNA synthesis, coupled with careful analysis of the fragments to be ligated into the
32 428 vector so that they also lack targets, may be an effective way forward to improve transformation of *S.*
33 429 *aureus* now that so many target specificities have been determined. Obviously, the avoidance of the
34 430 sequence AN₆₋₉T, although difficult to achieve without altering protein coding sequences in a vector,
35 431 would be a general method to negate the effect of the Type I RM systems in *S. aureus* and other
36 432 prokaryotes.
37 433

38 434 Lastly, the determination of so many recognition sequences of Type I RM systems in different
39 435 lineages of *S. aureus*, in effect a “Rosetta Stone”, means that now the population structure of *S.*
40 436 *aureus* can be investigated from an epigenetic/evolutionary perspective (4) as performed previously
41 437 with, for example, *H. pylori* (71) and *S. pneumoniae* (72).
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43 439

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49 445

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3 453 **Figure legends**

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5 455 **Figure 1.** *S. aureus* genomes showing the genes and the typical organisation of TRDs in the HsdS DNA
6 456 sequence specificity subunit. (a) Strain ED133 (CC133) has two *hsdS*; strain 11819-97 (CC80) has
7 457 three *hsdS* (CC80) and strain JKD6159 (CC93) contains an extra Type I RM system from a different
8 458 Type I RM family. From top to bottom: ED133, 11819-97, JKD6159. *hsdR* (red), *hsdM* (blue), *hsdS*
9 459 (yellow). (b) The structural organisation of the HsdS specificity subunit. The conserved regions (cr)
10 460 are common to all S subunits within a family. The two target recognition domains (TRD1 and TRD2)
11 461 define the target sequences recognised by the RM enzyme and can be swapped between S subunits
12 462 of the same family to generate new specificities.

13 463

14 464 **Figure 2.** Amino acid sequence and secondary structure alignment of two pairs of TRDs recognising
15 465 the same DNA target. The TRD sequences are highlighted in yellow. Consensus secondary structure
16 466 shows “h” for alpha helix and “e” for beta sheet. (a) TRDs B and P are examples of a first and a
17 467 second TRD respectively recognising 5'-AGG-3'. (b) TRDs U and c* are both examples of second TRDs
18 468 with the same specificity, 5'-GAY-3'. The long predicted alpha helices at the start and the end of the
19 469 sequences are the conserved helical spacer regions in the HsdS subunits while the sequence
20 470 between these helices makes up the TRD.

21 471

22 472 **Figure 3.** General protection from endonuclease activity using M.EcoGII MTase to methylate all
23 473 adenines. Plasmid without M.EcoGII treatment is digested (- lanes) but plasmid with M.EcoGII
24 474 treatment is protected from digestion (+ lanes). Panel (a) uses Sau347I (CC45-1, TRDs C and L)
25 475 restriction enzyme against plasmids E2, E5 and E10 described in (31). Panel (b) uses SauNY (TRDs N
26 476 and Y) against plasmids E10, E11 and E12 described in (31). Panel (c) uses three different enzymes,
27 477 SauN315I (CC5-1, TRDs B and D), SauN315II (CC5-2, TRDs A and H) and SauMRSII (CC30-1, TRDs C and
28 478 D), against plasmid E10. In each panel EcoRI restriction enzyme was used as a control and markers
29 479 (M) are in kb.

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Figure 1.

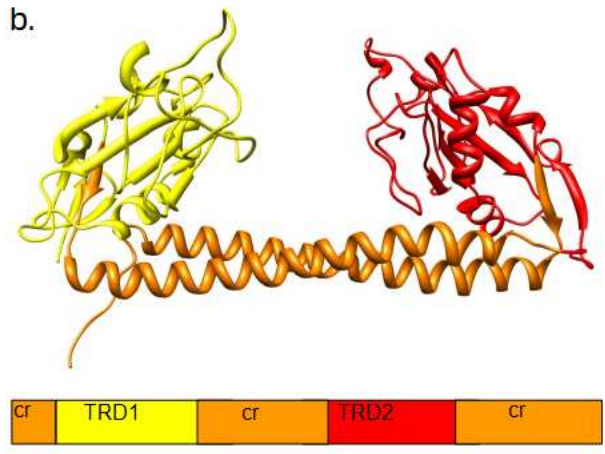
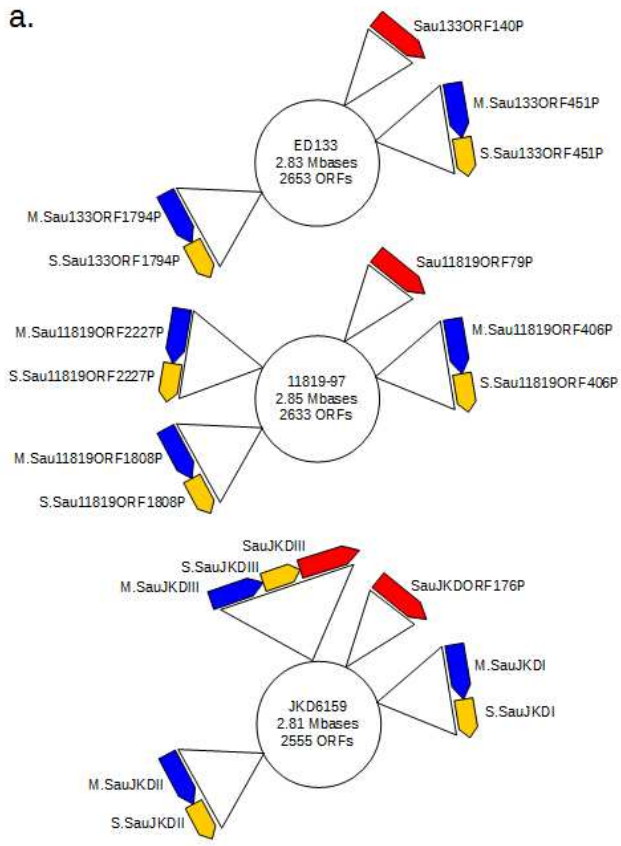


Figure 3.

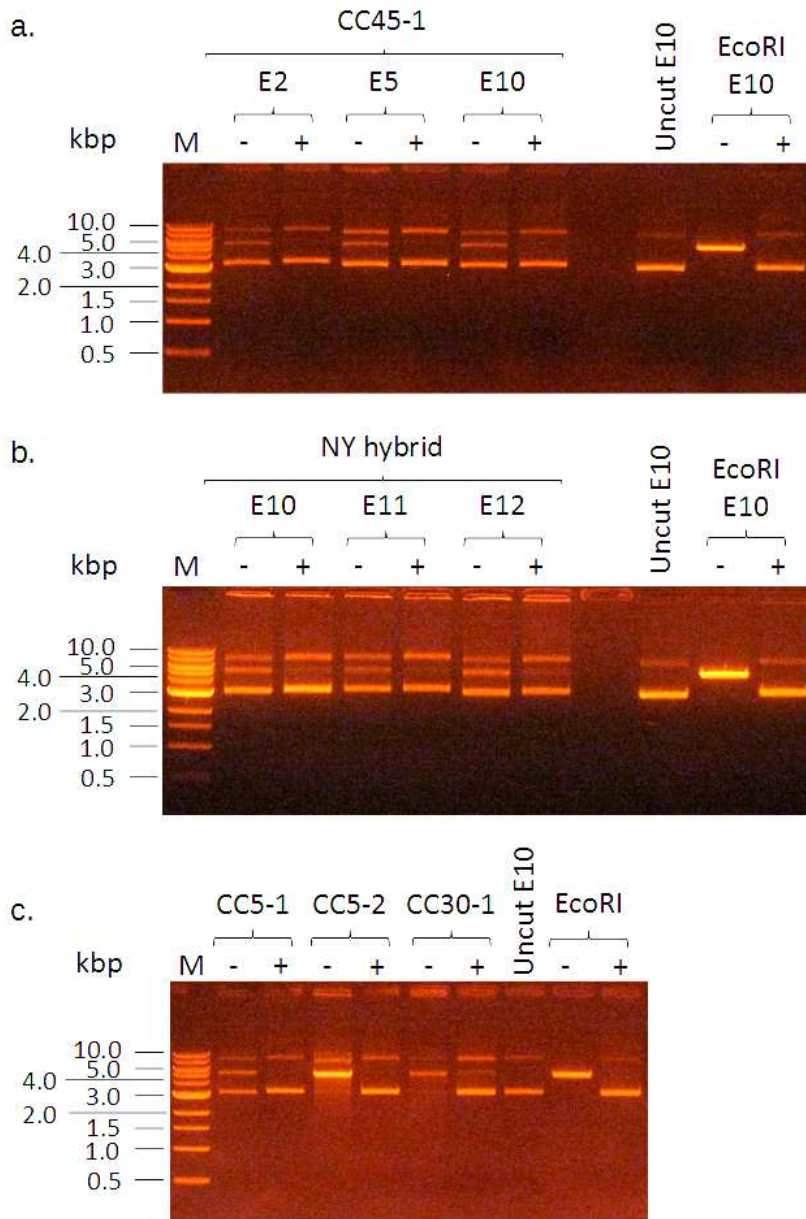


Table 1. TRD targets shown from 5' to 3'. The underlined A indicates the site of adenine methylation. TRD pair B and P, and TRD pair U and c* recognise the same DNA sequence and are highlighted in red. In the targets, Y = C or T; R = A or G; W = A or T; H = A, T or C; N = A, G, T or C.

TRD1 code letter and target		TRD2 code letter and target	
A	CC <u>A</u> Y	D	<u>A</u> TC
B	<u>A</u>GG	E	TC <u>A</u> Y
C	GW <u>A</u> G	F	TT <u>A</u> A
J	GG <u>A</u>	G	AC <u>A</u>
M	C <u>A</u> G	H	T <u>A</u> C
N	<u>A</u> CC	I	YT <u>C</u> A
O	CA <u>A</u> C	K	CG <u>A</u>
R	GAR <u>A</u>	L	TTT <u>A</u>
T	CA <u>A</u> G	P	<u>A</u>GG
V	CNG <u>A</u>	Q	AC <u>A</u> Y
X	TCT <u>A</u>	S	G <u>C</u> A
Z	G <u>A</u> C	U	<u>G</u>AY
b*	GGH <u>A</u>	W	CRA <u>A</u>
e*	G <u>A</u> G	Y	CT <u>A</u>
		a*	G <u>A</u> A
		c*	<u>G</u>AY
		d*	CY <u>A</u> A
		f*	GA <u>A</u> Y

Table 2. The *Sau1* RM systems with published recognition sequences. Target sites are shown from 5' to 3' with the length of the non-specific spacer shown as a number. Underlined A or T indicates the site of adenine methylation on the top or bottom strands respectively. The experimental methods used are indicated as g = target obtained by DNA cleavage with a purified enzyme, s = target obtained by SMRT sequencing of *E. coli* ER2796 genomic DNA, a = target obtained by ATPase assay with a purified enzyme. Full details are given in the supplementary information. *S.Sau133ORF1794P* is characterised in this work but is included here as it is part of the RM system found in strain ED133. *SauMRSI* and *SauMRSII* characterised by Monk et al. and *S.SauSTORF499P* characterised by Chen et al. are also further characterised in this work.

Strain name and genome reference	Clonal Complex or Sequence Type	S subunit name in REBASE	Recognition sequence	TRDs assigned	Suggested generic name	Experimental method	Reference for target specificity and method
MW2 (53)	CC1	S.SauMW2I	CC <u>A</u> Y-5-T <u>I</u> AA	AF	CC1-1	g, s, a	g (31) a (36) s (CC8-1 and CC8-2 in strain NRS384 are from ref. 35)
		S.SauMW2II	CC <u>A</u> Y-6- <u>I</u> GT	AG	CC1-2 (CC8-2)	g, s, a	
N315 (54)	CC5	S.SauN315II	CC <u>A</u> Y-6-G <u>I</u> A	AH	CC5-2	g, s, a	s (35) g, s (this work)
		S.SauN315I	<u>A</u> GG-5-G <u>A</u> <u>T</u>	BD	CC5-1 (CC8-1)	g, s, a	
MRSA252 (55)	CC30	S.SauMRSII	GW <u>A</u> G-5-G <u>A</u> <u>T</u>	CD	CC30-1	g, s	s (35) g, s (this work)
		S.SauMRSI	G <u>G</u> <u>A</u> -7- <u>I</u> CG	JK	CC30-2	s	
JKD6159 (56)	CC93	S.SauJKDIII	GA <u>A</u> G-5- <u>T</u> AC or complement	Not a <i>Sau1</i> system	CC93-3	s	s (35) Note the ambiguity in assigning CC93-1 and CC93-3 is clarified with strains ED133 and 32320 and from Table 3.
		S.SauJKDII	GG <u>H</u> A-7- <u>I</u> CG	b*K	CC93-2	s	
ED133 (57)	CC133	S.SauJKDI	C <u>A</u> G-6- <u>T</u> TC	Ma*	CC93-1	s	g (36) s (this work)
		S.Sau133ORF451P	C <u>A</u> G-5-R <u>I</u> G <u>A</u>	ME	CC133-1	g	
32320 (58)	CC133	S.Sau133ORF1794P	G <u>G</u> <u>A</u> -7- <u>T</u> TRG	Jd*	CC133-2	s	g (36) s (this work)
		S.Sau32320ORFAP	C <u>A</u> G-5-R <u>I</u> G <u>A</u>	ME	CC133-1	g	
S0385 (59)	CC398	S.SauSTORF499P	<u>A</u> CC-5-R <u>I</u> G <u>A</u>	NE	CC398-1	g, s	g (36) s (this work)

Table 3. The “artificial” *Sau1* systems containing novel pairings of TRDs. Target sites are shown from 5’ to 3’ with the length of the non-specific spacer shown as a number. Underlined A or T indicates the site of adenine methylation on the top or bottom strands respectively. The experimental methods used are indicated as g = target obtained by DNA cleavage with a purified enzyme, s = target obtained by SMRT sequencing of *E. coli* ER2796 genomic DNA, a = target obtained by ATPase assay with a purified enzyme. Full details are given in the supplementary information.

“Artificial” <i>Sau1</i> RM systems.					
Recognition sequence	TRDs assigned	Experimental method	Recognition sequence	TRDs assigned	Experimental method
<u>A</u> GG-5-R <u>I</u> GA	BE	a	<u>A</u> CC-6- <u>T</u> TC	Na*	s
GGA-6-R <u>I</u> GA	JE	g, s	<u>A</u> CC-6-R <u>I</u> C	Nc*	s
<u>A</u> CC-6- <u>I</u> GAR	NI	g	<u>A</u> CC-6- <u>I</u> TRG	Nd*	g, s
<u>A</u> CC-6- <u>I</u> CG	NK	g	GARA-6-R <u>I</u> GA	RE	s
<u>A</u> CC-6- <u>I</u> AAA	NL	g	CA <u>A</u> G-5-R <u>I</u> GA	TE	s
<u>A</u> CC-5- <u>C</u> CI	NP	s	CNGA-6-R <u>I</u> GA	VE	s
<u>A</u> CC-5-R <u>I</u> GT	NQ	g, s	TCTA-6-R <u>I</u> GA	XE	g, s
<u>A</u> CC-6- <u>I</u> GC	NS	s	G <u>A</u> C-5-R <u>I</u> GA	ZE	a
<u>A</u> CC-5-R <u>I</u> C	NU	g, s	G <u>A</u> C-6- <u>I</u> GC	ZS	a
<u>A</u> CC-6- <u>I</u> TYG	NW	g, s	GGHA-6-R <u>I</u> GA	b*E	s
<u>A</u> CC-6- <u>I</u> AG	NY	g, s	G <u>A</u> G-6-R <u>I</u> GA	e*E	g, s

Table 4. The *Sau1* RM systems investigated in this project. Target sites are shown from 5' to 3' with the length of the non-specific spacer shown as a number. Underlined A or T indicates the site of adenine methylation on the top or bottom strands respectively. TRD pair e*f* in strain LGA251 was not cloned in *E. coli* while TRD pair XY was cloned. However, no target modification was observed using SMRT on genomic DNA from either *E. coli* or *S. aureus* for these TRD pairs. If the genes are translated, their target is inferred from other TRDs in this table although the spacer length remains undefined. The experimental methods used are indicated as g = target obtained by DNA cleavage with a purified enzyme, s = target obtained by SMRT sequencing of *E. coli* ER2796 genomic DNA, s* = target obtained by SMRT sequencing of *S. aureus* genomic DNA, a = target obtained by ATPase assay with a purified enzyme. Full details are given in the supplementary information.

Strain name and genome reference	Clonal Complex or Sequence Type	S subunit name in REBASE	Recognition sequence	TRDs assigned	Suggested generic name	Experimental method
CO1791 (58)	CC97	S.SauC01791ORFAP	CC <u>A</u> Y-6-R <u>T</u> C	Ac*	CC97-1	s
HO5096 (60)	CC22	S.Sau5096I	<u>A</u> GG-6-I <u>G</u> AR	BI	CC22-1	g, s
LGA251 (61)	ST425	S.Sau251I	GW <u>A</u> G-5-R <u>T</u> GA	CE	ST425-1	g, s*
		S.Sau251ORF16900P	G <u>A</u> G-?-R <u>T</u> TC	e*f*	ST425-2	Not expressed, no signature with s*.
		S.Sau251II	GA <u>A</u> G-5- <u>T</u> AC or complement	Not a <i>Sau1</i> system	Same as CC93-3	s*
Isolate 3 (19)	CC51	S.SauL3ORFAP	GGA-6-C <u>C</u> T	JP	CC51-1	s
Isolate 3067 (19)	CC45	S.Sau347I	GW <u>A</u> G-6-I <u>A</u> AA	CL	CC45-1	g
Isolate 3150 (19)	CC15	S.SauL315ORFAP	CA <u>A</u> C-5-R <u>T</u> GA	OE	CC15-1	s
SA40 (62)	CC59	S.SauSA40ORF370P	GGA-6-R <u>I</u> GT	JQ	CC59-1	a
CN1 (63) MSHR1132 (64) NCTC13435 NCBI Biosample identifier: SAMEA2479566	CC72	S.SauCN1ORF415P	GAR <u>A</u> -6-R <u>T</u> GT	RQ	CC72-1	a
		S.SauCN1ORF1757P	GGA-7-I <u>G</u> C	JS	CC72-2	a
	CC75	S.Sau1132ORF3780P	CA <u>A</u> G-5-R <u>T</u> C	TU	CC75-1	g
		S.Sau1132ORF16570P	CNG <u>A</u> -7-I <u>T</u> YG	VW	CC75-2	s
	ST80	S.Sau13435ORF394P	TCT <u>A</u> -?-I <u>A</u> G	XY	ST80-1	Not expressed, no signature with s or s*.
		S.Sau13435ORF1751P	G <u>A</u> C-6-I <u>T</u> YG	ZW	ST80-2	
S.Sau13435ORF2165P	TCT <u>A</u> -6-R <u>T</u> TC	Xf*	ST80-3	s, s*		
32326 (58)	CC873	S.Sau32326ORFAP	G <u>A</u> G-6-G <u>A</u> I	e*D	CC873-1	a

Table 5. Further TRD pairs found in sequenced strains of *S. aureus*. Every pair of TRD1 with TRD2 in table 1 was used in a BLASTP sequence search to identify HsdS subunit sequences in publicly accessible databases. Examples of strains containing these TRD pairs are shown. ST and CC are from the PATRIC database (69) or derived using www.cbs.dtu.dk/services/MLST (73). Some TRD pairs are present in many strains while others are rare.

TRD pair	Example Strain	Clonal Complex or Sequence Type of example strain	REBASE name
AD	FDAARGOS_159	ST5	S.Sau159ORF12345P
AL	K12S0375	ST692	S.Sau375ORFDP
AU	<i>S. schweitzeri</i> FSA084	-	S.SauFSA084ORF355P
AW	FDA209P	ST464	S.Sau209ORF1697P
BG	MRSN8611	ST8	S.Sau8611ORF11430P
BH	PLAC6019	ST5	S.Sau6019ORF851P
BU	SA-083	ST101	S.Sau083ORF9680P
BY	<i>S. argenteus</i> M260-MSHR	-	S.SarM260ORF2316P
Bf*	SA-083	ST101	S.Sau083ORF1720P
JE	Tager 104	ST49	S.Sau104ORF1102P
JL	W56227	ST45	S.Sau56227ORF970P
JW	CIG290	ST45	S.SauCIG290ORF2408P
JW	APS211	ST45	S.SauAPS211ORF9230P
MW	FSA037	ST1872	S.SauFSA037ORF2487P
NQ	KPL1845	ST96	S.Sau1845ORF2596P
Of*	USA300-TCH959	ST1159	S.SauTCH959ORF2844P
Rf*	Tager 104	ST49	S.Sau104ORF2433P
TY	M21126	ST2250	S.Sau21126ORF1065P
XF	21334	ST109, CC9	S.Sau21334ORF1353P
XF	RKI4	ST27	S.SauRKI4ORF1905P
XW	103564	ST80-PVL carrier	S.Sau103564ORF678P
ZY	D139	ST145	S.SauD139ORF2470P
b*W	ST20130941	CC15	S.Sau941ORF4310P
e*f*	SA-120	ST425	S.Sau120ORF4875P

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Table 6. New TRD pairs associated with pairs shown in Tables 2, 3 and 4. The new TRDs of unknown specificity are termed NOVEL 1, NOVEL 2, NOVEL 3 and NOVEL 4. TRD NOVEL 3 is a second TRD while the others are first TRDs in the HsdS amino acid sequence. Subspecies 21343 and species KPL1845 also contain S.SauNQ (S.Sau21343ORF1169P and S.Sau1845ORF2596P respectively).

Subspecies 21343 Bioproject accession: PRJNA53699
> S.Sau21343ORF2597P TRD NOVEL 1 + TRD K MSNTQKKNVPELRFPGFEGEWEEKLGEVATFAKGKLGAKKDVSONGVPVILYGELYTKYGAIIVSKIFSKTDIPENKLMMAKNDVLIIPSSGETAIDIATASCIYLNKGVAVGGDINILTPQKQDGRFISLSIN GINKNELSKYAQGKTVVHLYNNDIKNLKIAFPSEFEEQVRIGNFFSKLDRQIELEEQLLELLQQQKGYMQKIFSQELRFKDENGNDYPKWEEKKIEDIASQVYGGGTPNTKIKEFWNGDIPWIQSSDVKVNDL ILQQCNKFISKNSIELSSAKLIPANSIAIVTRVGVGKLCLEVEFDYATSQDFLSLSSLYDKLYSLYSLYTMKKISANLQGTSIKGITKKELLDSEIKIPHNLEEQQKIGDLFYKIDKYISFNKCKIEILKSLK QGLLKKMFI
Species KPL1845. Bioproject accession: PRJNA169473
> S.Sau1845ORF1619P TRD NOVEL 2 + NOVEL 3 MTEQINTPELRFPEFKNEWSYDLVSDVVTNKSKKFDPKKEEAKKDIELDSIEQNTGRLLDITYISNDFTSQKNKFNKGNVLYSKLRPYLNKYYYATIDGVCSSSEIWLNTLNKDVLANKFLLYFIQTNRFSSTVN KSAGSKMPRADWELVKNIRLYKGSIEEQEKIGYFFSKLDRQIELEEKKLELLEQQKGYMQKIFAQELRFKDENGNDYPDWVTKKLGDIGKVAMNKRIYKNETTENGEIPFYKIGNFGKNADFTITREKFDEYK EKYPYPNVGDILISASGSIGRTIEYTGEDAYYQDSNIVLWLNHNDEVINKYLKYFYKIVKWSGIEGTTIKRLYNKNLNTKIELPTVEEQYKMANFLSKLDKIIDIQIEKIELLKQRKQGLLQKMFV
> S.Sau1845ORF2199P TRD NOVEL 4 + TRD f* MSNTQKKNVPELRFPEFEGEKDVKFVSIFQEVSNKTSDLAKYPLFSLTVEKGITPKTERYKRDFLVKSDNFKIVEPRDIVYNPMNVTLGAIKIDLSKYNIDIALSGYHVMKIINSFNPDIKSNFLKTEKMIH YKKIATGSLMEKQRVHFSEFKNI IKKFPNTKEQQKIGDFFSKLDRQIELOVQKLELLQQQKGYMQKIFSQELRFKDENGEDYPDWKEKKLGDITEQSMYIGASATRFDSKNIIYIRITDIDEKSRKLNQNL TPDELNNKYKLRNDILFARTGASTGKSYIHKEEKDIYNYFAGFLIKFEIDEQNNPLFIYQFTLTSKFNKWKVMSVRSQPGINSEYAKLPLVLPNKLEQQKIAEFLDRFDQQIELEKQKIEILQQQKGL LQSMFI

1
2 SUPPLEMENTARY INFORMATION

3 DNA target recognition domains in the Type I
4 restriction/modification systems of *Staphylococcus aureus*.
5

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7 Laurie P. Cooper, Gareth A. Roberts, John H. White, Yvette Luyten,
8 Edward K.M. Bower, Richard D. Morgan, Richard J. Roberts, Jodi A.
9 Lindsay, David T.F. Dryden.

10
11 Pages 2 and 3: SUPPLEMENTARY INFORMATION FOR TABLE 1.
12

13
14 Pages 4 to 9: Supplementary information for MATERIALS AND METHODS
15 SECTION "Construction of further MTases with further combinations
16 of TRDs using synthetic genes."
17

18
19 Pages 10 to 16: SUPPLEMENTARY INFORMATION FOR TABLE 2.
20

21
22 Pages 17 to 57: SUPPLEMENTARY INFORMATION FOR TABLE 3.
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25 Pages 58 to 85: SUPPLEMENTARY INFORMATION FOR TABLE 4

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27 Pages 86 to 91: SUPPLEMENTARY INFORMATION FOR TABLES 5 AND 6.
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Pages 92 to 97: PROMALS ALIGNMENT OF TRD AMINO ACID SEQUENCES WITH
SECONDARY STRUCTURE PREDICTIONS.

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2 **SUPPLEMENTARY INFORMATION FOR TABLE 1.**

3 **The amino acid sequences in FASTA format of the first TRD with its**
4 **letter code, DNA target (5' to 3') and methylation site underlined.**
5 **The TRDs labelled as NOVEL 1, NOVEL 2 and NOVEL 4 were found once**
6 **all of the other TRDs had been analysed but are included in this**
7 **list for completeness.**

8 **The TRD sequences are flanked by the conserved regions so to**
9 **obtain the amino acid sequence of any HsdS subunit simply paste**
10 **the sequence for the second TRD directly on to the end of the**
11 **sequence for the first TRD.**

12
13
14 >A CCAY

15 MSNTQKKNVPELRFPGFEGEWEEKQLGDLTTKIGSGKTPKGGSENYTNKGI PFLRSQNI RNGLNLDLVYISKDIDDEMKNRSRTYYGDVLLNITGASIG
16 RTAINSIVETHANLNQHVCIRLKKYIYFFGQYLLSRKGRKIFLAQSGSREGLNFKEITANLKIFTPTTFEEQQKIGKFFSKLDRQIELEEQKLELL
17 QQQ

18 >B AGG

19 MSNTQKKNVPELRFPGFEGEWEEKQLGDLTDVIRKKNKLESKKPLTISGQLGLIDQTEYFSKSVSKNLENYTLIKNGEFAYNKSYNGYPLGAIKRLT
20 RYDSGVLSSLYICFSIKSEMSKDFMEAYFDSTHWYREVSGI AVEGARNHGLLNVSVNDFTTLIKYPSSLEEQQKIGKFFSKLDRQIELEEQKLELLQQQ

21 >C GWAG

22 MSNTQTKNVPELRFPGFEGEWEEKQVGELELFKNGLNKGEYFGSGSSIVNFKDVFNNRSINTNNLTGKVVNSKELKNYSVEKGDVFFTRTSEVIGEIG
23 YPSVILNDPENTVFSGFVLRGRPKSGIDLINNNFKRYVFFTNSFRKEMITKSSMTTRALTSGTAINKMKVIYPVSAKEQQKIGDFFSKLDRQIELEEQKLE
24 LLQQQ

25 >J GGA

26 MSNTQKKNVPELRFPEFEGEWEEKLGDLIKVNSGKDYKHLDKGDI PVYGTGGYMTSVSEPLSEIDAVGIGRKGITINKPYLLEAPFWTVDTLFYCTPEKE
27 ADILFILSLFRKINWKLYDESTGVPVLSKQITINKINRLVPTNKEQQKIGEFFSKLDRQIELEEQKLELLQQQ

28 >M CAG

29 MSNTQTKNVPELRFPGFEGEWEEKLEDLGLFQKSYFSRAKEGNGKTKHIHYGDIHSHKFKTVLSDGNIPNI IEKAVFELIQKGDIVFADASEDYSDLG
30 KAVMIDFEPNSLISGLHHLFRPLNNAISNFLIFYTKTLSYKFFIRQQGTGISVLGISKKSLNLDLVIPRSELEQQKIGQFFSKLDRQIELEEQKLELL
31 QQQ

32 >N ACC

33 MSNTQKKNVPELRFPGFEGEWEEKLGEFAGKVTQKNVDKXYIETLTNSAELGIIISQKDYFDKEISNIDNIKKYVVEENDFVYNPRMSNYAPFGPVNRN
34 KLGKKGVMSPLYTVFKIQNIDLNFIEFYFKSSKWRFMALNGDSGARADRFISIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQQQ

35 >O CAAC

36 MSNKQKKNVPELRFPGFEGEWEEKLGEVGTFTSGGTPLKSKSEYWNWDIPIWITTDGIHNIKRENI TNFITEKGLNESSAKLITNEALIAMYGQKTRG
37 MSAI LNFEATNQACAIYQTNQININVFVQYFQKLYEFLRSLNENGSQKNLSLSLLEKEITLNY PNEQEQQKIGDFFSKLDRQIELEEQKLELLQQQ

38 >R GARA

39 MSNTQKKNVPELRFPGFEGEWEEKLGEVAKIYDGTHTQTPKYTNEGIFL SVENIKTLNSSKYISEEAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNE
40 KFAYYVSLALLKTKNLSYFLKNLILSSSIQNELWRKTLHVAFPPKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIELEEQKLELLQQQ

41 >T CAAG

42 MSNTQTKNVPELRFPGFEGEWEEKELGEIFQIISGSTPLKSNKEFYENGNINWVKTDDLNNKSVTHSKEKITEYAMKSLKLVKPNVLIAMYGGFNQI
43 GRTGLLKIDATINQAI SALLMNHETNPEFIQAFNLN YQVKGWRYAASSRKDPNITKKDIEQFKVPYVVSINEQQKIGEFFSKIDHQIELEEQKLELLQQQ

44 >V CNGA

45 MSNTGKMNVPELRFPGFEGEWEEKLRELNRNPKDKYSYTGPFPGSDLKSDYTTDGIQI IQLQNIGDGYFYNSNKVFTSNEKAEVLKSCNVFPGDIVIAK
46 MADPIARA AIVPDNNIGKYL MASD GIRLSVDTVHFNTK FVLECI NRKSRKVEDN SSGSTRMRIGLSTLGS LTLKTTTLKEQQKIGQFFSKLDRQIVLE
47 EQKLELLQQQ

48 >X TCTA

49 MSNTQKKNVPELRFPGFEGEWEEKQFADFTKINQGLQIAINERKTEYSPELYFITNEFLRPN SQTKYFIENPPQSVIANKEDILMTRTGNTGKVVTNVF
50 GAFHNNFFKIKFDKNLYDRLFLVEVLN SSKI QNKILSLAGSSTIPDLNHSD FYSISSYPLLEQQKIGKFFSKLDRQIELEEQKLELLQQQ

51 >Z GAC

52 MSNTQTKNVPELRFPGFEGEYSLDIFGNLATNKSEKFNPNENASIDIELDCIEQNTGR LIKIYNSKEFSSQKNKFNPNQVLYGKLRPYLNKYFFTKKSG
53 VCSSEIWLKSTKEDKLLNLFYFIQTKRYS DVASKSAGSKMPRADWGLIENIRVYFPELCEQQKIGQFFSKLDRQIELEEQKLELLQQQ

54 >b* GGHA

55 MSNTQKKNAPELRFPEFEGEWEEKLEDTFEIKDGTHTGTHENVNNGP WLLSAKNIKNNKIIISSDDRKISESDYKKIYKNYKLEKGDLLLTIVGTIGRA
56 AIVKNPNNIAFQRSVAI LKTKATYDVGFIFQLFQTKYFKNLLLRKQV VSAQPLYLGDIRKIKISITNIEEQKIGIFFSKLDRQIELEEQKLELLQQQ

57 >e* GAG

58 MSNTQKKNVPELRFPGFEGEWEEKSIS SFLKESKIKSGNSHAKKLT VKLWGKGVVPK KETFKGSDNTQYYKRKAGQ LMYGKLDLNFCAFGIVPDSLNNY
59 ESTIDSPSFD FINGDSKFLLE RIKLKS FYKFKGDIANGSRKAKRINQDTF LSLPVFAPKYDEQLRIGEFFSKLDRQIELQKQKLELLQQQ

60 >NOVEL 1

MSNTQKKNVPELRFPGFEGEWEEKLGEVATFAKGLGAKKDV SQNGVPVILY GELYTKYGAIVSKI FSKTDIPENKLMKAKNDVLI PSSGETAIDIAT
ASCIYLNKGVAVGGDINILTPQKQDGRFISLSINGINKNELSKYAQKGTVVHLYNNDIKNLKIAFPSEFEEQVRIGNFFSKLDRQIELEEQKLELLQQQ

>NOVEL 2

MSNTQKKNVPELRFPEFEGEWKDVKFVSI FQEVSNKTS DLAKYPLFSLTVEKGITPKTERYKRDFLVKSDNFKIVEPRDIVNPMNVTLG AIDL SKYNY
DIALSGYYHVMKIINSFNPDFISNFLKTEKMI IHYKKIATGSLMEKQRVHFSEFKNI IKKFP TNKEQQKIGDFFSKLDRQIELQVQKLELLQQQ

>NOVEL 4

MTEQINTPELRFPEFKNEWSYDLVSDVVTNKS KKFDPKKEEAKKDIELDSIEQNTGRLLD TYISNDFTSQKNKFNKGNVLYSKLRPYLNKYFYATIDGVC
SSEIWLNTLNKDVLANKFLYFIQTNRFSSV TNKSAGSKMPRADWELVKNIRLYKGSIEEQEKIGYFFSKLDRQIELEEKLELLEQQQ

1
2 **The amino acid sequences in FASTA format of the second TRD with**
3 **its letter code, DNA target (5' to 3') and its methylation site**
4 **underlined. The TRD labelled as NOVEL 3 was found once all of the**
5 **other TRDs had been analysed but is included in this list for**
6 **completeness.**
7

8
9 >D ATC
10 KKGYMQKIFSQELRFKDENGEDYPHWENSKIEKYLKERNERSDKGQMLSVTINSGIIFSELDRKDNSSKNKSNYKVRKNDIAYNSMRMQGASGKSNY
11 NGIVSPAYTVLYPTQNTSSLFIFYGKFKTHRMIHKFKINSQGLTSDTWNLKYQLKNNINIDI PVLEEQEKIGDFFKMDILISKQKIKIEILEKEKQSFLO
12 KMFL
13 >E TCAY
14 KKGYMQKIFSQELRFKDENGNDYPEWEETTIKEIAQINXGKKDTKDAITNGSYDFYVRSPIVYKINTFSYEGEAILTVGDGVGVGVFHYVNGKFDYHQR
15 VYKISDFKNYYGLLLFFYYFSQNFLETKKYSAKTSVDSVRKDMIANKVPRPIYIEQKKIGQFIKRVNDNKTIKQKQVIELLQKQKALLQKMF I
16 >F TAA
17 KKGYMQKIFSQELRFKDEEGKDPDWKSKSIQEIFENKGGTALETEFNFQDGNKVISIGSYSINSTYNDQNIQIRVNNKKNKTEKYILSKGDLAMVLNDKTKD
18 GKIIIGRSIFIDKDNQYIYNQRTERLIPFAENDNKFLWFLMNTDLIRNKIKGMMQGATQVYINYSIKLISIQPLPLEEQKIRGFLEVLVSGITTKQLHKI
19 DQLKERKKAFLQKMF I
20 >G ACA
21 KKGYMQKIFTQELRFKDENGEYPEWENKFIKDIFIFENRRRPITSSLRKGLYPYGGATGIDYVVDYLFNNEERLLIGEDGAKWQGFETSSFIANGQ
22 YWVNNHAHVKSNDHNLFFMNYLNFKELRAFVTGNAPAKLTHANLCNINLKIPLTEQDKVSALLKSIDNKMNNQMNRIELKERRKELLQKMF I
23 >H TAC
24 KKCYYQKIFSQELRFKDEEGNYKGNKQKLDVLEFSNKRTINENYVPLTSSRQGLILQSDYYKDRKTFAESNIGYFILPKNHITVRSRSDDGIFKFN
25 LNLMDVGIISKYYPVFKGIDANQYYLTLHLNYQLKKEYIKYATGTSQVLVLSQKDLQNIKTKLPSYEEQKIGDFSEIDRLVEKQSSKVGRLKVRKEL
26 LQKMFV
27 >I YTCA
28 KKGYMQKIFSQELRFKNENNDYPDWERIKFFDVIDKVIDFRGRTPKKLNMEWSDEGYLALSAVNVKKGYIDFNVEAKYGNLDLYTRWMRGNELYKQVVL
29 FTTEAPMGNAQVDPDNKGYILSQRITAFNSNEKITDNFLASLLSSENVYNDLLKLCGATAGVSVQKNLNRLYVTIPHSISEQEEIAEFFRKINQLVELQ
30 KYKIEHTKSQKQVFLQKMF I
31 >K CGA
32 KKGYMQKIFSQELRFKDENGNDYPKWEKKIEDIASQVYGGTPTNTKIKFVWNGDI PWIQSSDVKVNLDLILQOCNKFISKNSIELSSAKLIPANSIAIVT
33 RVGVGKLCLEVFYDAYSQDFLSLSSLYDKLYSLYLLYTMKKISANLQGTISIKGITKKELLDSIIKIPHNLEEQQKIGDLFYKIDKYISFNKCKIEMLK
34 SLKQGLLKKMF I
35 >L TTA
36 KKGYMQKIFSQELRFKDENGNDYPNVRTIELKNILENIVDNRGKTPDNAPSEKYPLLEVNALGYRPAIKVSKFVSENTYNNWFREHLKENDILFSTVVG
37 NTGIVSLMDNYKAVIAQNIQVGLRVNNNNLPSFIYYMLSYKGNQKKIKRIQMGAVQPSVKVSQFKFIKYLVPKIKDEQEKVAKLLIEIDKLVNKQLIKIELL
38 QQRKALLKSMFI
39 >P AGG
40 KKGYMQKIFSQELRFKDESGNDYPDWEKELGEVADRVIRKKNKSFESKPLTISGQLGLIDQTEYFYSKSVSSKNLENYTLIKNGEFAYNKSYNGYPLGA
41 IKRLTRYDSGVLSSLYICFSIKSEMSKDFMEAYFDSTHWYREVSGIAVEGARNHGLLNISVNDFFTTILIKYPSLEEQRKIGDFFIKLDRQIELEEQLLEL
42 LQQRKALLKSMFI
43 >Q ACAY
44 KKGYMQKIFSQELRFKDENGEDYSEWEERRFADIFKFNKLRKPIKENLRVKSYPYGGATGIDYVDDFIFDGNVLLIGEDGANIITRSAPLVYLVNGK
45 FVWNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAPKLNINQNLKI INVVI STNLEEQQKIGSFLSKLDRQIDLEEQLLELQQRKALLKSMFV
46 >S GCA
47 KKGYMQKIFSQELRFKDENGNDYPDWTNERLGEVTTVTMGQSPKSVNYTDNSNDTVLIQGNADIENGLINPRIYTREVTKLIQKDEIILTVRAPVGLAM
48 AQINACIGRVCSIKGDKFLYFLEWFATQNKWIRFSQGSTFESISGNDIRNIHIKIPVEDERTKI ILLNSLDVLSNKTDLKIQNLKQRKQSLQKIFV
49 >U GAY
50 KKGYMQKIFSQELRFKDENGEDYDWEVTTIQNITKYTSSKSSNQYADKDNKSGYPYDAVQEQIGKDSNYDIEESYISILKDAGVGRNLNLRPGKSSVI
51 GTMGIYQSNNDIEFLYRMRVVDKFKYIIGSTIPHLYFKDYSKETLYIPSSIQEQAKIGMFISNLDKLIENKLNKLNCKLQKQGLLQSMFI
52 >W CRAA
53 KKGYMQKIFSQELRFKDENGNDYPDWEKQLGELSQIVRGASPRPIKDPKWFNKESDIGNLWLRISDVNTNQNKIYHLEQKLSIEGQEKTRVLVTTLLLSI
54 AASIGKPVMMNFVKTGVHDGFLIFLKPKNLFFMYWLEFYFKDKWSKYQPGSQVNLNSEIVKSQTLNMPNSNHEQEKVGFQFNREKLIELQEKIMYIKR
55 CKQVLLQKMF I
56 >Y CTA
57 KKGYMQKIFSQELRFKDENGNDYPDWEKKLKEIACVYTGNTSPKKNENIYWNKGEYVWVTPDINNKNIEYSENKLTQEGYKARQLPENTLLVTCIAS
58 IGKNAILRKQSGCNQINAVVPFENINIDYLYIISDSLSTFMKSIAGKTATQIVNKNFTFENLEIYLAPFEEQNKIADLISSELELIEKQASKLIKMKSRK
59 QGMLQIMFI
60 >a* GAA
KKGYMQKIFSQELRFKDENGNDYPWENKRIEDIANVNGKFTPTSTNNNEYWNNNDKNWLSIAGMNOQYLYKGNKGISKDAAKNYMKVKNNDTLIMSFKLTI
GKLAIVKAPLYTNEAICHFIWKVNKINTEFIYYLNSLNISTFGVQAVKGVTLNNDINSIIIVKLPNEEQNI IAKFLLVEDKTVNNQLVKTLLKQRKK
GLLQRMFV
>c* GAY
KKGYLQKIFSQELRFKDENGNDYPWRFARFKDFMYKPINIRPAINISKSELLTVKLHCKGIEKANINRVLKLGGATNYKRFEGQFIYKQNFNFNGAFDI
VPKFDGLYSSSDVPAFEINTEKIEPNYFISYISRPSFYKSKKEYSTGTGSKRIHENTVLFNLSLHPLCNEQLKIASFVCFNLNRIELLERKIYLIKQK
QALLQKMF I
>d* CYAA
KKGYMQKIFSQELRFKDENGNDYPWENVMKQVLKDKTEGKRGPFPGALKKDIFVESGYAVYEQRNAIYDISNFRYINENKYKEMQSFVQPNDIIM
SCSGTIGRLALIPHNYTKGIINQALIRFRTNHKIRSEFFLI FMRSNQMQRKILEANPGSAITNLVVPKELKLI PFPLPVKFEQDKISQFIHI INRRIEQS
EKKIESLKNRQGFQKLFV
>f* GAAY
KKGYMQKIFSQELRFKDENGEDYDWEKELGDIQSMYIGIGASATRFDSKNYIIRITDIDEKSRKLNQNLTPDELNNKYKLRNDILFARTGASTG
KSYIHKEEKDIYNYFAGFLIKFKINEQNSPLFIYQFTLTSKFNKWKVMSVRSQPGINSEYAKLPLVLPNKLEQQKIAKFLDRDRQIELEKQKIEI
LQQQKGLLQSMFI
>NOVEL 3
KKGYMQKIFAQELRFKDENGNDYPDWVTKKLDIGKAMNKRIYKNETTENGEIPFYKIGNFGKNADFTITREKFDEYKEYYPNVGDILISASGSIGR
TIEYTGEDAYYQDSNIVLWLNHNDEVINKYLYFYKIVKWSGIEGTTIKRLYNKLNLTKIELPTVEEQYKMANFLSKLDKIIDIQIEKIELLKQRKQGLL
QKMFV

**SUPPLEMENTARY INFORMATION FOR MATERIALS AND METHODS SECTION
 "CONSTRUCTION OF FURTHER MTASES WITH FURTHER COMBINATIONS OF TRDS
 USING SYNTHETIC GENES."**

The DNA sequence is followed by the predicted amino acid sequence for each TRD. Some synthetic sequences encoded two TRDs.

CC15 TRD O

CCCGGGTTTGAAGGCGAATGGGAGGAAAAAACTGGGTGAAGTTGGCACCTTTACCAGCGGTGGC
 ACTCCGCTGAAAAGCAAAGCGAATATTGGAATGGTGATATTCCGTGGATTACCACAGGCGATATT
 CATAACATTAACCGCGAAAACATCACCAACTTTATCACCGAAAAAGGCCTGAATGAAAGCAGCGCA
 AAACTGATTACCAATGAAGCAATTCTGATTGCCATGTATGGTCAGGGTAAAACCCGTGGTATGAGC
 GCCATTCTGAATTTTGAAGCAACCACCAATCAGGCCTGTGCAATTTATCAGACAAACCAGAACATC
 AACTTCGTGTTCCAGTATTTCCAGAACTGTATGAATTTCTGCGTAGCCTGAGCAATGAAGGTAGC
 CAGAAAAATCTGAGCCTGAGCCTGCTGAAAGAAATTACCCTGAATTATCCGAACGAGCAAGAACAG
 AAAAAATCGGCGATTTCTTCAGCAAACCTGGATCGTCAAATTGAATTAGAAGAACAGAAG

CC15 TRD O

PGFEGEWEEKKLGEVGTFTSGGTPLKSKSEYWNGDIPWITTTGDIHNIKRENI TNFITEKGLNESSA
 KLITNEAILIAMYGQGKTRGMSAILNFEATTNQACAIYQTNQNINVFVQYFQKLYEFLRSLSNEGS
 QKNLSLSLLKEITLNPNEQEQQKIGDFFSKLDRQIELEEQK

CC51 TRD P

CAAATTGAATTAGAAGAACAGAAGCTGGAAGCTGTTTCAGCAGCAGAAAAAAGGCTATATGCAGAAA
 ATCTTTAGCCAAGAGCTGCGCTTTAAAGATGAAAGCGGTAATGATTATCCGGATTGGGAAGAAAAA
 GAACTGGGTGAAGTTGCAGATCGTGTGATTCGTAAAAACAAAACTTTGAAAGCAAAAAACCGCTG
 ACCATTAGCGGTCAGCTGGGTCTGATTGATCAGACAGAATATTTTCAGCAAAAGCGTGAGCAGCAAA
 AACCTGGAAAACATAACCTGATTAACAAACGGCGAGTTTCGCTATAACAAAAGCTATAGCAATGGT
 TATCCGCTGGGTGCAATTAACGTCTGACCCGTTATGATAGCGGTGTTCTGAGCAGCCTGTATATT
 TGCTTTAGCATCAAAGCGAGATGAGCAAAGATTTTCATGGAAGCCTATTTTGATAGCACCCATTGG
 TATCGTGAAGTTAGCGGTATTGCAGTTGAAGGTGCACGTAATCATGGTCTGCTGAATATTAGCGTG
 AACGATTTTTTACCATCCTGATCAAATATCCGAGCCTGGAAGAACAGCGTAAAATCGGTGATTTT
 TTCATTAAACTGGATCGCCAGATTGAGCTGGAAGAACAAAACTGGAAGTCTGCAACAGCGCAAA
 AAAGCACTGCTGAAAAGTATGCTGATCCCCGGGGGATCCGATCGATC

CC51 TRD P

QIELEEQKLELFQQQKKGVMQKIFSQELRFKDESGNDYPDWEEKELGEVADRVIRKNKNFESKKPL
 TISGQLGLIDQTEYFSKSVSSKNLENYTLIKNGEFAYNKSYNNGYPLGAIKRLTRYDSGVLSLYI
 CFSIKSEMSKDFMEAYFDSTHWYREVSGI AVEGARNHGLLNI SVNDFFTILIKYPSLEEQRKIGDF
 FIKLDRQIELEEQKLELLQQRKKALLKSMIL

CC72-1 TRD R + CC59-1 TRD Q

CCCGGGTTTGAAGGCGAATGGGAGGAAAAAACTGGGTGAAGTTGGCCAAAATCTATGATGGCACC
 CATCAGACCCCGAAATATAACCAATGAAGGTATCAAATTTCTGAGCGTGGAAAACATCAAACCCCTG
 AATAGCAGCAAATACATTAGCGAAGAAGCCTTCGAGAAAGAATTCAAATTCGTCCGGAATTTGGC
 GATATTCTGATGACCCGTATTGGTGATATTGGCACCCCGAATATTGTTAGCAGCAATGAAAAATTC
 GCCTACTATGTTAGCCTGGCACTGCTGAAAACCAAAAATCTGAACAGCTACTTCTGAAAAACCTG
 ATTCTGAGCAGCAGCATTGAGAAATGAACTGTGGCGTAAAACCCCTGCATGTTGCATTTCCGAAAAA
 ATCAACAAAAACGAGATCGGCAAAATCAAATCAACTACCCGAAAAAACAAGAACAGCAGAAAAATC
 GGTGAGTTTTTTCAGCAAACCTGGATCGCCAAATTAATTAAGAAGAACAGAAGCTGGAAGTCTGCAA
 CAGCAGAAAAAAGGTTATATGCAGAAAATCTTCAGCCAAGAGCTGCGCTTTAAAGATGAAAATGGT
 GAAGATTATAGCGAGTGGGAAGAACGTCGTTTTGCCGATATTTTCAAATTTTCAACAAACTGCGC
 AAACCGATCAAAGAAAATCTGCGTGTAAAGGCAGCTATCCGTATTATGGTGCAACCGGCATTATT
 GATTATGTGGATGATTTTTATCTTCGATGGCAACTATCTGCTGATTGGCGAAGATGGTGCAACATT
 ATTACCCGTAGCGCACCGCTGGTTTTATCTGGTTAATGGTAAATTTTGGGTGAACAACCATGCCCAT
 ATTCTGAGTCCGCTGAATGGTAATATTCAGTATCTGTATCAGGTTGCCGAAGTGGTGAAGTATGAA

1
2 AAATACAATACCGGCACCGCACAGCCGAAACTGAACATTCAGAATCTGAAAATTATCAACGTGGTG
3 ATCAGCACCAATCTGGAAGAACAGCAAAAAATTGGTAGCTTCTGAGCAAACCTGGATCGTCAGATT
4 GACCTGGAAGAACAAAACTGGAAGTCTGCAACAACGTAAAAAAGCACTGCTGAAAAGCATGTTC
5 GTGCCCGGGGATCCGATCGATC
6

CC59-1 TRD Q

7
8 QIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGEDYSEWEERRFADIFKFHNKLRKPIKENLRV
9 KGSYPYYGATGIIDYVDDFIFDGNLLIGEDGANIITRSAPLVYLNGKFWVNNHAHILSPLNGNI
10 QYLYQVAELVNYEKYNTGTAQPKLNIQNLKIINVVIISTNLEEQQKIGSFLSKLDRQIDLEEQKLEL
11 LQQRKALLKSMFV
12

CC72-1 TRD R

13
14 PGFEGEWEEKLGEVAKIYDGTHQTPKYTNEGIFLSVENIKTLNSSKYISEEAFEKEFKIRPEFG
15 DILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYFLKNLILSSSIQNELWRKTLHVAFPKK
16 INKNEIGKIKINYPPKQEQQKIGQFFSKLDRQIELEEQK
17

CC75-1 TRD T and TRD U

18
19 CCCGGGTTTTGAAGGCGAATGGGAGGAAAAAGAACTGGGCGAAATCTTTCAGATTATTAGCGGTAGC
20 ACACCGCTGAAAAGCAACAAAGAATTTTATGAGAACGGCAACATCAACTGGGTAAAACCACCGAT
21 CTGAATAATAGCAAAGTGACCCATAGCAAAGAAAAATCACCGAGTATGCAATGAAAAGCCTGAAA
22 CTGAAACTGGTGCCGAAAAATAGCGTTCTGATTGCAATGTATGGTGGCTTTAATCAGATTGGTCGT
23 ACCGGTCTGCTGAAAATTGATGCAACCATTAATCAGGCAATTAGCGCACTGCTGATGAATCATGAA
24 ACCAACCCGGAATTTATTTCAGGCCTTTCTGAATTATCAGGTGAAAGGTTGGAAACGTTATGCAGCA
25 AGCAGCCGTAAAGATCCGAATATCACCAAAAAAGATATCGAACAGTTCAAAGTGCCGTACGTGAGC
26 ATTAATGAACAGCAGAAAAATTGGCGAGTTTTTTAGCAAAATCGATCATCAAATGAAATTAGAAGAA
27 CAGAAGCTGGAAGTCTGCAACAGCAGAAAAAGGTTATATGCAGAAAATCTTCAGCCAAGAGCTG
28 CGCTTTAAAGATGAAAATGGTGAAGATTATCCGGATTGGGAAGTTACCACCATTGAGAACATTACC
29 AAATACACCAGCAGCAAAAAAGCAGCAATCAGTATGCCGATAAAGACAACAGCAAAGGTTATCCG
30 GTTTATGATGCCGTTCAAGAAATTGGCAAAGATAGCAACTATGACATCGAAGAGAGCTATATCAGC
31 ATTCTGAAAGATGGTGCCGGTGTGGTCTGCTGAATCTGCGTCCGGGTAAAAGCAGCGTTATTGGC
32 ACCATGGGTTATATTCAGAGCAACAACGTGGATATCGAGTTCCTGTATTATCGTATGAAAGTGGTG
33 GACTTCAAAAATACATTATCGGTAGCACCATTCCGCACCTGTATTTCAAAGATTATAGCAAAGAA
34 ACCCTGTACATTCCGAGCAGCATTCAAGAACAGGCAAAAAATTGGTATGTTTCATCAGCAACCTGGAT
35 AAAGTATCGAGAACAAAAACCTGAAACTGAACTGTCTGAAACAACCTGAAACAGGGATTGCTACAA
36 TCTATGTTTTATTCCCGGGGGATCCGATCGATC
37

CC75-1 TRD T

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39 PGFEGEWEEKELGEIFQIISGSTPLKSNKEFYENGNINWVKTDLNNSKVTHSKEKITEYAMKSLK
40 LKLVPKNSVLIAMYGFNQIGRTGLLKIDATINQAI SALLMNHETNPEFIQAFLNYQVKGWKRYAA
41 SSRKDPNITKKDIEQFKVPYVSINEQQKIGEFFSKIDHQIELEEQK
42

CC75-1 TRD U

43
44 QIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGEDYDWEVTTIQNITKYTSSKSSNOYADKD
45 NSKGYPVYDAVQIEIGKDSNYDIEESYISILKDGAGVGRNLNLRPGKSSVIGTMGYIQSNNVDIEFLY
46 YRMKVVDFFKYYIIGSTIPHLFYFKDYSKETLYIPSSIQEQAKIGMFI SNLDKLIENKNLKNLCLKQL
47 KQGLLQSMFI
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CC75-2 TRD V

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54 CCCGGGTTTTGAAGGCGAATGGGAGGAAAAAGAACTGCGTGAAGTGGCAATCCGAAAGATAAATAC
55 AGCTATAACCGGTGGTCCGTTTGGTAGCGATCTGAAAAAAGCGATTATACCACCGATGGCATTAG
56 ATTATTCAGCTGCAGAATATTGGTGACGGCTATTTCTATAACAGCAACAAAGTGTTCACAGCAAC
57 GAAAAAGCCGAAGTCTGAAAAGCTGTAATGTTTTTCCGGGTGATATTGTGATTGCCAAAATGGCA
58 GATCCGATTGCACGTGCCGCAATTGTTCCGGATAATAACATTGGTAAATACCTGATGGCCAGTGAT
59 GGTATTCGTCTGAGCGTTGATACCGTTTCAATTTAACACCAAATTTGTGCTGGAATGCATCAACCGT
60 AAAAGCTTTCGTAAAAAAGTCGAGGATAATAGCAGCGGTAGCACCCGTATGCGTATTGGTCTGAGT
ACCCTGGGTAGCCTGACCTGAAAACCACCACCTGAAAGAACAGCAGAAAAATTGGTCAGTTTTTC

1
2 AGCAAACCTGGATCGTCAAATTGAATTAGAAGAACAGAAG

3 **CC75-2 TRD V**

4 PGFEGEWEEKELRELRNPKDKYSYTGPFSGDLKSDYTTDGIQIIQLQNIQDGYFYNSNKVFTSN
5 EKAEVLKSCNVFPGDIVIAKMADPIARAAIVPDNNIGKYLMSDGIKLSVDTVHFNTKRVLECI
6 KSFRRKVEDNSSGSTRMRIGLSTLGLSLTLKTTTLKEQQKIGQFFSKLDRQIELEEOK
7

8
9 **CC75-2 TRD W**

10 CAAATTGAATTAGAAGAACAGAAGCTGGAACCTGCTGCAACAGCAGAAAAAAGGTTATATGCAGAAA
11 ATCTTCAGCCAAGAGCTGCGCTTTAAAGATGAAAATGGTAACGATTATCCGGATTGGGAAGAAAA
12 CAGCTGGGTGAACTGAGCCAGATTGTTCTGTTGGTGCAAGTCCGCGTCCGATTAAAGATCCGAAATGG
13 TTTAACAAAGAAAGCGATATTGGTTGGCTGCGCATTAGTGATGTTACCAATCAGAATGGCAAAATC
14 TATCATCTGGAACAGAACTGAGCATCGAAGGTCAAGAAAAAACCCGTGTTCTGGTTACCACCCAT
15 CTGCTGCTGAGCATTGCAGCAAGCATTGGTAAACCGGTTATGAACTTTGTGAAAACCGGTGTGCAT
16 GATGGCTTTCTGATTTTTCTGAAACCGAAATCAACCTGTTCTTTATGTACTATTGGCTGGAATAT
17 TTCAAAGATAAATGGTCCAAATATGGTCAGCCTGGTAGCCAGGTTAATCTGAATAGCGAAATGTT
18 AAAAGCCAGACCCTGAATATGCCGAGCAATCATGAACAAGAAAAAGTGGGCCAGTTTTTTAACCGC
19 AACGAAAAACTGATTGAACTGCAGCAAGAGAAAAATCATGTATATCAAACGTTGCAAACAGGTGCTG
20 CTGCAAAAAATGTTTATTCCCGGGGGATCCGATCGATC
21

22 **CC75-2 TRD W**

23 QIELEEOKLELLLQQQKKGVMQKIFSQELRFKDENGNDYPDWEEKQLGELSQIVRGASPRPIKDPKW
24 FNKESDIGWLRISDVTNQNNGKIYHLEQKLSIEGQEKTRVLVTTTHLLLSIAASIGKPVMMNFVKTGVH
25 DGFLIFLKPKNLFFMYWLEYFKDKWSKYGQPGSQVNLNSEIVKSQTLNMPSPNHEQEKVQGFNR
26 NEKLIELQQEKIMYIKRCKQVLLQKMF I
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30 **CC80-1 TRD X and TRD Y**

31 CCCGGGTTTTGAAGGCGAATGGGAGGAAAAACAGTTTTGCCGACTTCACCAAAATTAACCAGGGTCTG
32 CAGATTGCCATTAATGAACGTAAAACCGAATATAGCCCTGAGCTGTATTTCTATATCACCAACGAA
33 TTTCTGCGTCCGAATAGCCAGACCAAATATTTTCATTGAAAATCCGCCTCAGAGCGTGATTGCCAAC
34 AAAGAAGATATTCTGATGACCCGCACCGGTAATACCGGCAAAGTTGTTACCAATGTTTTTGGTGCC
35 TTCCACAACAACCTTTTTCAAATCAAATTCGATAAAAACCTGTATGATCGCCTGTTTCTGGTTGAA
36 GTTCTGAACAGCAGCAAAAATCCAGAACAAAATTCAGCCTGGCAGGTAGCAGCACCATTCCGGAT
37 CTGAATCATAGCGATTTCTATAGCATTAGCAGCAGCTATCCGCTGCTGCGCGAACAGCAAAAAATT
38 GGCAAATTTCTTAGCAAACTGGATCGTCAAATTGAATTAGAAGAACAGAAGCTGGAACCTGCTGCAA
39 CAGCAGAAAAAAGGTTATATGCAGAAAATCTTCAGCCAAGAGCTGCGCTTTAAAGATGAAAATGGT
40 AACGATTATCCGGATTGGGAGAAAAAAAACCTGAAAGAAATTGCCTGCGTGTATACCGGTAATACC
41 CCGAGCAAAAAAGAAAACATCTATTGGAACAAAGGCGAGTATGTTTGGGTTACCCCGACCGATATT
42 AACAAACAGCAAAAACATTTATGAAAGCGAAAACAACTGACCCAAGAAGGCTACAAAAAAGCACGT
43 CAGCTGCCGGAAAAATACCTGCTGGTTACCTGTATTGCAAGCATTGGTAAAAATGCCATTCTGCGT
44 AAACAGGGTAGCTGTAATCAGCAGATTAATGCAGTTGTGCCGTTTGAGAACATCAACATCGATTAT
45 CTGTATTATATCAGCGATAGCCTGAGCACCTTCATGAAAAGCATTGCAGGTAAAACCGCAACCCAG
46 ATTGTGAACAAAAACACCTTTGAAAACCTGAAATTTACCTGGCACCTTTTGAGGAACAGAACAAA
47 ATTGCAGATCTGATTAGCAGCCTGGAAGAACTGATTGAAAAACAGGCAAGCAAACTGATCAAAATG
48 AAAAGCCGCAAACAGGGCATGCTGCAGATTATGTTTATTCCCGGGGGATCCGATCGATC
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52 **CC80-1 TRD X**

53 PGFEGEWEEKQFADF TKINQGLQI AINERKTEYSPELYFYITNEFLRPNSQTKYFIENPPQSVIAN
54 KEDILMTRTGNTGKVVTVNFVGFHNNFFKIKFDKNLYDRFLVEVLNSSKIQNKILSLAGSSTIPD
55 LNHSDFYSISSYPLLREQQKIGKFFSKLDRQIELEEOK
56

57 **CC80-1 TRD Y**

58 QIELEEOKLELLLQQQKKGVMQKIFSQELRFKDENGNDYPDWEKKLKEIACVYTGNTPSKKENIYW
59 NKGEYVWVTPDINNSKNIYESENKLTQEGYKKARQLPENTLLVTCIASIGKNAILRKQSGCNQOI
60 NAVVPFENINIDYLYIISDSLSTFMKSIAGKTATQIVNKNTFENLEIYLAPFEEQNKIADLISSLE
ELIEKQASKLIKMKSRKQGMLOIMFI

CC80-2 TRD Z + CC72-2 TRD S

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CCCCGGTTTTGAAGGCGAATATTCTCTGGATATTTTTGGTAATCTGGCCACCAACAAAAGCGAAAA
TTCAATCCGCAGAATGAAAACGCCAGCATTGATATTTGAACTGGATTGCATTGAACAGAATACCGGT
CGTCTGATCAAAATCTATAACAGCAAAGAATTTAGCAGCCAGAAAAACAAATTTAACCCGCAGAAC
GTGCTGTATGGTAAACTGCGTCCGTATCTGAACAAATATTACTTCACCAAAAAAAGTGGTGTGTGC
AGCAGCGAAATTTGGGTTCTGAAAAGCACCAAAGAAGATAAACTGCTGAACCTGTTCTGTACTAT
TTCATTCAGACCAAACGCTATAGTGATGTTGCAAGCAAAGCGCAGGTAGCAAATGCCTCGTGCA
GATTGGGGTCTGATTGAAAATATTCGTGTGTATTTTCCGGAACCTGTGCGAACAGCAGAAAATTGGT
CAGTTTTTTTAGCAAACCTGGACCGTCAAATTTGAATTAGAAGAACAGAAGCTGGAACCTGCTGCAACAG
CAGAAAAAAGGTTATATGCAGAAAATCTTCAGCCAAGAGCTGCGCTTTAAAGATGAAAATGGTAAC
GATTATCCGGACTGGACCAATGAACGTCTGGGTGAAGTTACCACCGTTACCATGGGTGAGAGCCCG
AAAAGCGTGAATTATACCGATAATAGCAATGACACCGTTCTGATTACAGGGTAATGCCGATATTGAA
AACGGTCTGATTAATCCGCGTATCTATAACCGTGAAGTGACCAAACCTGATTCAGAAAGATGAGATT
ATTCTGACCGTTCGTGCACCGGTTGGTAAACTGGCAATGGCACAGATTAATGCATGTATTGGTCGT
GGTGTGTTGCAGCATTAAAGGCGATAAATTTCTGTATTATTTCTGGAATGGTTCGCCACCCAGAAT
AAATGGATTCTGTTTTAGCCAGGGTAGCACCTTTGAAAGCATTAGCGGTAATGATATTCGCAACATC
CATATCAAATCCCGGTTGAAGATGAACGCACCAAATTTATCAAACCTGCTGAATAGCCTGGATGTG
CTGAATTCAAAACCGATCTGAAAATCCAGAATCTGAAACAGCGTAAACAGAGCCTGCTGCAAAAA
ATCTTTGTGCCCGGGGATCCGATCGATC

CC80-2 TRD Z

PGFEGEYSLDIFGNLATNKSEKFNPNQENASIDIELDCIEQNTGRLIKIYNSKEFSSQKNKFNPNQ
VLYGKLRPYLNKYYFTKKSVCSSSEI WVLKSTKEDKLLNLFLLYYFIQTKRYSVASKSAGSKMPRA
DWGLIENIRVYFPELCEQQKIGQFFSKLDRQIELEEQK

CC72-2 TRD S

QIELEEQKLELLLOQQKGYMQKIFSQELRFKDENGNDYPDWTNERLGEVTTVTMGQSPKSVNYTDN
SNDTVLIQGNADIENGLINPRIYTREVTKLIQKDEIILTVRAPVGLAMAQINACIGRGVCSIKGD
KFLYYFLEWFATQNKWIRFSQGSTFESISGNDIRNIHIKIPVEDERTKIKLLNSLDVLSKTDLK
IQNLKQRKQSLLOKIFV

CC93-2 TRD b*

CCCCGGTTTTGAAGGCGAATGGGAGGAGAAAAAACTGGAAGATACCCTGGAATTCATTAAAGATGGC
ACCCATGGTACACATGAAAATGTTAATAATGGTCCGTGGCTGCTGAGCGCCAAAAACATTA AAAAC
AACAAAATCATCATCAGCAGCGACGATCGCAAAATTAGCGAAAGCGATTACAAAAAATCTACAAA
AACTATAAACTGAAAAAGGCGATCTGCTGCTGACCATTGTTGGCACCATTGGTTCGTGCAGCAATT
GTTAAAAATCCGAACAATATTGCCTTTCAGCGTAGCGTTGCAATCCTGAAAACCAAAGCAACCTAT
GATGTGGGCTTTATCTTTTCAGCTGTTCCAGACCAAATACTTTAAAAACCTGCTGCTGCGTAAACAG
GTTGTTAGCGCACAGCCTGGTCTGTATCTGGGTGATATTCGTAAAATCAAATCAGCATTACCAAC
ATCATCGAAGAACAGCGCAAAATCGGTATCTTTTTCAGCAAACCTGGATCGTCAAATTGAATTAGAA
GAACAGAAG

CC93-2 TRD b*

PGFEGEWEEKLEDTFLEFIKDGTHGTHENVNNGPWLLSAKNIKNNKIISSDDRKISESDYKKIYK
NYKLEKGDLLLITIVGTIGRAAIVKNPNNIAFQRSVAILKTKATYDVGFIFQLFQTKYFKNLLLRKQ
VVSAQPGLYLGDIRKIKISITNIEEQRKIGIFFSKLDRQIELEEQK

C93-3 TRD a*

CAAATTGAATTAGAAGAACAGAAGCTGGAACCTGCTGCAACAGCAGAAAAAAGGTTATATGCAGAAA
ATCTTCAGCCAAGAGCTGCGCTTTAAAGATGAAAATGGTAACGATTATCCGGAATGGGAAAACAAA
CGCATTGAAGATATTGCCAATGTGAACAAAGGTTTTACCCCGAGCACCAACAATAACGAATATTGG
GATAACAACGATAAAAACTGGCTGAGCATTGACAGGCATGAATCAGAAATATCTGTATAAAGGCAAC
AAAGGCATCAGCAAAGATGCAGCCAAAAACTATATGAAAGTGAAAAACGACACCCTGATCATGTCC
TTTAAACTGACCATTGGTAAACTGGCGATTGTTAAAGCACCGCTGTATACCAATGAAGCCATTTGT
CATTTTATCTGGAAAGTGAACAAAATCAACACCGAGTTCATCTACTATTACCTGAACAGCCTGAAC
ATTAGCACCTTTGGTGTTCAGGCAGTTAAAGGTGTTACCCTGAATAACGATAGCATCAACAGCATT

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ATTGTGAAACTGCCGAATGAAGAGGAACAGAACATTATCGCAAAATTTCTGCTGGAAGTGGACAAA
ACCGTTAATAATCAGCTGGTAAAACCAAACCTGCTGAAACAACGTAATAAAGGCCTGCTGCAGCGT
ATGTTTGTTCCTCCGGGGGATCCGATCGATC

CC93-3 TRD a*

QIELEEQKLELLQQQKKGMYMOKIIFSQELRFKDENGNDYPEWENKRIEDIANVKNKGFPTSTNNNEYW
DNNDKNWLSIAGMNQKYLYKGNKGISKDAAKNYMKVKNDTLIMSFKLTIGKLAIVKAPLYTNEAIC
HFIWKVNKINTEFIYYLNSLNIISTFGVQAVKGVTLNNSINSIIVKLPNEEEQNI IAKFLLEVVK
TVNNQLVKTKLLKQRKKGLLQRMFV

CC873 TRD e* + CC97 TRD c*

CCCGGGTTTTGAAGGCGAATGGGAGGAAAAATCGATCAGCAGCTTTCTGAAAGAAAGCAAATCAAA
GGTAGCAATGGTAGCCATGCAAAAAAACTGACCGTTAAACTGTGGGGTAAAGGTGTTGTTCCGAAA
AAAGAAACGTTTAAAGGCAGCGATAACACCCAGTATTACAAACGTAAGCAGGTCAGCTGATGTAT
GGCAAACCTGGATTTTCTGAATTGCGCCTTTGGTATTGTTCCGGATAGCCTGAATAACTATGAAAGC
ACCATTGATAGCCCCGAGCTTTGATTTTATTAATGGCGATAGCAAATTTCTGCTGGAACGCATTAAA
CTGAAAAGCTTCTACAAAAAATTCGGCGATATTGCAAATGGCAGCCGTAAAGCAAACGTATTAAT
CAGGATACCTTTCTGAGCTGCCGGTTTTTGCACCGAAATATGATGAACAGCTGCGTATTGGTGAA
TTTTTTCAGTAAACTGGATCGTCAAATTGAATTAGAAGAACAGAAGCTGGAACCTGCTGCAACAGCAG
AAAAAAGGTTATCTGCAGAAAATCTTTAGCCAAGAGCTGCGCTTTAAAGATGAAAACGGTAATGAT
TATCCGGAATGGCGTTTTGCCCCGTTTCAAAGATTTTATGTACAAACCGATTAATATCCGTCCGGCA
ATCAACATTAGCAAAAAGCGAACTGCTGACCGTTAAACTGCATTGCAAAGGTATTGAAAAGCCAAC
ATTAACCGTGTGCTGAAACTGGGTGCAACCAATTATTACAAACGTTTTGAAGGCCAGTTTATCTAT
GGCAAACAGAACCTTTTTTAAACGGTGCCTTTGATATCGTGCCGAAAAAATTCGATGGTCTGTATAGC
AGCAGTGATGTTCCGGCATTGAAATCAATACCGAGAAAATTGAGCCGAACCTACTTCATCAGCTAT
ATTAGCCGTCCGAGCTTCTATAAAAGCAAAGAGAAATATAGCACCGGCACCGGTAGCAAACGTATT
CATGAAAATACCGTGCTGAACTTTAGCCTGCATCTGCCGTGTCTGAATGAACAGCTGAAAATTGCA
AGCTTTGTGTGCTTTCTGAACCGTAAAATTGAACTGCTGGAACGCAAATCTATCTGATCAAAAAA
CAGAAACAGGCCCTGCTGCAGCAAATGTTTATTCCCGGGGGATCCGATCGATC

CC873 TRD e*

PGFEGEWEEKSISSEFLKESKIKGSNGSHAKKLTVKLWKGKGVVPPKETFKGSNDTQYYKRKAGQLMY
GKLDFLNCAFGIVPDSLNNYESTIDSPSDFDINGDSKFLLERIKLSFYKKGFDIANGSRKAKRIN
QDTFLSLPVFAPKYDEQLRIGEFFSKLDRQIELEEQK

CC97 TRD c*

QIELEEQKLELLQQQKKGMYLQKIFSQELRFKDENGNDYPEWRFARFKDFMYKPINIRPAINISKSE
LLTVKLVHCKGIEKANINRVLKLGATNYYKRFEGQFIYKQNFNGAFDIPKFDGLYSSSDVPAF
EINTEKIEPNYFISYISRPSFYKSKEKYSTGTGSKRIHENTVNLNFSLHLPCLNEQLKIASFVCFLN
RKIELLERKIYLIKKQKQALLQQMFI

CC133-2 from ED133 TRD d*

CAAATTGAATTAGAAGAACAGAAGCTGGAACCTGCTGCAACAGCAGAAAAAAGGTTATATGCAGAAA
ATCTTCAGCCAAGAGCTGCGCTTTAAAGATGAAAATGGTAACGATTATCCGGAATGGGAAAATGTG
ATGCTGCAGAAAGTTCTGAAAGATAAAACCGAAGGTATTAAACGTTGGTCCGTTTGGTGGTGCACCTG
AAAAAAGATATTTTTGTGGAAAGCGGCTATGCCGTTTATGAACAGCGTAATGCCATTTATGATATC
AGCAAACCTCCGCTACTATATCAACGAGAACAAATACAAAGAGATGCAGAGCTTTAGCGTTCAGCCC
AATGATATTATCATGAGCTGTAGCGGCACCATTGGTTCGTCTGGCACTGATTCCGCATAACTATAACC
AAAGGTATTATCAACCAGGCCCTGATTCGTTTTTCGTACCAATCATAAAATCCGCAGCGAATTCTTT
CTGATCTTTATGCGTAGCAATCAGATGCAGCGTAAAATTTCTGGAAGCAAATCCGGGTAGCGCAATT
ACCAATCTGGTTCGGTTAAAGAACTGAAACTGATCCCGTTTTCCGCTGCCGGTTAAATTTGAACAG
GATAAAATCAGCCAGTTCATCCACATTATTAACCGTCGTATTGAACAGAGCGAGAAAAAATCGAA
AGCCTGAAAAATCGCAAACAGGGTTTCTGCAGAACTGTTTTGTTCCCGGGGGATCCGATCGATC

CC133-2 from ED133 TRD d*

QIELEEQKLELLQQQKKGMYMOKIIFSQELRFKDENGNDYPEWENVMLQKVLKDKTEGIKRGPFGGAL
KKDIFVESGYAVYEQRNAIYDISNFRYYINENKYKEMQSFSVQPNDIIMSCSGTIGRLALIPHNYT

1
2 KGIINQALIRFRTNHKIRSEFFLI FMRSNQMQRKILEANPGSAITNLVPVKELKLI PFPLPVKFEQ
3 DKISQFIHI INRRIEQSEKKIESLKNRKQGFLQKLFV
4

5 **ST80-3 TRD X + TRD f***

6 CCCGGGTTTGAAGGCGAATGGGAGGAAAAACAGTTTGCCGATTTTACCAAATAACCAGGGTCTG
7 CAGATTGCCATTAATGAACGTAAAACCGAATATAGCCCTGAGCTGTATTTCTATATCACCAACGAA
8 TTTCTGCGTCCGAATAGCCAGACCAAATATTTTCATTGAAAATCCGCCTCAGAGCGTGATTGCCAAC
9 AAAGAAGATATTCTGATGACCCGCACCCGGTAATACCGGCAAAGTTGTTACCAATGTTTTTGGTGCC
10 TTCCACAACAACCTTTTTCAAATCAAATTCGATAAAAACCTGTATGATCGCCTGTTTTCTGGTTGAA
11 GTTCTGAACAGCAGCAAAAATCCAGAACAAAATTCTGAGCCTGGCAGGTAGCAGCACCATTCCGGAT
12 CTGAATCATAGCGATTTCTATAGCATTAGCAGCAGCTATCCGCTGCTGCGCGAACAGCAAAAAATT
13 GGCAAATTCTTTAGCAAACCTGGATCGCCAGATTGAACTGGAAGAACAGAACTGGAAGTCTGCAA
14 CAGCAGAAAAAAGGCTATATGCAGAAAATCTTTAGCCAAGAGCTGCGCTTTAAAGATGAAAACGGT
15 GAAGATTATCCGGATTGGAAAGAAAAAAACTGGGCGATATTACCGAGCAGAGCATGTATGGTATT
16 GGTGCAAGCGCAACCGTTTTGATAGCAAAAATATCTATATCCGCATCACCGACATCGATGAAAAAA
17 GCCGTAAACTGAATTATCAGAATCTGACCACACCGGATGAACTGAACAATAAATACAAACTGAAAC
18 GCAACGACATCCTGTTTTGCACGTACCGGTGCAAGTACCGGTAAAAGCTATATTCATAAAGAAGAGA
19 AAGACATCTACAACACTACTTTTGCGGGTTTTCTGATCAAATTCAAAATTAACGAACAGAACAGTC
20 CGCTGTTTCTATCTATCAGTTTACCCTGACCAGCAAATTCACAAAATGGGTAAAGTTATGAGCGTGC
21 GTAGCGGTCAGCCTGGTATTAATAGCGAAGAATATGCAAAACTGCCGCTGGTTCTGCCGAATAAAC
22 TGGAACAACAAAAAATCGCGAAATTCCTGGATCGTTTTGATCGTCAGATCGAGCTGGAAAAACAAA
23 AAATTGAAATTCTGCAGCAACAAAAAAAAGGCCTGCTGCAGAGTATGTTTATTTCCCGGGGGATCCG
24 ATCGATC
25

26 **ST80-3 TRD X**

27 MSNTQKKNVPELRFPGFEGEWEEKQFADF TKINQGLQI AINERKTEYSPELYFYITNEFLRPNSQT
28 KYFIENPPQSVIANKEDILMTRTGNTGKVVTNVFGAFHNNFFKIKFDKNLYDRLFLVEVLNSSKIQ
29 NKILSLAGSSTIPDLNHSDFYSSISSYPLLREQQKIGKFFSKLDR
30

31 **ST80-3 TRD f***

32 QIELEEQKLELLQOQKKGVMQKIFSQELRFKDENGEDYPDWKEKKLGDITEQSMYGIGASATRFDS
33 KNIYIRITDIDEKSRKLNQNLTPDELNNKYKLRNDILFARTGASTGKSYIHKEEKDIYNYFYFA
34 GFLIKFKINEQNSPLFIYQFTLTSKFNKWKVMSVRSQPGINSEEYAKLPLVLPNKLEQQKIAKF
35 LDRFDRQIELEKQKIEILQOQKGLLQSMFIPGGSHHHHHH
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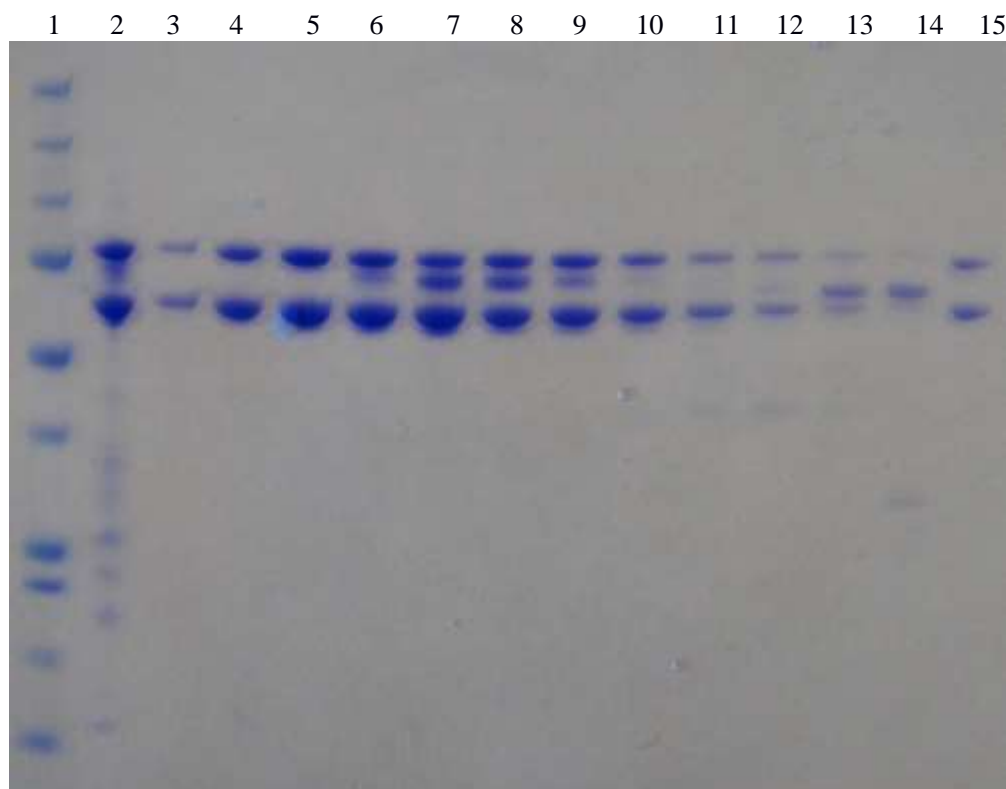
SUPPLEMENTARY INFORMATION FOR TABLE 2.

S. SauCD-EGFP

CC30-1 GWAG-5-GAT

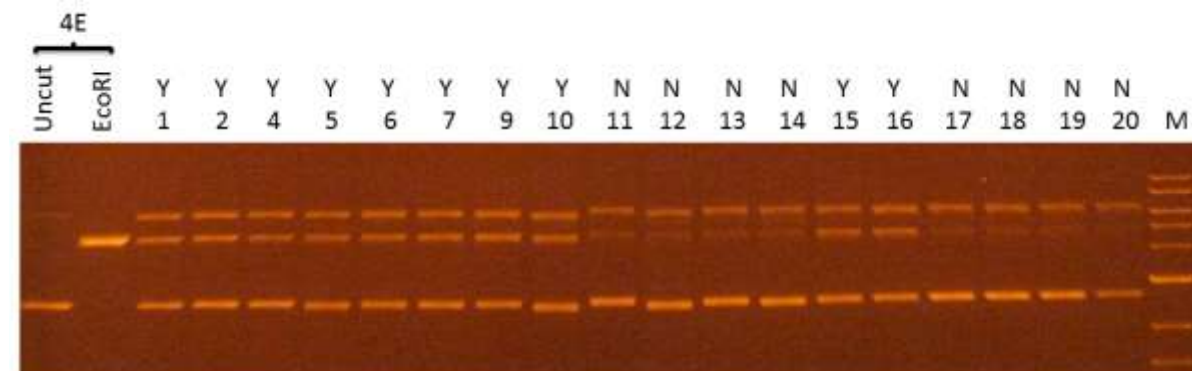
This MTase was expressed and purified as a fusion with EGFP.

MSNTQTKNVP~~ELR~~FPGFEGEWEEKKVGELLEFKNGLNKGKEYFGSGSSIVNFKDVFNNRSLNTNNL
 TGKVVNSKELKNYSVEKGDVFFTRTSEVIGEIGYPSVILNDPENTVFSGFVLRGRPKSGIDLINN
 NFKRYVFFFTNSFRKEMITKSSMTTRALTS~~GS~~AINKMKVIYPVSAKEQRKIGDFFSKLDRQIELEEQ
 KLELLQQQKKGYMQKIFSQELRFKDENSEDYPHWENSKIEKYLKERNERSDKGQMLSVTINSGIIK
 FSELDRKDNSSKDKSNYKVVRKNDIAYNSMRMQGASGRSNYNGIVSPAYTVLYPTQNTSSLFIGY
 KFKTHRMIHKFKINSQGLTSDTWNLYKQQLKNINIDIPVLEEQEKIGDFFKKMDILISKQKIKIEI
 LEKEKQSFLQKMFLGSMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGLTLKFICT
 TGKLPVPWPTLVTTLTLYGVQCFSRYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFE
 GDTLVNRIELKGIDFKEDGNILGHKLEYNNSHNVYIMADKQKNGIKVNFKIRHNIEDGSVQLADH
 YQQNTPIGDGPVLLPDNHYLSTQSALS~~KDP~~NEKRDHMLLEFVTAAGITLGMDELYKHHHHHH



1- marker 2- Nickel column eluate 3-14 Fractions from gel filtration column
 15- CC5-1 Purified protein marker

Nuclease assay on the plasmid library.



S. SauCD-EGFP

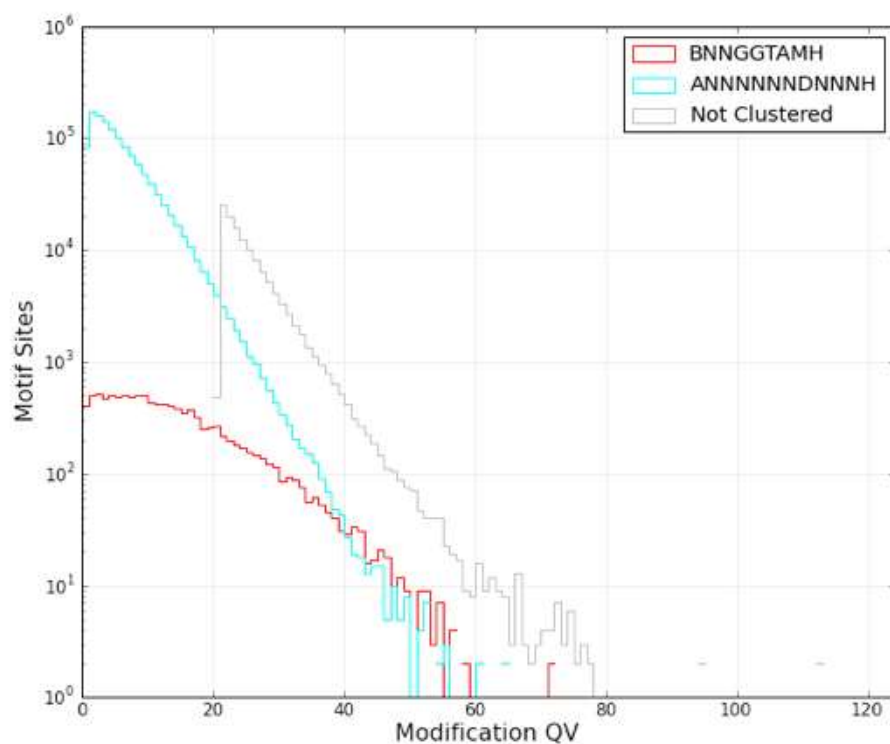
CC30-1 GWAG-5-GAT

SMRT did not work for the CC30-1 system when looking for methylation of genomic DNA from *E. coli*.

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
BNNGGTAMH	4	unknown	7.05	780	11058	37.7	88.6	
ANNNNNNDNNNH	1	unknown	0.11	1312	1235059	36.0	100.7	
Not Clustered	0		0.19	14583	7880091	36.1	107.4	

Modification QV Histogram By Motif

Modification QV Histogram



S. SauJK-EGFP**CC30-2 GGA-7-TCG****This MTase was a fusion with EGFP.**

MSNTQKKNVPELRFPEFEGEWEERKLGDLIKVNSGKDYKHLDKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPEKEADILFILSLFRKINWKLYDESTGVPSSLKQTI
 NKINRLVPTNKEQQKIGEFFSKLDRQIELEEQKLELLQQQKGYMOKIFSQELRFKDENGNDYPKW
 EEKKIEDIASQVYGGGTPNTKIKEFWNGDIPWIQSSDVKVNDLILQQCNKFISKNSIELSSAKLIP
 ANSIAIVTRVGVGKLCLEFVDYATSQDFLSLSSLYKDYKLYSLYSLLYTMKKISANLQGTSIKGITK
 KELLDSI IKIPHNLEEQQKIGDLFYKIDKYISFNKCKIEMLSLQGLLKKMFIGSMVSKGEELFT
 GVVPIVLVELDGDVNGHKFSVSGEGEGDATYGLTLKFICTTGKLPVPWPPTLVTTLTLYGVQCFSRYP
 DHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGI DFKEGDNILGHKL
 EYNYNSHNVIIMADKQKNGIKVNFKIRHNIEDGSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSAL
 SKDPNEKRDHMLLEFVTAAGITLGMDELYKHHHHHH

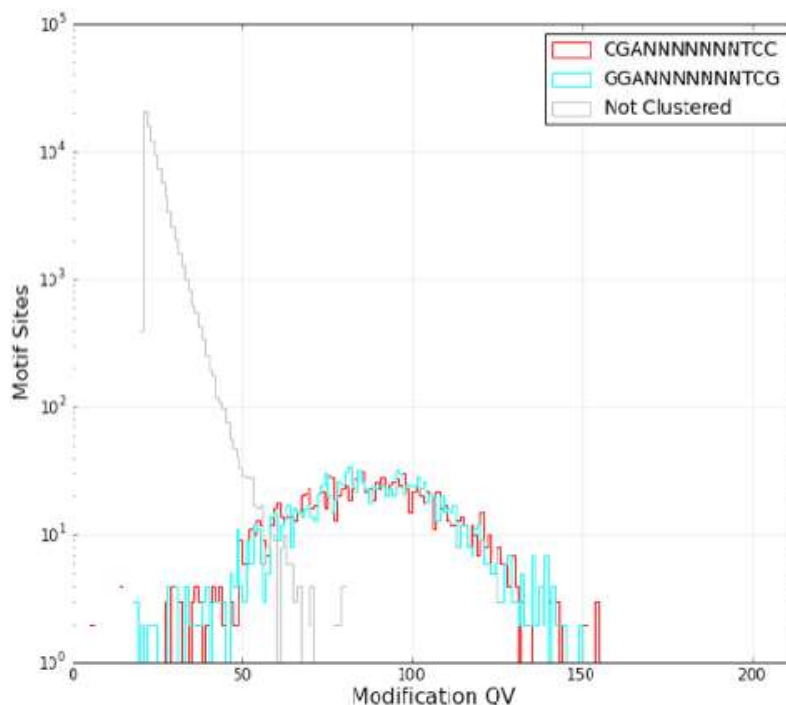
This system could not be expressed so was used for SMRT sequencing only.

SMRT analysis of genomic DNA from *E. coli*.

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs in Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CGANNNNNNTCC	3	m6A	98.76	1439	1457	89.3	76.9	GGANNNNNNTCCG
GGANNNNNNTCCG	3	m6A	98.56	1436	1457	91.2	76.8	CGANNNNNNTCC
Not Clustered	0		0.09	8260	9123294	35.7	87.7	

Modification QV Histogram By Motif

Modification QV Histogram



S.SauJd*

CC133-2 from ED133 GGA-7-TTRG

This enzyme was studied using the SMRT assay. There are minor variations in S subunit sequence in CC133-2.

Recombinant S.SauJd* CC133-2

MSNTQKKNVPELRFPGFEGEWEEKKLGDLIKVNSGKDYKHLEKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLFYCTPKKETDILFILSLFRKINWKVYDESTGVP SLSKQTI
 NKINRFVPSNKEQQKIGEFFIKLDRQIELEEQKLELLQQQKKG YMQKIFSQELRFKDENGNDYPEW
 ENVMLQKVLKDKTEG I KRGPF GGALKKDI FVESGYAVYEQRNAIYDISNFRYYINENKYKEMQSFS
 VQPNDIIMSCSGTIGRLALIPHNYTKGIINQALIRFRTNHKIRSEFFLIFMRSNQMQRKILEANPG
 SAITNLVPVKELKLIPFPLPVKFEQDKISQFIHIINRRIEQSEKKIESLKNRKQGFLQKLFVPGGS
 HHHHHH

Wild type S.SauJd*

MSNTQKKNVPELRFPGFEGEWEEKKLESIIKVNSGKDYKHLDKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLFYCTPKKETDILFILSLFRKINWKVYDESTGVP SLSKQTI
 NKINRFVPTNKEQQKIGKFFSKLDRQIELQE QKLELLQQQKKG YMQKIFSQELRFKDENGNDYPEW
 ENVMLQKVLKDKTEG I KRGPF GGALKKDI FVESGYAVYEQRNAIYDISNFRYYINENKYKEMQSFS
 VQPNDIIMSCSGTIGRLALIPHNYTKGIINQALIRFRTNHKIRSEFFLIFMRSNQMQRKILEANPG
 SAITNLVPVKELKLIPFPLPVKFEQDKISQFIHIINRRIEQSEKKIESLKNRKQGFLQKLFV*

Reports for Job Dryden_J_delta_MODs

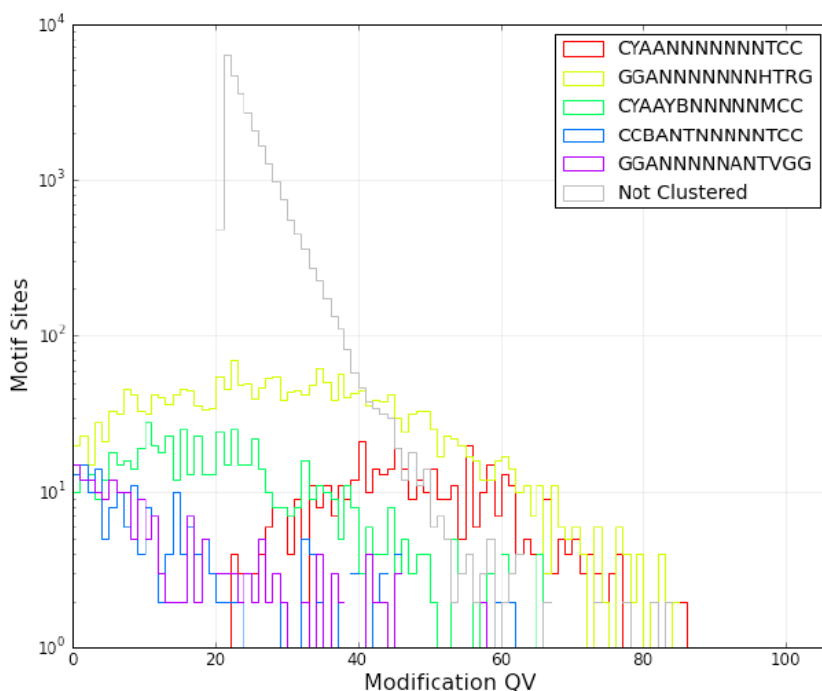


SMRT Cells: 1 Movies: 1

Motif Summary

Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CYAANNNNNNTCC	4	m6A	90.36%	422	467	51.19	30.33	
GGANNNNNNHTRG	3	m6A	47.67%	1114	2337	45.72	32.24	
CYAAYBNNNNMCC	4	m6A	25.68%	169	658	42.89	34.51	
CCBANTNNNNNTCC	4	m6A	20.39%	42	206	44.40	32.14	GGANNNNNANTVGG
GGANNNNNANTVGG	3	m6A	18.45%	38	206	44.76	31.37	CCBANTNNNNNTCC

Modification QVs



S. SauNE**CC398-1 ACC-5-RTGA**

The clone obtained contained a single amino acid substitution A50S which did not affect activity. The enzyme was expressed using plasmid pSauNE-XmaI.

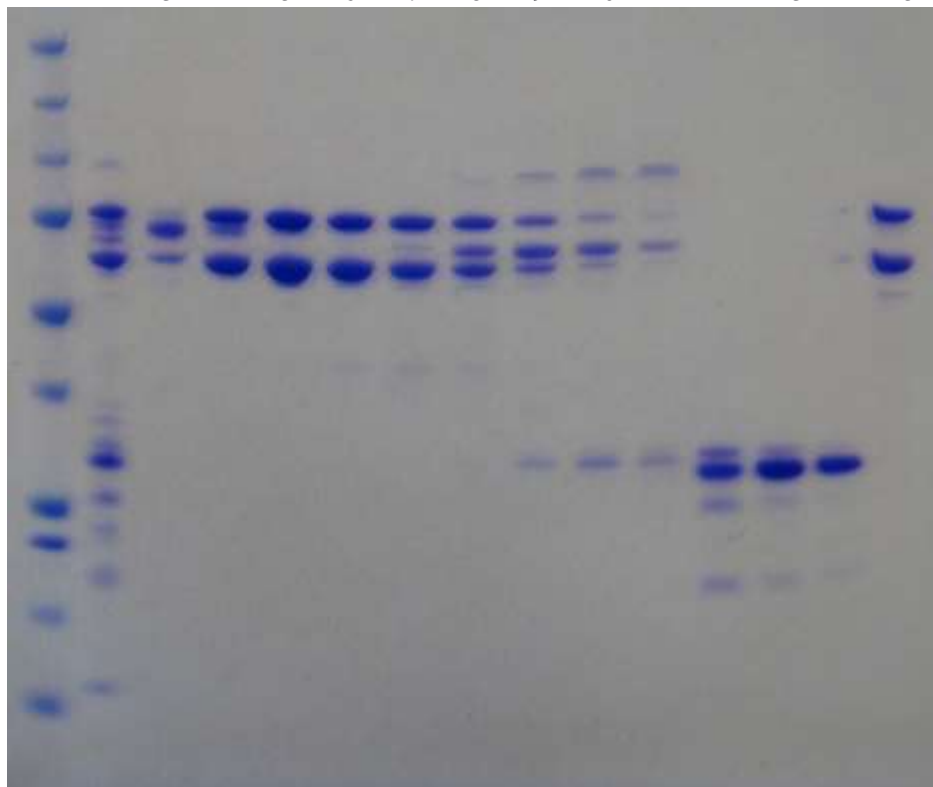
S. SauNEXmaI "Expected" sequence

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLGGKGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGDSGARADRFSIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQQQ
 KKGVMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKIN
 TFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYSA
 KTSVDSVRKDMIANKVPRPIYIEQKKIGQFIKRVDNKTKIQQQVIELLKQRKKSLLQKMFIPGGS
 HHHHHH*

S. SauNEXmaI "Actual" sequence

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSSELGIIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLGGKGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGDSGARADRFSIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQQQ
 KKGVMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKIN
 TFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYSA
 KTSVDSVRKDMIANKVPRPIYIEQKKIGQFIKRVDNKTKIQQQVIELLKQRKKSLLQKMFIPGGS
 HHHHHH*

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15



1- marker 2- Nickel column eluate 3-14 Fractions from gel filtration column

15- CC5-1 purified protein marker

S. SauNE

CC398-1 ACC-5-RTGA

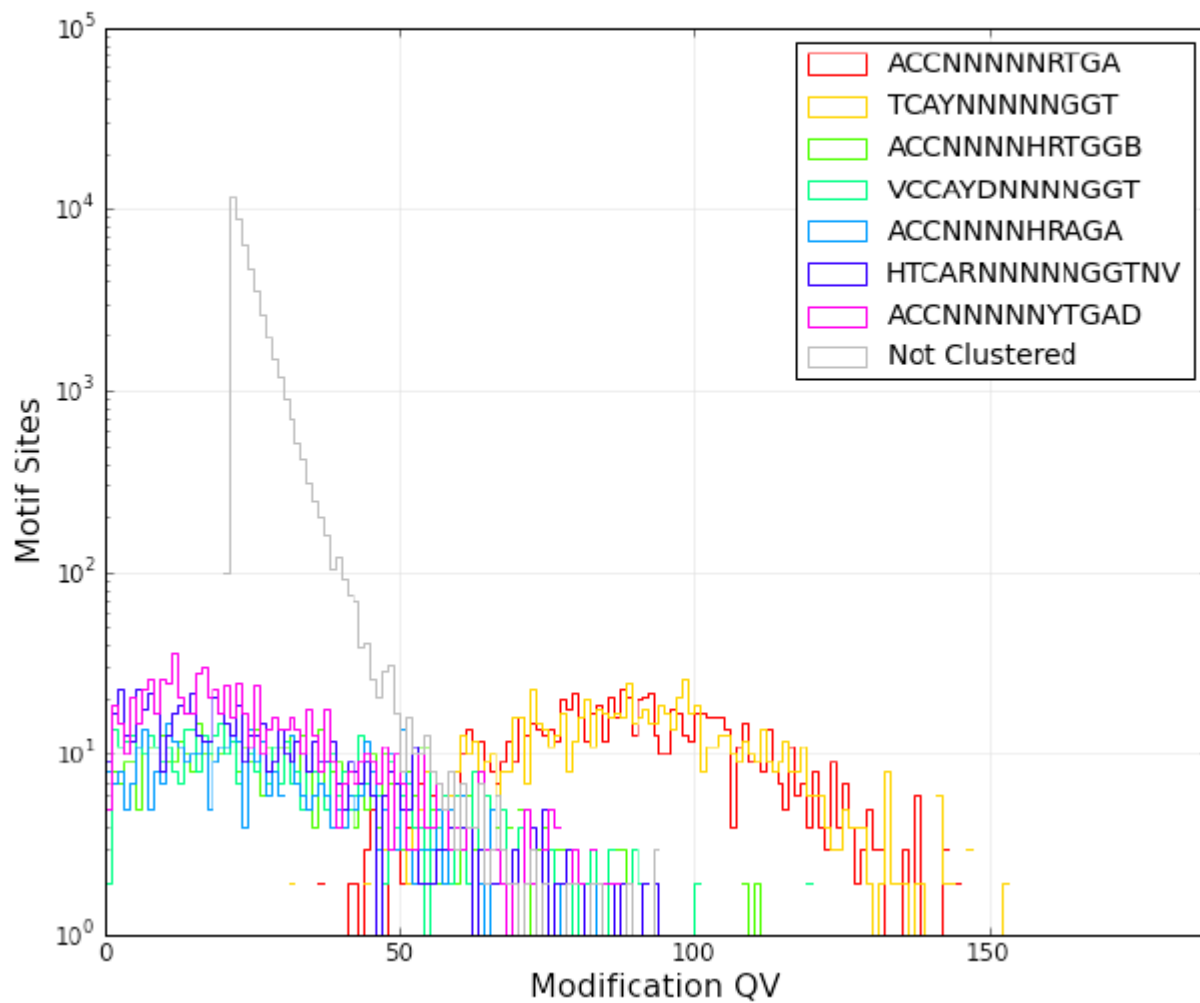
Reports for Job Ed_1_Dryden_MODS



SMRT Cells: 1 Movies: 1

Motif Summary								
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNRTGA	1	m6A	99.69%	971	974	89.04	57.17	TCAYNNNNNGGT
TCAYNNNNNGGT	3	m6A	99.69%	971	974	90.00	57.86	ACCNNNNNRTGA
ACCNNNNHRTGGB	1	m6A	49.07%	291	593	54.17	60.71	VCCAYDNNNNGGT
VCCAYDNNNNGGT	4	m6A	45.36%	269	593	54.62	61.85	ACCNNNNHRTGGB
ACCNNNNHRAGA	1	m6A	41.75%	200	479	48.38	61.76	
HTCARNNNNNGGTV	4	m6A	36.31%	264	727	51.22	62.33	
ACCNNNNNYTGAD	1	m6A	34.9%	320	917	49.93	60.88	

Modification QVs

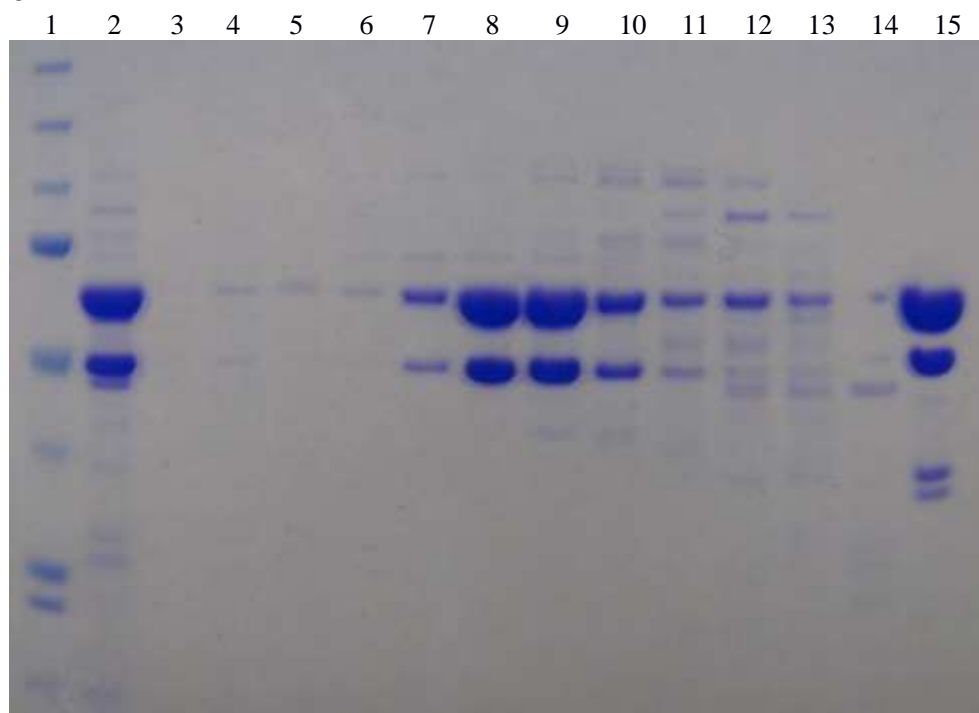


1
2 **SUPPLEMENTARY INFORMATION FOR TABLE 3.**
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S.SauBE AGG-5-RTGA

This MTase was purified but cut all the plasmids in the nuclease assay. Therefore once the targets for each TRD had been determined from other MTases, we used the ATPase assay to verify the length of the non-specific spacer.

MSNTQKKNVPELRFPGFEGEWEEKKLGDLTDRVIRKKNKNLESKKPLTISGQLGLIDQTEYFSKSVS
SKNLENYTLIKNGEFAYNKSYNGYPLGAIKRLTRYDSGVLSSLYICFSIKSEMSKDFMEAYFDST
HWYREVSGIAVEGARNHGLLNVS VNDFFTTILIKYPSLEEQQKIGKFFSKLDRQIELEEQKLELLQQ
QKKGVMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKI
NTFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYS
AKTSVDSVRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKTKIQKQVIELLKQRKKSLLQKMFIPGG
SHHHHHH



1- marker 2- Nickel column eluate 3-14 Fractions from gel filtration column
15- CC398-1 purified protein marker

Oligonucleotides for checking BE target site using ATPase assay.

Underlined refers to methylated bases.

5' -AGG-N-RTGA-3'

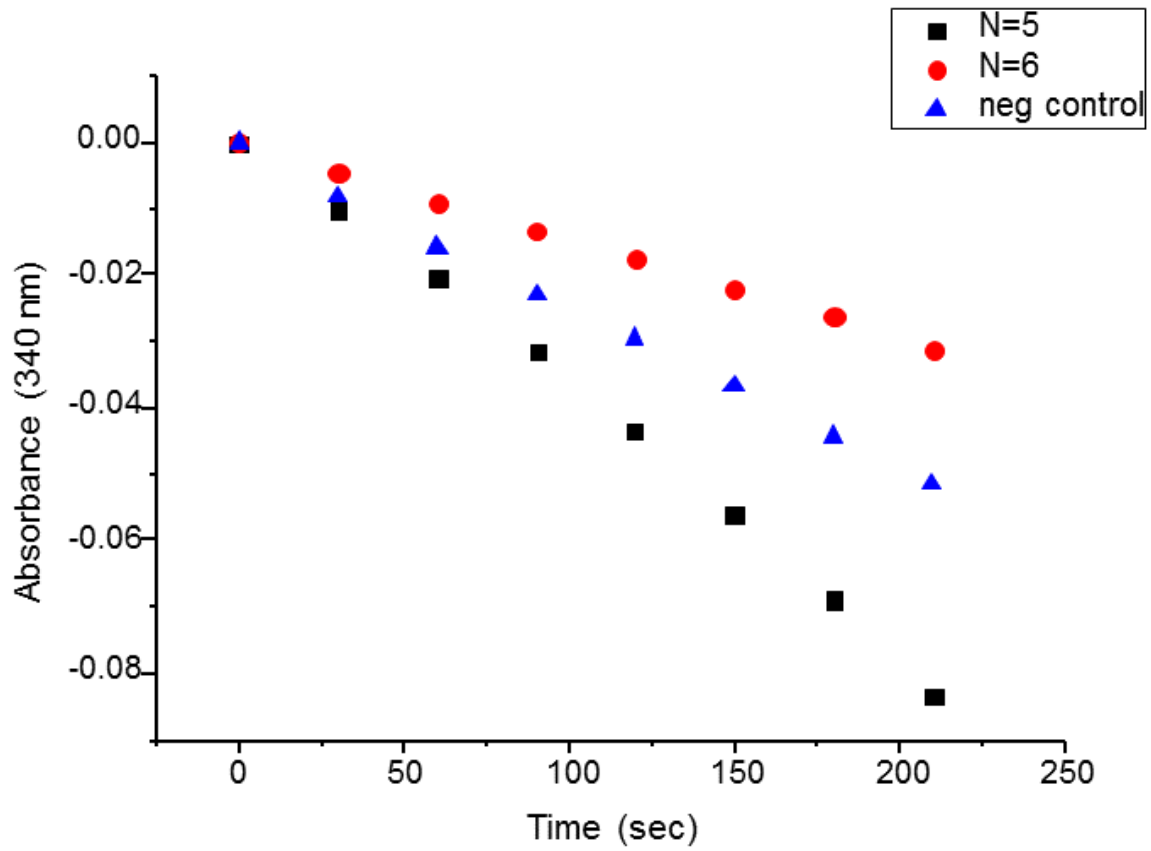
3' -TCC-N-YACT -5'

N values may be 4-6 i.e., number of base pairs between methylated adenines of 7-9. However, DNA digests show that pUC19 contains the site. This rules out the possibility of N=4 (i.e., no site in pUC19 for N value of 4). Therefore we checked for N5 and N6 only.

Oligonucleotide name	DNA sequence (5' to 3')
BE5for	AGATGATGGAATCAATGCAGGTTCCAGTGAGCCCTATACGATATAA
BE5rev	TTATATCGTATAGGGCTCACTGGAACCTGCATTGATTCCATCATCT
BE6for	AGATGATGGAATCAATGCAGGTTACAGTGAGCCCTATACGATATAA
BE6rev	TTATATCGTATAGGGCTCACTGTGAACCTGCATTGATTCCATCATCT

S. SauBE AGG-5-RTGA

N=5 gives the most activity therefore we conclude from the ATPase assay that the site for the BE TRD combination is AGG-5-RTGA.

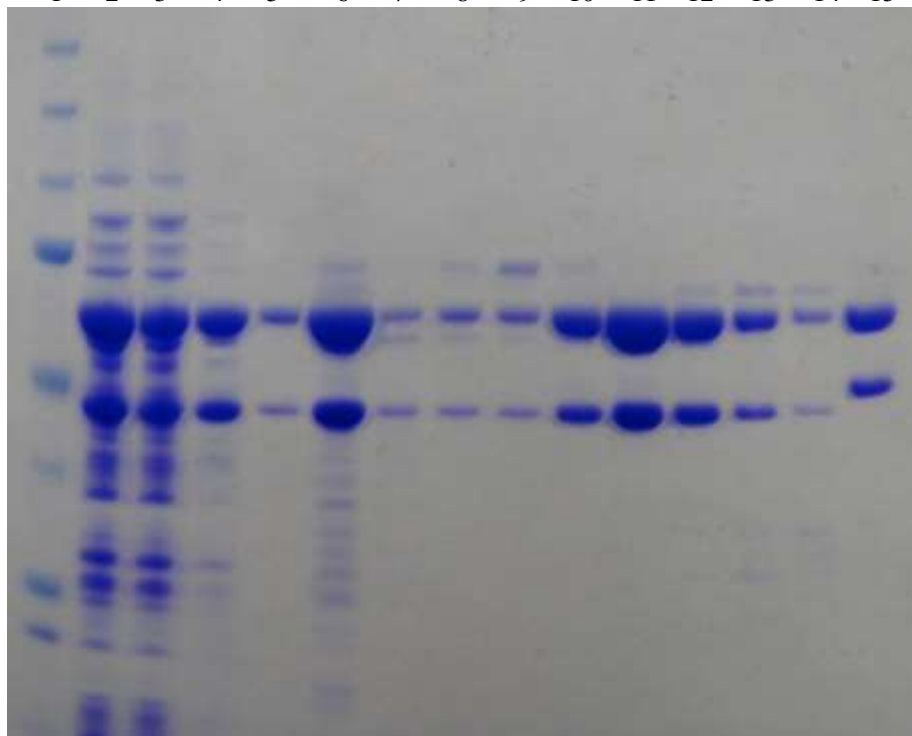


S. SauJE GGA-6-RTGA

This MTase was used in both nuclease and SMRT assays. The TRD pair JE occurs in other ST groups namely ST49 and ST50.

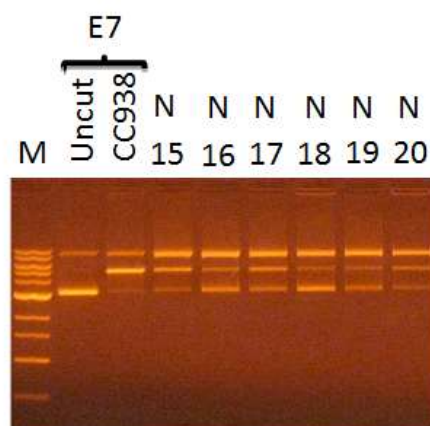
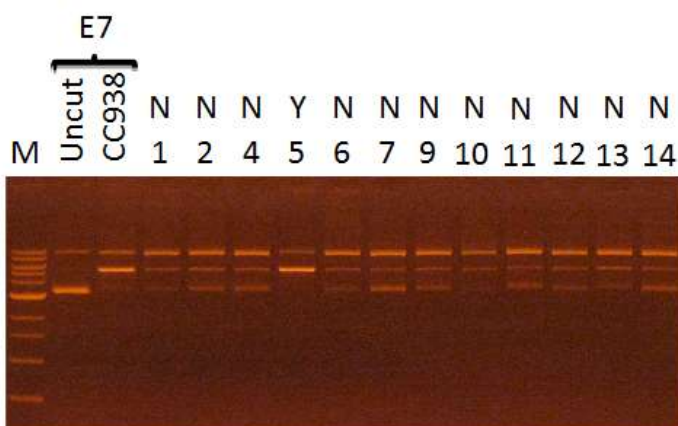
MSNTQKKNVPELRFPGFEGEWEEKLLGLDLIKVNSGKDYKHLEKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPKKETDILFILSLFRKINWKVYDESTGVPSSLKQTI
 NKINRFVPSNKEQQKIGEFFIKLDRQIELEEOKLELLQOQKKGVMQKIFSQELRFKDENGKDYPEW
 EETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKINTFSYEGEAILTVGDGVGVGKVFHYVN
 GKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYSAKTSVDSVRKDMIANMKVPRPIYIEQK
 KIGQFIKRVDNKTKIQKQVIELLKQRKKSLLQKMFIPGGSHHHHHH

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15



1- marker 2- soluble cell extract 3- Nickel column flow through 4- Nickel column wash 1
 5- Nickel column wash 2 6- Nickel column eluate 7-14 Fractions from gel filtration column
 15- CC398-1 purified protein marker

Possible site: GGANNNNNNRTGA Note that the background linearisation may be due to the enzyme displaying star activity against a similar site (i.e., a single GGAN7RTGA site is found in pUC19) to the real site (GGAN6RTGA). Repeated digests generate an identical pattern of digestion.



S. SauJE GGA-6-RTGA

SMRT data showed only the N=6 spacer giving modification.

SMRT Cells: 2		Movies: 2				Print	Download	Email
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTCC	3	m6A	31.44	305	970	38.3	17.1	GGANNNNNRTGA
GGANNNNNRTGA	3	m6A	24.43	237	970	38.3	17.5	TCAYNNNNNTCC
Not Clustered	0		0.00	324	9,124,268	34.2	15.8	

S. SauNI ACC-6-TGAR

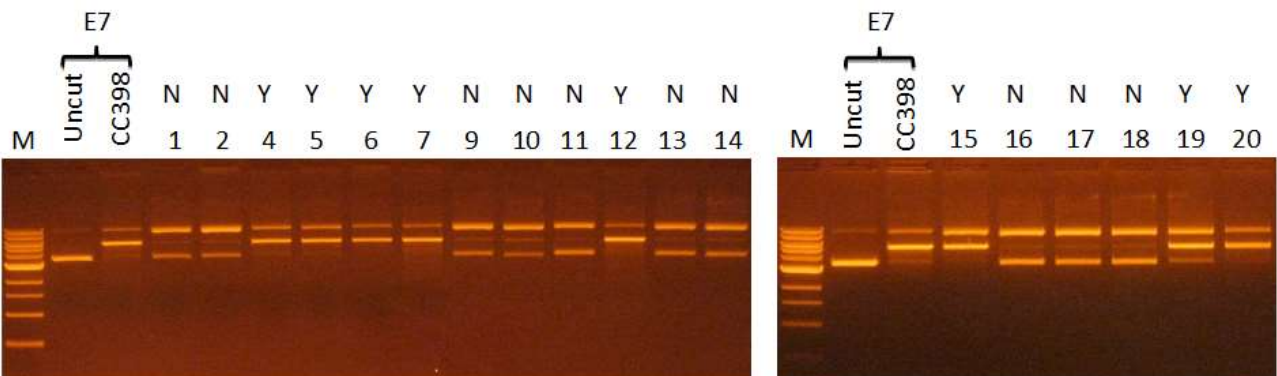
MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLGKKGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGD SGARADRF SIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQQQ
 KKGYMQKIFSQELRFKNENGN DYPDWERIKFFDVIDKVIDFRGRTPKKLNMEWSDEGYLALS AVNV
 KKG YIDFNVEAKYGNLDLYTRWMRGNELYKGQVLF TTEAPMGNVAQVPDNKGYILSQR TIAFNSNE
 KITDNFLASLLSSENVYNDLLKLC SGATAKGV SQKNLNRLYVTIPHSISEQEEIAEFFR KINQLVE
 LQKYKIEHTKSQKQVFLQKMFIPGGSHHHHHH

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15



1- marker 2- soluble cell extract 3- Nickel column flow through 4- Nickel column wash 1
 5- Nickel column wash 2 6- Nickel column eluate 7-14 Fractions from gel filtration column
 15- CC398-1 purified protein marker

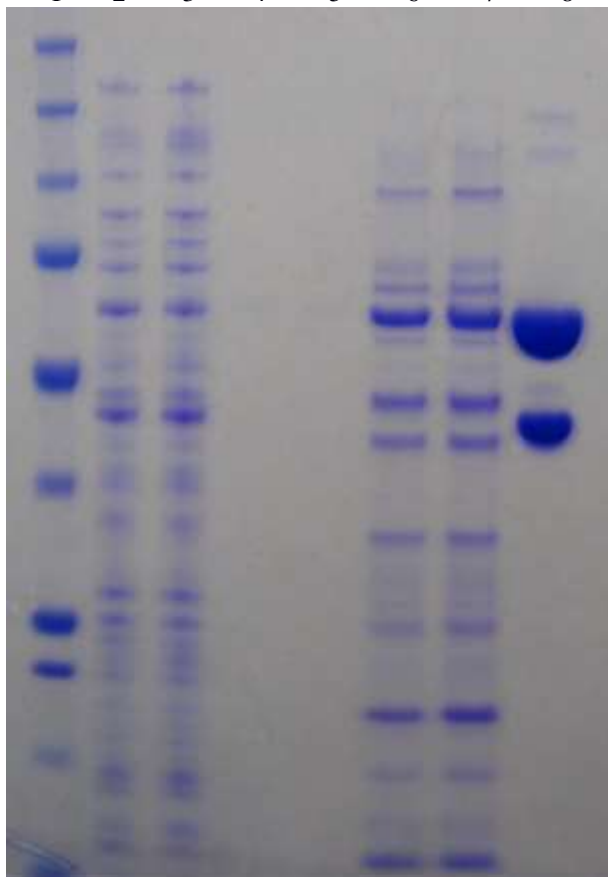
Nuclease assay on the plasmid library gave a clear result.



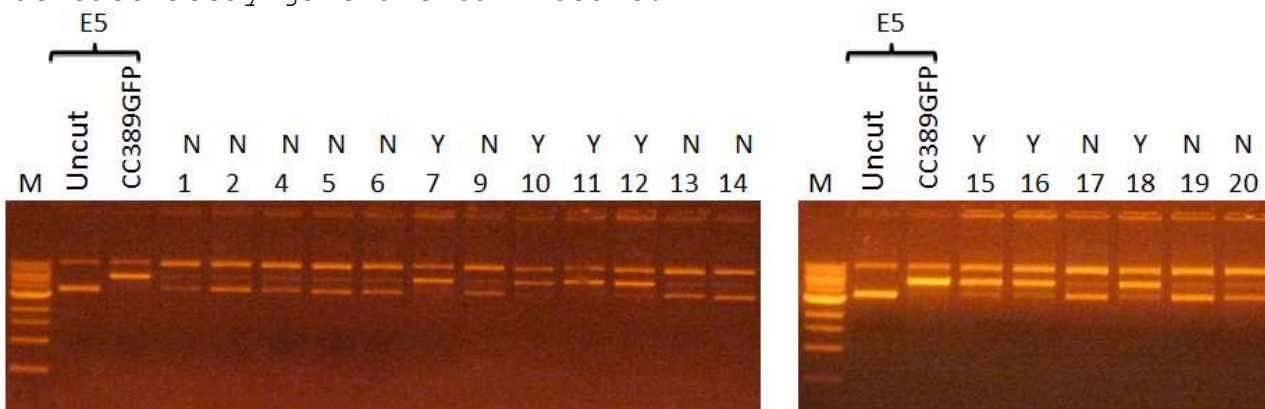
S. SauNK ACC-6-TCG

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVRNKLKGGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGDSGARADRFSIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQQQ
 KKGYMQKIFSQELRFKDENGNDYPNWEKKIEDIASQVYGGGTPNTKIKEFWNGDIPWIQSSDVKV
 NDILILRQCNKFISKNSIELSSAKLIPANSIAIVTRVGVGKLCLEVEFDYATSQDFLSLSLKYDKLY
 SLYSLLYTMKKISANLQGTSIKGITKKELLDSIIKIPHNLEEQQKIGDLFYKIDKYISFNKCKIEI
 LKSLKQGLLQKIFIPGGSHHHHHH

1 2 3 4 5 6 7 8



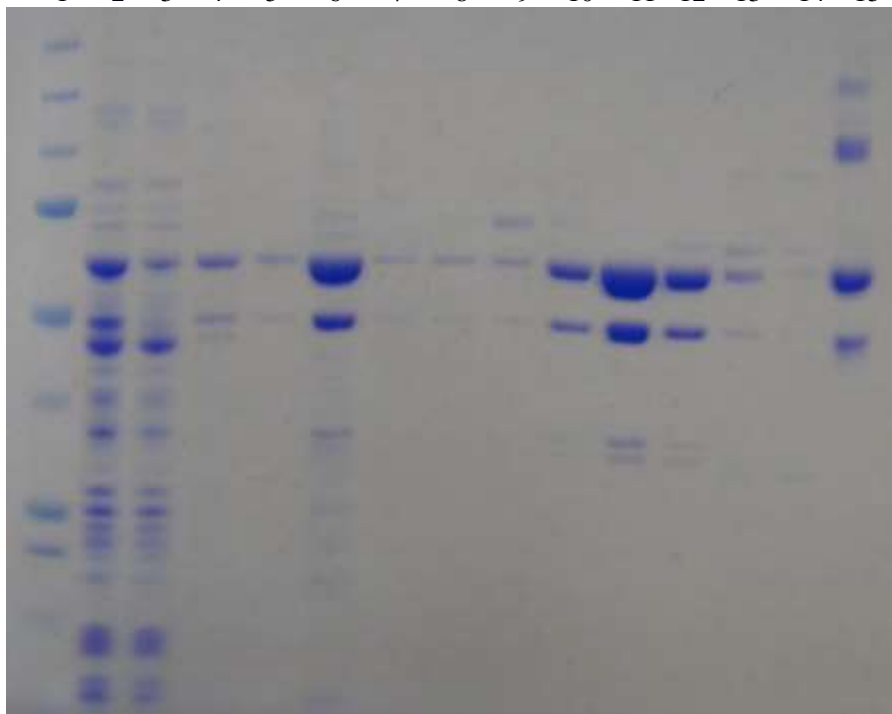
1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- Eluate after PD10 desalting 8- CC398-1 purified protein marker
 Nuclease assay gave a clear result.



S. SauNI ACC-6-TAAA

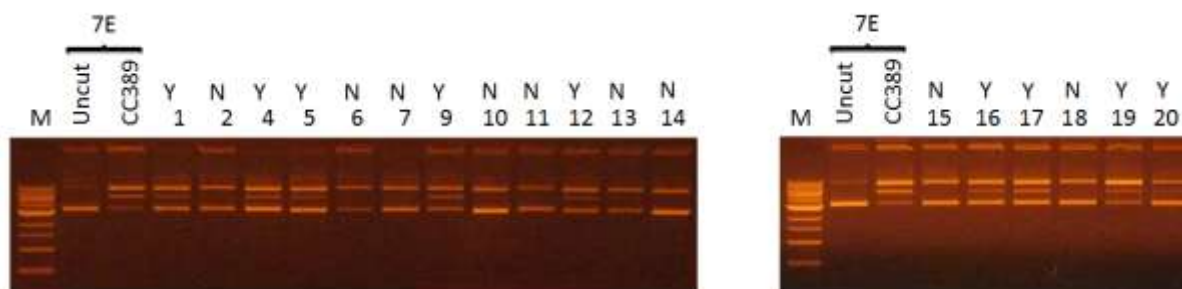
MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVRNKLKGGKGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGD SGARADRF SIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQLLELLQQQ
 KKGYMQKIFSQELRFKDENGNDYPNWRTIELKNIENIVDNRGKTPDNAPSEKYPLLEVNALGYR
 PAYIKVSKFVSENTYNNWFREHLKENDILFSTVGNTGIVSLMDNYKAVIAQNIVGLRVNNNNLPSF
 IYYMLS YKGNQKKIKRIQMGA VQPSVKVSQFKFIKYLVP IKDEQEKVAKLLIEIDKLVNKQLIKIE
 LLQQQRKALLKSMFIPGGSHHHHHH

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15



1- marker 2- soluble cell extract 3- Nickel column flow through 4- Nickel column wash 1
 5- Nickel column wash 2 6- Nickel column eluate 7-14 Fractions from gel filtration column
 15- CC398-1 purified protein marker

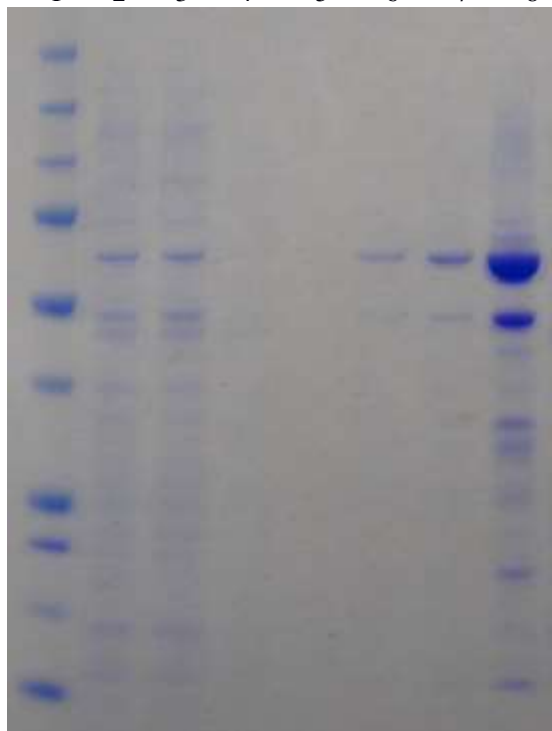
Nuclease assay gave a clear result.



S. SauNP ACC-5-CCT

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLGKKGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGDSGARADRFSIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQLLELFQQQ
 KKGVMQKIFSQELRFKDESGNDYPDWEEKELGEVADR VIRKNKNFESKKPLTISGQLGLIDQTEYF
 SKSVSSKNLENYTLIKNGEFAYNKSY SNGYPLGAIKRLTRYDSGVLSSLYICFSIKSEMSKDFMEA
 YFDSTHWYREVSGIAVEGARNHGLLNISVNDFF TILIKYPSLEEQRKIGDFFIKLDRQIELEEQL
 ELLQQRKKALLKSMLIPGGSHHHHHH

1 2 3 4 5 6 7 8



1- marker, 2- soluble cell extract, 3- Nickel column flow through, 4- Nickel column wash 1, 5- Nickel column wash 2, 6- Nickel column eluate, 7- eluate after conc. and PD10 desalting, 8- Final concentrated protein

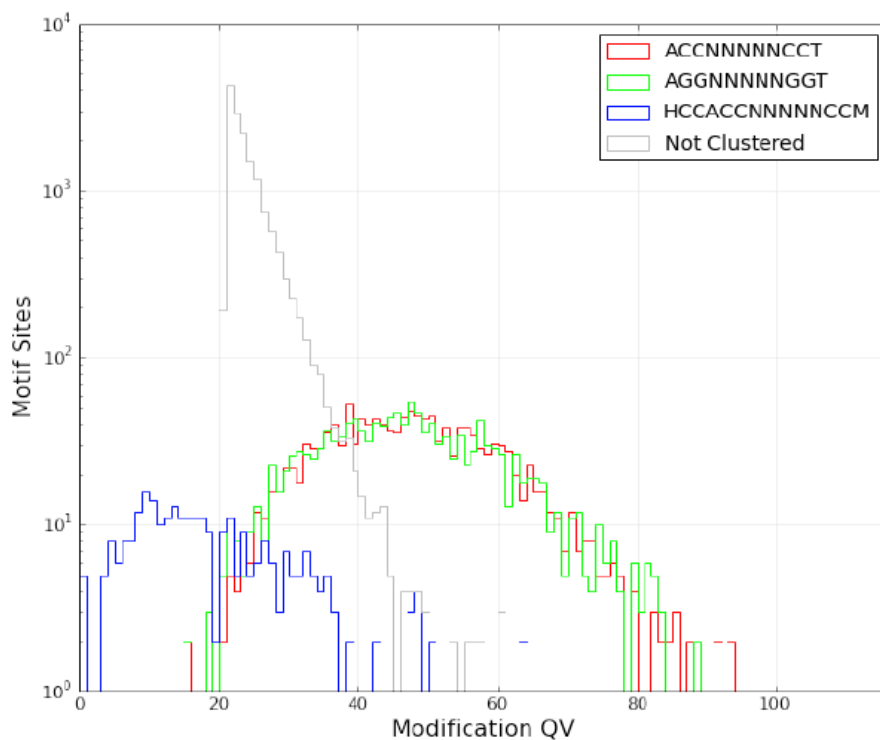
Although purified this MTase was only assayed via SMRT.

S. SauNP ACC-5-CCT

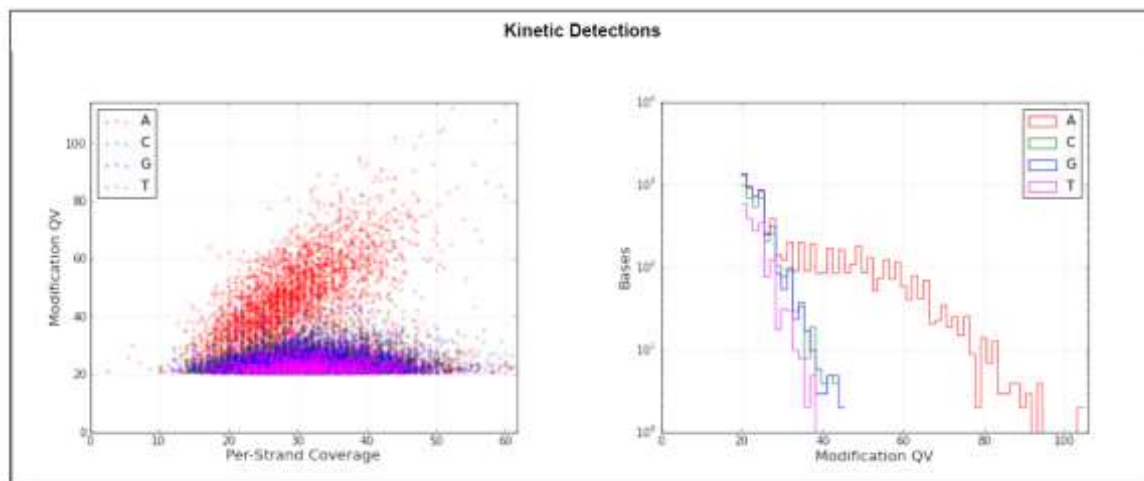
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNCCT	1	m6A	91.03	1320	1450	49.8	29.6	AGGNNNNNGGT
AGGNNNNNGGT	1	m6A	89.79	1302	1450	50.0	29.7	ACCNNNNNCCT
HCCACNNNNNCCM	4	m6A	17.39	52	299	40.0	34.2	
Not Clustered	0		0.01	737	9114127	34.7	34.2	

Modification QV Histogram By Motif



Kinetic Detections

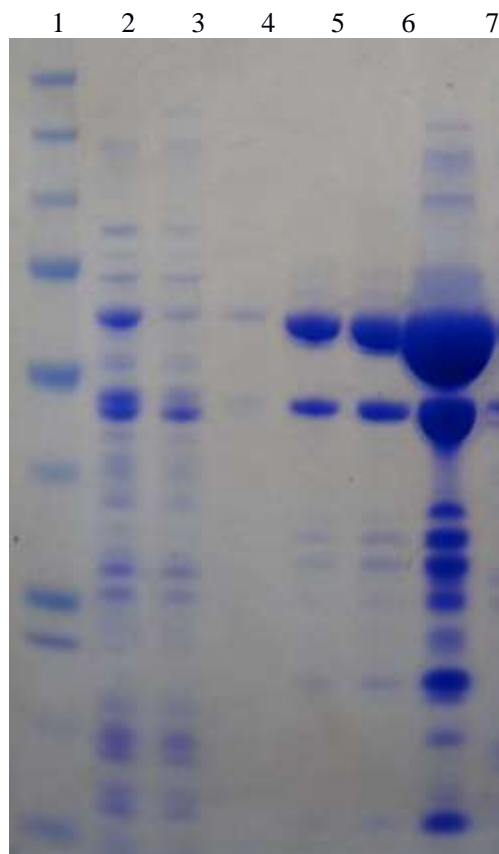


Motifs

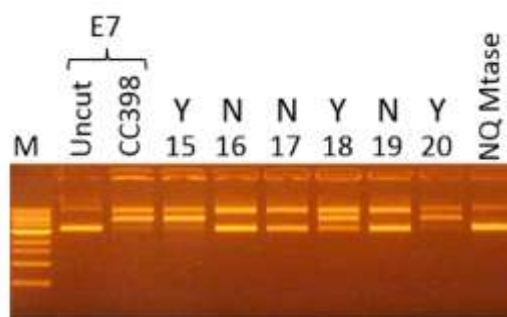
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNCCT	1	m6A	91.03	1320	1450	49.8	29.6	AGGNNNNNGGT
AGGNNNNNGGT	1	m6A	89.79	1302	1450	50.0	29.7	ACCNNNNNCCT
HCCACNNNNNCCM	4	m6A	17.39	52	299	40.0	34.2	
Not Clustered	0		0.01	737	9114127	34.7	34.2	

S. SauNQ ACC-5-RTGT

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLKGGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGDSGARADRFSIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQLLELLQQQ
 KKGYMQKIFSQELRFKDENGEDYSEWEERRFADIFKFHNKLRKPIKENLRVKGSYPYGGATGIIDY
 VDDFI FDGN YLLIGEDGANIITRSAPLVYLVNGKFWVNNHAHILSPLNGNIQYLYQVAELVNYEKY
 NTGTAQPKLNIQNLKIINVVISTNLEEQQKIGSFLSKLDRQIDLEEQLLELLQQRKKALLKSMFVP
 GGSHHHHHH



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 5- Nickel column eluate 6- eluate after conc. and PD10 desalting
 7- Final concentrated protein
 DNA cleavage assay and SMRT assay agreed.

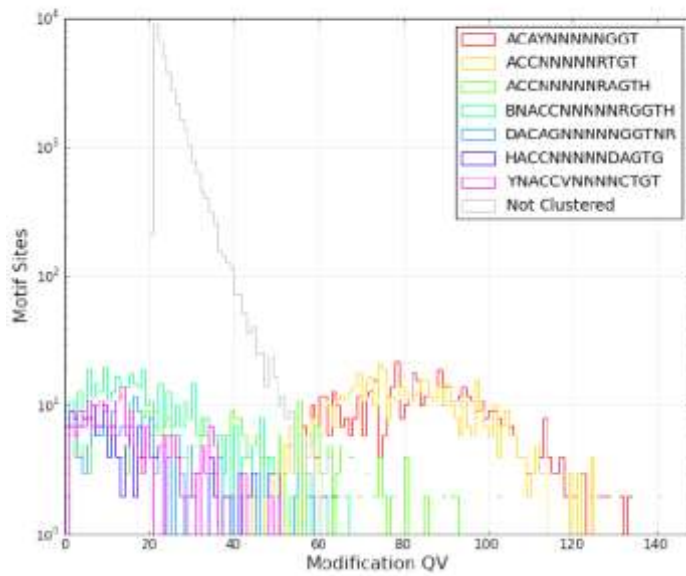


S. SauNQ ACC-5-RTGT

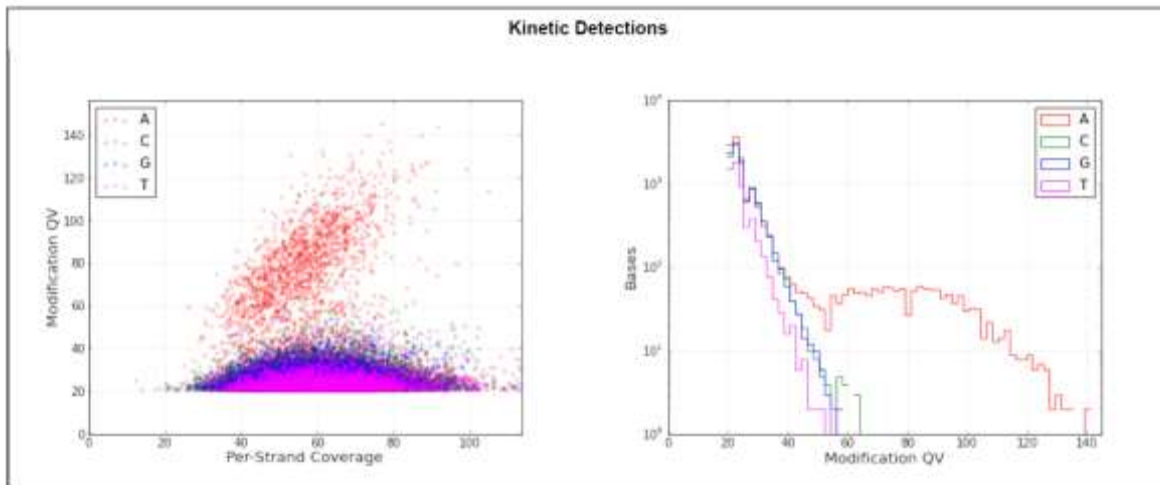
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACAYNNNNNGGT	3	m6A	99.85	654	655	83.5	56.3	ACCNNNNRRTGT
ACCNNNNRRTGT	1	m6A	99.85	654	655	80.7	55.5	ACAYNNNNNGGT
ACCNNNNRAGTH	1	m6A	55.56	215	387	54.3	56.5	
BNACCNNNNRGGTH	3	m6A	23.74	118	497	45.8	57.6	
DACAGNNNNGGTNR	4	m6A	21.65	50	231	42.8	55.5	
HACCNNNNNDAGTG	2	m6A	21.03	41	195	44.8	57.9	
YNACCVNNNNCTGT	3	m6A	20.52	47	229	42.9	57.7	
Not Clustered	0		0.03	3095	9114477	35.3	62.5	

Modification QV Histogram By Motif



Kinetic Detections



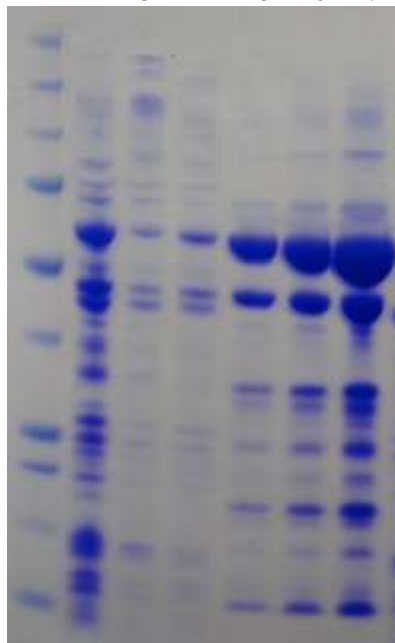
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACAYNNNNNGGT	3	m6A	99.85	654	655	83.5	56.3	ACCNNNNRRTGT
ACCNNNNRRTGT	1	m6A	99.85	654	655	80.7	55.5	ACAYNNNNNGGT
ACCNNNNRAGTH	1	m6A	55.56	215	387	54.3	56.5	
BNACCNNNNRGGTH	3	m6A	23.74	118	497	45.8	57.6	
DACAGNNNNGGTNR	4	m6A	21.65	50	231	42.8	55.5	
HACCNNNNNDAGTG	2	m6A	21.03	41	195	44.8	57.9	
YNACCVNNNNCTGT	3	m6A	20.52	47	229	42.9	57.7	
Not Clustered	0		0.03	3095	9114477	35.3	62.5	

S. SauNS ACC-6-TGC

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLGGKGVMSPLYTVFKIQNIDLNFIEFYFKSS
 KWYRFMALNGDSGARADRFSIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQQQ
 KKGYMQKIFSQELRFKDENGNDYPDWTNERLGEVTTVTMGQSPKSVNYTDNSNDTVLIQGNADIEN
 GLINPRIYTRVTKLIQKDEIILTVRAPVVGKLAMAQINACIGRGVCSIKGDKFLYYFLEWFATQNK
 WIRFSQGSTFESISGNDIRNIHIKIPVEDERTKIIKLLNSLDVLSKTDLKIQLKQRKQSSLQKI
 FVPPGGSHHHHHH

1 2 3 4 5 6 7



1- marker 2- soluble cell extract

3- Nickel column flow through 4- Nickel column wash

5- Nickel column eluate 6- eluate after conc. and PD10 desalting

7- final protein after concentration

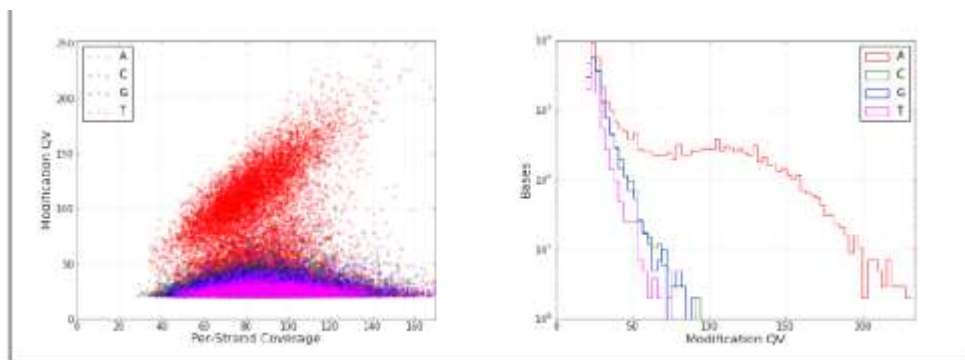
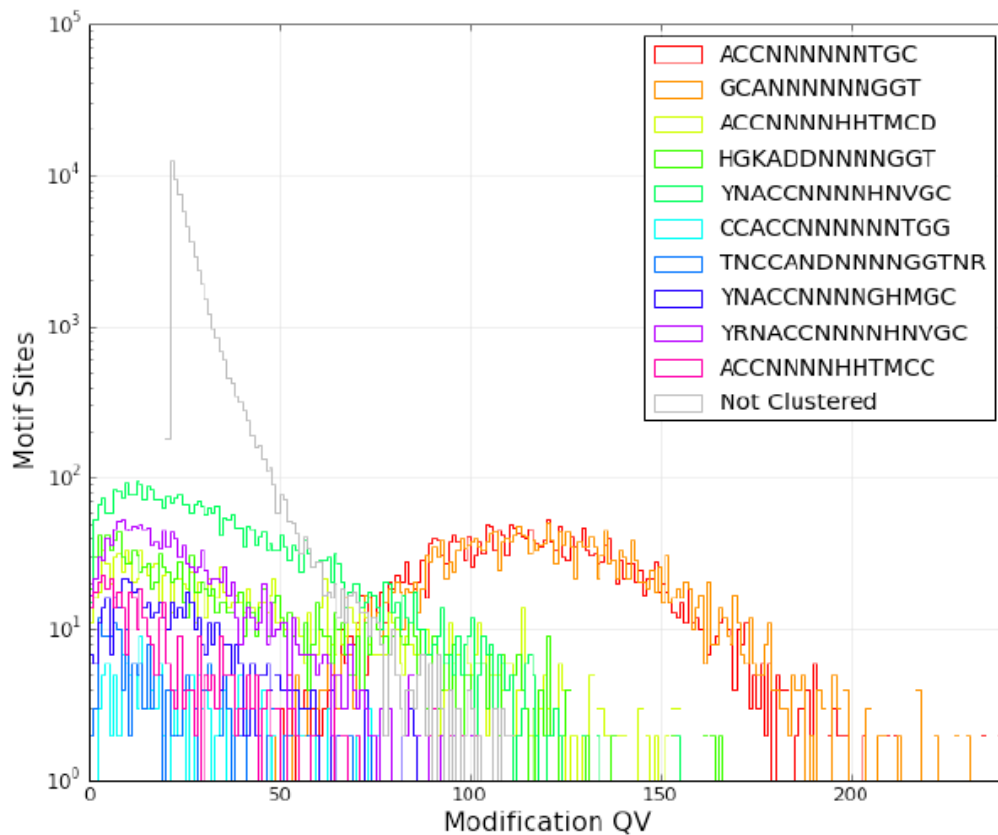
Although purified this MTase was only assayed via SMRT.

Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNTGC	1	m6A	100.00	2938	2938	118.8	81.7	GCANNNNNGGT
GCANNNNNGGT	3	m6A	99.90	2935	2938	120.7	83.9	ACCNNNNNTGC
ACCNNNHHTMCD	1	m6A	57.03	925	1622	71.1	83.7	HGKADDNNNGGT
HGKADDNNNGGT	4	m6A	48.83	792	1622	68.8	86.4	ACCNNNHHTMCD
YNACCNNNHVGC	3	m6A	46.49	1925	4141	57.7	84.6	
CCACCNNNNNTGG	3	m6A	39.15	74	189	55.7	85.9	
TNCCANDNNNGGTNR	5	m6A	31.60	73	231	53.5	82.5	
YNACCNNNGHMGC	3	m6A	31.35	195	622	53.2	87.4	
YRNACCNNNHVGC	4	m6A	28.65	465	1623	48.9	86.3	
ACCNNNHHTMCC	1	m6A	27.58	131	475	58.1	84.3	
Not Clustered	0		0.09	8284	9100925	38.6	92.5	

S. SauNS ACC-6-TGC

Modification QV Histogram By Motif

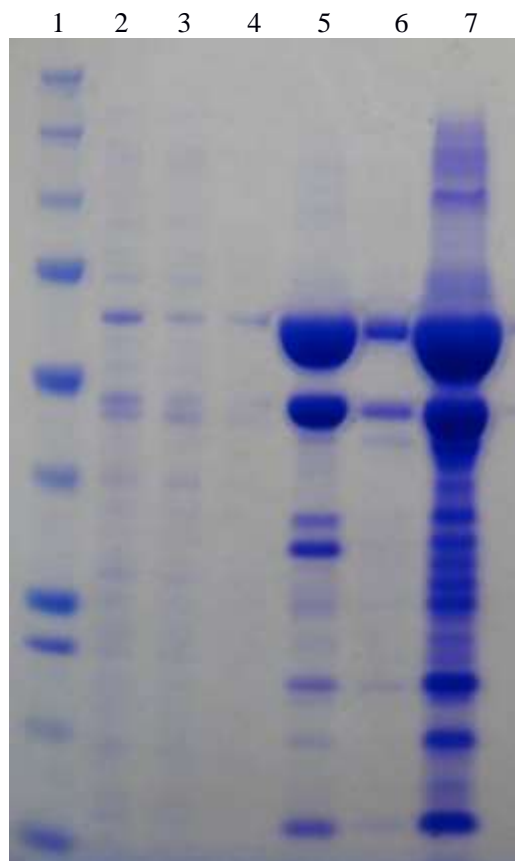


Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs in Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNTGC	1	m5A	100.00	2930	2930	118.8	81.7	GCANNNNNGGT
GCANNNNNGGT	3	m5A	99.90	2935	2938	120.7	83.9	ACCNNNNNTGC
ACCNNNNHHTMCD	1	m5A	57.03	925	1622	71.1	83.7	HGKADDNNNNGGT
HGKADDNNNNGGT	4	m5A	48.83	792	1622	68.8	86.4	ACCNNNNHHTMCD
YNACCNNNNHNVGC	3	m5A	46.48	1928	4141	57.7	84.6	
CCACCNNNNNTGG	3	m5A	39.15	74	189	65.7	85.9	
TNCCANDNNNNGGTNR	5	m5A	31.60	73	231	53.5	82.5	
YNACCNNNNGHMGC	3	m5A	31.35	196	622	53.2	87.4	
YRNACCNNNNHNVGC	4	m5A	28.65	465	1623	48.9	86.3	
ACCNNNNHHTMCC	1	m5A	27.55	131	475	58.1	84.3	
Not Clustered	0		0.09	8204	9100925	38.6	92.5	

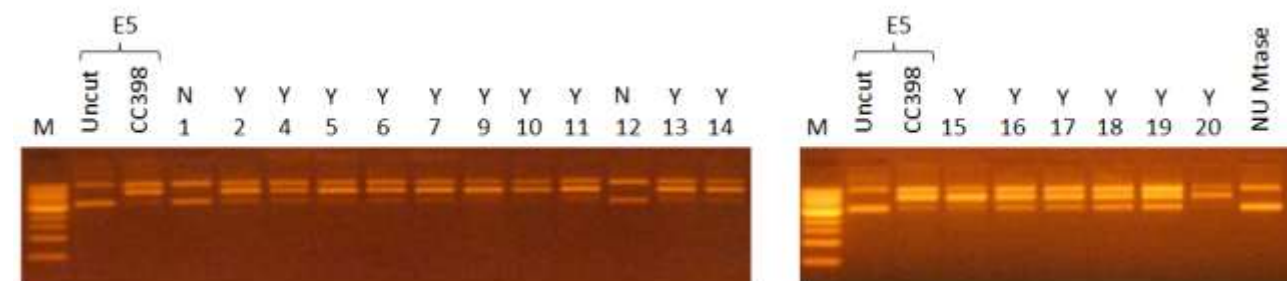
S. SauNU ACC-5-RTC

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 HHHHH



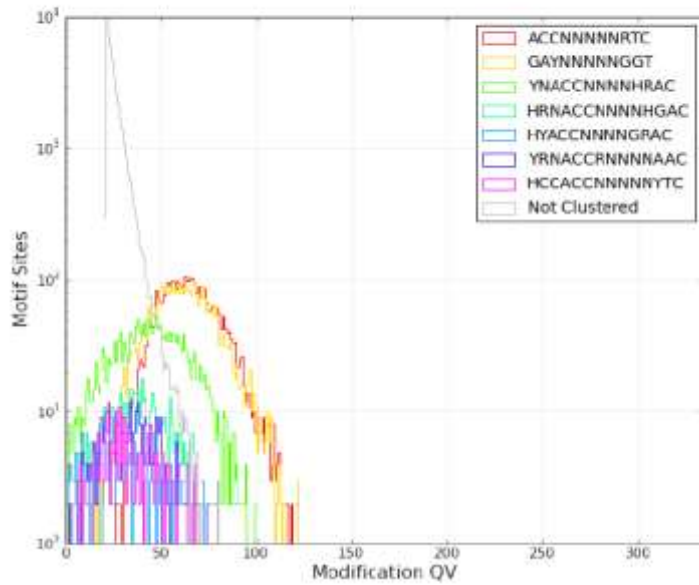
1- marker 2- soluble cell extract
 3- Nickel column flow through 4- Nickel column wash
 5- Nickel column eluate 6- eluate after PD10 desalting
 7- final protein after concentration

DNA cleavage assay worked despite there being one site in pUC19
 but this site was subject to dam methylation and therefore not cut.



S. SauNU ACC-5-RTC

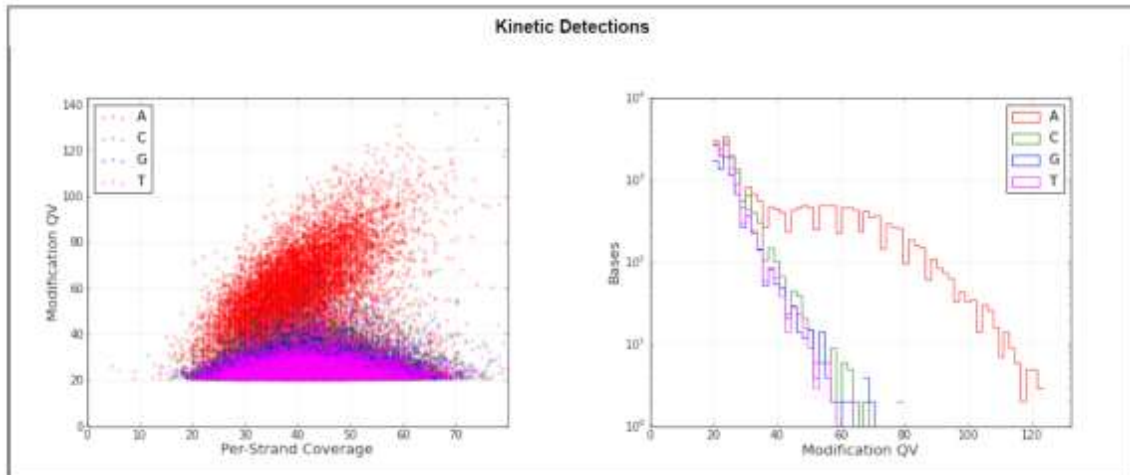
Modification QV Histogram By Motif



Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNRTC	1	m6A	81.87	3162	3862	70.1	41.5	GAYNNNNGGT
GAYNNNNGGT	2	m6A	75.14	2902	3862	70.8	41.9	ACCNNNNRTC
YNACCNNNNHRAC	3	m6A	37.48	820	2188	64.9	43.3	
HRNACCNNNNHGAC	4	m6A	26.22	140	534	63.7	43.0	
HYACCNNNNGRAC	3	m6A	19.23	50	260	63.3	44.4	
YRNACCRNNNNAAC	4	m6A	17.15	59	344	63.7	45.8	
HCCACCNNNNNYTC	4	m6A	16.81	39	232	61.9	45.9	
Not Clustered	0		0.00	229	9106044	58.3	48.5	

Kinetic Detections



Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNRTC	1	m6A	81.87	3162	3862	70.1	41.5	GAYNNNNGGT
GAYNNNNGGT	2	m6A	75.14	2902	3862	70.8	41.9	ACCNNNNRTC
YNACCNNNNHRAC	3	m6A	37.48	820	2188	64.9	43.3	
HRNACCNNNNHGAC	4	m6A	26.22	140	534	63.7	43.0	
HYACCNNNNGRAC	3	m6A	19.23	50	260	63.3	44.4	
YRNACCRNNNNAAC	4	m6A	17.15	59	344	63.7	45.8	
HCCACCNNNNNYTC	4	m6A	16.81	39	232	61.9	45.9	
Not Clustered	0		0.00	229	9106044	58.3	48.5	

S. SauNW ACC-6-TTYG

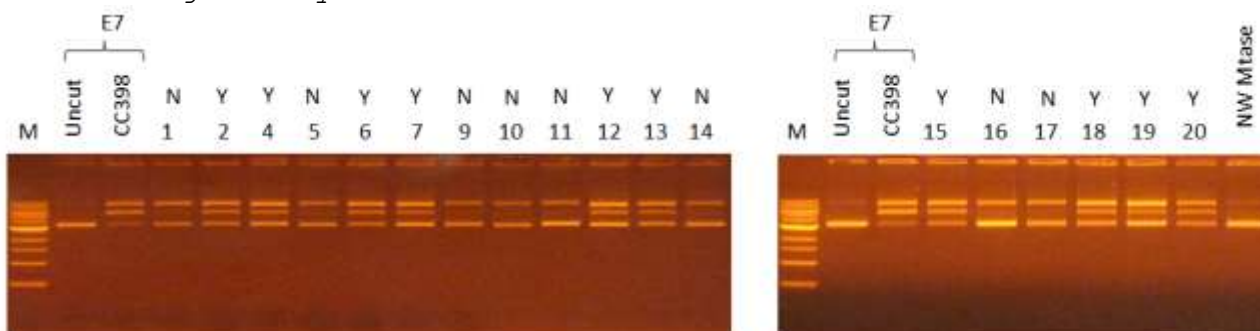
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 MYYWLEYFKDKWSKYGQPGSQVNLNSEIVKSQTLNMP SNHEQEKVGQFFNRNEKLI ELQQEKIMYI
 KRCKQVLLQKMFIPGGSHHHHHH

1 2 3 4 5 6 7



1- marker 2- soluble cell extract
 3- Nickel column flow through 4- Nickel column wash
 5- Nickel column eluate 6- eluate after PD10 desalting
 7- final protein after concentration

DNA cleavage assay.

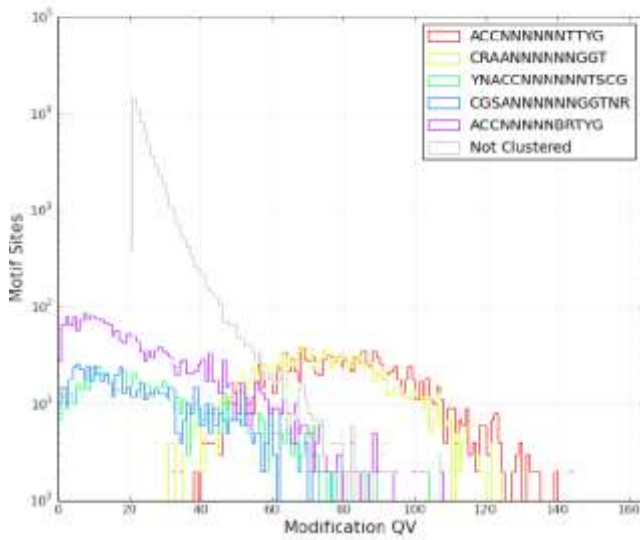


S. SauNW ACC-6-TTYG

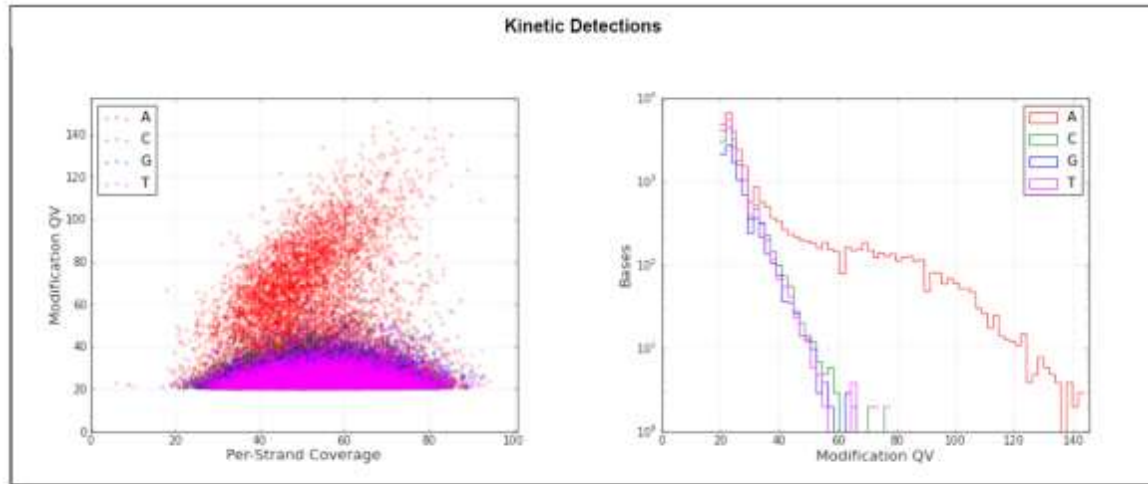
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACNNNNNNNTTYG	1	m6A	99.86	1461	1463	80.7	49.6	CRAANNNNNNNGGT
CRAANNNNNNNGGT	4	m6A	99.59	1457	1463	74.7	48.6	ACNNNNNNNTTYG
YNACNNNNNNNTSCG	3	m6A	39.52	313	792	52.0	52.7	CGSANNNNNNGGTNR
CGSANNNNNNGGTNR	4	m6A	35.23	279	792	50.1	52.3	YNACNNNNNNNTSCG
ACNNNNNNBRTYG	1	m6A	28.16	680	2415	49.3	51.3	
Not Clustered	0		0.08	6917	9110401	37.6	55.8	

Modification QV Histogram By Motif



Kinetic Detections



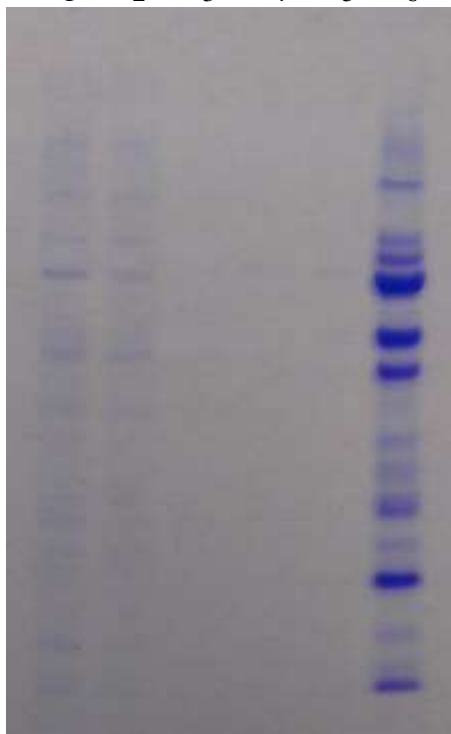
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACNNNNNNNTTYG	1	m6A	99.86	1461	1463	80.7	49.6	CRAANNNNNNNGGT
CRAANNNNNNNGGT	4	m6A	99.59	1457	1463	74.7	48.6	ACNNNNNNNTTYG
YNACNNNNNNNTSCG	3	m6A	39.52	313	792	52.0	52.7	CGSANNNNNNGGTNR
CGSANNNNNNGGTNR	4	m6A	35.23	279	792	50.1	52.3	YNACNNNNNNNTSCG
ACNNNNNNBRTYG	1	m6A	28.16	680	2415	49.3	51.3	
Not Clustered	0		0.08	6917	9110401	37.6	55.8	

S. SauNY ACC-6-TAG

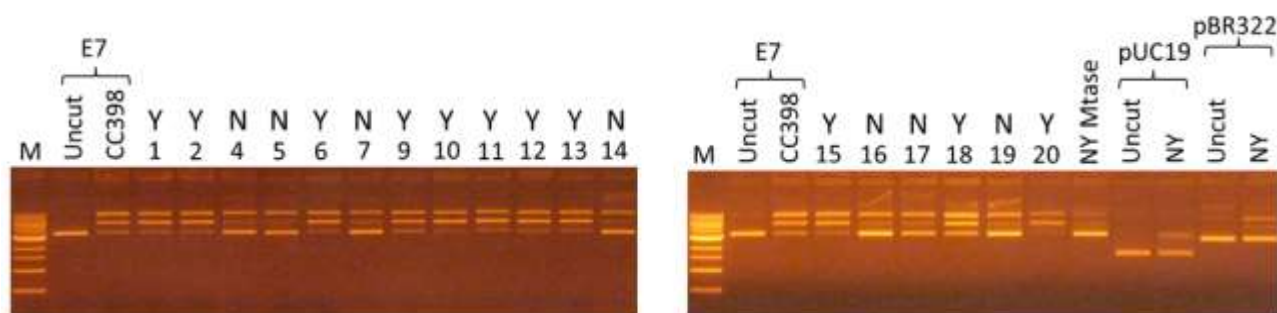
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 KWYRFMALNGD SGARADRF SIKDRTFMEMPLHIPCMDEQIKIGQFFSKLDRQIELEEQKLELLQOQ
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 YISDSLSTFMKSIAGKTATQIVNKNTFENLEIYLAPFEEQNKIADLISSLEELIEKQASKLIKMS
 RKQGMLQIMFIPGGSHHHHHH

1 2 3 4 5 6

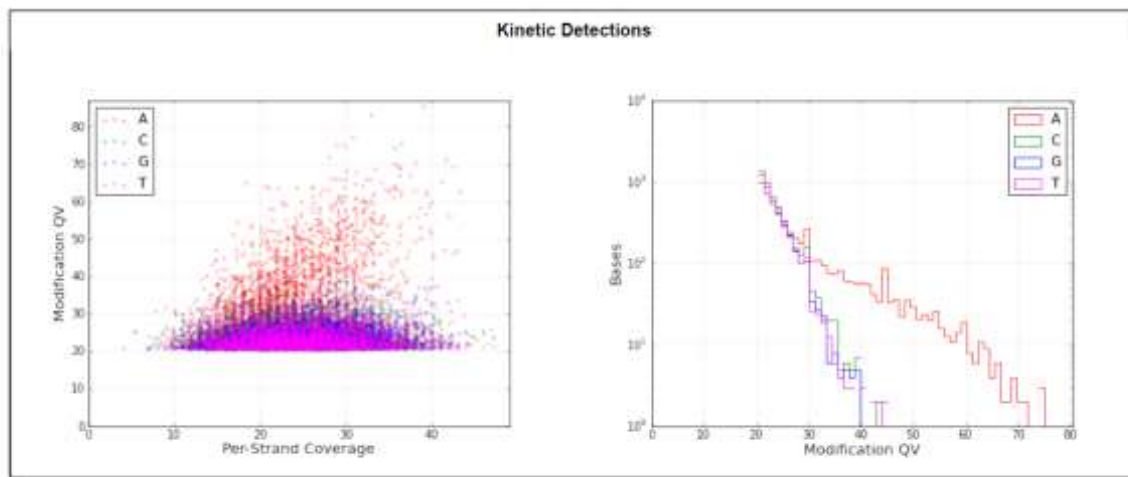


1- soluble cell extract 2- Nickel column flow through
 3- Nickel column wash 1 4- Nickel column wash 2 5- Nickel column eluate
 6- final protein after PD10 desalting and concentration of eluate

DNA cleavage assay.



S. SauNY ACC-6-TAG



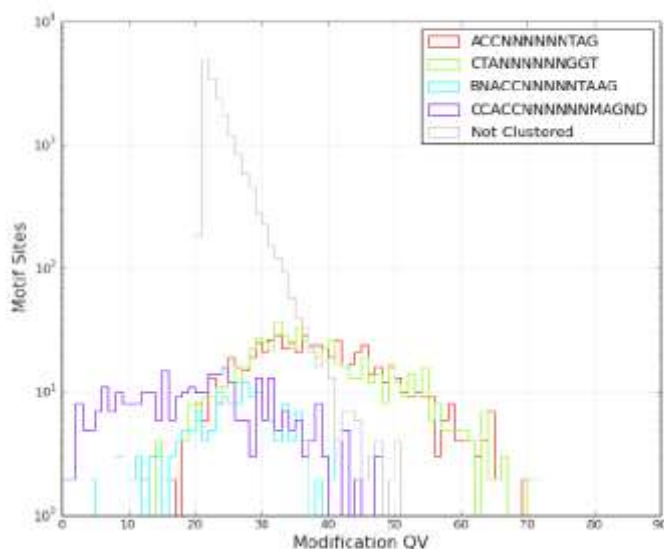
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNTAG	1	m6A	75.92	539	710	43.3	24.7	CTANNNNNGGT
CTANNNNNGGT	3	m6A	72.39	514	710	42.9	24.7	ACCNNNNNTAG
BNACCNNNNNTAAG	3	m6A	34.00	68	200	39.3	25.0	
CCACCNNNNNMAGND	3	m6A	23.68	85	359	38.3	26.1	
Not Clustered	0		0.01	622	9115347	34.4	27.6	

Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNTAG	1	m6A	75.92	539	710	43.3	24.7	CTANNNNNGGT
CTANNNNNGGT	3	m6A	72.39	514	710	42.9	24.7	ACCNNNNNTAG
BNACCNNNNNTAAG	3	m6A	34.00	68	200	39.3	25.0	
CCACCNNNNNMAGND	3	m6A	23.68	85	359	38.3	26.1	
Not Clustered	0		0.01	622	9115347	34.4	27.6	

Modification QV Histogram By Motif

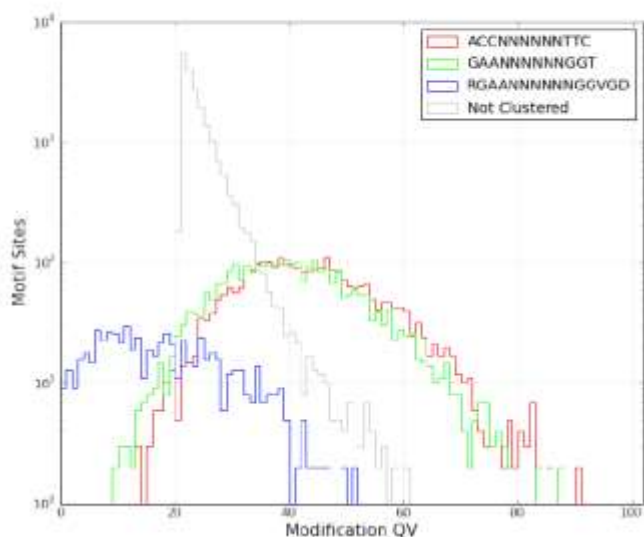


S. SauNa* ACC-6-TTC

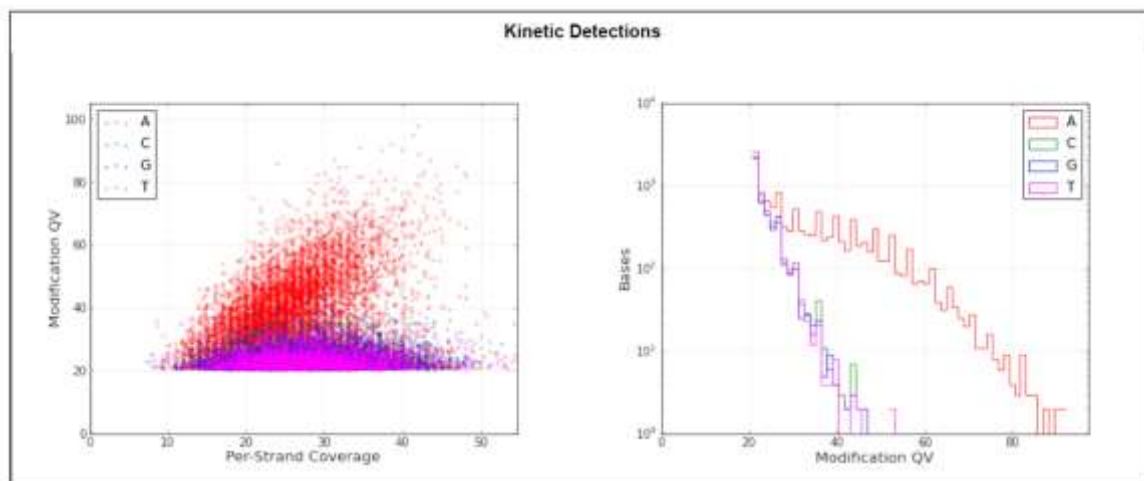
Motifs								
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNTTC	1	m6A	85.91	2567	2988	46.4	26.2	GAANNNNNGGT
GAANNNNNGGT	3	m6A	78.11	2334	2988	44.7	26.2	ACCNNNNNTTC
RGAANNNNNGGVGD	4	m6A	16.09	107	665	37.1	27.8	
Not Clustered	0		0.01	1034	9110685	35.6	31.8	

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
 NIDNIKKYYVVEENDFVYNPRMSNYAPFGPVNRNKLKGGVMSPLYTVFKIQNIDLNFIEFYFKSS
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 YYLNSLNISTFGVQAVKGVTLNNDINSIIVKLPNEEEQNI IAKFLLEVDKTVNNQLVKTLLKQR
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Modification QV Histogram By Motif



Kinetic Detections

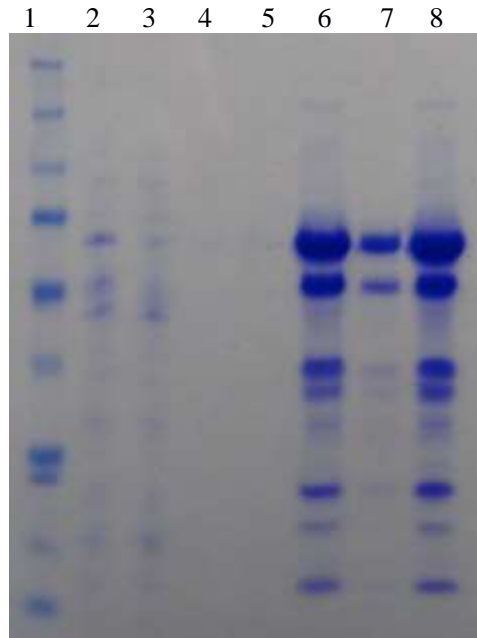


Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNTTC	1	m6A	85.91	2567	2988	46.4	26.2	GAANNNNNGGT
GAANNNNNGGT	3	m6A	78.11	2334	2988	44.7	26.2	ACCNNNNNTTC
RGAANNNNNGGVGD	4	m6A	16.09	107	665	37.1	27.8	
Not Clustered	0		0.01	1034	9110685	35.6	31.8	

S. SauNc* ACC-6-RTC

MSNTQKKNVPELRFPGFEGEWEEKKLGEFAGKVTQKNVDKKYIETLTNSAELGIISQKDYFDKEIS
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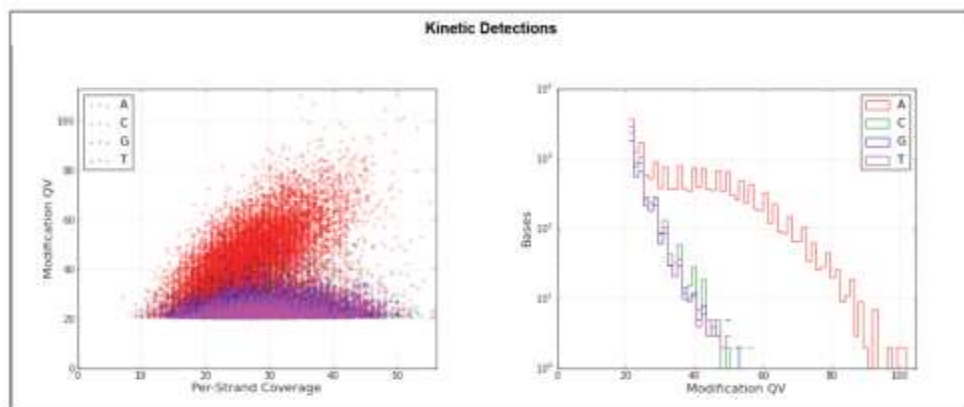
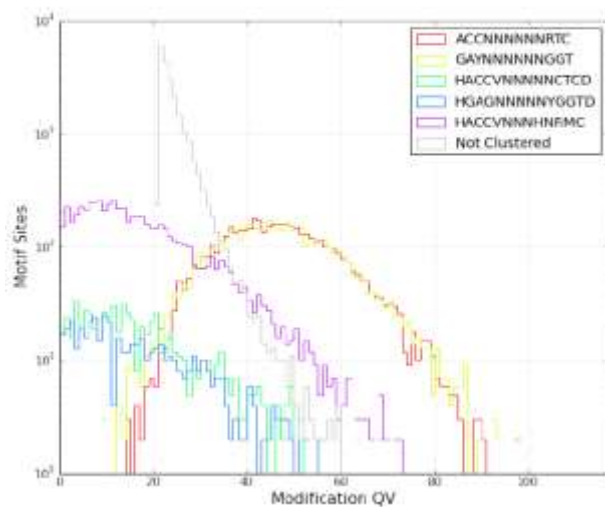
- 1- marker 2- soluble cell extract
 3- Nickel column flow through 4- Nickel column wash 1
 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after PD10 desalting
 8- final protein after concentration

Although purified, this MTase was only assayed by SMRT.

Motifs								
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNRTC	1	m5A	90.98	4680	5144	48.9	27.4	GAYNNNNNGGT
GAYNNNNNGGT	2	m5A	90.18	4639	5144	50.0	27.8	ACCNNNNNRTC
HACCVNNNNCTCD	2	m5A	16.64	117	703	40.7	30.2	
HGAGNNNNYGGTD	3	m5A	16.60	86	518	41.5	29.5	
HACCVNNNHRMC	2	m5A	14.94	936	6265	40.2	29.1	
Not Clustered	0		0.01	1163	9099552	35.6	31.2	

S. SauNc* ACC-6-RTC

Modification QV Histogram By Motif



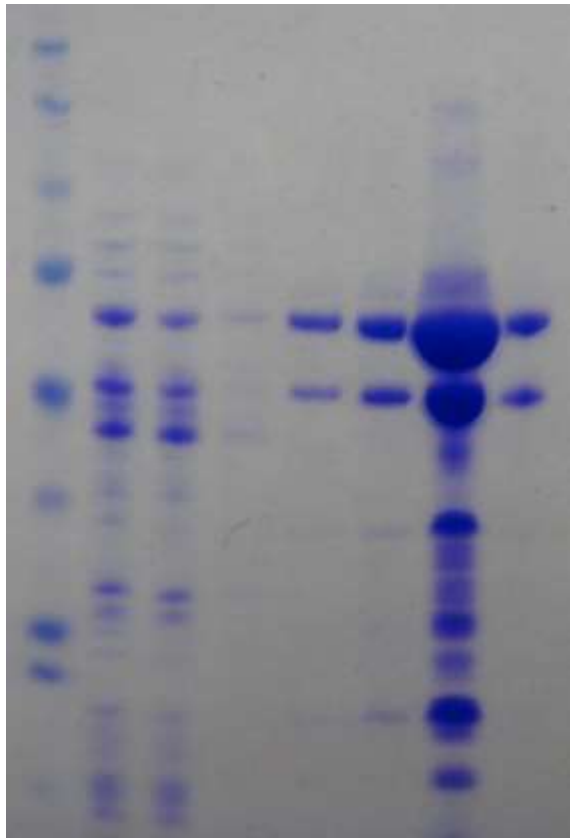
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs in Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
ACCNNNNNRTC	1	m6A	90.98	4690	5144	48.9	27.4	GAYNNNNNGGT
GAYNNNNNGGT	2	m6A	90.18	4638	5144	50.0	27.8	ACCNNNNNRTC
HACCNNNNCTCD	2	m6A	16.64	117	703	40.7	30.2	
HGACNNNNYGGTD	3	m6A	16.06	86	518	41.5	29.5	
HACCNNNNHFMTC	2	m6A	14.94	836	6205	40.2	29.1	
Not Clustered	0		0.01	1163	909502	35.6	31.2	

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60**S. SauNd* ACC-6-TTRG**

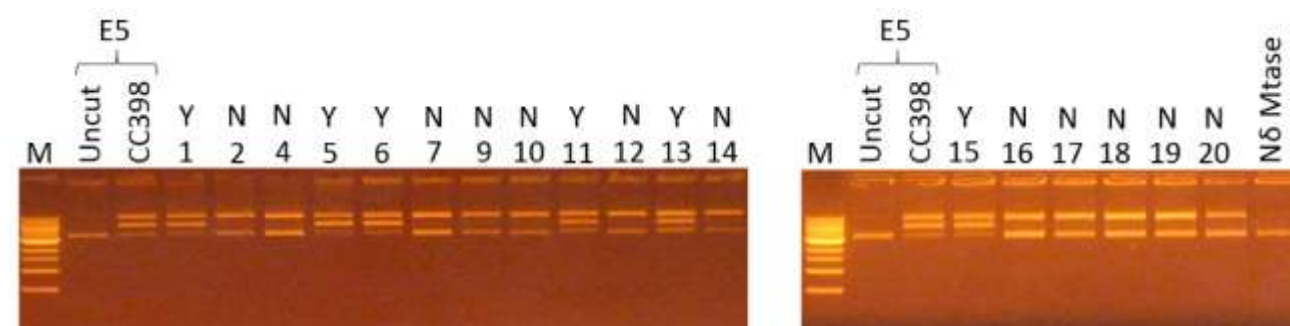
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 KKGYMQKIFSQELRFKDENGNDYPEWENVMLQKVLKDKTEGIKRGPFGGALKKDI FVESGYAVYEQ
 RNAIYDISNFRYYINENKYKEMQSFVQPNDIIMSCSGTIGRLALIPHNYTKGIINQALIRFRTNH
 KIRSEFFLIFMRSNQMQRKILEANPGSAITNLVPVKELKLI PFPLPVKFEQDKISQFIHIINRRIE
 QSEKKIESLKNRKQGFLOKLFVPPGGSHHHHHH

1 2 3 4 5 6 7 8



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 5- Nickel column eluate
 6- eluate after conc. and PD10 desalting
 7- final protein after concentration 8- NP purified protein marker

DNA cleavage assay.



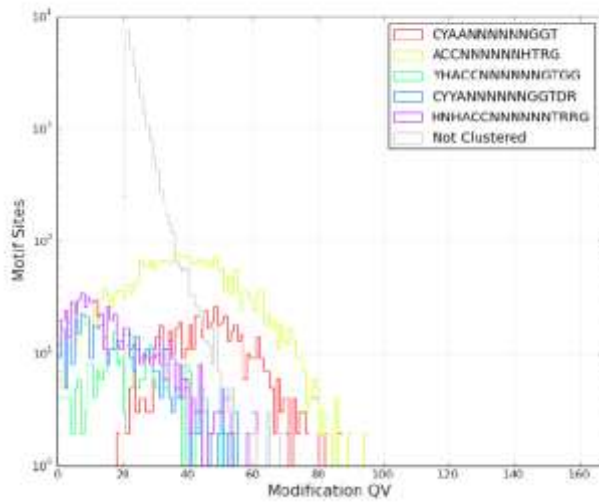
Site determined to be ACC-6-TTRG or ACC-6-YTRG. Note that the underlined site was determined by SMRT and is accepted since if Y is a cytosine, then it can't be methylated.

S. SauNd* ACC-6-TTRG

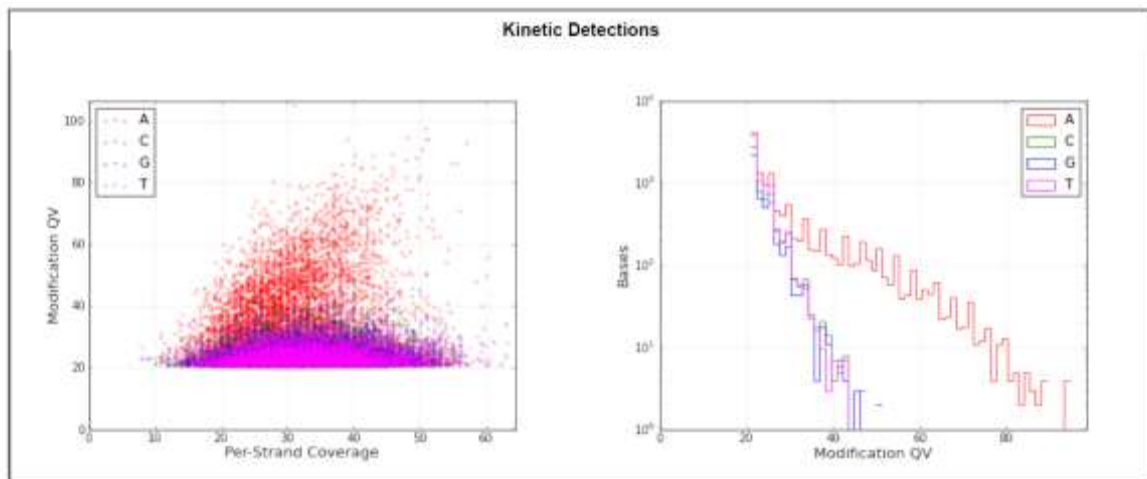
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CYAANNNNNGGT	4	m6A	89.98	557	619	50.0	29.9	
ACCNNNNNHTRG	1	m6A	67.06	2013	3002	47.5	31.0	
YHACCNNNNNGTGG	3	m6A	30.03	88	293	40.5	34.0	
CYANNNNNGGTDR	4	m6A	20.24	102	504	42.0	32.9	
HNHACCNNNNNTRRG	4	m6A	17.94	127	708	41.5	32.6	
Not Clustered	0		0.02	1435	9112200	35.8	37.4	

Modification QV Histogram By Motif



Kinetic Detections

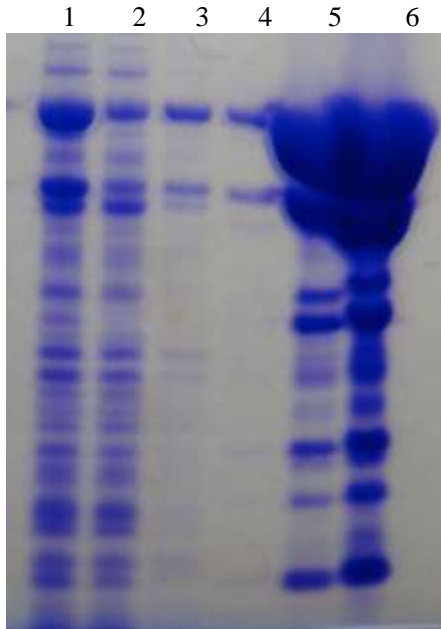


Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CYAANNNNNGGT	4	m6A	89.98	557	619	50.0	29.9	
ACCNNNNNHTRG	1	m6A	67.06	2013	3002	47.5	31.0	
YHACCNNNNNGTGG	3	m6A	30.03	88	293	40.5	34.0	
CYANNNNNGGTDR	4	m6A	20.24	102	504	42.0	32.9	
HNHACCNNNNNTRRG	4	m6A	17.94	127	708	41.5	32.6	
Not Clustered	0		0.02	1435	9112200	35.8	37.4	

S. SauRE GARA-6-RTGA

MSNTQKKNVPELRFPGFEGEWEEKKLGEVAKIYDGTHQTPKYTNEGIFLSVENIKTLNSSKYISE
 EAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYFLKNLILSSSIQN
 ELWRKTLHVAFPKKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIELEEQKLELLQQQKGYMQ
 KIFSQELRFKDENGKDYPEWEETTKEIAQINTGKKDKDAITNGSYDFYVRSPIVYKINTFSYEG
 EAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFSONFLKETKKYSAKTSVDS
 VRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKTQKQVIELLKQRKKSLLQKMFIPGGSHHHHHH

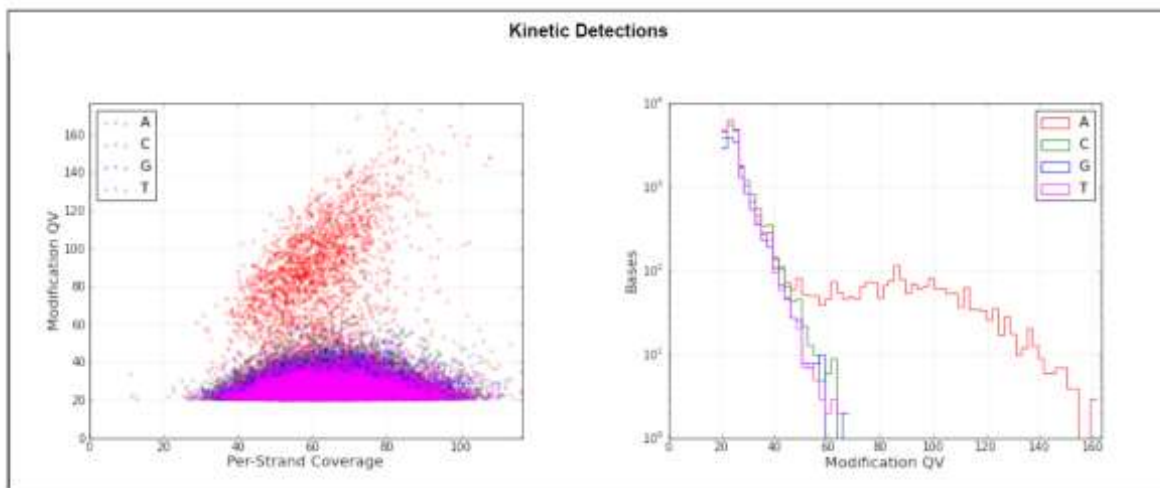


1- soluble cell extract, 2- Nickel column flow through, 3-Nickel column wash 1, 4- Nickel column wash 2, 5- Nickel column eluate, 6- Final protein after PD10 desalting and concentration

Although purified, this MTase was only used in SMRT.

Motifs								
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTYTC	3	m6A	91.88	792	862	104.6	62.5	GARANNNNNRTGA
GARANNNNNRTGA	4	m6A	79.58	686	862	96.7	61.7	TCAYNNNNNTYTC
CCACDNNNNTYTC	3	m6A	15.87	30	189	90.4	65.5	
TCAGNNTNNTNNTCNB	3	m6A	15.03	29	193	91.0	67.7	
Not Clustered	0		0.00	129	9115220	85.4	67.9	

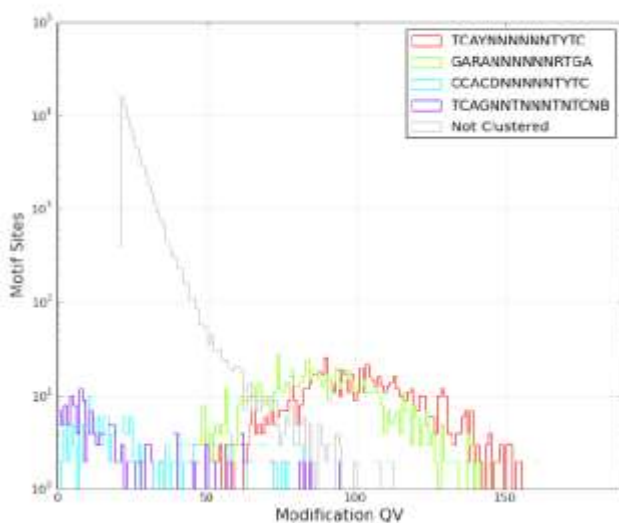
S. SauRE GARA-6-RTGA



Motifs

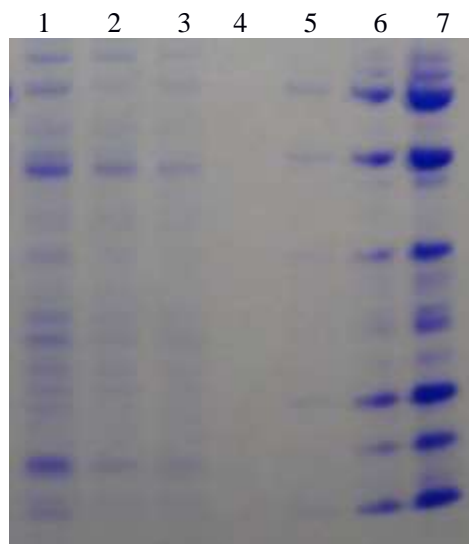
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTYTC	3	m6A	91.88	792	862	104.6	62.5	GARANNNNNRTGA
GARANNNNNRTGA	4	m6A	79.58	686	862	96.7	61.7	TCAYNNNNNTYTC
CCACDNNNNNTYTC	3	m6A	15.87	30	189	90.4	65.5	
TCAGNNTNNNTNCNB	3	m6A	15.03	29	193	91.0	62.7	
Not Clustered	0		0.00	129	9115220	85.4	67.9	

Modification QV Histogram By Motif



S. SauTE CAAG-5-RTGA

MSNTQKKNVPELRFPGFEGEWEEKELGEIFQIISGSTPLKSNKEFYENGNINWVKTTDLNNSKVTH
 SKEKITEYAMKSLKLLKLVPKNSVLIAMYGGFNQIGRTGLLKIDATINQAISALLMNHETNPEFIQA
 FLNYQVKGWKRYAASSRKDPNITKKDIEQFKVPYVSINEQQKIGEFFSKIDHQIELEEQKLELLQQ
 QKKGVMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKI
 NTFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYS
 AKTSVDSVRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKTKIQKQVIELLKQRKSSLLQKMFIPGG
 SHHHHHH

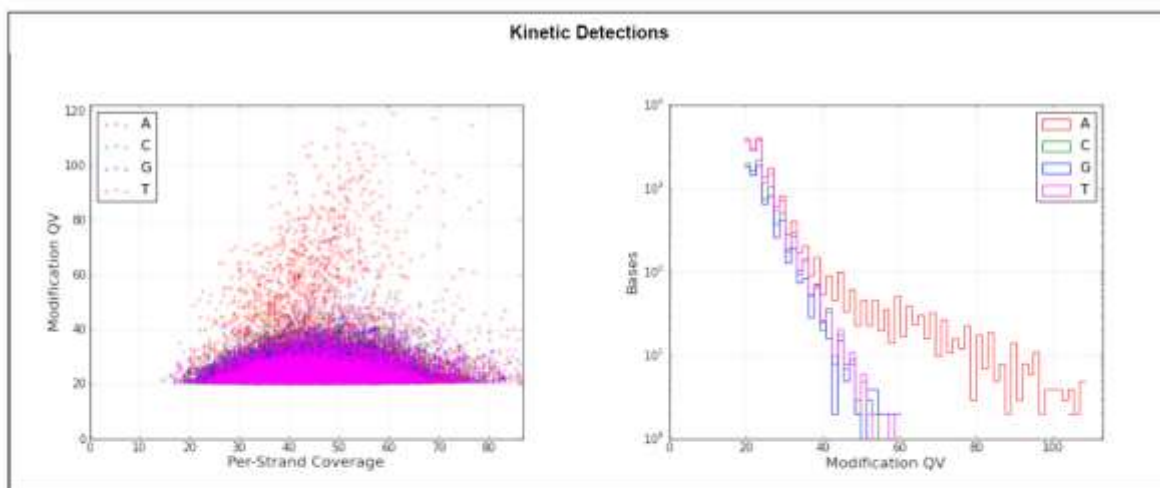
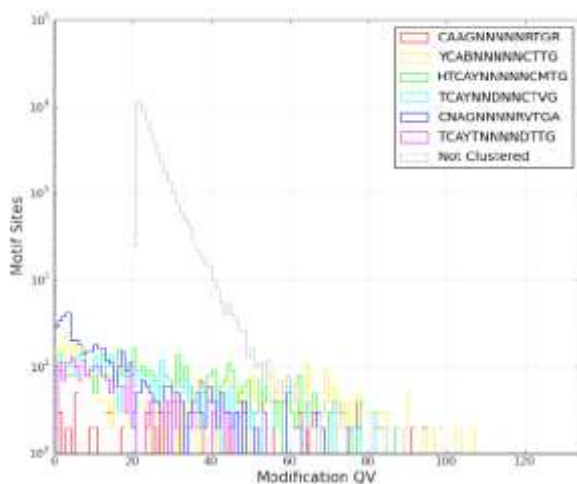


1- soluble cell extract 2- Nickel column flow through
 3-Nickel column wash 1 4- Nickel column wash 2
 5- Nickel column eluate 6- eluate after concentrating and PD10 step
 7- Final concentrated protein

This MTase did not purify well and was only analysed by SMRT sequencing. The degeneracy in the target determined by SMRT sequencing can be removed using results from other systems.

Motifs								
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CAAGNNNNRTGR	3	m6A	80.15	214	267	63.8	42.7	
YCABNNNNCTTG	3	m6A	55.08	260	472	65.1	43.1	
HTCAYNNNNCMTG	4	m6A	47.41	238	502	51.7	44.4	
TCAYNNDNCTVG	3	m6A	31.07	119	383	51.0	44.3	
CNAGNNNNRVTGA	3	m6A	28.62	170	594	59.5	43.6	
TCAYTNNNDTTG	3	m6A	16.89	38	225	45.4	44.3	
Not Clustered	0		0.04	3962	9114883	35.8	50.6	

S. SauTE CAAG-5-RTGA
Modification QV Histogram By Motif



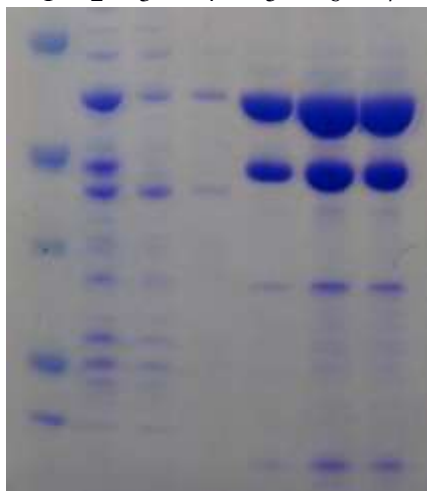
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CAAGNNNNRTGR	3	m6A	80.15	214	267	63.8	42.7	
YCABNNNNCTTG	3	m6A	55.08	260	472	65.1	43.1	
HTCAYNNNNCMTG	4	m6A	47.41	238	502	51.7	44.4	
TCAYNNDNNCTVG	3	m6A	31.07	119	383	51.0	44.3	
CNAGNNNNRTGA	3	m6A	28.62	170	594	59.5	43.6	
TCAYTNNNDTTG	3	m6A	16.89	38	225	45.4	44.3	
Not Clustered	0		0.04	3062	9114883	35.8	50.6	

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60**S. SauVE CNGA-6-RTGA**

MSNTQKKNVPELRFPGFEGEWEEKELRELNRNPKDKYSYTGPFSGDLKKS DYTTDGIQIIQLQNI
 DGYFYNSNKVFTSNEKAEVLKSCNVFPGDIVIAKMADPIARAAI VPDNNIGKYL MASDGIRLSVDT
 VHFNTK FVLE C INRKSFRKKVEDNSSGSTRMRIGLSTLGSLTLKTTTLKEQQKIGQFFSKLDRQIE
 LEEQKLELLQQQKGYM QKIFSQELRFKDENGKDYPEWEETT IKEIAQINTGKKDTKDAITNGSYD
 FYVRSPIVYKINTFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYYS
 QNFLKETKKYSAKTSVDSVRKDMIANMKVPRPIYIEQKKIGQFIKRV DNKTKIQKQVIELLKQRKK
 SLLQKMFIPGGSHHHHHH

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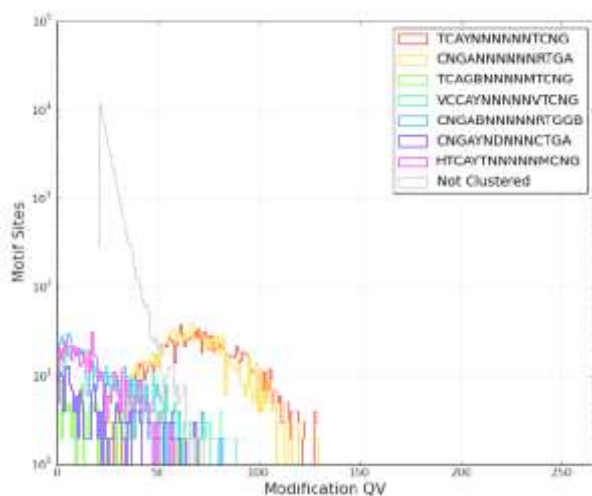
1- marker, 2- soluble cell extract, 3- Nickel column flow through, 4- Nickel column wash, 5- Nickel column eluate, 6- eluate after conc. and PD10 desalting, 7- Final protein after concentration
 Although the MTase was purified, it was only analysed via SMRT sequencing.

Motifs

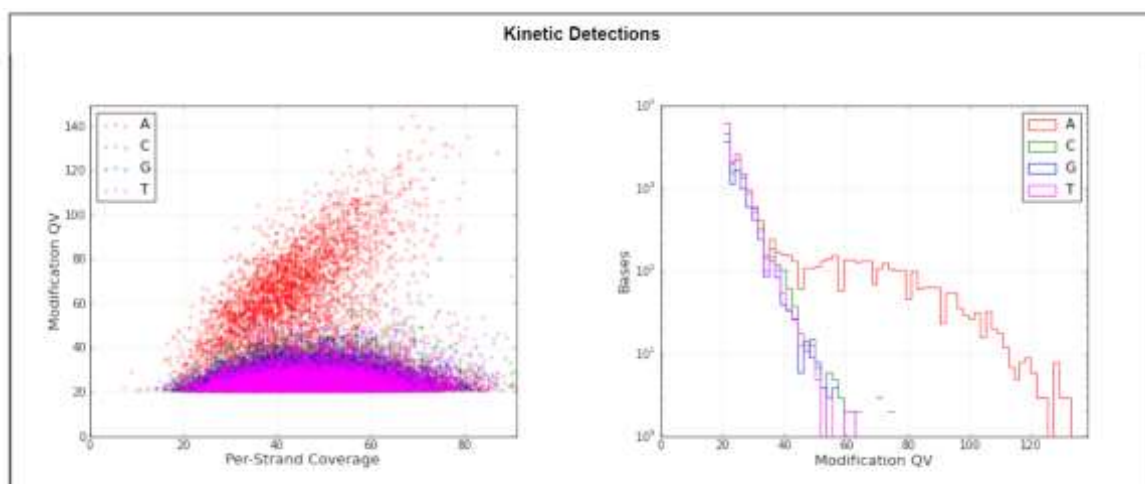
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTCNG	3	m6A	99.34	1354	1363	73.0	43.2	CNGANNNNNRTGA
CNGANNNNNRTGA	4	m6A	98.09	1337	1363	71.4	43.4	TCAYNNNNNTCNG
TCAGBNNNNMTCNG	3	m6A	41.58	79	190	53.7	43.2	
VCCAYNNNNVTCNG	4	m6A	33.12	211	637	50.8	44.7	CNGABNNNNRTGGB
CNGABNNNNRTGGB	4	m6A	29.98	191	637	50.2	45.8	VCCAYNNNNVTCNG
CNGAYNDNNCTGA	4	m6A	31.90	74	232	53.0	44.9	
HTCAYTNNNNMCNG	4	m6A	23.31	131	562	44.6	45.5	
Not Clustered	0		0.05	4345	9112342	35.9	49.9	

S. SauVE CNGA-6-RTGA

Modification QV Histogram By Motif



Kinetic Detections



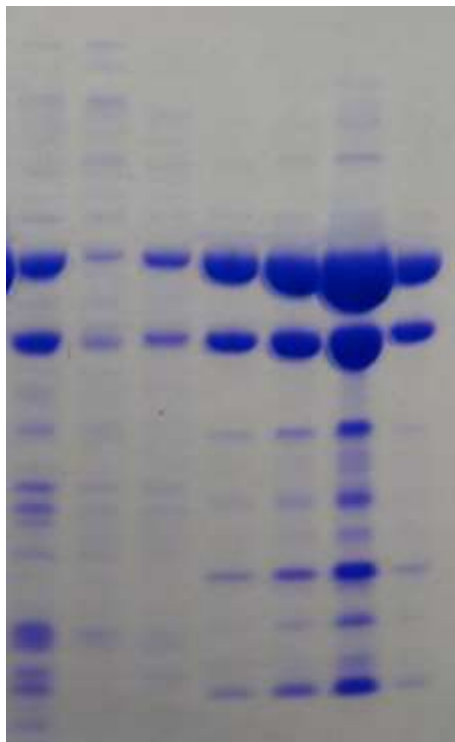
Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTCNG	3	m6A	99.34	1354	1363	73.0	43.2	CNGANNNNNRTGA
CNGANNNNNRTGA	4	m6A	98.09	1337	1363	71.4	43.4	TCAYNNNNNTCNG
TCAGBNNNNMTCNG	3	m6A	41.58	79	190	53.7	43.2	
VCCAYNNNNVTCNG	4	m6A	33.12	211	637	50.8	44.7	CNGABNNNNRTGGB
CNGABNNNNRTGGB	4	m6A	29.98	191	637	50.2	45.8	VCCAYNNNNVTCNG
CNGAYNDNNCTGA	4	m6A	31.90	74	232	53.0	44.9	
HTCAYTNNNNMCNG	4	m6A	23.31	131	562	44.6	45.5	
Not Clustered	0		0.05	4345	9112342	35.9	49.9	

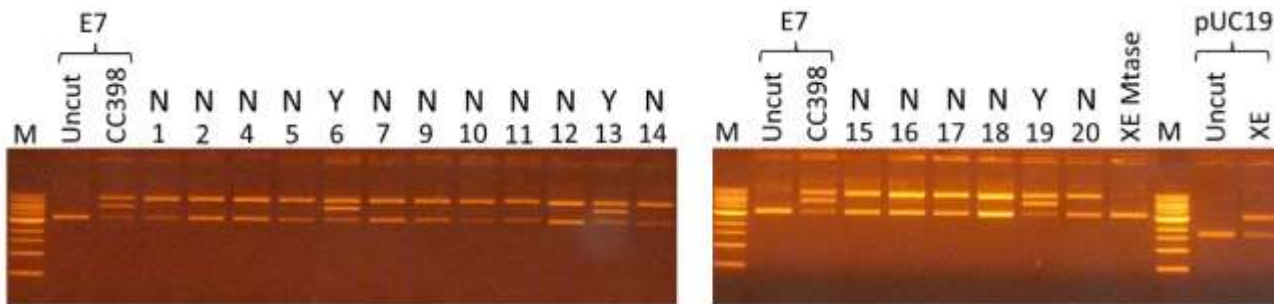
S. SauXE TCTA-6-RTGA

MSNTQKKNVPELRFPGFEGEWEEKQFADF^TKINQGLQIAINERKTEYSPELYFYITNEFLRPNSQT
 KYFIENPPQS^VIANKEDILMTRTGN^TGKVVTNVFGAFHNNFFKIKFDKNLYDRLFLVEVLN^SSKIQ
 NKILSLAGSSTIPDLNHSDFYSISSSYPLLREQQKIGKFFSKLDRQIELEE^QKLELLQQQKGYMQ
 KIFSQELRFKDENGKDYPEWEETT^IKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKINTFSYEG
 EAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYYSQNFLKETKKYSAKTSVDS
 VRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKT^KIQKQVIELLKQRKKSLLQKMFIPGGSHHHHHH

1 2 3 4 5 6 7



1- soluble cell extract 2- Nickel column flow through
 3- Nickel column wash 4- Nickel column eluate
 5- eluate after conc. and PD10 desalting
 6- final protein after concentration
 7- CC398-1 purified protein marker
 DNA cleavage assay.



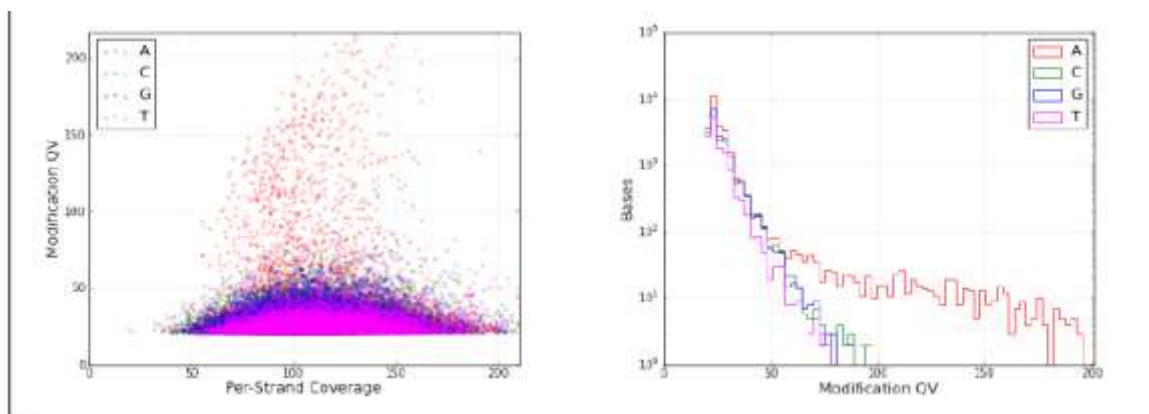
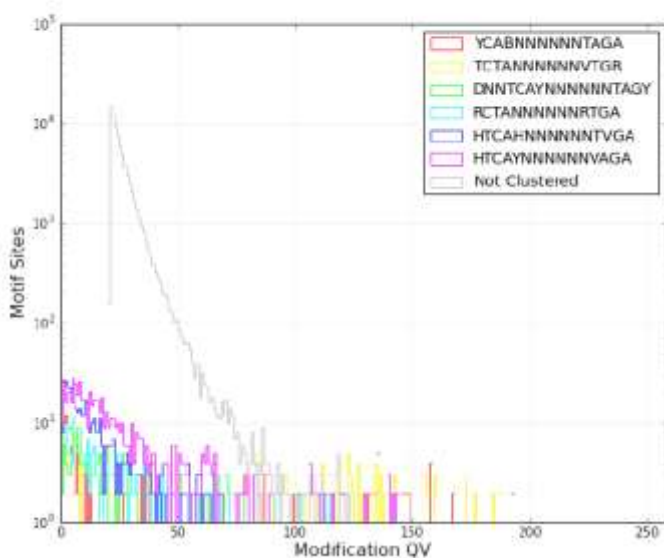
S. SauXE TCTA-6-RTGA

The degeneracy in the target determined by SMRT sequencing can be resolved by reference to targets from other systems.

Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
YCABNNNNNTAGA	3	m6A	68.77	196	285	121.6	104.1	TCTANNNNNVTGR
TCTANNNNNVTGR	4	m6A	68.07	194	285	120.0	106.6	YCABNNNNNTAGA
DNNTCAYNNNNNTAGY	6	m6A	44.57	82	184	79.3	103.0	
RCTANNNNNRTGA	4	m6A	40.41	99	245	71.8	104.1	
HTCAHNNNNNTVGA	4	m6A	32.43	156	481	103.9	104.2	
HTCAYNNNNNVAGA	4	m6A	32.07	211	658	67.2	107.8	
Not Clustered	0		0.12	10813	9115188	38.1	115.4	

Modification QV Histogram By Motif

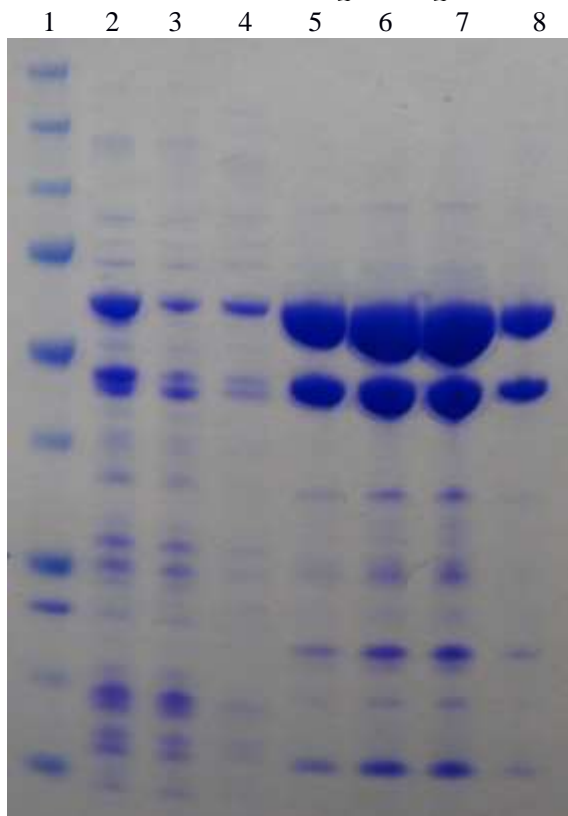


Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
YCABNNNNNTAGA	3	m6A	68.77	196	285	121.6	104.1	TCTANNNNNVTGR
TCTANNNNNVTGR	4	m6A	68.07	194	285	120.0	106.6	YCABNNNNNTAGA
DNNTCAYNNNNNTAGY	6	m6A	44.57	82	184	79.3	103.0	
RCTANNNNNRTGA	4	m6A	40.41	99	245	71.8	104.1	
HTCAHNNNNNTVGA	4	m6A	32.43	156	481	103.9	104.2	
HTCAYNNNNNVAGA	4	m6A	32.07	211	658	67.2	107.8	
Not Clustered	0		0.12	10813	9115188	38.1	115.4	

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60**S. SauZE GAC-5-RTGA**

MSNTQKKNVPELRFPGFEGEYSLDIFGNLATNKSEKFNPNQENASIDIELDCIEQNTGRLIKIYNS
 KEFSSQKNKFNPNQNVLYGKLRPYLNKYYFTKKSVCSSSEIWLKSTKEDKLLNLFYFIQTKRYS
 DVASKSAGSKMPRADWGLIENIRVYFPELCEQQKIGQFFSKLDRQIELEEQLLELLQQKKGVMQK
 IFSQELRFKDENGKDYPEWEETTIKEIAQINTGKDKTKDAITNGSYDFYVRSPIVYKINTFSYEGE
 AILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYSAKTSVDSV
 RKDMIANMKVPRPIYIEQKKIGQFIKRVDNKTKIQKQVIELLKQRKKSLLQKMFIPGGSHHHHHH*

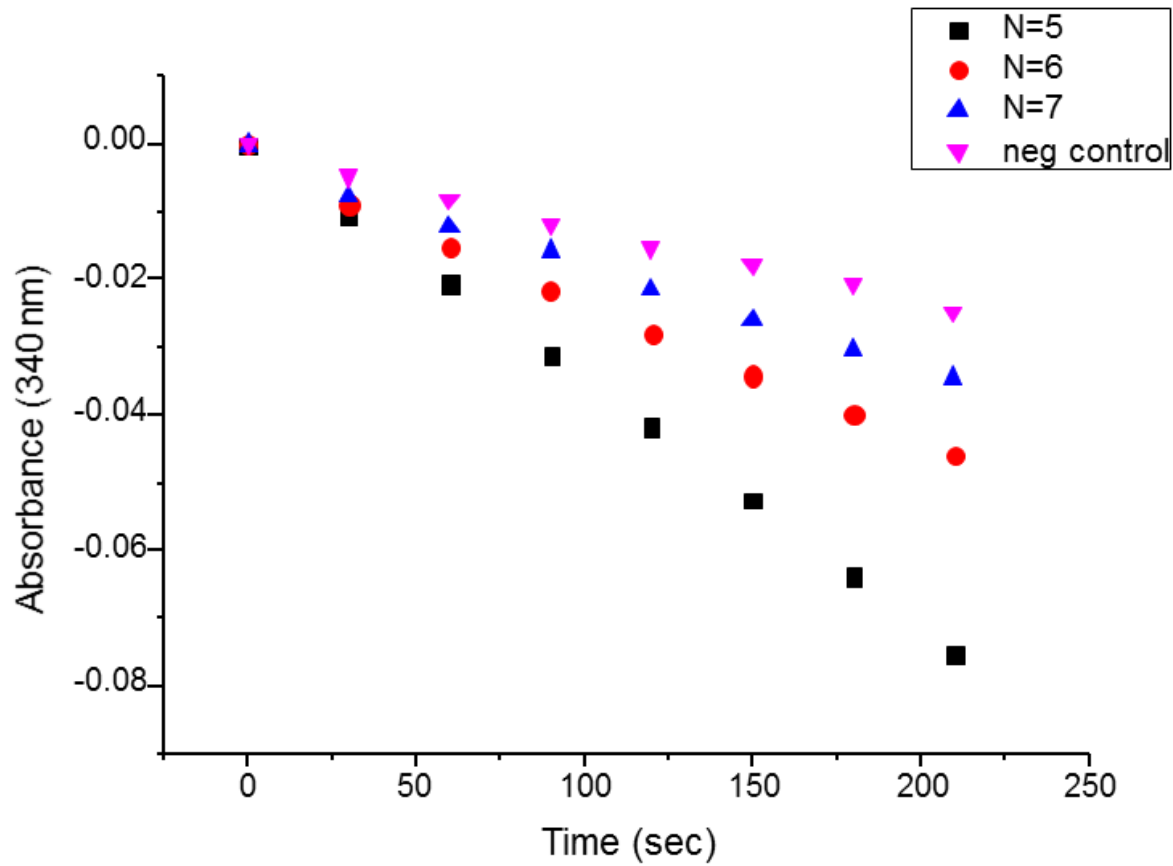


1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 5- Nickel column eluate 6- eluate after conc. step and PD10 desalting
 7- final concentrated protein 8- CC398-1 purified protein marker

DNA cleavage assay showed cutting of all plasmids so the ATPase assay was used given that we knew the individual TRD specificities.

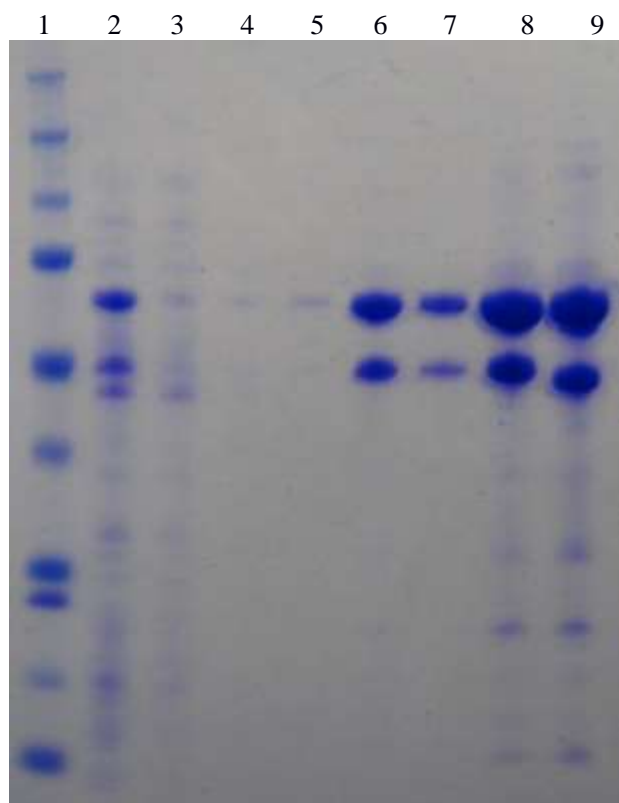
Oligonucleotide name	DNA sequence (5' to 3')
ZE5for	AGATGATGGAATCAATGCGACTTCCAGTGAGCCCTATACGATATAA
ZE5rev	TTATATCGTATAGGGCTCACTGGAAGTCGCATTGATTCCATCATCT
ZE6for	AGATGATGGAATCAATGCGACTTCCATGTGAGCCCTATACGATATAA
ZE6rev	TTATATCGTATAGGGCTCACATGGAAGTCGCATTGATTCCATCATCT
ZE7for	AGATGATGGAATCAATGCGACTTCACATGTGAGCCCTATACGATATAA
ZE7rev	TTATATCGTATAGGGCTCACATGTGAAGTCGCATTGATTCCATCATCT

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2 **S. SauZE** **GAC-5-RTGA**
3 N=5 gives the clearest signal.
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60**S. SauZS GAC-6-TGC**

MSNTQKKNVPELRFPGFEGEYSLDIFGNLATNKSEKFNPNQENASIDIELDCIEQNTGRLIKIYNS
 KEFSSQKNKFNPNQNVLYGKLRPYLNKYYFTKKSGVCSSEIWVLKSTKEDKLLNLFYFYFIQTKRYS
 DVASKSAGSKMPRADWGLIENIRVYFPELCEQQKIGQFFSKLDRQIELEEQLLELLQQQKKGVMQK
 IFSQELRFKDENGNDYPDWTNERLGEVTTVTMGQSPKSVNYTDNSNDTVLIQGNADIENGLINPRI
 YTREVTCLIQKDEIILTVPVPGKLAMAQINACIGRGVCSIKGDKFLYYFLEWFATQNKWIRFSQG
 STFESISGNDIRNIHIKIPVEDERTKI IKLLNSLDVLSKTDLKIQLNKQRKQSLLOKIFVPPGSH
 HHHHH*



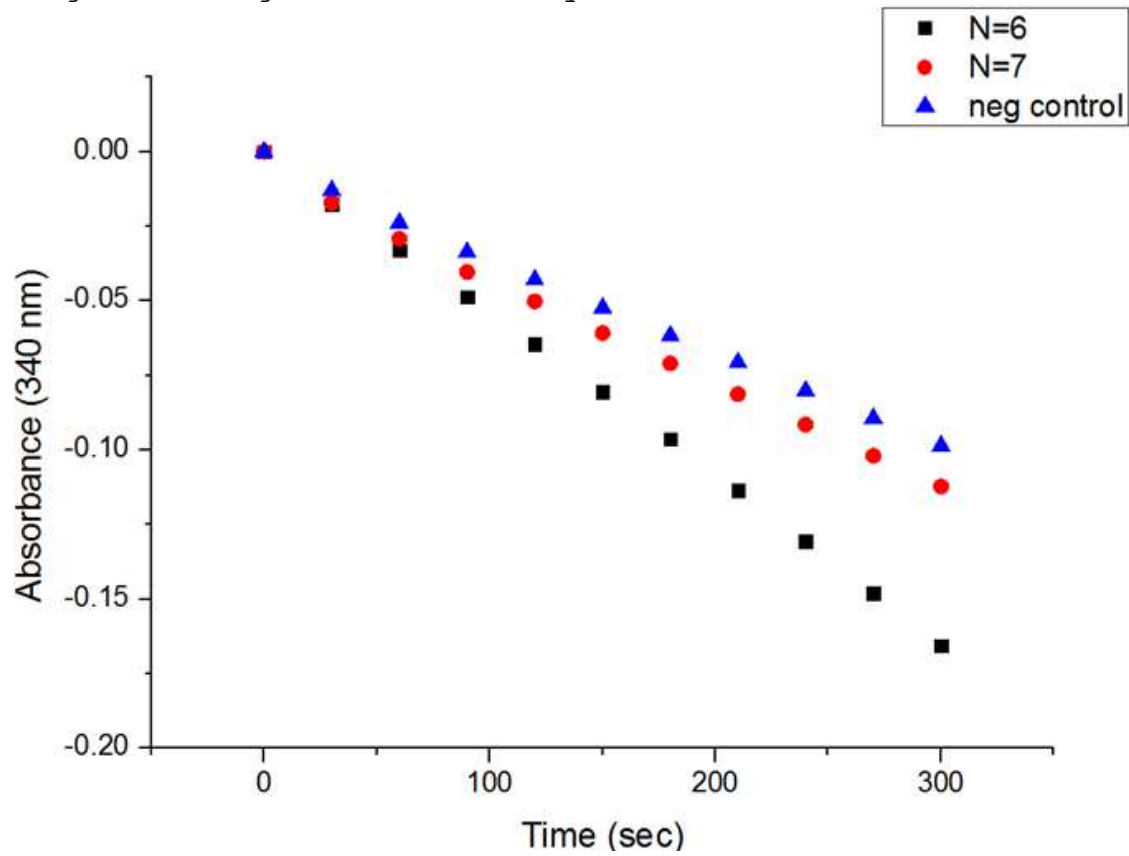
1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after PD10 desalting 8- final protein after concentration
 9- NQ purified protein marker

The DNA cleavage assay showed cutting of all plasmids so the ATPase assay was used since we knew the TRD specificities.

Oligonucleotide name	DNA sequence (5' to 3')
ZS6for	AGATGATGGAATCAATGCGACTTCCATTGCGCCCTATACGATATAA
ZS6rev	TTATATCGTATAGGGCGCAATGGAAGTCGCATTGATTCCATCATCT
ZS7for	AGATGATGGAATCAATGCGACTTCACATTGCGCCCTATACGATATAA
ZS7rev	TTATATCGTATAGGGCGCAATGTGAAGTCGCATTGATTCCATCATCT

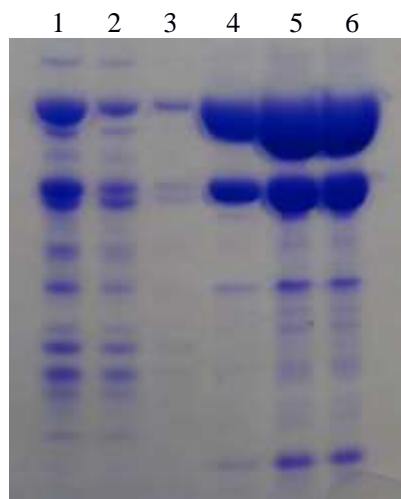
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2 **S. SauzS** **GAC-6-TGC**

3 N=6 gives the greatest activity.
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60**S. Saub*E GGHA-6-RTGA**

MSNTQKKNVPELRFPGFEGEWEEKKLEDTLLEFIKDGTHGTHENVNNGPWLLSAKNIKNNKIISSD
 DRKISESDYKKIYKNYKLEKGDLLLTIVGTIGRAAIVKNPNNIAFQRSVAILKTKATYDVGFIFQL
 FQTKYFKNLLLRKQVVSQAQPLYLGDIRKIKISITNIIIEQRKIGIFFSKLDRQIELEEOKLELLQ
 QQKKGVMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYK
 INTFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFSONFLKETKKY
 SAKTSVDSVRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKTQKIQKQVIELLKQRKKSLLQKMFIPG
 GSHHHHHH

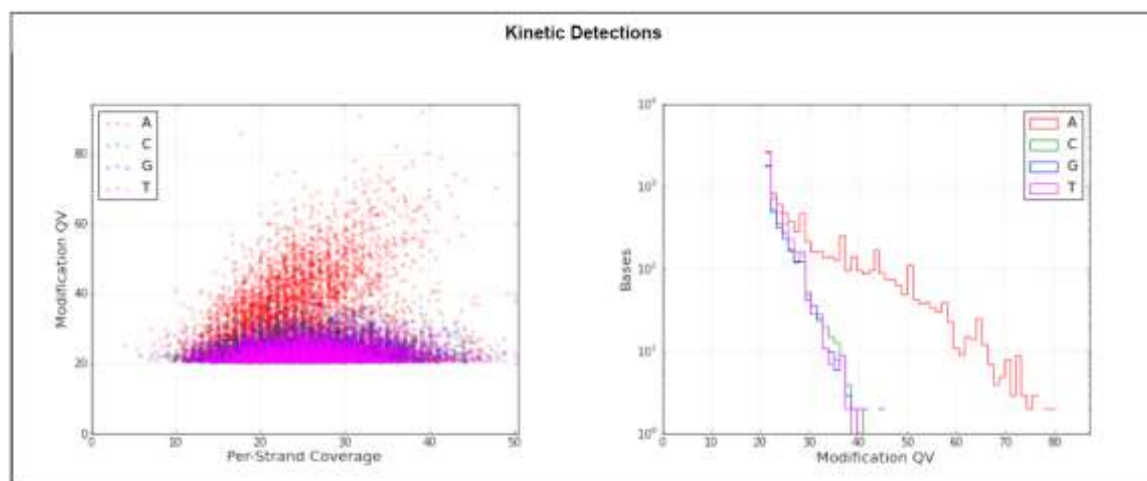
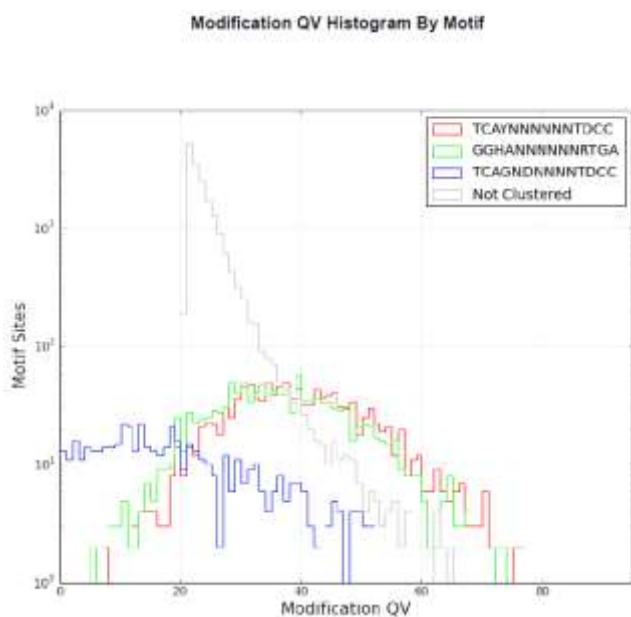


1- soluble cell extract, 2- Nickel column flow through, 3- Nickel column wash, 4- Nickel column eluate, 5- eluate after conc. step and PD10 desalting, 6- Final concentrated protein

Although this MTase was purified, it was only used in the SMRT sequencing assay.

Motifs								
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTDCC	3	m6A	77.39	989	1278	44.5	24.6	GGHANNNNNRTGA
GGHANNNNNRTGA	4	m6A	67.68	865	1278	43.2	24.7	TCAYNNNNNTDCC
TCAGNDNNNTDCC	3	m6A	21.57	110	510	41.0	25.2	
Not Clustered	0		0.01	928	9114260	36.7	27.3	

S. Saub*E GGHA-6-RTGA

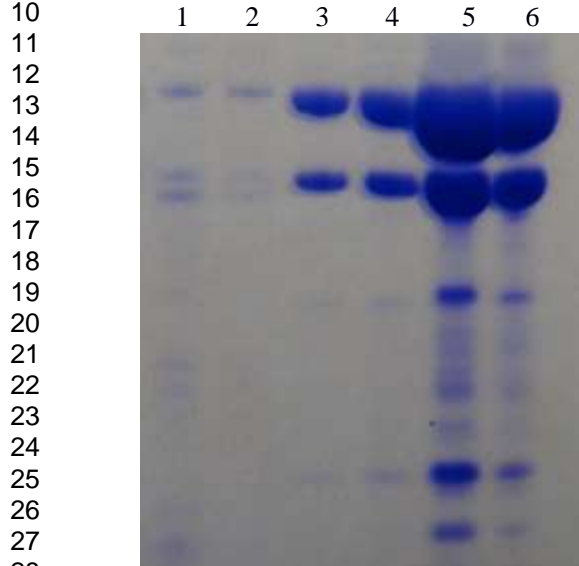


Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNTDCC	3	m6A	77.39	889	1278	44.6	24.6	GGHANNNNNRTGA
GGHANNNNNRTGA	4	m6A	67.68	885	1278	43.2	24.7	TCAYNNNNNTDCC
TCAGNDDNNNTDCC	3	m6A	21.57	110	510	41.0	25.2	
Not Clustered	0		0.01	928	9114260	36.7	27.3	

1
2 **S. Saue*E GAG-6-RTGA**

3 MSNTQKKNVPELRFPGFEGEWEEKSISSEFLKESKIKGSNGSHAKKLTVKLWGKGVVPPKETFKGSD
4 NTQYYKRKAGQLMYGKLDLDFLNCAFGIVPDSLNNYESTIDSPSFDFFINGDSKFLLERIKLKSFYKFF
5 GDIANGSRKAKRINQDTFLSLPVFAPKYDEQLRIGEFFSKLDRQIELEEQKLELLQQQKKGVMQKI
6 FSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKINTFSYEGEA
7 ILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYYFSQNFLKETKKYSAKTSVDSVR
8 KDMIANMKVPRPIYIEQKKIGQFIKRVDNKTKIQQQVIELLKQRKKSLLQKMFIPGGSHHHHHH



28 1- soluble cell extract, 2- Nickel column flow through, 3- Nickel column eluate, 4- eluate after
29 PD10 desalting, 5- Final concentrated protein, 6- RE purified protein as marker

31 DNA cleavage assay.

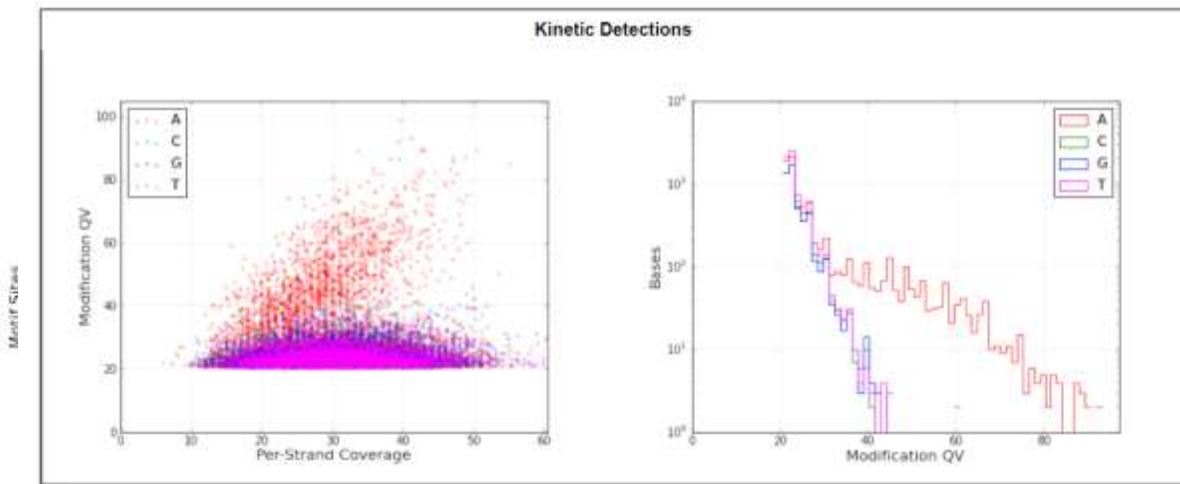


S. Saue*E GAG-6-RTGA

Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNCTC	3	m6A	88.60	762	860	49.4	28.5	GAGNNNNNRTGA
GAGNNNNNRTGA	2	m6A	87.33	751	860	50.0	28.2	TCAYNNNNNCTC
GAGNDNNNGTGGB	2	m6A	20.22	37	183	40.9	29.5	
DNNGAGNDNNNGAGA	5	m6A	18.56	36	194	39.8	32.7	
Not Clustered	0		0.01	914	9115229	34.6	36.3	

Kinetic Detections



Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNCTC	3	m6A	88.60	762	860	49.4	28.5	GAGNNNNNRTGA
GAGNNNNNRTGA	2	m6A	87.33	751	860	50.0	28.2	TCAYNNNNNCTC
GAGNDNNNGTGGB	2	m6A	20.22	37	183	40.9	29.5	
DNNGAGNDNNNGAGA	5	m6A	18.56	36	194	39.8	32.7	
Not Clustered	0		0.01	914	9115229	34.6	36.3	

1
2 SUPPLEMENTARY INFORMATION FOR TABLE 4.
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S. SauAc* CCAY-6-RTC

The Ac* TRD combination is found in CC97-1. The MTase was not purified but instead used to methylate the genome of E. coli ER2796 for SMRT analysis. The target is CCAYNNNNNNRTC. There are a few minor amino acid differences in the S.SauAc* between members of CC97.

CC97

Recombinant S.SauAc* CC97-1

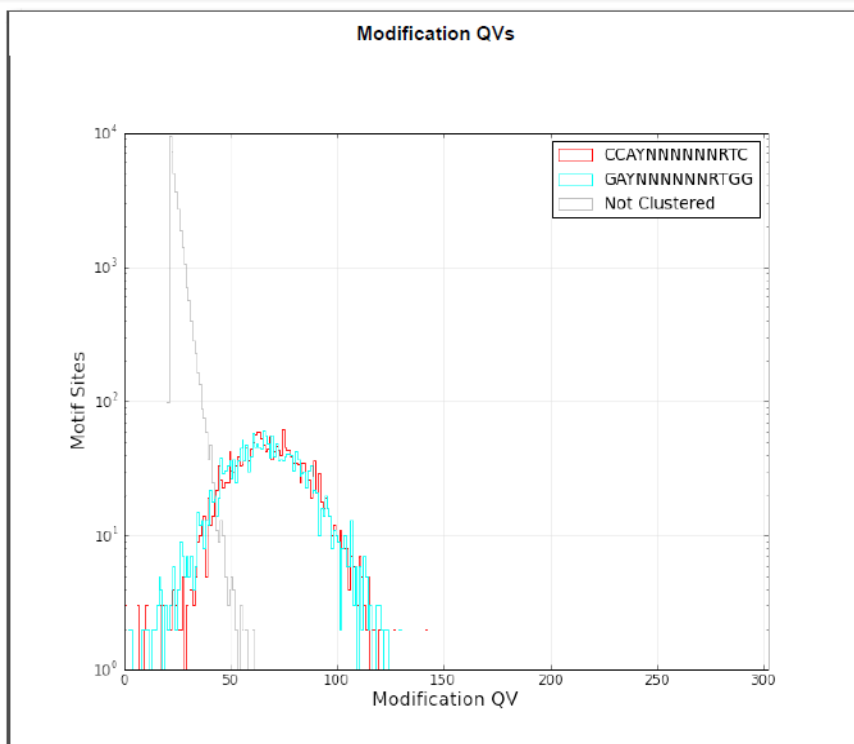
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 KNSRYYGDVLLNITGASIGRTAINSIVEIHANLNQHVCIIRLKKEYYYNFFGQYLLSRKGKRKIFLAQSGGSREGLNFK
 EIANLKI FTPTIFEEQQKIGEFISKDRQIELEEQKLELLQQQKGYLQKIFSQELRFKDENGNDYPEWRFARFKDFMYK
 PINIRPAINISKSELLTVKLHCKGIEKANINRVLKLGATNYYKRFEGQFIYKQNFNGAFDIVPKKFDGLYSSSDVPAF
 EINTEKIEPNYFISYISRPSFYKSKEKYSTGTGSKRIHENTVNLNFSLHLPCLNEQLKIASFVCFNLNKIELLERKIYLIK
 KQKQALLQQMFI PGGSHHHHHH

Wild Type S.SauAc*

MSNTQKKNVPELRFPGFEGEWEEKQLGLDLTTKIGSGKTPKGGSENYTNKGIPFLRSQNIRNGKLNLDLVYISKDIDDEM
 KNSRYYGDVLLNITGASIGRTAINSIVETHANLNQHVCIIRLKKEYYYIFFGQYLLSRKGKRKIFLAQSGGSREGLNFK
 EIANLKI FTPTIFEEQQKIGKFFSKLDRQIELEEQKLELLQQQKGYLQKIFSQELRFKDENGNDYPEWRFARFKDFMYK
 PINIRPAINISKSELLTVKLHCKGIEKANINRVLKLGATNYYKRFEGQFIYKQNFNGAFDIVPKKFDGLYSSSDVPAF
 EINTEKIEPNYFISYISRPSFYKSKEKYSTGTGSKRIHENTVNLNFSLHLPCLNEQLKIASFVCFNLNKIELLERKIYLIK
 KQKQALLQQMFI*

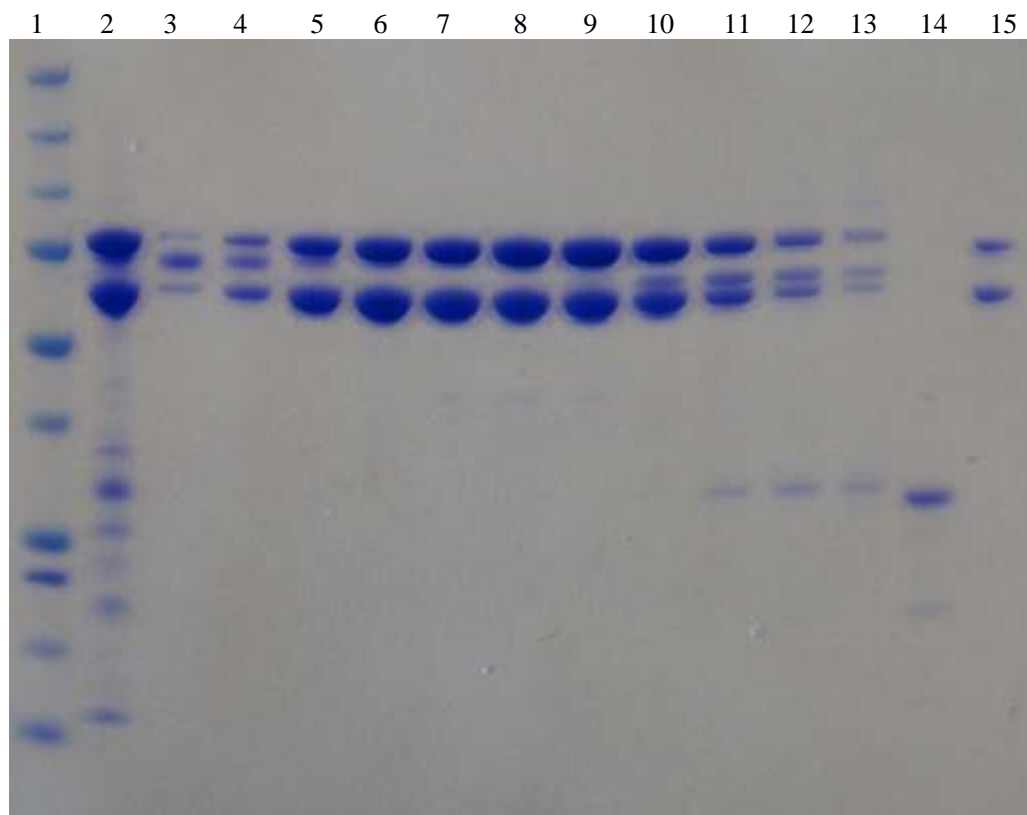
SMRT Cells: 1 Movies: 1

Motif Summary								
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CCAYNNNNNNRTC	3	m6A	97.52%	2199	2255	68.95	50.78	GAYNNNNNNRTGG
GAYNNNNNNRTGG	2	m6A	96.01%	2165	2255	68.50	51.01	CCAYNNNNNNRTC



S. SauBI-EGFP**CC22-1** **AGG-6-TGAR****This MTase was expressed and purified as a fusion with EGFP.****Nuclease assays and SMRT analysis gave the same target site.**

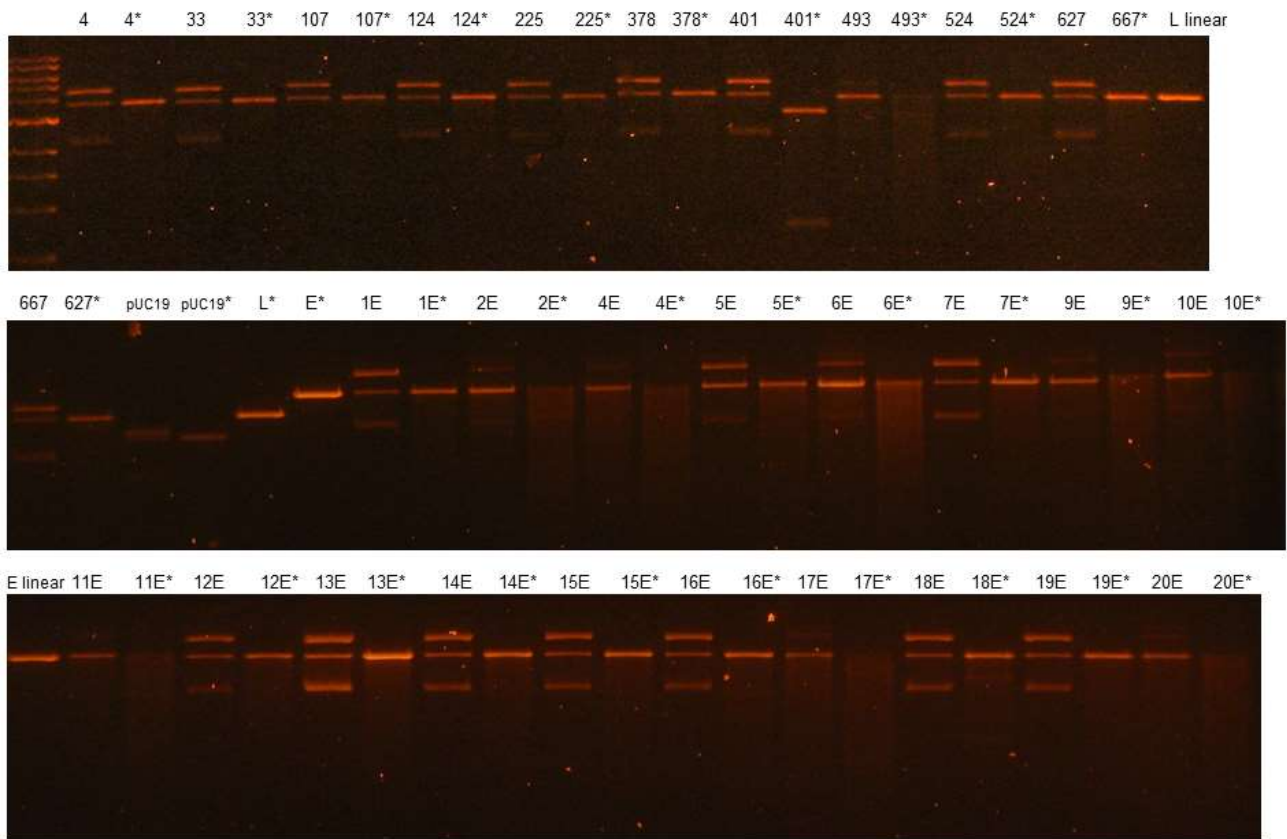
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 HWYREVSGIAVEGARNHGLLNVSVDFFTTILIKYPSLEEQQKIGKFFSKLDRQIELEEOKLELLQO
 QKKGYMOKIIFSQELRFKNENGDYPDWERIKFFDVIDKVIDFRGRTPKKNMEWSDEGYLALSAVN
 VKKGYIDFNVEAKYGNLDLYTRWMRGNELYKQVLFTEAPMGNVAQVPDNKGYILSQRTIAFNSN
 EKITDNFLASLLSSENVYNDLLKLCGATAKGVSQKNLNRLYVTI PHSISEQEEIAEFFRKINQLV
 ELQKYKIEHTKSQKQVFLQKMFIGSMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYG
 KLTLKFICTTGKLPVPWPTLVTTTLTYGVQCFSRYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNY
 KTRAEVKFEGDTLVNRIELKGIIDFKEDGNILGHKLEYNYNSHNVYIMADKQKNGIKVNFKIRHNIE
 DGSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSALS KDPNEKRDH MVLLFVTAAGITLGMDELYK
 HHHHHH



1- marker 2- Nickel column eluate 3-14 Fractions from gel filtration column
 15- CC5-1 Purified protein marker

S. SauBI-EGFP
CC22-1 **AGG-6-TGAR**

DNA cleavage assay



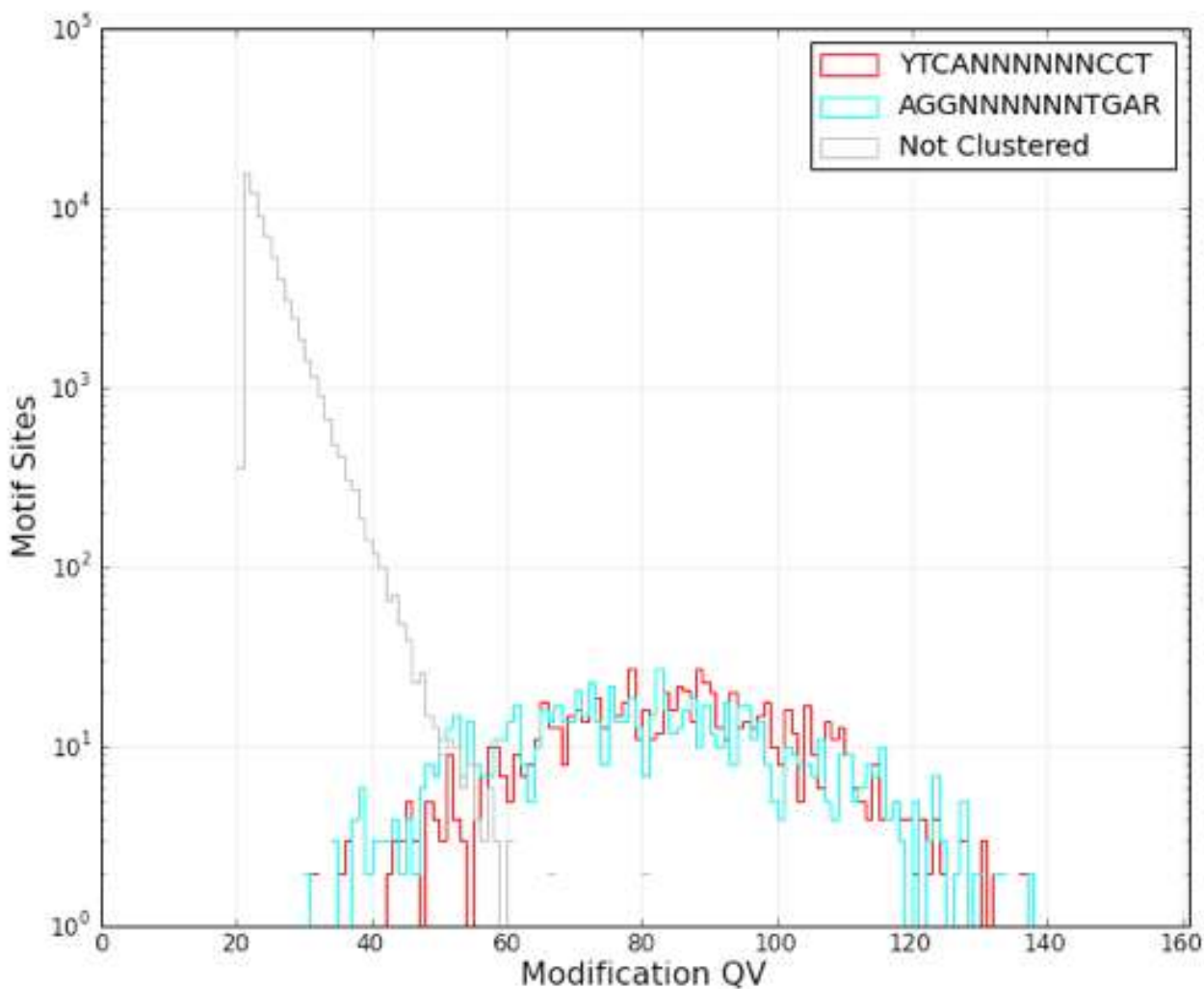
S. SauBI-EGFP

CC22-1 AGG-6-TGAR

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
YTCANNNNNNCCT	4	m6A	99.24	919	926	86.2	56.3	AGGNNNNNNTGAR
AGGNNNNNNTGAR	1	m6A	99.24	919	926	83.9	55.7	YTCANNNNNNCCT
<i>Not Clustered</i>	0		0.06	5230	9124356	34.8	61.5	

Modification QV Histogram By Motif

Modification QV Histogram



S. SauCE**ST425-1 GWAG-5-RTGA**

The recombinant enzyme with TRDs C and E was purified and used in the nuclease assay. There are minor differences in amino acid sequence between members of ST425-1.

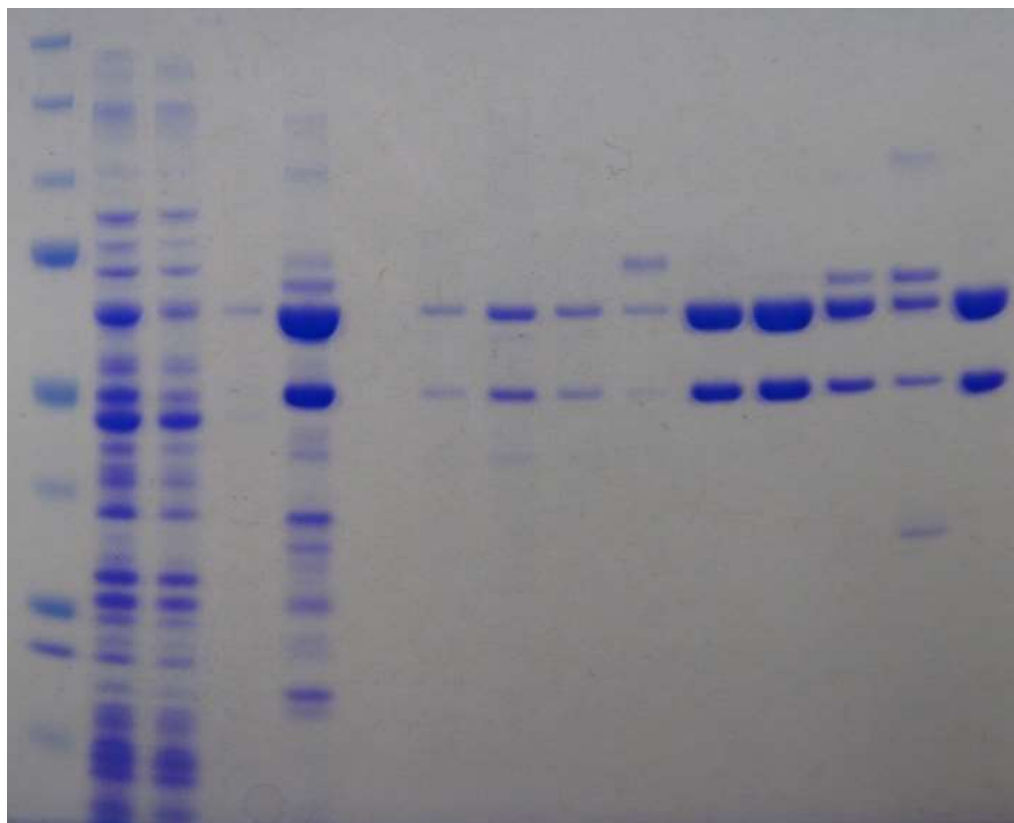
Recombinant S. SauCE CC425-1 GWAG-5-RTGA

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 TGKVVNNSKELKNYSVEKGDVFFTRTSEVIGEIGYPSVILNDPENTVFSGFVLRGRPKSGIDLIN
 NFKRYVFFFTNSFRKEMITKSSMTTRALTSGTAINRMKVIYPVSAKEQKKIGDFFSKLDRQIELEEQ
 KLELLQQQKKGMYQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVR
 SPIVYKINTFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYYFSQNFL
 KETKKYSAKTSVDSVRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKTQKQVIELLKQRKKSLLQ
 KMFIPGGSHHHHHH

Wild type S. SauCE

MSNTQTKNVPELRFPGFEGEWEEKQVGELLEFKNGLNKGKEYFGSGSSIVNFKDVFNNRSINTNNL
 TGKVVNNSKELKNYSVEKGDVFFTRTSEVIGEIGYPSVILNDPENTVFSGFVLRGRPKSGIDLIN
 NFKRYVFFFTNSFRKEMITKSSMTTRALTSGTAINKMKVIYPVSAKEQKKIGDFFSKLDRQIELEEQ
 KLELLQQQKKGMYQKIFTQELRFKDENGNDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVR
 SPIVYKINTFSYEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYYFSQNFL
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 KMF I *

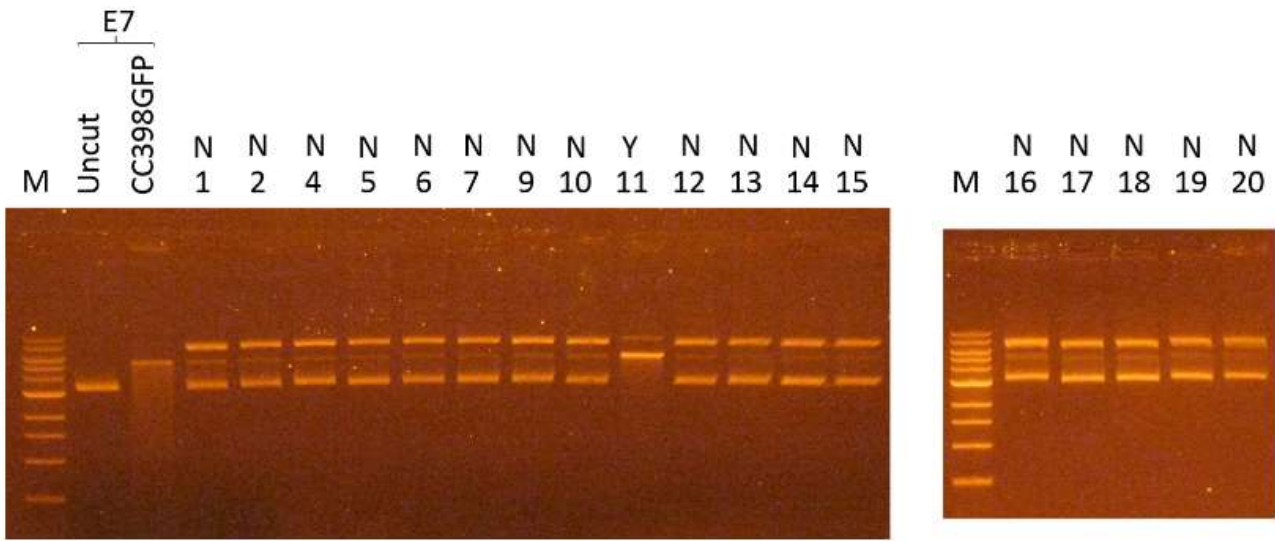
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15



1- marker 2- soluble cell extract 3- flow through from Nickel column 4- wash from Nickel column
 5- eluate from Nickel column 6-14 Fractions from gel filtration column 15- CC398-1 purified protein marker

S. SauCE
ST425-1 GWAG-5-RTGA

DNA cleavage assay.



S. SauJP**CC51** **GGA-6-CCT****This MTase was used in the SMRT analysis of *E. coli* ER2796.****There are minor variations in the sequences of the S subunits in CC51.****Recombinant S.SauJP** **CC51-1**

MSNTQKKNVPELRFPGFEGEWEEKKLGDLIKVNSGKDYKHLEKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPKKETDILFILSLFRKINWKVYDESTGVPSLSKQTI
 NKINRFVPSNKEQQKIGEFFIKLDRQIELEEQKLELFQQQKKGVMQKIFSQELRFKDESGNDYPDW
 EEKELGEVADR VIRKNKNFESKKPLTISGQLGLIDQTEYFSKSVSSKNLENYTLIKNGEFAYNKSY
 SNGYPLGAIKRLTRYDSGVLSSLYICFSIKSEMSKDFMEAYFDSTHWYREVSGIAVEGARNHGLLN
 ISVNDFFTILIKYPSLEEQRKIGDFFIKLDRQIELEEQKLELLQQRKKALLKSMLIPGGSHHHHHH

Wild Type S.SauJP

MSNTQTKNVPELRFPGFEGEWEEKKLEDIIKVNSGKDYKHLDKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPKKETDILFILSLFRKINWKVYDESTGVPSLSKQTI
 NKINRFVPTNKEQQKIGKFFSKLDRQIELEEQKLELFQQQKKGVMQKIFSQELRFKDESGNDYPDW
 EEKELGEVADR VIRKNKNFESKKPLTISGQLGLIDQTEYFSKSVSSKNLENYTLIKNGEFAYNKSY
 SNGYPLGAIKRLTRYDSGVLSSLYICFSIKSEMSKDFMEAYFDSTHWYREVSGIAVEGARNHGLLN
 ISVNDFFTILIKYPSLEEQRKIGDFFIKLDRQIELEEQKLELLQQRKKALLKSMLI

Reports for Job Dryden_J_P_MODs

SMRT Cells: 1 Movies: 1

Motif Summary

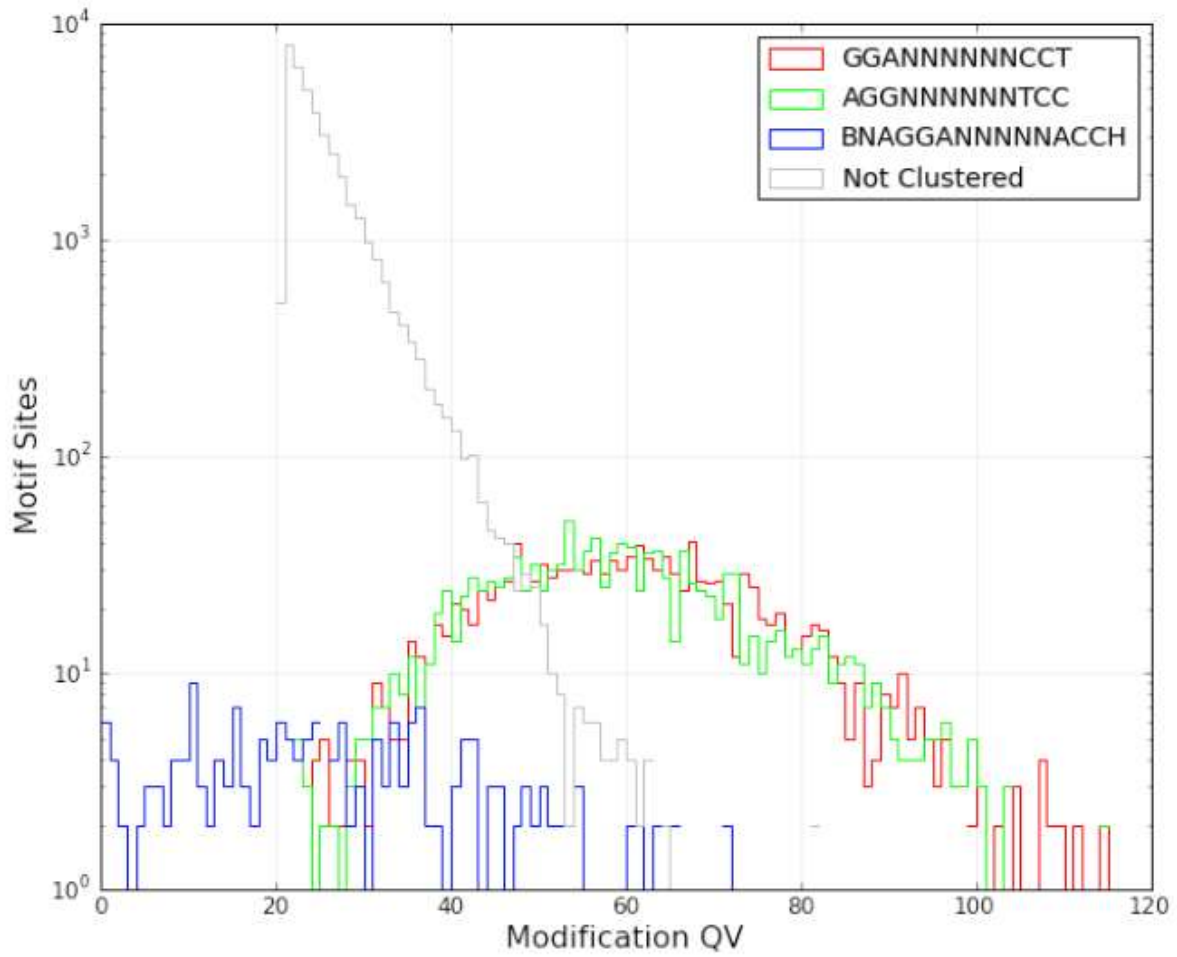
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
GGANNNNNCCT	3	m6A	98.1%	1340	1366	62.31	39.31	AGGNNNNNTCC
AGGNNNNNTCC	1	m6A	97.58%	1333	1366	61.04	39.18	GGANNNNNCCT
BNAGGNNNNACCH	3	m6A	46.26%	99	214	47.92	39.46	

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S. SauJP
CC51

GGA-6-CCT

Modification QVs



S. SauCL-EGFP**CC45-1 GWAG-6-TAAA**

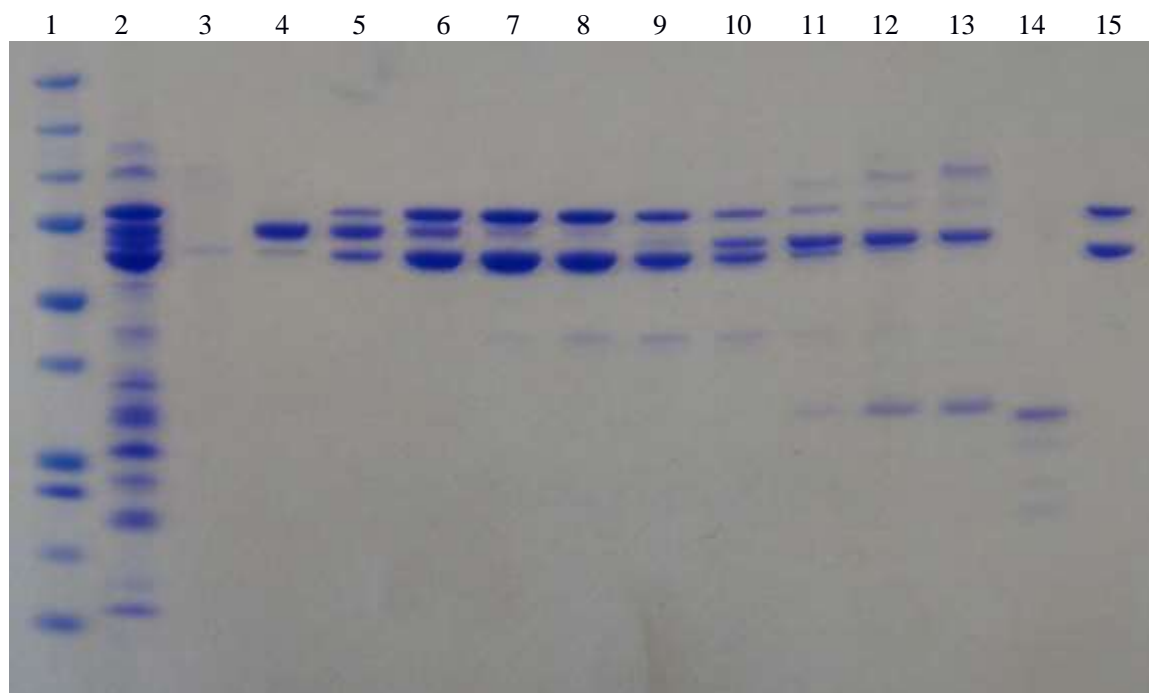
Two separate clones of pSauCL-EGFP encode residue 167 as Lysine (K) instead of arginine (R), but this does not affect the specificity as identical sequences are recognised in Trd C from CC30-1.

S. SauCL-EGFP "Expected" sequence

```
MSNTQKKNVPELRFPGFEGEWEEKKVGELLEFKNGLNKGKEYFGSGSSIVNFKDVFNNRSINTNNL
TGKVVNNSKELKNYSVEKGDVFFTRTSEVIGEIGYPSVILNDPENTVFSGFVLRGRPKSGIDLIN
NFKRYVFFFTNSFRKEMITKSSMTTRALTSGTAINRMKVIYPVSAKEQKKIGDFFSKLDRQIELEEQ
KLELLQQQKKGGMQKIFSQELRFKDENGNDYPNWRTEIELKNILENIVDNRGKTPDNAPSEKYPLLE
VNALGYRRPAYIKVSKFVSENTYNNWFREHLKENDILFSTVGNTGIVSLMDNYKAVIAQNIVGLRV
NNNNLPSFIYYMLS YKGNQKKIKRIQMGA VQPSVKVSQFKFIKYLVP IKDEQEKVAKLLIEIDKLV
NKQLIKIELLQQRK KALLKSMFIGSMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYG
KLTLKFICTTGKLPVPWPPTLVTTLT YGVQCFSRYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNY
KTRAEVKFE GDTLVNRIELKGIDFKEDGNILGHKLEYNYN SHNVYIMADKQKNGIKVNFKIRHNIE
DGSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSALS KDPNEKRDHMLLEFVTAAGITLGMDELYK
HHHHHH
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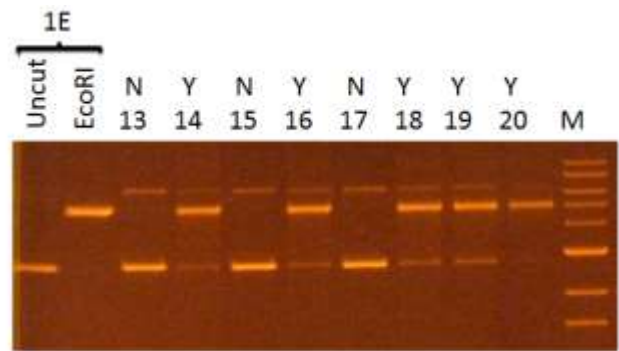
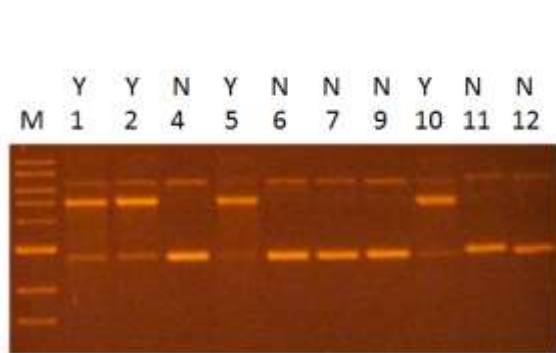
S. SauCL-EGFP "Actual" sequence

```
MSNTQKKNVPELRFPGFEGEWEEKKVGELLEFKNGLNKGKEYFGSGSSIVNFKDVFNNRSINTNNL
TGKVVNNSKELKNYSVEKGDVFFTRTSEVIGEIGYPSVILNDPENTVFSGFVLRGRPKSGIDLIN
NFKRYVFFFTNSFRKEMITKSSMTTRALTSGTAINKMKVIYPVSAKEQKKIGDFFSKLDRQIELEEQ
KLELLQQQKKGGMQKIFSQELRFKDENGNDYPNWRTEIELKNILENIVDNRGKTPDNAPSEKYPLLE
VNALGYRRPAYIKVSKFVSENTYNNWFREHLKENDILFSTVGNTGIVSLMDNYKAVIAQNIVGLRV
NNNNLPSFIYYMLS YKGNQKKIKRIQMGA VQPSVKVSQFKFIKYLVP IKDEQEKVAKLLIEIDKLV
NKQLIKIELLQQRK KALLKSMFIGSMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYG
KLTLKFICTTGKLPVPWPPTLVTTLT YGVQCFSRYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNY
KTRAEVKFE GDTLVNRIELKGIDFKEDGNILGHKLEYNYN SHNVYIMADKQKNGIKVNFKIRHNIE
DGSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSALS KDPNEKRDHMLLEFVTAAGITLGMDELYK
HHHHHH
```



1- marker 2- Nickel column eluate 3-14 Fractions from gel filtration column
15- CC5-1 purified protein marker

1
2 **S. SauCL-EGFP**
3 **CC45-1 GWAG-6-TAAA**
4 DNA cleavage assay.
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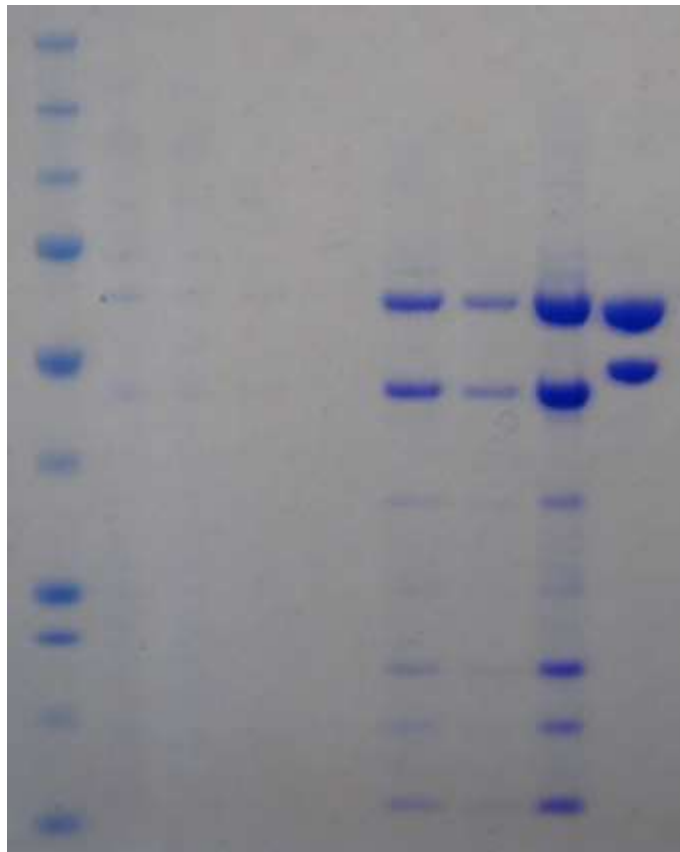
S. SauOE**CC15****Recombinant S. SauOE****CC15-1****CAAC-5-RTGA**

MSNTQKKNVPELRFPGFEGEWEEKKLGEVGTFTSSGGTPLKSKSEYWNGDIPWITTTGDIHNIKRENI
 TNFITEKGLNESSAKLITNEAILIAMYGQGKTRGMSAILNFEATTNQACAIYQTNQNINVFVFQYFQ
 KLYEFLRSLSNESQKNLSLSLLKEITLNYPNQEQQKIGDFFSKLDRQIELEEOKLELLQQQKKG
 YMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDTKDAITNGSYDFYVRSPIVYKINTFS
 YEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYSAKTS
 VDSVRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKT~~TKIQKQV~~IELLKQRKKSLQKMFIPGGSHHH
 HHH

Wild Type S. SauOE

MSNKQKKNVPELRFPGFEGEWEEKKLGEVGTFTSSGGTPLKSKSEYWNGDIPWITTTGDIHNIKRENI
 TNFITEKGLNESSAKLITNEAILIAMYGQGKTRGMSAILNFEATTNQACAIYQTNQNINVFVFQYFQ
 KLYEFLRSLSNESQKNLSLSLLKEITLNYPNQEQQKIGDFFSKLDRQIELEEOKLELLQQQKKG
 YMQKIFSQELRFKDENGNDYPEWEETTIKEIAQINXGKKDTKDAITNGSYDFYVRSPIVYKINTFS
 YEGEAILTVGDGVGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETKKYSAKTS
 VDSVRKDMIANMKVPRPIYIEQKKIGQFIKRVDNKT~~TKIQKQV~~IELLKQRKKALLQKMF I

1 2 3 4 5 6 7 8 9



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after PD10 desalting 8- Final concentrated protein
 9- CC398-1 purified protein marker

Although purified, this MTase was only used in SMRT sequencing.

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S. SauOE

CC15

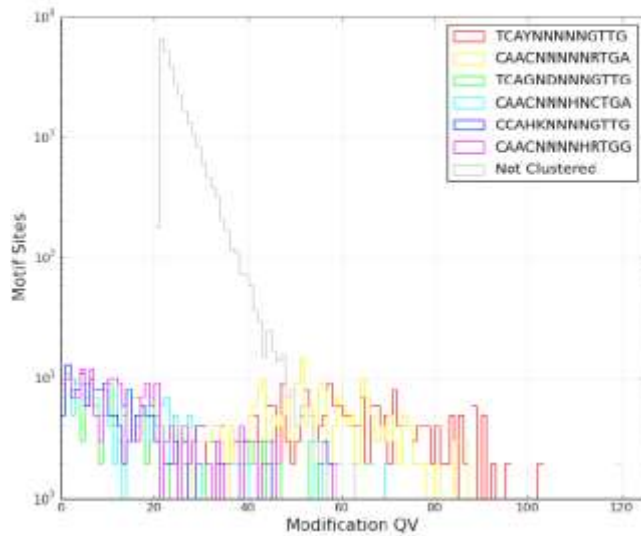
Recombinant S. SauOE

CC15-1

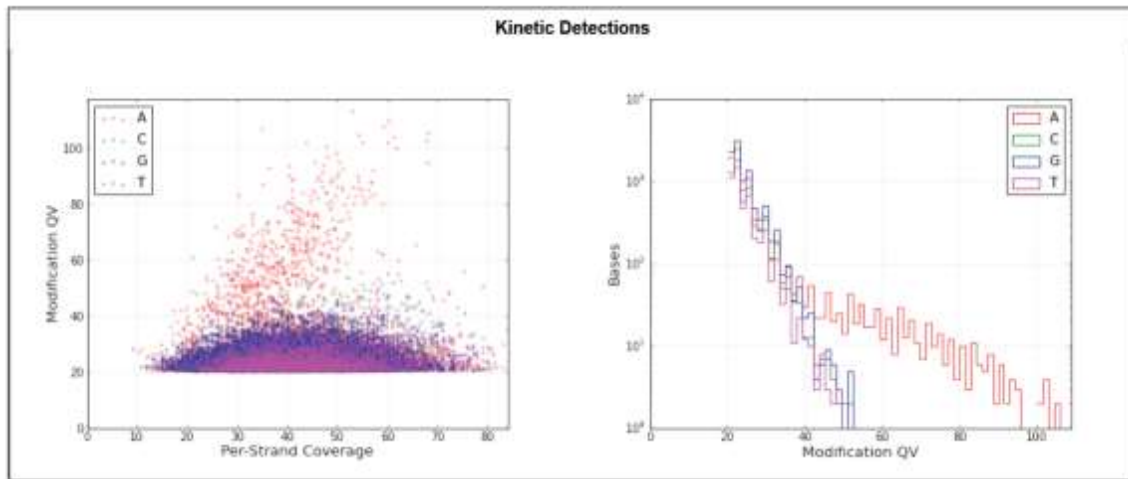
CAAC-5-RTGA

Motifs								
Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNGTTG	3	m6A	96.67	261	270	63.4	38.3	CAACNNNNRTGA
CAACNNNNRTGA	3	m6A	92.22	249	270	57.7	36.1	TCAYNNNNNGTTG
TCAGNDNNNGTTG	3	m6A	23.86	47	197	48.5	38.3	CAACNNNNHCTGA
CAACNNNNHCTGA	3	m6A	17.77	35	197	49.9	38.6	TCAGNDNNNGTTG
CCAHKNNNNGTTG	3	m6A	16.93	32	189	50.6	40.4	
CAACNNNNHRTGG	3	m6A	16.28	35	215	45.3	38.2	
Not Clustered	0		0.02	2201	9115988	34.9	47.5	

Modification QV Histogram By Motif



Kinetic Detections



Motifs

Motif	Modified Position	Modification Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNNGTTG	3	m6A	96.67	261	270	63.4	38.3	CAACNNNNRTGA
CAACNNNNRTGA	3	m6A	92.22	249	270	57.7	36.1	TCAYNNNNNGTTG
TCAGNDNNNGTTG	3	m6A	23.86	47	197	48.5	38.3	CAACNNNNHCTGA
CAACNNNNHCTGA	3	m6A	17.77	35	197	49.9	38.6	TCAGNDNNNGTTG
CCAHKNNNNGTTG	3	m6A	16.93	32	189	50.6	40.4	
CAACNNNNHRTGG	3	m6A	16.28	35	215	45.3	38.2	
Not Clustered	0		0.02	2201	9115988	34.9	47.5	

S. SauJQ**CC59**

This enzyme was purified and analysed using the ATPase assay as both TRD specificities were known and the DNA cleavage assay showed cutting of all plasmids.

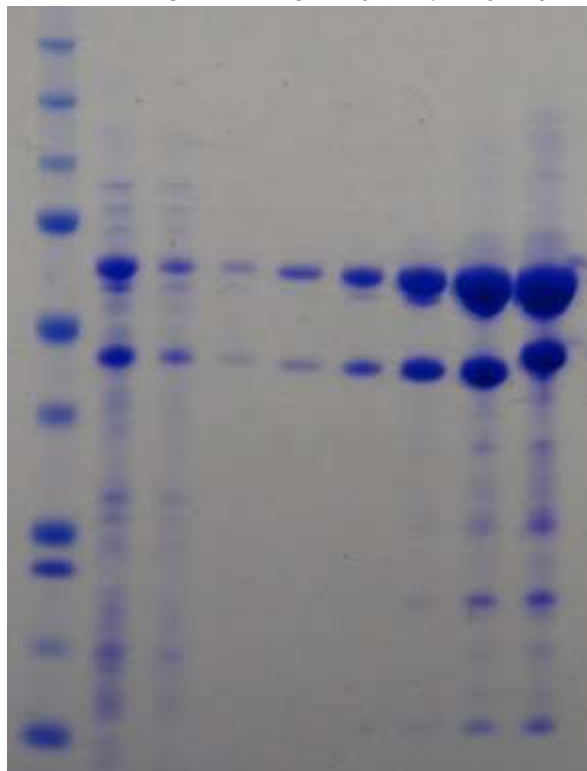
Recombinant S.SauJQ CC59-1 GGA-6-RTGT

MSNTQKKNVPELRFPGFEGEWEEKLDGLIKVNSGKDYKHLEKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPKKETDILFILSLFRKINWKVYDESTGVPSLSKQTI
 NKINRFVPSNKEQQKIGEFFIKLDRQIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGEDYSEW
 EERRFADIFKFHNKLRKPIKENLRVKGSYPYYGATGIIDYVDDFIFDGNLYLLIGEDGANIITRSAP
 LVYLVNGKFWVNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAQPKLNIQNLKIINVVISTNLE
 EQQKIGSFLSKLDRQIDLEEQKLELLQQRKKALLKSMFVPGGSHHHHHH

Wild type S.SauJQ

MSNTQKKNVPELRFPEFEGEWEEERKLDGLIKVNSGKDYKHLDKGDIPVYGTGGYMTSVSEPLSEID
 AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPEKEADILFILSLFRKINWKLYDESTGVPSLSKQTI
 NKINRLVPTNKEQQKIGEFFSKLDRQIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGEDYSEW
 EERRFADIFKFHNKLRKPIKENLRVKGSYPYYGATGIIDYVDDFIFDGNLYLLIGEDGANIITRSAP
 LVYLVNGKFWVNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAQPKLNIQNLKIINVVISTNLE
 EQQKIGSFLSKLDRQIDLEEQKLELLQQRKKALLKSMFV*

1 2 3 4 5 6 7 8 9



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after conc. and PD10 desalting
 8- final concentrated protein 9- XE purified protein marker

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2 **S. SauJQ**

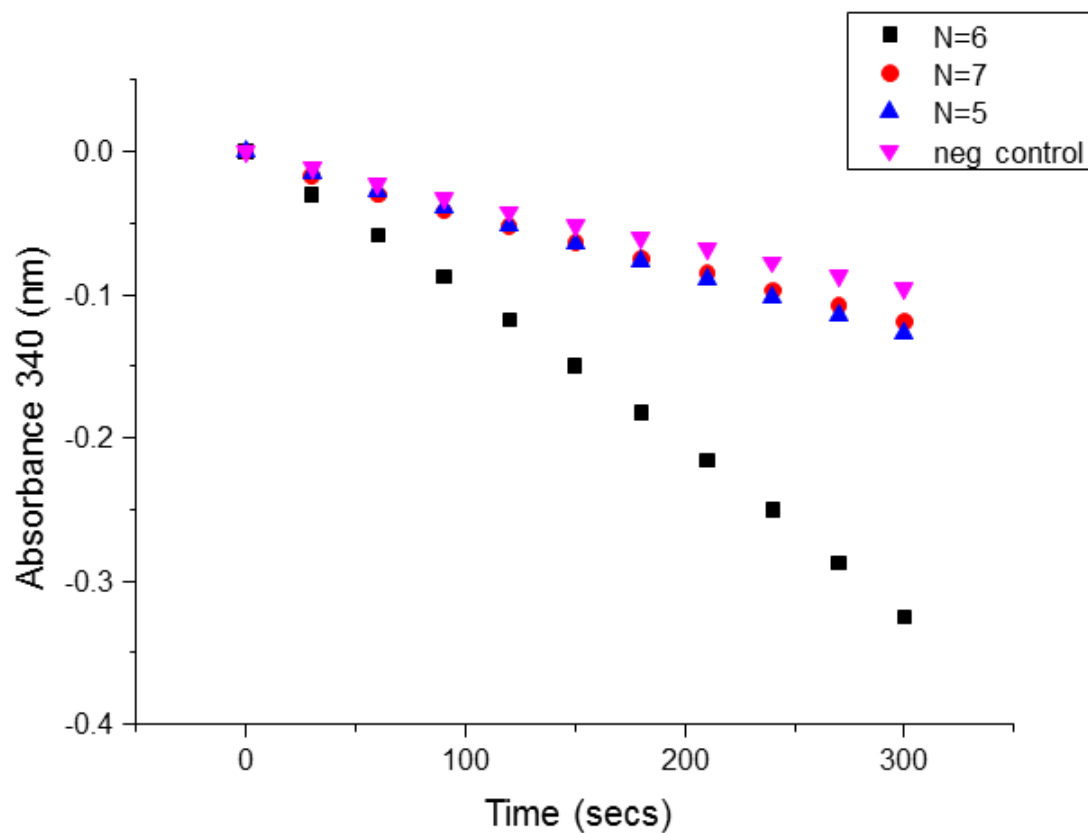
3 **CC59**

4 **Recombinant S.SauJQ**

5 **CC59-1**

6 **GGA-6-RTGT**

7 ATPase assay shows that N=6.



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Oligonucleotide name	DNA sequence (5' to 3')
JQ5for	AGATGATGTCATCAATGCGGATTACAGTGTGCCCTATACGATATAA
JQ5rev	TTATATCGTATAGGGCACACTGTAATCCGCATTGATGACATCATCT
JQ6for	AGATGATGTCATCAATGCGGATTGACAGTGTGCCCTATACGATATAA
JQ6rev	TTATATCGTATAGGGCACACTGTCAATCCGCATTGATGACATCATCT
JQ7for	AGATGATGTCATCAATGCGGATTAGACAGTGTGCCCTATACGATATAA
JQ7rev	TTATATCGTATAGGGCACACTGTCTAATCCGCATTGATGACATCATCT

S. SauRQ**CC72**

This enzyme was purified and analysed using the ATPase assay as both TRD specificities were known and the DNA cleavage assay showed cutting of all plasmids.

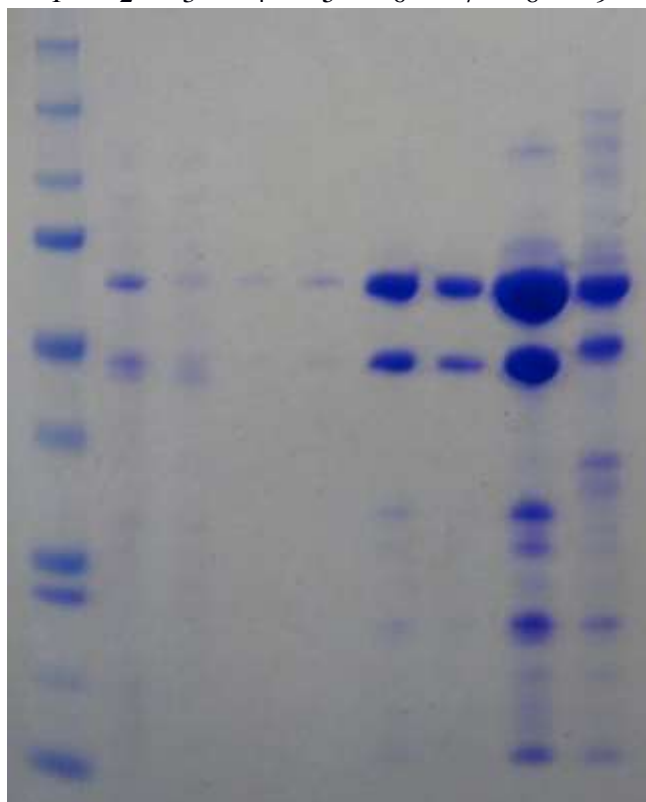
Recombinant S. SauRQ CC72-1 GARA-6-RTGT

MSNTQKKNVPELRFPGFEGEWEEKLGVAKIYDGTHQTPKYTNEGIFLSVENIKTLNSSKYISE
EAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYFLKNLILSSSIQN
ELWRKTLHVAFPKKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIELEEOKLELLQQQKKGYMQ
KIFSQELRFKDENGEDYSEWEERRFADIFKFHNKLRKPIKENLRVKGSYPYYGATGIIDYVDDFIF
DGNYLIGEDGANIITRSAPLVYLVNGKFWVNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAQ
PKLNIQNLKIINVVISTNLEEQQKIGSFLSKLDRQIDLEEOKLELLQQRKKALLKSMFVPGGSHHH
HHH

Wild type S. SauRQ

MSNTQKKNVPELRFPGFEGEWEEKLGVAKIYDGTHQTPKYTNEGIFLSVENIKTLNSSKYISE
EAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYFLKNLILSSSIQN
ELWRKTLHVAFPKKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIELEEOKLELLQQQKKGYMQ
KIFSQELRFKDENGNDYPEWEERRFADIFKFHNKLRKPIKENLRVKGSYPYYGATGIIDYVDDFIF
DGNYLIGEDGANIITRSAPLVYLVNGKFWVNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAQ
PKLNIQNLKIISVVISSTNLEEQQKIGSFLSKLDRQIDLEEOKLELLQQRKKALLKSMFV*

1 2 3 4 5 6 7 8 9



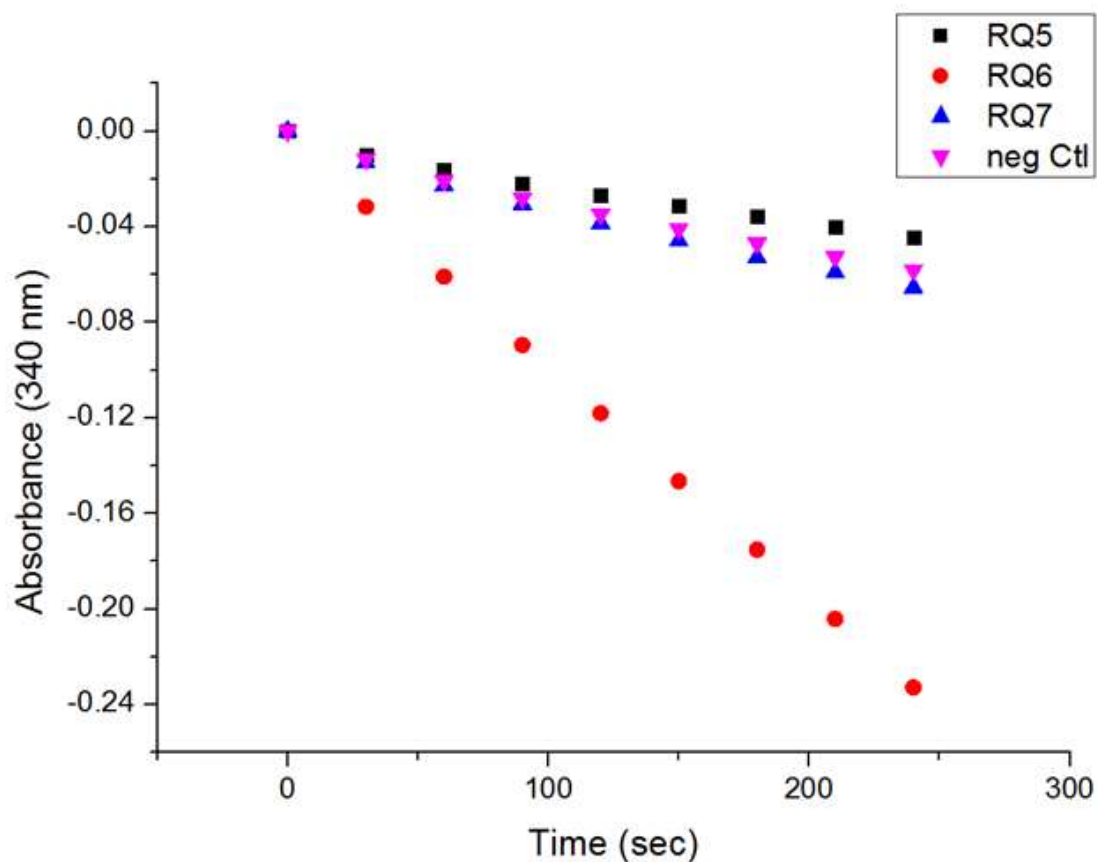
1- marker 2- soluble cell extract 3- Nickel column flow through
4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
7- eluate after PD10 desalting 8- Final protein after concentration
9- NP purified protein as marker

S. SauRQ

CC72

Recombinant S. SauRQ

GARA-6-RTGT



N=6 shows activity.

Oligonucleotide name	DNA sequence (5' to 3')
RQ5for	AGATGATGGAATCAATGCGAGATTCCAGTGTGCCCTATACGATATAA
RQ5rev	TTATATCGTATAGGGCACACTGGAATCTCGCATTGATTCCATCATCT
RQ6for	AGATGATGGAATCAATGCGAGATGTCCAGTGTGCCCTATACGATATAA
RQ6rev	TTATATCGTATAGGGCACACTGGACATCTCGCATTGATTCCATCATCT
RQ7for	AGATGATGGAATCAATGCGAGATGTACCAGTGTGCCCTATACGATATAA
RQ7rev	TTATATCGTATAGGGCACACTGGTACATCTCGCATTGATTCCATCATCT

S.SauJS

This second enzyme from CC72 was purified and analysed using the ATPase assay. There are minor variations between S subunit sequences in CC72-2.

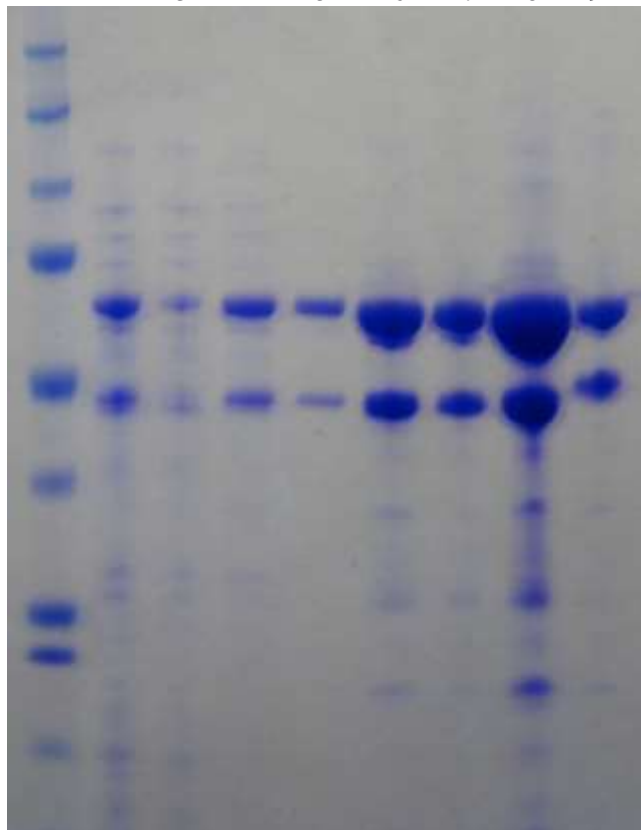
CC72**Recombinant S.SauJS****CC72-2****GGA-7-TGC**

```
MSNTQKKNVPELRFPGFEGEWEEKKLGDLIKVNSGKDYKHLEKGDIPVYGTGGYMTSVSEPLSEID
AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPKKETDILFILSLFRKINWKVYDESTGVP SLSKQTI
NKINRFVPSNKEQQKIGEFFIKLDRQIELEEQKLELLQQQKKG YMQKIFSQELRFKDENGNDYPDW
TNERLGEVTTVTMGQSPKSVNYTDNSNDTVLIQGNADIENGLINPRIYTREVTKLIQKDEIILT VR
APVGKLAMAQINACIGRGVCSIKGDKFLYYFLEWFATQNKWIRFSQGSTFESISGNDIRNIHIKIP
VEDERTKIIKLLNSLDV LNSKTDLKIQNLKQRKQSL LQKIFVPGGSHHHHHH
```

Wild Type S.SauJS

```
MSNTQKKNVPELRFPEFEGEWEEKQLGNI IKVNSGKDYKHLDKGDIPVYGTGGYMTSVSEPLSEID
AVGIGRKGKTINKPYLLEAPFWTVDTLIFYCTPKKETDILFILSLFRKINWKVYDESTGVP SLSKQTI
NKINRFVPTNKEQQKIGKFFSKLDRQIELEEQKLELLQQQKKG YMQKIFSQELRFKDENGNDYPDW
TNERLGEVTTVTMGQSPKSVNYTDNSNDTVLIQGNADIENGLINPRIYTREVTKLIQKDEIILT VR
APVGKLAMAQINACIGRGVCSIKGDKFLYYFLEWFATQNKWIRFSQGSTFESISGNDIRNIHIKIP
VEDERTKIIKLLNSLDV LNSKTDLKIQNLKQRKQSL LQKIFV
```

1 2 3 4 5 6 7 8 9



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after PD10 desalting 8- final protein after concentration
 9- NP purified protein as marker

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2 **S. SauJS**

3 **CC72**

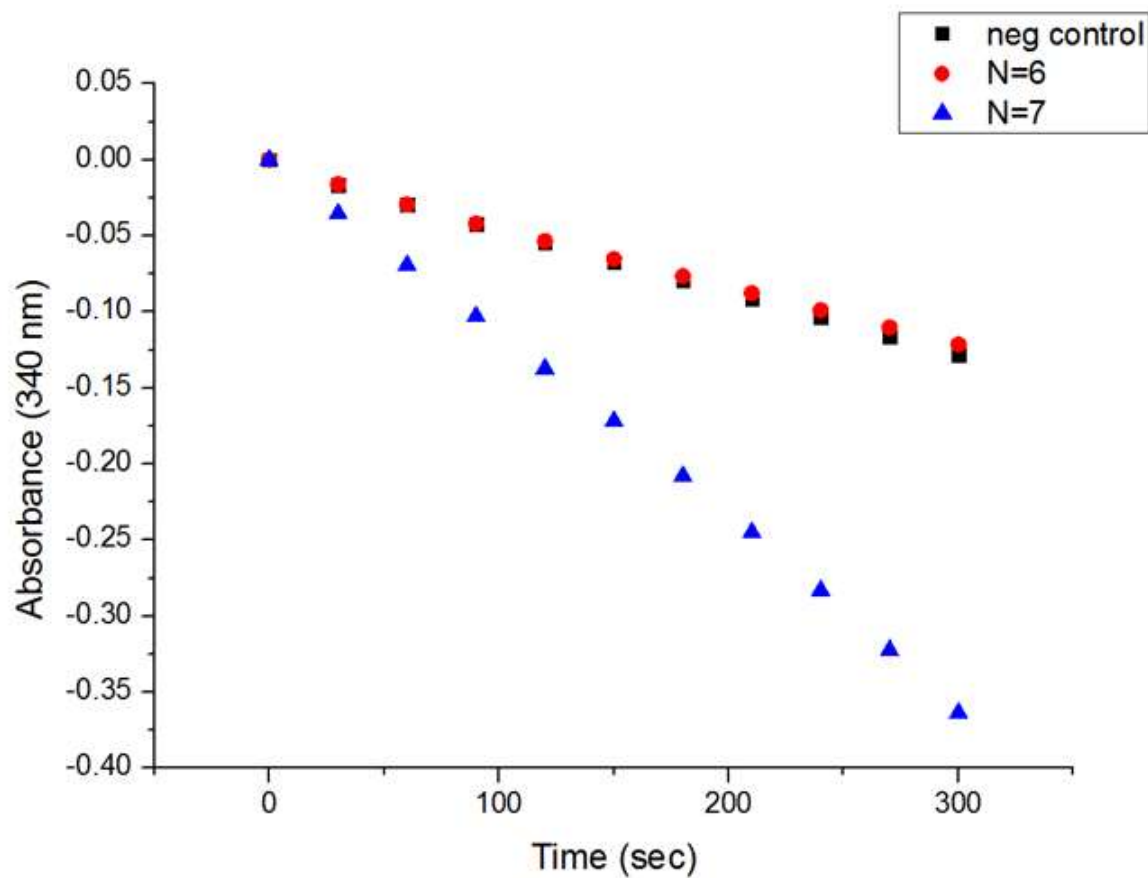
4 **Recombinant S.SauJS**

5 **CC72-2**

6 **GGA-7-TGC**

7 N=7 shows activity.

Oligonucleotide name	DNA sequence (5' to 3')
JS6for	AGATGATGGCATCAATGCGGATTACATTGCGCCCTATACGATATAA
JS6rev	TTATATCGTATAGGGCGCAATGTAATCCGCATTGATGCCATCATCT
JS7for	AGATGATGGCATCAATGCGGATTGACATTGCGCCCTATACGATATAA
JS7rev	TTATATCGTATAGGGCGCAATGTCAATCCGCATTGATGCCATCATCT



S. SauTU

CC75

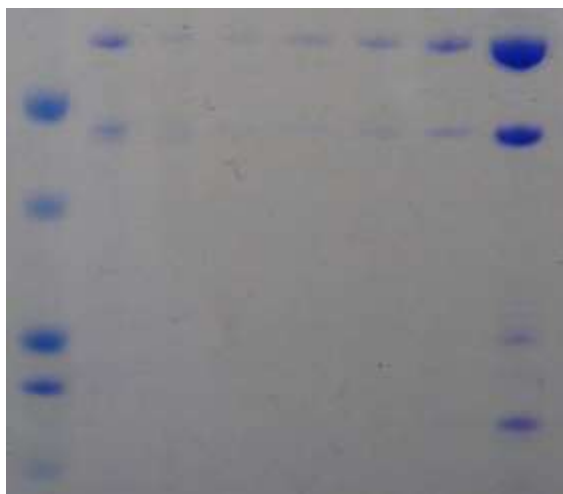
Recombinant S. SauTU CC75-1 CAAG-5-RTC

MSNTQKKNVPELRFPGFEGEWEEKELGEIFQIISGSTPLKSNKEFYENGNINWVKTTDLNNSKVTH
 SKEKITEYAMKSLKLLKLVPKNSVLIAMYGGFNQIGRTGLLKIDATINQAISALLMNHETNPEFIQA
 FLNYQVKGWKRYAASSRKDPNITKKDIEQFKVPYVSINEQQKIGEFFSKIDHQIELEEOKLELLQO
 QKKGVMQKIFSQELRFKDENGEDYPDWEVTTIQNITKYTSSKKSSNQYADKDNSKGYPVYDAVQEI
 GKDSNYDIEESYISILKDGAGVGRNLNLRPGKSSVIGTMGYIQSNNVDIEFLYRMMKVVDFFKYYIIG
 STIPHLYFKDYSKETLYIPSSIQEQAKIGMFISNLDKLIENKNLKLNLNCLKQLKQGLLQSMFIPGGS
 HHHHHH

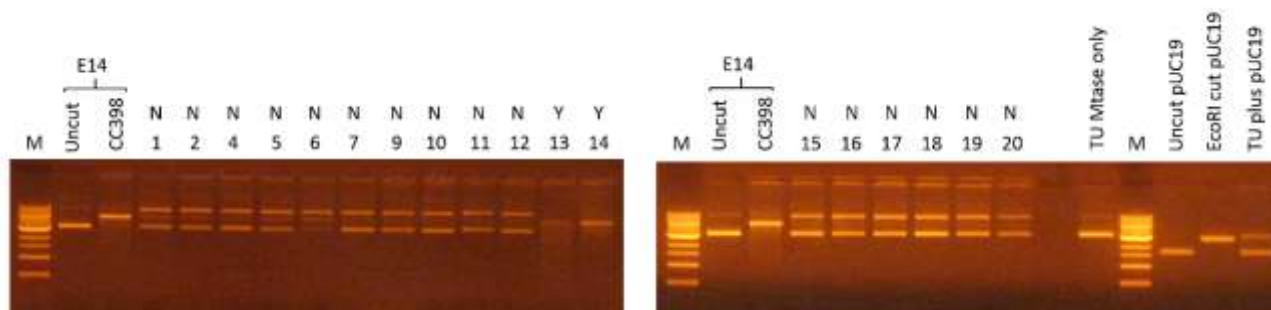
Wild type S. SauTU

MSNTQTKNVPELRFPGFEGEWEEKELGEIFQIISGSTPLKSNKEFYENGNINWVKTTDLNNSKVTH
 SKEKITEYAMKSLKLLKLVPKNSVLIAMYGGFNQIGRTGLLKIDATINQAISALLMNHETNPEFIQA
 FLNYQVKGWKRYAASSRKDPNITKKDIEQFKVPYVSINEQQKIGEFFSKIDHQIELEEOKLELLQO
 QKKGVMQKIFSQELRFKDENGEDYPDWEVTTIQNITKYTSSKKSSNQYADKDNSKGYPVYDAVQEI
 GKDSNYDIEESYISILKDGAGVGRNLNLRPGKSSVIGTMGYIQSNNVDIEFLYRMMKVVDFFKYYIIG
 STIPHLYFKDYSKETLYIPSSIQEQAKIGMFISNLDKLIENKNLKLNLNCLKQLKQGLLQSMFI

1 2 3 4 5 6 7 8



1- marker 2- soluble cell extract 3- Nickel column flow through 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate 7- eluate after conc. and PD10 desalting 8- final protein after concentration
 DNA cleavage assay.



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60**S. SauVW****CC75****Recombinant S. SauVW CC75-2 CNGA-7-TTYG**

MSNTQKKNVPELRFPGFEGEWEEKELRELRNPKDKYSYTGPFSGDLKKS DYTTDGIQIIQLQNI
 DGYFYNSNKVFTSNEKAEVLKSCNVFPGDIVIAKMADPIARAAIVPDNNIGKYL MASDGIRLSVDT
 VHFNTK FVLECI NRKSF RKKVEDNSSGSTRMRIGLSTLGSLTLKTTTLKEQQKIGQFFSKLDRQIE
 LEEQKLELLQQQKGYMQKIFSQELRFKDENGNDYPDWEEKQLGELSQIVRGASPRPIKDPKWFNK
 ESDIGWLRISDVTNQNNGKIYHLEQKLSIEGQEKTRVLVTTHLLLSIAASIGKPV MNFVKTGVHDGF
 LI FLKPKFNLFMYWLEYFKDKWSKYGQPGSQVNLNSEIVKSQTLNMP SNHEQEKV GQFFNRNEK
 LIELQQEKIMYIKRCKQVLLQKMFIPGGSHHHHH

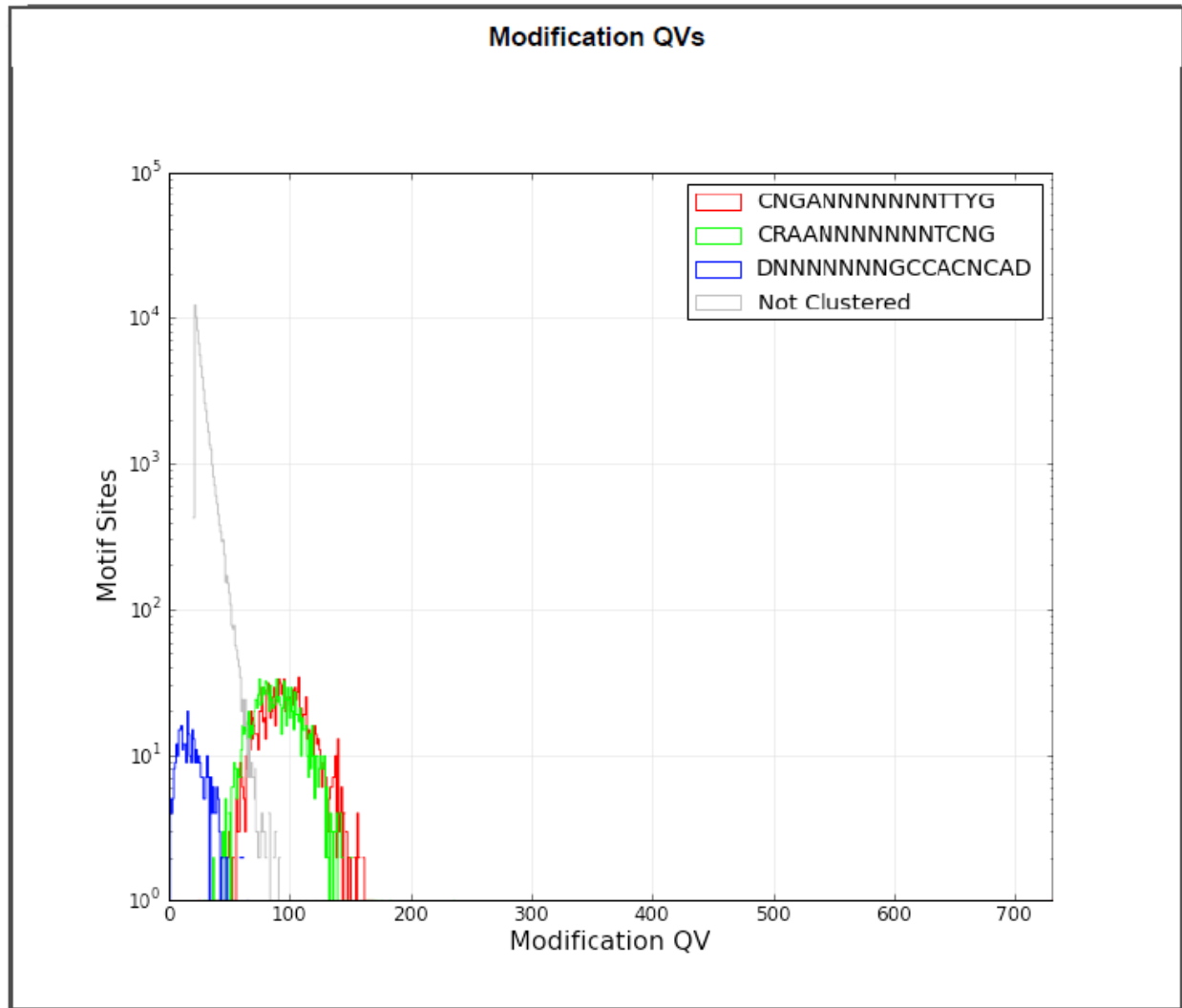
Wild Type S. SauVW

MSNTGKMNPELRFPGFEGEWEEKELRELRNPKDKYSYTGPFSGDLKKS DYTTDGIQIIQLQNI
 DGYFYNSNKVFTSNEKAEVLKSCNVFPGDIVIAKMADPIARAAIVPDNNIGKYL MASDGIRLSVDT
 VHFNTK FVLECI NRKSF RKKVEDNSSGSTRMRIGLSTLGSLTLKTTTLKEQQKIGQFFSKLDRQIV
 LEEQKLELLQQQKGYMQKIFSQELRFKDENGNDYPDWEEKQLGELSQIVRGASPRPIKDPKWFNK
 ESDIGWLRISDVTNQNNGKIYHLEQKLSIEGQEKTRVLVTTHLLLSIAASIGKPV MNFVKTGVHDGF
 LI FLKPKFNLFMYWLEYFKDKWSKYGQPGSQVNLNSEIVKSQTLNMP SNHEQEKV GQFFNRNEK
 LIELQQEKIMYIKRCKQVLLQKMF I *

Reports for Job Dryden_V_W_MODs

SMRT Cells: 1 Movies: 1

Motif Summary								
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CNGANNNNNTTYG	4	m6A	99.93%	1442	1443	97.87	66.11	CRAANNNNNTCNG
CRAANNNNNTCNG	4	m6A	99.86%	1441	1443	89.76	63.95	CNGANNNNNTTYG
DNNNNNGCCACNCA	9	unknown	19.1%	72	377	38.56	67.49	

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2 **S. SauVW**3 **CC75**4 **Recombinant S. SauVW**5 **CC75-2**6 **CNGA-7-TTYG**
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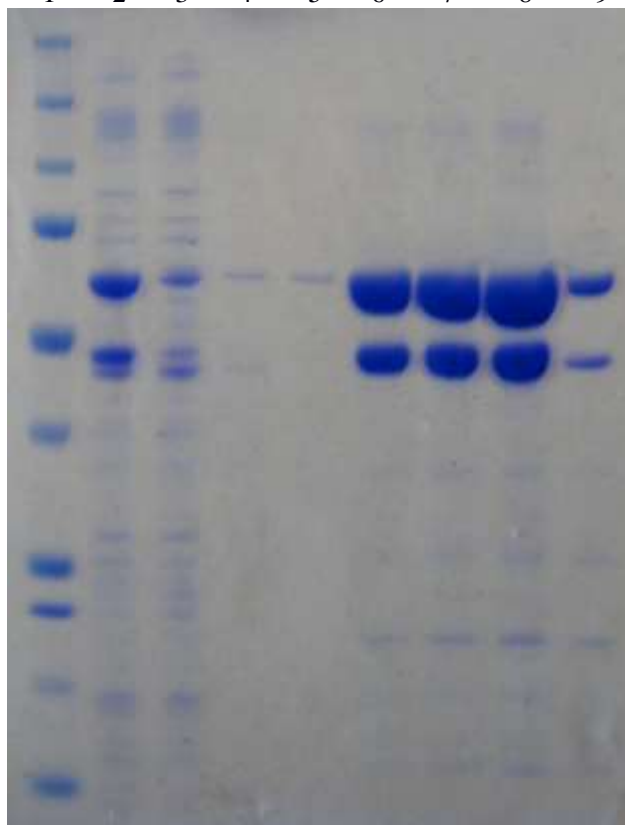
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60**S. SauZW****CC80****Recombinant S. SauZW CC80-2 GAC-6-TTYG**

MSNTQKKNVPELRFPGFEGEYSLDIFGNLATNKSEKFNPQENASIDIELDCIEQNTGRLIKIYNS
 KEFSSQKNKFNPNQNVLYGKLRPYLNKYYFTKKSGVCSSEIWLKSTKEDKLLNLFYFIQTKRYS
 DVASKSAGSKMPRADWGLIENIRVYFPELCEQQKIGQFFSKLDRQIELEEQKLELLQQQKGYMOK
 IFSQELRFKDENGNDYPDWEEKQLGELSQIVRGASPRPIKDPKWFNKE SDIGWLRISDVTNONGKI
 YHLEQKLSIEGQEKTRVLVTTHLLLSIAASIGKPV MNFVK TGVDHGFLIFLKPKNLFFMYWLEY
 FKDKWSKYGQPGSQVNLNSEIVKSQTLNMP SNHEQE KVGQFFNRNEKLI ELQQEKIMYIKRCKQVL
 LQKMFIPGGSHHHHHH

Wild Type S. SauZW

MSNTQTKNVPELRFPGFEGEYSLDIFGNLATNKSEKFNPQENASIDIELDCIEQNTGRLIKIYNS
 KEFSSQKNKFNPNQNVLYGKLRPYLNKYYFTKKSGVCSSEIWLKSTKEDKLLNLFYFIQTKRYS
 DVASKSAGSKMPRADWGLIENIRVYFPELCEQQKIGQFFSKLDRQIELEEQKLELLQQQKGYMOK
 IFSQELRFKDENGNDYPDWEEKQLGELSQIVRGASPRPIKDPKWFNKE SDIGWLRISDVTNONGKI
 YHLEQKLSIEGQEKTRVLVTTHLLLSIAASIGKPV MNFVK TGVDHGFLIFLNPKNLFFMYWLEY
 FKDKWSKYGQPGSQVNLNTEIVKSQTLNMP SNHEQE KVGQFFNRNEKLI ELQQEKIMYLKRRKQVL
 LQKMF I *

1 2 3 4 5 6 7 8 9



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after conc. and PD10 desalting
 8- final protein after concentration 9- CC75-1 purified protein marker

Although purified, this enzyme cut all plasmids in the DNA cleavage assay so the ATPase assay was used as we knew the specificities of the TRDs.

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2 **S. SauZW**

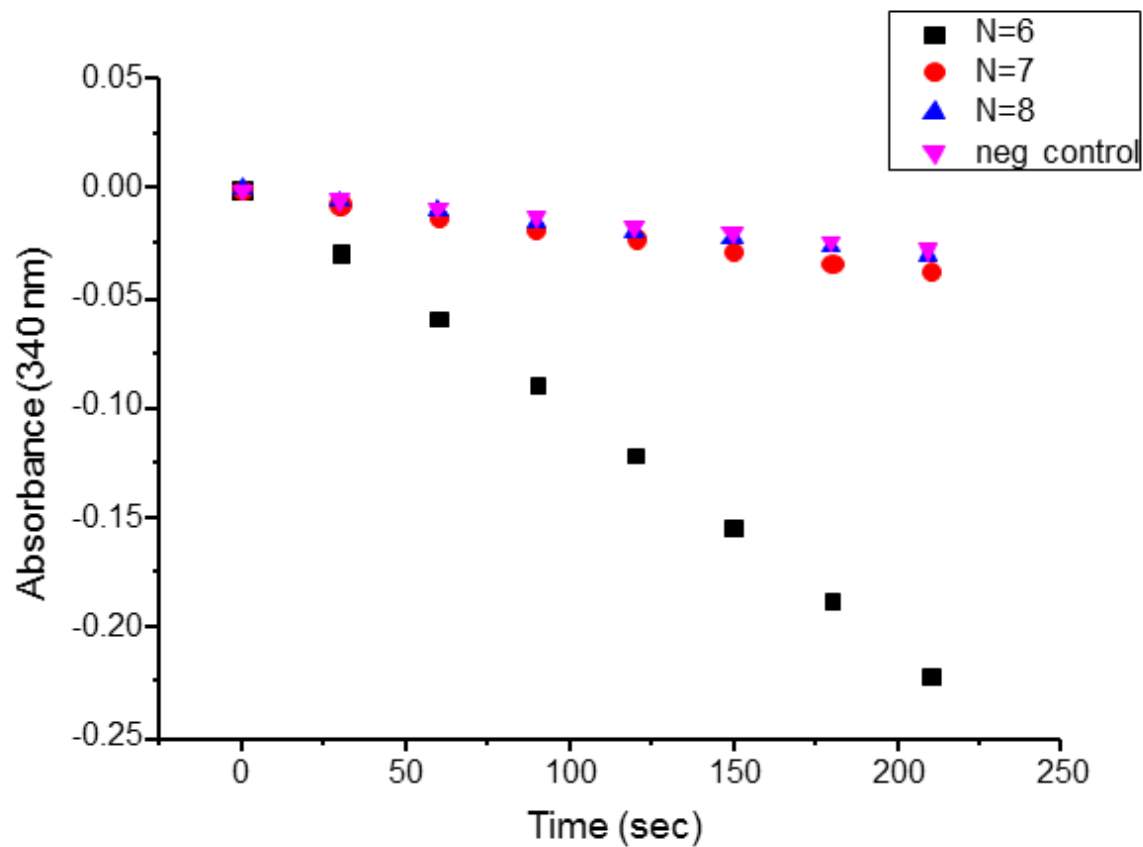
3 **CC80**

4 **Recombinant S. SauZW**

5 **CC80-2**

6 **GAC-6-TTYG**

7 N=6 shows activity.



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Oligonucleotide name	DNA sequence (5' to 3')
ZW6for	AGATGATGGAATCAATGCGACTTCCATTTTCGGCCCTATACGATATAA
ZW6rev	TTATATCGTATAGGGCCGAAATGGAAGTCGCATTGATTCCATCATCT
ZW7for	AGATGATGGAATCAATGCGACTTCTCATTTTCGGCCCTATACGATATAA
ZW7rev	TTATATCGTATAGGGCCGAAATGAGAAGTCGCATTGATTCCATCATCT
ZW8for	AGATGATGGAATCAATGCGACTTCTACATTTTCGGCCCTATACGATATAA
ZW8rev	TTATATCGTATAGGGCCGAAATGTAGAAGTCGCATTGATTCCATCATCT

S. SauXf*

ST80

Recombinant S. SauXf*

CC80-3

TCTA-6-RTTC

MSNTQKKNVPELRFPGFEGEWEEKQFADF^TTKINQGLQIAINERKTEYSPELYFYITNEFLRPNSQT
 KYFIENPPQSVIANKEDILMTRTGNTGKVVTNVFGAFHNNFFKIKFDKNLYDRFLFLVEVLNSSKIQ
 NKILSLAGSSTIPDLNHSDFYSISSSYPLLREQQKIGKFFSKLDRQIELEEQKLELLQQQKKG^YMQ
 KIFSQELRFKDENGEDYDPDWKEKKLGDITEQSMYGIGASATRFDSKNIYIRITDIDEKSRKLN^YQN
 LTPDELNNKYKLRNDILFARTGASTGKSYIHKEEKDIYNYFAGFLIKFKINEQNSPLFIYQFT
 LTSKFNKVVKMSVRSQPGINSEYAKLPLVLPNKLEQQKIAKFLDRFDRQIELEKQKIEILQQQ
 KKGLLQSMFI PGGSHHHHHH

Wild Type S. SauXf*

MSNTQKKNVPELRFPEFEGEWEEKQFADF^TTKINQGLQIAINERKTEYSPELYFYITNEFLRPNSQT
 KYFIENPPQSVIANKEDILMTRTGNTGKVVTNVFGAFHNNFFKIKFDKNLYDRFLFLVEVLNSSKIQ
 NKILSLAGSSTIPDLNHSDFYSISSSYPLLREQQKIGKFFSKLDRQIELEEQKLELLQQQKKG^YMQ
 KIFSQELRFKDENGEDYDPDWKEKKLGDITEQSMYGIGASATRFDSKNIYIRITDIDEKSRKLN^YQN
 LTPDELNNKYKLRNDILFARTGASTGKSYIHKEEKDIYNYFAGFLIKFKINEQNSPLFIYQFT
 LTSKFNKVVKMSVRSQPGINSEYAKLPLVLPNKLEQQKIAKFLDRFDRQIELEKQKIEILQQQ
 KKGLLQSMFI

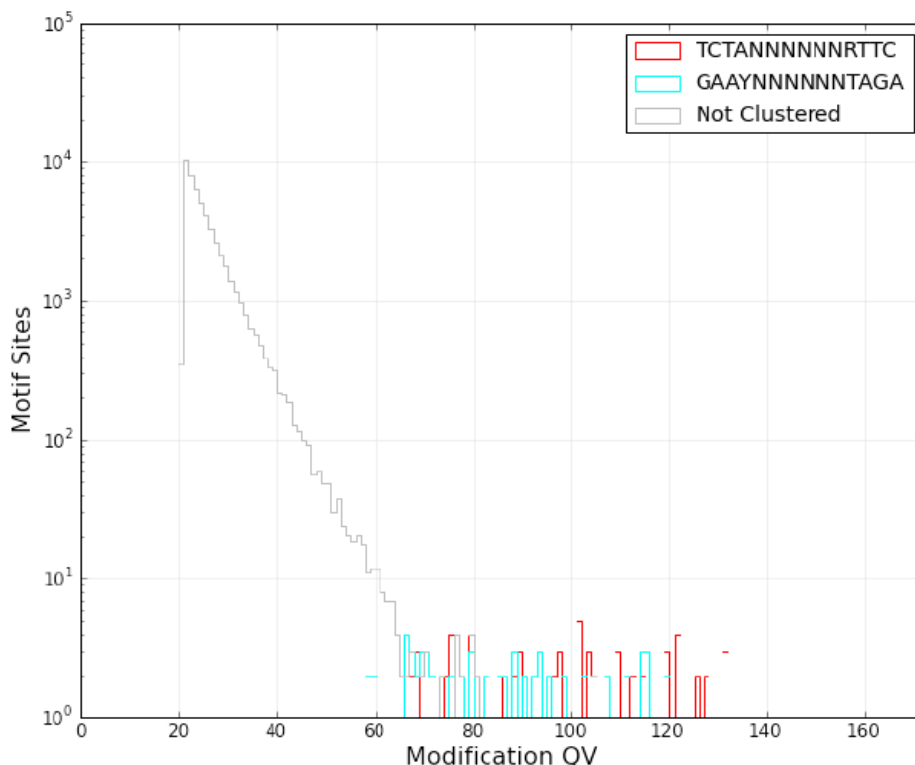
Reports for Job Dryden_X_zeta_MODs



SMRT Cells: 1 Movies: 1

Motif Summary								
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs in Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCTANNNNNR ^T TTC	4	m6A	100.0%	92	92	96.27	61.85	GAAYNNNNNTAGA
GAAYNNNNNTAGA	3	m6A	100.0%	92	92	90.82	60.21	TCTANNNNNR ^T TTC

Modification QVs



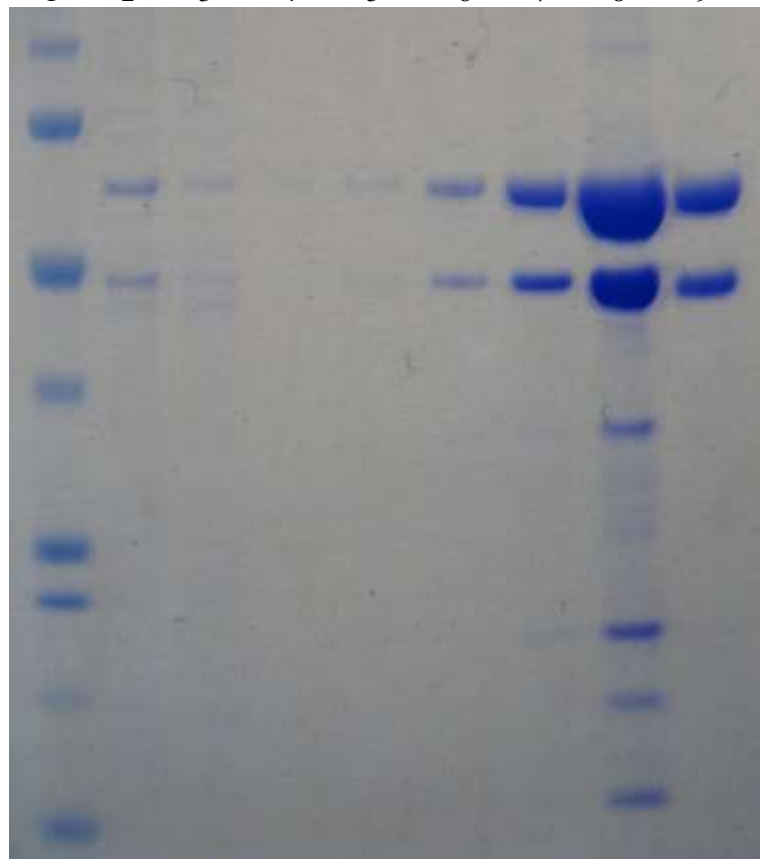
S. Saue*D**CC873****Recombinant S.Saue*D****CC873-1****GAG-6-GAT**

MSNTQKKNVPELRFPGFEGEWEEKSISSFLKESKIKGSHAKKLTVKLWGKGVVPKKE^TFKGSD
 NTQYYKRKAGQLMYGKLDLDFLNCAFGIVPDSLNNYESTIDSPSFD^FINGDSKFLLERIKLKSFYK^KF
 GDIANGSRKAKRINQDTFLSLPVFAPKYDEQLRIGEFFSKLDRQIELEE^QKLELLQQQKKG^YMQKI
 FSQELRFKDENSEDYPHWENSKIEKYLKERNERSDKGQMLS^VTINSGIIKFSELDRKDNSSK^DKSN
 YKVVRKNDIAYNSMRMWQGASGRSNYNGIVSPAYTVLYPTQNTSSLFIGYK^FKTHRMIHK^FKINSQ
 GLTSDTWNLKYKQLKNINIDIPVLEE^QEKIGDFFKMDILISKQIKIEILEKE^QSFLQKMFLPG
 GSHHHHHH

Wild Type S.Saue*D

MSNTQKKNVPELRFPGFEGEWEEKSISSFLKESKIKGSHAKKLTVKLWGKGVVPKKE^TFKGSD
 NTQYYKRKAGQLMYGKLDLDFLNCAFGIVPDSLNNYESTIDSPSFD^FINGDSKFLLERIKLKSFYK^KF
 GDIANGSRKAKRINQDTFLSLPVFAPKYDEQLRIGEFFSKLDRQIE^LQKQKLELLQQQKKG^YMQKI
 FSQELRFKDENGEDYPHWENSKIEKYLKERNERSDKGQMLS^VTINSGIIKFSELDRKDNSSK^NKSN
 YKVVRKNDIAYNSMRMWQGASGKSNYNGIVSPAYTVLYPTQNTSSLFIGYK^FKTHRMIHK^FKINSQ
 GLTSDTWNLKYKQLKNINIDIPVLEE^QEKIGDFFKMDILISKQIKIEILEKE^QSFLQKMFL*

1 2 3 4 5 6 7 8 9



1- marker 2- soluble cell extract 3- Nickel column flow through
 4- Nickel column wash 1 5- Nickel column wash 2 6- Nickel column eluate
 7- eluate after PD10 desalting and concentration
 8- Final concentrated protein 9- CC398-1 purified protein marker

1
2 **S. Saue*D**

3 **CC873-1**

4 **Recombinant S.Saue*D**

GAG-6-GAT

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6
7 e*D clearly digests pUC19 so the ATPase assay was used as we knew
8 the specificities of both TRDs.

9
10 Likely site: GAG-N_x-GAT

11
12 GAG-4-GAT 2 sites in pUC19

13
14 GAG-5-GAT 0 sites in pUC19

15
16 GAG-6-GAT 2 sites in pUC19

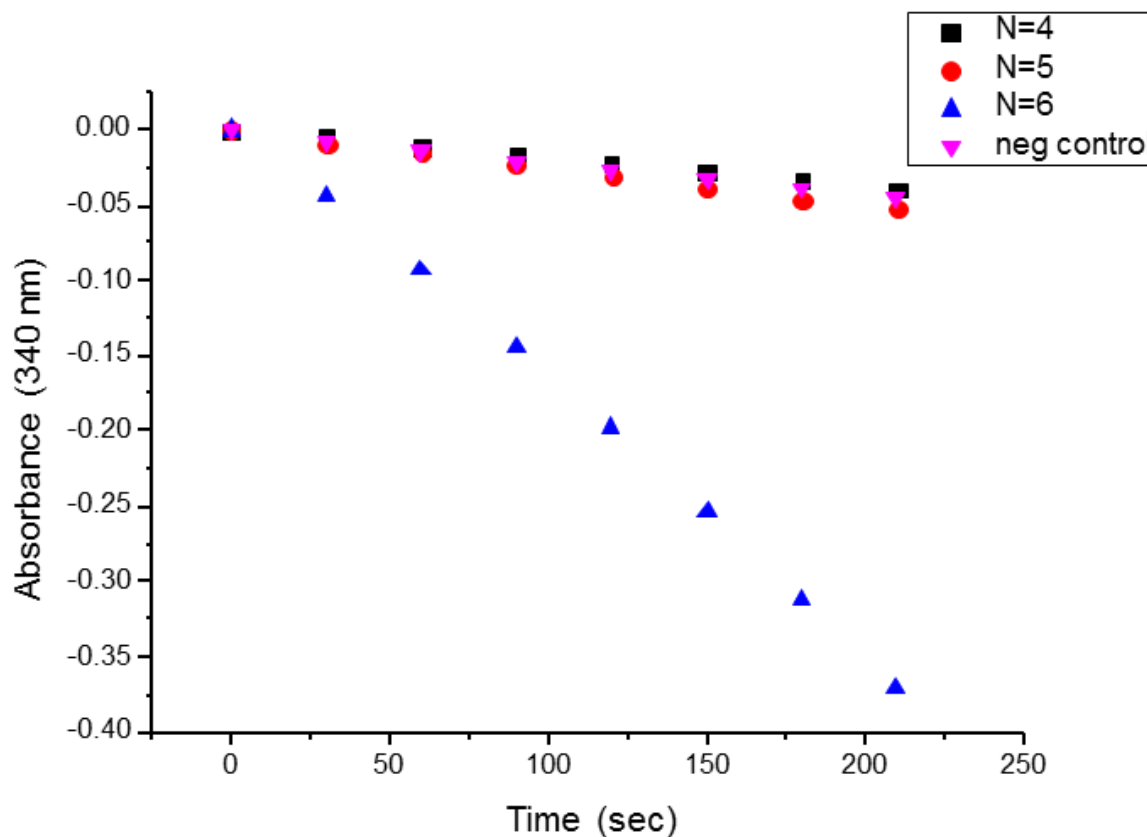
17
18 GAG-7-GAT 0 sites in pUC19

19
20
21

Oligonucleotide name	DNA sequence (5' to 3')
e*D6for	AGATGATGGAATCAATGCGAGTTCATGATGCCCTATACGATATAA
e*D6rev	TTATATCGTATAGGGCATCATGGAACCTCGCATTGATTCCATCATCT
e*D5for	AGATGATGGAATCAATGCGAGTTCAGATGCCCTATACGATATAA
e*D5rev	TTATATCGTATAGGGCATCTGGAACCTCGCATTGATTCCATCATCT
e*D4for	AGATGATGGAATCAATGCGAGTTCAGATGCCCTATACGATATAA
e*D4rev	TTATATCGTATAGGGCATCTGAACTCGCATTGATTCCATCATCT

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33 N=6 shows activity.



SMRT results for *S. aureus* strains LGA251 and NCTC13435

LGA251

SMRT® Portal Print

Reports for Job Dryden_LGA_Mods PACIFIC BIOSCIENCES™

SMRT Cells: 2 Movies: 2

Motif Summary								
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
TCAYNNNNCTWC	3	m6A	100.0%	391	391	352.92	251.20	GWAGNNNNRTGA
GWAGNNNNRTGA	3	m6A	100.0%	391	391	349.86	243.53	TCAYNNNNCTWC
GTANNNNCTTC	3	m6A	99.59%	245	246	349.71	251.31	GAAGNNNNNTAC
GAAGNNNNNTAC	3	m6A	99.59%	245	246	349.91	237.85	GTANNNNCTTC
BTTGGTAVY	2	unknown	26.29%	127	483	38.52	249.23	

NCTC13435

SMRT® Portal Print × Close

Reports for Job Dryden_NTCT_Mods PACIFIC BIOSCIENCES™

SMRT Cells: 2 Movies: 2

Motif Summary								
Motifs	Modified Position	Type	% Motifs Detected	# Of Motifs Detected	# Of Motifs In Genome	Mean Modification QV	Mean Motif Coverage	Partner Motif
CRAANNNNNGTC	4	m6A	100.0%	422	422	275.90	212.58	GACNNNNNTTYG
GACNNNNNTTYG	2	m6A	100.0%	422	422	315.18	220.50	CRAANNNNNGTC
GAAYNNNNNTAGA	3	m6A	100.0%	260	260	307.05	214.54	TCTANNNNNR TTC
TCTANNNNNR TTC	4	m6A	100.0%	260	260	318.07	218.98	GAAYNNNNNTAGA
GGATG	3	m6A	100.0%	2818	2818	327.54	220.52	CATCC
CATCC	2	m6A	100.0%	2818	2818	324.65	220.07	GGATG
DNNNNNNASNGGATG	9	m6A	26.07%	67	257	141.42	225.84	

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2 SUPPLEMENTARY INFORMATION FOR TABLES 5 AND 6.
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2 **SUPPLEMENTARY INFORMATION FOR TABLES 5 AND 6.**

3
4 **By combining all TRD 1 with all TRD 2 amino acid sequences and**
5 **searching sequence databases, we found that some of our**
6 **"artificial hybrids" described in Table 3 were actually present in**
7 **real strains of *S. aureus*. We present several examples below.**
8
9

10
11 **S. SauAU**

12 A plasmid expressing S.SauAU with the M subunit was prepared but
13 not analysed further. The S.SauAU sequence matches that of the S
14 subunit of the Type I RM system in *S. schweitzeri* FSA084.
15

16 >S.SauAU

17 MSNTQKKNVPELRFPGFEGEWEEKKLGDLTTKIGSGKTPKGGSENYTNKGIPFLRSQNIRNGKLN
18 NDLVYISKDIDDEMKNRRTYYGDVLLNITGASIGRTAINSIVEIHANLNQHVCIIIRLKKEYYYNFF
19 GQYLLSRKGRKIFLAQSGGSREGLNFKEIANLKIFTPTIFEEQQKIGEFISKLDRQIELEEQKLE
20 LLQQQKKGVMQKIFSQELRFKDENGEDYPDWEVTTIQNITKYTSSKKSSNQYADKDNSKGYPVYDA
21 VQEIGKDSNYDIEESYISILKDGAGVGRNLNLRPGKSSVIGTMGYIQSNNVDIEFLYYRMKVVDFFK
22 YIIIGSTIPHLYFKDYSKETLYIPSSIQEQAKIGMFISNLDKLIENKNLKNLCLKQLKQGLLQSMFI
23 PGGSHHHHHH
24
25
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27

28 **S. schweitzeri FSA084**

29 CLUSTAL O(1.2.1) multiple sequence alignment
30

31
32 FSA084 msn-tqkkvpelrfpgfegeweekklgevttkigsgktpkggseinytnkgipflrsqnir
33 S.SauAU MSNTQKKNVPELRFPGFEGEWEEKKLGDLTTKIGSGKTPKGGSENYTNKGIPFLRSQNIR
34 *** :*:*****:*****:*****:*****:*****:*****:*****:*****
35
36 FSA084 ngklnlnldlvyiskdiddemknsrtyygdvllnitgasigrtainsivethanlnqhvc
37 S.SauAU NGKLNLDLVYISKDIDDEMKNRRTYYGDVLLNITGASIGRTAINSIVEIHANLNQHVC
38 *****:*****:*****:*****:*****:*****:*****:*****:*****
39
40 FSA084 irlkkeyyydffeqyllsrkgkrkiflaqsggsreglnfkeianlkiftstifeeqqkv
41 S.SauAU IRLKKEYYYNFFGQYLLSRKGRKIFLAQSGGSREGLNFKEIANLKIFTPTIFEEQQKIG
42 *****:*****:*****:*****:*****:*****:*****:*****:*****:
43
44 FSA084 kffskldrqielleeqklellqqqkkgymqkifsqelrfkdengneypewkvtsiqdvtky
45 S.SauAU EFISKLDRQIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGEDYPDWEVTTIQNITKY
46 :*:*****:*****:*****:*****:*****:*****:*****:*****:
47
48 FSA084 tsskkssnyadkidskgyvydavreigkdsnydieesyisilkdgagvgrlnlrpeks
49 S.SauAU TSSKKSSNQYADKDNSKGYPVYDAVQEIGKDSNYDIEESYISILKDGAGVGRNLNLRPGKS
50 *****:*****:*****:*****:*****:*****:*****:*****:*****
51
52 FSA084 svigtmgylqannidleflyyrmkivdfkkyiigstiphlyfkdyketlyipssiqeqa
53 S.SauAU SVIGTMGYIQSNNVDIEFLYYRMKVVDFFKYYIIIGSTIPHLYFKDYSKETLYIPSSIQEQ
54 *****:*****:*****:*****:*****:*****:*****:*****:*****:
55
56 FSA084 kigkfisnldkmiengktrklnclqklqgllqgmfi-----
57 S.SauAU KIGMFISNLDKLIENKNLKNLCLKQLKQGLLQSMFIPGGSHHHHHH
58 *** *****:*****:*****:*****:*****:*****:*****:*****:
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S.SauJE GGA-6-RTGA
S.SauJE against subspecies 21262, a member of ST49

HsdS sequences from strain 21262.
>EH091218 This has TRD R + f*
MSNTQKKNVPELRFPGFEGEWEEKKLGEVAKIYDGTHQTPKYTNEGIFLSVENIKTLNS
SKYISEEAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYF
LKNLILSSSIQNELWRKTLHVAFPKKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIE
LEEQKLELLQQQKGYMQKIFSQELRFKDENGEDYPDWKEKKLGDITEQSMYGIGASATR
FDSKNIYIRITDIDEKSRKLNYNLTTPDELNNKYKLKRNLDILFARTGASTGKSYIHKEE
KDIYNYFAGFLIKFEIDEQNNPLFIYQFTLTSKFNKWKVMSVRSVSGQPGINSEYAKLP
LVLPNKLEQQKIAEFLDRFDQIELEKQKIEILQQQKGLLQSMFI

>EH092010 This has TRD J + E
MSNTQKKNVPELRFPGFEGEWEEKLEDIKVNNGKDYKHLDKGDI PVYGTGGYMTSVSE
PLSEIDAVGIGRGTINKPYLLEAPFWTVDTLfyCTPKKETDILFILSLFRKINWKVYDE
STGVPSLSKQTINKINRFVPTNKEQQKIGKFFSKLDRQIELEEQKIELLQQQKGYIQKI
FSQELRFKDENGDDYPEWEETTIQIEIAQINTGKKDKDAITNGSYDFYVRSPIVYKINTF
SYEGEAILTVGDGVGKVFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKETK
KYSAKTSVDSVRKDMVANMKVPRPIYIEQEKIGQFIKKVDNKIKIQQVIPELLQKQKAL
LQKMF I

CLUSTAL O(1.2.1) multiple sequence alignment

EH091218	msntqkknvpelrfpgfegeweekklgevakiydgthqtpkytnegikflsveniktlns
S.SauJE	MSNTQKKNVPELRFPGFEGEWEEKKLDLIKVNNGKDYKHL-----EKGDI PVYGT
EH092010	msntqkknvpelrfpgfegeweeklediikvnngkdykhl-----dkgdipvygt
	*****::*:.*. . . : * . :

EH091218	skyiseeafekefkirpefgdilmtrigdigtpnivssnekfayyvslallk--tknlns
S.SauJE	GGYMTSVS-----EPLSEIDAVGIGRGTINKPYLLEAP---FWTVDTLfyCTPKKETDI
EH092010	ggymtsvs-----eplseidavgigrkgtinkpylleap---fwtvdtlfyctpkketdi
	.*::: . : * : : * * * . * :::: : * . : . * : :

EH091218	yflknlilsssiqnelwrkthlvafpkkinkneigkikinyppkkqeqqkigqffskldrq
S.SauJE	LF-----ILSLFRKINWKVYDESTGVPSLSKQTINKINRFVPSNKEQQKIGEFFIKLDRQ
EH092010	lf-----ilslfrkinwkvydestgvpslskqtinkinrfvptnkeqqkigkffskldrq
	* : * ::: * : . : . . : * * * : * : : * * * : * * * :

EH091218	ieleeqklellqqkkgymqkifsqelrfkdengedypdwkekkkgditeqsmygigas-
S.SauJE	IELEEQKLELLQQQKGYMQKIFSQELRFKDENGKDYPEWEETTIKEIAQINTGKKDKD
EH092010	ieleeqkiellqqkkggyiqkifsqelrfkdengddypeweettiqeiaqintgkkdkd
	*****:*****:*****.***:*:*::: : : . : .

EH091218	atrfdskniyiritdideksrklnynlttpeLnnkyklkrndilfartgastgksyih
S.SauJE	AITNGSYDFYVRSPIV-YK-----INTFSYEGEAILTVGDGVGKVFHY
EH092010	aitngsydfyvrspiv-yk-----intfsyegailtvgdgvvgkvhfy
	* * ::* * : * * . . . : : * * . * . . * * : :

EH091218	keekdiynnyfagflikfeideqnnplfiyqftltskfnkvwkmsvrsvsgqpginseya
S.SauJE	VNGK--FDYHQRVYKIS-DFKNYYGLLLFYF--SQNFLKETKYSAKTSVDSVRKDMIA
EH092010	vngk--fdyhrvykis-dfknnygllyfyf--sqnflketkysaktsvdsvrkdmdva
	: * ::* : * . : : * : * * : * * * . * * : : . . . : *

EH091218	klplvlpnklegqkiaefldrfdqqielekqkieilqqqkglqsmfi-----
S.SauJE	NMKVPRPIYIEQKIGQFIKRVNDNKTIQKQVIPELLQKQKSSLLQKMFIPGGSHHHHHH
EH092010	nmkvprpiyieqekigqfikkvdnkikiqkqviellqkrkallqkmf-----
	:: : * :**:*:*:*. . : : : * : * * : * * : * * * . * * *

The above alignment shows that SauJE is identical to the EH092010 sequence from this strain.

1
2 **S. SauJE GGA-6-RTGA**
3 **Sub species 21262, a member of ST49**
4

5 CLUSTAL O(1.2.1) multiple sequence alignment
6 TRD R and TRD f* against EHO91218, the second HsdS in this strain.
7

8 CC80-3 -----
9 EHO91218 msntqkknvpelrfpgfegeweekklgevakiydgthqtpkynegikflsveniktlns
10 CC72-1 MSNTQKKNVPELRFPGFEGEWEEKKLGEVAKIYDGTHQTPKYTNEGIFLSVENIKTLNS
11

12 CC80-3 -----
13 EHO91218 skyiseeafekefkirpefgdilmtrigdigtpnivssnekfayyvslallktnlnsyf
14 CC72-1 SKYISEEAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYF
15

16 CC80-3 -----
17 EHO91218 lknlilsssiqnelwrkthlvafppkinkneigkikinyppkkqeqqkigqffskldrqi
18 CC72-1 LKNLILSSSIQNELWRKTHLVAFPPKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIE
19

20 CC80-3 -----QELRFKDENGEDYPDWKEKKLGDITEQSMYGIGASATR
21 EHO91218 leeqklellqqqkkgymqkifsqelrfkdengedypdwkekklgditeqsmygigasatr
22 CC72-1 LEEQKLELLQQQKKGYMQKIFS-----
23

24 CC80-3 FDSKNIYIRITDIDEKSRKLNQNLTPDELNNKYKLRNDILFARTGASTGKSYIHKEE
25 EHO91218 fdskniyiritdideksrklnyqnlttpdelnnkyklkrndilfartgastgksyihkee
26 CC72-1 -----
27

28 CC80-3 KDIYNYFAGFLIKFKINEQNSPLFIYQFTLTSKFNKWKVMSVRSQPGINSEYAKLP
29 EHO91218 kdiynnyfagflikfeideqnnplfiyqftltskfnkwwkmsvrsqpginseeyaklp
30 CC72-1 -----
31

32 CC80-3 LVLPNKLEQQKIAKFLDRFDRQIELEKQKIEILQQQKGLLQSMFI
33 EHO91218 lvlpnkleqqkiaefldrfdqqielekqkieilqqqkglqsmfi
34 CC72-1 -----
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60**S.SauJE GGA-6-RTGA****S.SauJE against ST49 strain "Tager 104"**

The ST49 Tager genome has the same TRD combinations as the ST49 strain 21262.

PATRIC db

>fig|1381115.3.peg.1063|VBIStaAur301678_1063| Type I restriction-modification system, specificity subunit S (EC 3.1.21.3) [Staphylococcus aureus subsp. aureus Tager 104 | 1381115.3] This is TRD R+f*

MSNTQKKNVPELRFPGFEGEWEEKKLGEVAKIYDGHQTPKYTNEGIFLSVENIKTLNS
SKYISEEAFEKEFKIRPEFGDILMTRIGDIGTPNIVSSNEKFAYYVSLALLKTKNLNSYF
LKNLILSSSIQNELWRKTLHVAFPPKINKNEIGKIKINYPKKQEQQKIGQFFSKLDRQIE
LEEQKLELLQQQKGYMQKIFSQELRFKDENGEDYPDWKEKKLGDITEQSMYGIGASATR
FDSKNIYIRITDIDEKSRKLNQNLTPDELNNKYKLRNDILFARTGASTGKSYIHKEE
KDIYNYFAGFLIKFEIDEQNNPLFIYQFTLTSKFNKWKVMSVRSQPGINSEEYAKLP
LVLPNKLEQQKIAEFLDRFDQIELEKQKIEILQQQKGLLQSMFI

>fig|1381115.3.peg.2628|VBIStaAur301678_2628| Type I restriction-modification system, specificity subunit S (EC 3.1.21.3) [Staphylococcus aureus subsp. aureus Tager 104 | 1381115.3] This is TRD J+E

MSNTQKKNVPELRFPGFEGEWEEKKLEDIIKVNKSGKDYKHLKGDIPVYGTGGYMTSVSE
PLSEIDAVGIGRGTINKPYLLEAPFWTVDTLFYCTPKKETDILFILSLFRKINWKVYDE
STGVPSLSKQTINKINRFVPTNKEQQKIGKFFSKLDRQIELEEQKIELLQQQKGYIQKI
FSQELRFKDENGDDYPEWEETTIQIEIAQINTGKDKTKDAITNGSYDFYVRSPIVYKINTF
SYEGEAILTVGDGVGVGVGFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKQK
KYSAKTSVDSVRKDMVANMKVPRPIYIEQEKIGQFIKKVDNKIKIQKQVIELLKQRKKAL
LQKMFI

S.SauJE against ST49 Tager 104**GGA-6-RTGA**

CLUSTAL O(1.2.1) multiple sequence alignment

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S.SauJE          MSNTQKKNVPELRFPGFEGEWEEKKLGDLIKVNSGKDYKHLKGDIPVYGTGGYMTSVSE
fig|1381115.3.peg.2628|VBIStaAur301678_2628| MSNTQKKNVPELRFPGFEGEWEEKKLEDIIKVNKSGKDYKHLKGDIPVYGTGGYMTSVSE
*****
S.SauJE          PLSEIDAVGIGRGTINKPYLLEAPFWTVDTLFYCTPKKETDILFILSLFRKINWKVYDE
fig|1381115.3.peg.2628|VBIStaAur301678_2628| PLSEIDAVGIGRGTINKPYLLEAPFWTVDTLFYCTPKKETDILFILSLFRKINWKVYDE
*****
S.SauJE          STGVPSLSKQTINKINRFVPSNKEQQKIGEFFIKLDRQIELEEQKLELLQQQKGYMQKI
fig|1381115.3.peg.2628|VBIStaAur301678_2628| STGVPSLSKQTINKINRFVPTNKEQQKIGKFFSKLDRQIELEEQKIELLQQQKGYIQKI
*****
S.SauJE          FSQELRFKDENGKDYPEWEETTIQIEIAQINTGKDKTKDAITNGSYDFYVRSPIVYKINTF
fig|1381115.3.peg.2628|VBIStaAur301678_2628| FSQELRFKDENGDDYPEWEETTIQIEIAQINTGKDKTKDAITNGSYDFYVRSPIVYKINTF
*****
S.SauJE          SYEGEAILTVGDGVGVGVGFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKQK
fig|1381115.3.peg.2628|VBIStaAur301678_2628| SYEGEAILTVGDGVGVGVGFHYVNGKFDYHQRVYKISDFKNYYGLLLFYFYSQNFLKQK
*****
S.SauJE          KYSAKTSVDSVRKDMIANMKVPRPIYIEQKKIGQFIKKVDNKIKIQKQVIELLKQRKKSL
fig|1381115.3.peg.2628|VBIStaAur301678_2628| KYSAKTSVDSVRKDMVANMKVPRPIYIEQEKIGQFIKKVDNKIKIQKQVIELLKQRKKAL
*****
S.SauJE          LQKMFI PGSSHHHHH
fig|1381115.3.peg.2628|VBIStaAur301678_2628| LQKMFI-----
*****

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S. SauNQ ACC-5-RTGT

This TRD pair was found in strains KPL1845 (ST96) and 21343 (ST88).
 Subspecies 21343 contains SauNQ and a novel TRD (NOVEL 1) paired with TRD K.

>EHQ67679 THIS IS TRD NOVEL 1 + TRD K

MSNTQKKNVPELRFPGFEGEWEEKKGGEVATFAKGLGAKKDVSQNGVVPVILYGELYTKY
 GAIVSKIFSKTDIPENKLMMAKKNVDLIPSSGETAIDIATASCIYLNKGVAVGGDINILT
 PQKQDGRFISLSINGINKNELSKYAQGKTVVHLYNNDIKNLKIAFPSEFEEQVRIGNFFS
 KLDRQIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGNDYPKWEKKIEDIASQVYGG
 GTPNTKIKEFWNGDIPWIQSSDVKVNDLILQQCNKFISKNSIELSSAKLIPANSIAIVTR
 VGVGKLCIVEFDYATSQDFLSLSSLYDKLYSLYSLLYTMKKISANLQGTSIKGITKKEL
 LDSIIKIPHNLEEQQKIGDLFYKIDKYISFNKCKIEILKSLKQGLLKKMFI

>EHQ71248 THIS IS TRD N+Q ACC-5-RTGT

MSNTQTKNVPELKFPEFEGEWEEKKGGEFAGKVTKKNVDKKYIETLTNSAELGIISQKDY
 FDKEISNIDNIKKYYVVEENDFVYNPRISNYAPFGPVNRNKLKGGVMSPLYTVFKIQNI
 DLNFIEFYFKSSKWYRFMALNGDSGARADRFSIKNRTFMEMPLHIPCMDEQIKIGQFFSK
 LDRQIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGNDYPEWEERRFADIFKFHNKLR
 KPIKENLRVKGSYPYGGATGIIDYVDDFIFDGNLYLLIGEDGANIITRSAPLVYLVNGKFW
 VNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAQPKLNIQNLKIIISVVIISTNLEEQQK
 IGSFLSKLDRQIDLEEQQKLELLQQRKKALLKSMFV

SPECIES KPL1845 CONTAINS THREE *SauI* S SUBUNITS.**>ETD06224 THIS IS TRD N+Q ACC-5-RTGT**

MSNTQTKNVPELKFPEFEGEWEEKKGGEFAGKVTKKNVDKKYIETLTNSAELGIISQKDY
 FDKEISNIDNIKKYYVVEENDFVYNPRISNYAPFGPVNRNKLKGGVMSPLYTVFKIQNI
 DLNFIEFYFKSSKWYRFMALNGDSGARADRFSIKNRTFMEMPLHIPCMDEQIKIGQFFSK
 LDRQIELEEQKLELLQQQKKGVMQKIFSQELRFKDENGNDYPEWEERRFADIFKFHNKLR
 KPIKENLRVKGSYPYGGATGIIDYVDDFIFDGNLYLLIGEDGANIITRSAPLVYLVNGKFW
 VNNHAHILSPLNGNIQYLYQVAELVNYEKYNTGTAQPKLNIQNLKIIISVVIISTNLEEQQK
 IGSFLSKLDRQIDLEEQQKLELLQQRKKALLKSMFV

>ETD11204 THIS HAS TWO NOVEL TRDS, NOVEL 2 + NOVEL 3.

MTEQINTPELRFPEFKNEWSYDLVSDVVTNKSCKFDPKKEEAKKDIELDSIEQNTGRLLD
 TYISNDFTSQKNKFNKGNVLYSKLRPYLNKYYYATIDGVCSSSEIWVNLNTLNKDVLANKFL
 YYFIQTNRFSVTNKSAGSKMPRADWELVKNIRLYKGSIEEQEKIGYFFSKLDRQIELEE
 KKLELLEQQKKGVMQKIFAQELRFKDENGNDYPDWVTKKLDIGKVMNKRIYKNETTEN
 GEIPFYKIGNFGKNADTFITREKFDEYKEKYPYPNVGDILISASGSIGRTIEYTGEDAYY
 QDSNIVWLNHNDEVINKYLKYFYKIVKWSGIEGTTIKRLYNKNILNTKIELPTVEEQYKM
 ANFLSKLDKIIDIQIEKIELLKQRKQGLLQKMFV

>ETD09130 THIS HAS A NOVEL TRD (NOVEL 4) PAIRED WITH TRD f*

1MSNTQKKNVPELRFPEFEGEWKDVKFVSIFQEVSNKTSDLAKYPLFSLTVEKGITPKTER
 61YKRDFLVKKSDFNKIVEPRDIVYNPMNVTLAGAIDLSKYNIDIALSGYYHVMKIINSFNPD
 121FISNFLKTEKMIHYKKIATGSLMEKQRVHFSEFKNIKKFPTNKEQQKIGDFFSKLDRQ
 181IELQVQKLELLQQQKKGVMQKIFSQELRFKDENGEDYPDWKEKKLGDITEQSMYGIGASA
 241TRFDSKNIYIRITDIDEKSRKLNQNLTPDELNNKYKLRNDILFARTGASTGKSYIHK
 301EEKDIYNYFAGFLIKFEIDEQNNPLFIYQFTLTSKFNKWKVMSVRSQPGINSEYAK
 361LPLVLPNKLEQQKIAEFLDRFDQQIELEKQKIEILQQQKGLLQSMFI

PROMALS ALIGNMENT OF TRD AMINO ACID SEQUENCES WITH SECONDARY STRUCTURE PREDICTIONS.

"e" means beta strand and "h" means alpha helix in the consensus secondary structure.

PROMALS alignment of all first TRDs.

Table with 3 columns: Conservation, sequence alignment, and line numbers. Includes entries for NOV_4_189, Z_GAC_191, NOV_2_194, NOV_1_199, R_GARA_192, J_GGA_172, N_ACC_198, O_CAAC_195, T_CAAG_199, C_GWAG_206, M_CAG_203, X_TCTA_192, B_AGG_199, A_CCAAY_203, e*_GAG_190, V_CNGA_210, b*_GGHA_200, and Consensus_ss: eee eeeeeeeeee eeeeeeeeee.

Table with 3 columns: Conservation, sequence alignment, and line numbers. Includes entries for NOV_4_189, Z_GAC_191, NOV_2_194, NOV_1_199, R_GARA_192, J_GGA_172, N_ACC_198, O_CAAC_195, T_CAAG_199, C_GWAG_206, M_CAG_203, X_TCTA_192, B_AGG_199, A_CCAAY_203, e*_GAG_190, V_CNGA_210, b*_GGHA_200, and Consensus_ss: e ee eeeee eeeee eeee eeeee.

Table with 3 columns: Conservation, sequence alignment, and line numbers. Includes entries for NOV_4_189, Z_GAC_191, NOV_2_194, NOV_1_199, R_GARA_192, J_GGA_172, N_ACC_198, O_CAAC_195, T_CAAG_199, C_GWAG_206, M_CAG_203, X_TCTA_192, B_AGG_199, A_CCAAY_203, e*_GAG_190, V_CNGA_210, b*_GGHA_200, and Consensus_ss: hhhhhhhh hhhhhhhh hhhh ee hhhhhhhhhhhhhhhh.

Table with 3 columns: Conservation, sequence alignment, and line numbers. Includes entries for NOV_4_189, Z_GAC_191, NOV_2_194, NOV_1_199, R_GARA_192, J_GGA_172, N_ACC_198, O_CAAC_195, T_CAAG_199, C_GWAG_206, M_CAG_203, X_TCTA_192, B_AGG_199, A_CCAAY_203, e*_GAG_190, V_CNGA_210, b*_GGHA_200, and Consensus_ss: hhhhhhhh.

PROMALS alignment of all second TRDs.

Conservation: 9989799997999999896797 9 5
 NOVEL3_205 1 KKGVMQKIFAQELRFKDENGNDYPDWVTKKGLDIGKIVAMNKKRIYKNE-----TTENGEIPFYKIGNFG 63
 S_GCA_200 1 KKGVMQKIFSQELRFKDENGNDYPDWVNERLGEVTTVTMGOQSPKSVN-----YTDSNDTVLIQGNADIE 65
 d*_CYAA_220 1 KKGVMQKIFSQELRFKDENGNDYPWENVMQKVLKDKTEGIRKGPFGGALKKDFVVESSGAYVYQRNAI 70
 a*_GAA_208 1 KKGVMQKIFSQELRFKDENGNDYPEWENKRIEDIANVNKGFTPSTNN-----NEYWDNNDKNWLISIAGNM 65
 E_TCA_194 1 KKGVMQKIFSQELRFKDENGNDYPEWETTIEIAQINXGKDKTD-----AITNGSYDFYVRSPIV 62
 W_CRAA_211 1 KKGVMQKIFSQELRFKDENGNDYPDWEKQLGELSQVIRGASPRPIK---DPKWFNKESDIQWLRISDV 67
 Q_ACAY_197 1 KKGVMQKIFSQELRFKDENGEDYSEWEERRFADIFKFHNLKRPIKE-----NLRVKGSGYPYATGATGII 64
 G_ACA_196 1 KKGVMQKIFSQELRFKDENGEYPEWENKFKIDIFIFENNRKRPKITS-----SLREKGLYPYATGATGII 64
 f*_GAA_224 1 KKGVMQKIFSQELRFKDENGEDYPDWEKQIDITEQSMYGI GASA-----TRFDSKNIYIRITDID 62
 L_TTTA_213 1 KKGVMQKIFSQELRFKDENGNDYPNWRTEIENLNLENIVDNRGKTPD-----NAPSEKYPLEVNALG 63
 Y_CTA_209 1 KKGVMQKIFSQELRFKDENGNDYPDWEKVKLKEIACVYTGNTPSKKE-----NIYWNKGCVVWVTFDIN 65
 U_GAY_193 1 KKGVMQKIFSQELRFKDENGEDYPDWEVTTIQNTKYSKSKSSNQY-----ADKNSKGYVYDAVQEI 65
 I_YTCA_220 1 KKGVMQKIFSQELRFKDENGNDYPDWERIKFDFVDKVIDFRGRTPK---KLNMEWSDEGALALSAVNVK 67
 K_GCA_212 1 KKGVMQKIFSQELRFKDENGNDYKWEKKEKIEDIASOVYGGGTPNTK-----IKEFWNGDIPWIOQSDVK 65
 D_ATC_204 1 KKGVMQKIFSQELRFKDENGEDYKWEKSKIEKYKERNERSDK-----GQMLSVTINSIGI 56
 c*_GAY_209 1 KKGVLQKIFSQELRFKDENGNDYPEWFRFARPKDFMYKPINIRPAINI-----SKSELLTVKLCHE 59
 P_AGG_214 1 KKGVMQKIFSQELRFKDESGNDYPDWEKQIDITEQSMYGI GASA-----KPLPLTISGQLGLIDITTEYF 66
 F_TTAA_216 1 KKGVMQKIFSQELRFKDEEGKDYDPAWKSSIQEIFENKGGTALETE-----FNFDGNKYKVISIGSYS 62
 H_TAC_206 1 KKCIVQKIFSQELRFKDEEGNYKGNWVKQKLDVLEFSNKRTINE-----NEYVLTSSRRQG 57
 Consensus_ss: **hhhhh** **eeeeeh** **eeee** **eeeeeeeee**

Conservation: 5 7
 NOVEL3_205 64 KNADTFITR--EKF----DEYKEKYPYPNVGD-ILISASG-----SIGRTIEYTG--EDA-YYQDSNIV 117
 S_GCA_200 66 NGLINPR-----IYTREVTKLIQKDE-IILTVRA-----PVGKLAMAQIN----ACIGRGGVC 112
 d*_CYAA_220 71 YDISNFRIY--INEN---KYKEMQSFVQPNPDI-IMSCSG-----TIGRLALIPH-NYTK-GIINQALI 126
 a*_GAA_208 66 QKLYL-KGN-KGIS---KDAAKNYMKVKNDT-LIMSFKL-----TIGKLAIVKAP----LYTNEAIC 117
 E_TCA_194 63 YK-----INTFSEGEAAILTVGDBGV-----GVGKVFHYVNGK---FDYHQRVY 102
 W_CRAA_211 68 NQNGKIHLEKQLS---IEGQEKTRVLTHTH-LLLSIAA-----SIGKPMVMFVK-----TGVDHGF 121
 Q_ACAY_197 65 DYV-----DDFIEDGNY-LLIGEDGANII TRSAPLVYLVNGK-----FWVNNHAH 108
 G_ACA_196 65 DYV-----KDYLFNNEERLLIGEDGAK-WGQFETSSFIANGQ-----YWVNNHAH 108
 f*_GAA_224 63 EKSRLKNYQ-NLTT---PDELNKNYKIKLRND-ILFARTGA-----STGKSYIHKEEKDIYNYFAGFLI 121
 L_TTTA_213 64 YRPAIKV-SKFSV-ENTYNNWFREHLKEND-ILFSTVG-----NTGIVSLMDNYK----AVIAQNI 120
 Y_CTA_209 66 NSKNIESE-NKLT---QEGYKARQLPENT-LLVTCIA-----SIGKNAILRQK-----GSCNQIN 118
 U_GAY_193 66 GK-----DSNYDIEESY-ISILKDGA-----GVGRNLNLRPGK-----SSVIGTM 104
 I_YTCA_220 68 KGIYDFNV-AYGNLDLYTRWMRGNELYKQ-VLFTTEA-----PMGNVAQVDPNKG---YILSRITI 126
 K_GCA_212 66 VNDLILQCNFKFISK--NSIELSSAKLIPANS-IAIVTRV-----GVGKLCVLEFD-----YATSQDF 121
 D_ATC_204 57 IKFSELDR--KDNS---SKNKSQYKVVRRND-IAYNSMRM-----WQGASGKSNYN---GIVSPAYT 109
 c*_GAY_209 60 KGIEK-ANI-NRVL---KLGATNYKRFEGQ-FYIGKQNF-----FNGAFDIVPKKFDG---LYSSDVP 115
 P_AGG_214 67 SKSVS-----SKNLNYTLTKNGE-FAYNKSYSN---GYPLGAIKRLTRYDS---GVLSLVI 117
 F_TTAA_216 63 INSTYNDQN-IRVN---KNKTEKYIILSKGD-LAMVLNDKTKDGKIGRSIFIDKDNQ---YIYNQRTE 123
 H_TAC_206 58 LILQSDYKDKRKT---AESNIGVILPKNH-TYRSRS-----DDGIKFNLNLMIDV-GIISKYYP 115
 Consensus_ss: **ee** **eeee** **eeeeee** **eeee** **ee**

Conservation: 6 5 7 98 76 7
 NOVEL3_205 118 WLNHND-EVINKYLKYFYKI-----VKWSGIEG---TTIKRILNKNILNNTKIELEPT-VEEQYKMANFLLS 176
 S_GCA_200 113 SIKG-----DKFLYFLWFFATQNKWIRFSQG---STFESISGNDIRNIHIIKIPV-EDERTKIIKLLN 171
 d*_CYAA_220 127 RFRTNH-KIRSEFFLI FMRSNQMRKILEANPG---SAITNLPVKELKLI PFPPLPV-KFEQDKISQFTH 191
 a*_GAA_208 118 HFIWKNVINTFEFYYLNSL---NISTFGQA---VKGVTLNNDNSINSIIVKLPN-EEEQNI IAKFLI 179
 E_TCA_194 103 KISDFK-NYYGLLLFYFYSQN-FLKETKYSAK---TSVDSVRKMIANMKVPRPI-YIEQKQIGQFIK 165
 W_CRAA_211 122 IFLKP--KFNLFYFLYWLEEF--KDKWSKYGQP---GSQVNLNSEIVKSQTLNMPNS-NHEQEKVQFPFN 182
 Q_ACAY_197 109 ILSPL--NGNIQYLVQVAEL---VNYEKYNTG---TAQPKLNIQNLI INVVISLTLNEEQKIGSFLS 168
 G_ACA_196 109 VVKSND--DHNLFFMYYLNF---KELRAVFTG---NAPAKLTHANLNCINLKI PC-LTEQDKVSALLK 167
 f*_GAA_224 122 KFKINE-QNSPLFYQFTLTSKFNKWKVMSVR---SQPGINSEEYAKLPLVLPN-KLEQQKIAKFLD 185
 L_TTTA_213 121 GLRVNN--NNSLPSFYIYMLSYKGNQKIKRIQMG---AVQPSVKSQPKFIKYLVI-KDEQEKVAKLLI 184
 Y_CTA_209 119 AVVPFE-NINIDYLYISDSL---STFMKSIAGK---TATQIVNKNTFENLEIYLAP-FEEQNKIADLI 180
 U_GAY_193 105 YIQSN--NVDIEFLLYRMKVV---DFPKYIIG---STIPLHYPKDYSKETLYIPSSIQEQAIGFMIS 164
 I_YTCA_220 127 AFDNSNE-KITDNFLASLLSSENVNDLKLCSG---ATAKGVSKNLDNLSYVTTIPHSISEQEEIAEF 191
 K_GCA_212 122 SLSSLK--YDKLISLGSY--LYTMKKSINLQG---TSIKGITKKEKLLNSI I KIPHNLEEQQKIGDLY 183
 D_ATC_204 110 VLYPTQ--NTSSLFYGYKFKTHRMIHKFINSOGL---TSDTWNLKYLKLNINIDIPV-LEEQEKIGDFFK 175
 c*_GAY_209 116 AFEINTEKIEPNYFISYSRPSFYKSKYSTG---TGSKRIHENTVLFSLHLPC-LNEQLKIASFVC 180
 P_AGG_214 118 CFSIKS-EMSKDFMEAYFDSTHWYVEVSGIAVEGARNHGLLNSVNDFFTILIKYPS-LEEQRKIGDFFI 185
 F_TTAA_216 124 RLIPFA-ENDNKFVFLMNTDLIRNKIKGMMGQ---ATQVYINYSIKLISIQPL-LEEQQKIRGFLE 187
 H_TAC_206 116 VFKGI--DANQYYTLHLNLYQ-LKKEYIKYATG---TSQLVLSQKLDQNIKTKLPS-YEEQKIGDFFS 177
 Consensus_ss: **eee** **hhhhhhh** **hhhhhhh** **hhhh** **ee** **hhhhhhhhh**

Conservation: 5 75 56 97 697 786
 NOVEL3_205 177 KLDKIIDIQIEKIELLQKQRKQLLQKMFV----- 205
 S_GCA_200 172 SLDVNSKTDLKIQNKKRQKQLLQKIFV----- 200
 d*_CYAA_220 192 IINRRIEQSEKKIESLKNRQKQGLQLKLFV----- 220
 a*_GAA_208 180 EVDKTVNQLVKTLLQKQRKQLLQRMFV----- 208
 E_TCA_194 166 RVDNKTQIQKQVIELLQKQRKALLQKMFV----- 194
 W_CRAA_211 183 RNEKLIELQKEKIMYIKRCKQVLLQKMFV----- 211
 Q_ACAY_197 169 KLDRQIDLEEQKLELLQKQRKALLKSMFV----- 197
 G_ACA_196 168 SIDNKMNQMNRIEILLKERRKELLQKMFV----- 196
 f*_GAA_224 186 RFDROIELEKQKIEILQQKQLLQSMFV----- 214
 L_TTTA_213 185 EIDKLVNKLKIELLQKQRKALLKSMFV----- 213
 Y_CTA_209 181 SLEELIEQKASKLIMKSRKQGMQLQIMFV----- 209
 U_GAY_193 165 NLDKLIENKLNKLNCLKQLKQLLQSMFV----- 193
 I_YTCA_220 192 KINQLVLELQYKIEHTKSQKQVFLQKMFV----- 220
 K_GCA_212 184 KIDKYISFNKCKIEMKSLKQGLLQKMFV----- 212
 D_ATC_204 176 KMDILISKQKIKIEILEKEKQSFQKMFV----- 204
 c*_GAY_209 181 FLNRKIELLERKIYLIKQKQKALLQSMFV----- 209
 P_AGG_214 186 KLDRQIELEEQKLELLQKQRKALLKSMLI----- 214
 F_TTAA_216 188 VLSGITTKQLHKIQKERRKAFLLQKMFV----- 216
 H_TAC_206 178 EIDRLVEKQSSKVGRKLVKRRKELLQKMFV----- 206
 Consensus_ss: **hhhhhhhhhhhhhhhhhhhhhhhhhhhh**

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2 PROMALS alignment of all TRDs.
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Conservation:	799997576	6	85	5		
CC80-3 f*	1	-----QELRFKDENGEDY	PDWKEKKLGDITEQSMY	GIGASA-----	TRFDSKNIYIRITDI	51
CC45-1L	1	-----QELRFKDENGNDY	PNWRTIELKNI	LENIVDNRGKTP-----	DNAPSEKYPLLEVNAL	52
CC97 c*	1	-----QELRFKDENGNDY	PEWRFARFKDFMYKPI	NIRPAIN-----	ISKSELLTVKHLHCK-GI	52
CC22-1I	1	-----QELRFKNENGNDY	PDWERIKFFDVIDKVID	FRGRTPKK----	LNMEWSDEGYLALSAVNV	56
CC873D	1	-----QELRFKDENGEDY	PHWENSKIEKYLKERNERS	SDKGQM-----	LSVTIN--SGIKFSEL	52
CC5-1D	1	-----QELRFKDENGEDY	PDWENSKIEKYLKERNERS	SDKGQM-----	LSVTIN--SGIKFSEL	52
CC30-1D	1	-----QELRFKDENSEDY	PHWENSKIEKYLKERNERS	SDKGQM-----	LSVTIN--SGIKFSEL	52
CC5-2H	1	-----QELRFKDEEGNY	KGNKKQLKDVLEFSNKRT	INE-----	NEYPVLTSSRQ	46
CC133-2fromED133 d*	1	-----QELRFKDENGNDY	PEWENVMLQKVLKDKTE	GIKRPFPGG-ALKKDI	FVESGGYAVYEQRNA	59
CC72-2S	1	-----QELRFKDENGNDY	PDWTNERLGEVTTVTMG	QSPKSVN-----	YTDNSNDTVLIQGNADI	54
CC93-3 a*	1	-----QELRFKDENGNDY	PEWENKRIEDIANV	NKGFPTSTNN-----	NEYWDDNDKNWLSIAGM	54
CC93-2K	1	-----QELRFKDENGNDY	PKWEEKKIEDIASQVY	GGGTPNTK-----	IKEFWNGDIPWIQSSDV	54
CC30-2K	1	-----QELRFKDENGNDY	PNWEEKKIEDIASQVY	GGGTPNTK-----	IKEFWNGDIPWIQSSDV	54
CC80-2W	1	-----QELRFKDENGNDY	PDWEEKQLGELSQIV	RGASPRPIK----	PKWFNKESDIGWLRISDV	56
CC75-2W	1	-----QELRFKDENGNDY	PDWEEKQLGELSQIV	RGASPRPIK----	PKWFNKESDIGWLRISDV	56
CC59Q	1	-----QELRFKDENGDY	SEWEERRFADIFKFHN	KLRKPIK-----	ENLVRKGSYPYIGATGI	53
CC72-1Q	1	-----QELRFKDENGNDY	PEWEERRFADIFKFHN	KLRKPIK-----	ENLVRKGSYPYIGATGI	53
CC1-2G	1	-----QELRFKDENGEY	PEWENKFIKIDIFIFEN	NRKRPIT-----	SSLREKGLYPYIGATGI	53
ST425-1E	1	-----QELRFKDENGDY	PEWEETTIKEIAQINT	GKDKTKD-----	AITNGSYDFYVRSPI	51
CC15TRD2E	1	-----QELRFKDENGNDY	PEWEETTIKEIAQIN	XGKDKTKD-----	AITNGSYDFYVRSPI	51
CC133_771E	1	-----QELRFKDENGDDY	PEWEETTIKEIAQINT	GKDKTKD-----	AITNGSYDFYVRSPI	51
CC398-1E	1	-----QELRFKDENGDY	PEWEETTIKEIAQINT	GKDKTKD-----	AITNGSYDFYVRSPI	51
CC80-1Y	1	-----QELRFKDENGNDY	PDWEEKKLEIACVYT	GNTPSKKE-----	NIYWNKGEYVWVTPDI	54
CC75-1U	1	-----QELRFKDENGEDY	PDWEVTTIQNITKY	SSKSSNQY-----	ADKDNSKGYPVYDAVQE	54
CC1-1F	1	KKGYMQKIFSQELRFK	DEEGKDYPDWKSQIQE	IFENKGGTALETE-----	FNFDGNYKVISIGSY	61
CC873 e*	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKS	ISSFLKESKIKGSNGS-----	HAKKLTVKLWKGKGVV	56
CC80-2Z	1	-MSNTQTKNVPELRF	PPGFE---GEYSLDIF	GNLATNKSEKFNPNQ-----	ENASIDIELDCIEQNTG	58
CC80-3XS.Saul1819ORF2227P	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKQ	FADFTKINQGLQIAINE-----	RKTEYSPELYFYITNEF	59
CC80-1X	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKQ	FADFTKINQGLQIAINE-----	RKTEYSPELYFYITNEF	59
CC75-1T	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKEL	GELIFQIISGSTPLKSN-----	KEFYENGINNWKTDDL	59
ST130-1T	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKL	GELIFQIISGSTPLKSN-----	KKFYENGINNWKTDDL	59
CC93-3M	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKL	LEDLGLFQKSYFSRA-----	KEGNGKTKHIHYGDI	56
CC133_771-1strain32320Hsd	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKL	GDGLGLFQKSYFSRA-----	KEGNGKTKHIHYGDI	56
CC133-2fromED133J	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKLES	IIKVNSGKDYKH-----	LDKGDIPYVGTGGY	54
CC72-2J	1	-MSNTQKKNVPELRF	PEFE---GEWEEKL	GNIIKVNSGKDYKH-----	LDKGDIPYVGTGGY	54
CC51TRD1J	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKL	EDIKVNSGKDYKH-----	LDKGDIPYVGTGGY	54
CC30-2strainMRSa252HsdSJ	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKL	GDGLIKVNSGKDYKH-----	LEKGDIPYVGTGGY	54
CC59-1J	1	-MSNTQKKNVPELRF	PEFE---GEWEERK	LDGLIKVNSGKDYKH-----	LDKGDIPYVGTGGY	54
CC72-1R	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKL	GEVAKIYDGTHTPK-----	YTNEGIKFLSVENI	55
CC15TRD1O	1	-MSNKQKKNVPELRF	PPGFE---GEWEEKL	GEVGTFTSGGTPLKS-----	KSEYWNGDIPWITTDGI	58
CC398-1strain398HsdSN	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKL	GEFAGKVTQKNVDKDY-----	IETLTSNSELGIIISQKDY	60
ST425-1C	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKQ	VGELLEFKNGLNKGKE-----	YFGSGSSIVNFKDV	55
CC30-1strainMRSa252HsdSC	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKV	GELLEFKNGLNKGKE-----	YFGSGSSIVNFKDV	55
CC45-1strain3067HsdSC	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKV	GELLEFKNGLNKGKE-----	YFGSGSSIVNFKDV	55
CC97A	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKQ	LGDLTTKIGSGKTPKGG-----	SENYTNKGIPLFRSQNI	59
CC1-2strainMW2HsdSA	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKL	GNLTTKIGSGKTPKGG-----	SENYTNKGIPLFRSQNI	59
CC1-1strainMW2HsdSA	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKL	GDLTTKIGSGKTPKGG-----	SENYTNKGIPLFRSQNI	59
CC5-2strainN315HsdSA	1	-MSNTQTKNVPELRF	PPGFE---GEWEEKL	GNLTTKIGSGKTPKGG-----	SENYTNKGIPLFRSQNI	59
CC75-2V	1	-MSNTGKMNVPELRF	PPGFE---GEWEEKL	RELRLNPKDKYSYTG	GGPFGSDLKSDYTTDGIQI	65
CC22-1strain5096HsdSB	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKL	GDLDLDRVIRKKNLES----	KKPLTISGQLGLIDQTEY	60
CC51TRD2P	1	-----QELRFKDESNDY	PDWEEKLGEVADR	VIRKKNLES----	KKPLTISGQLGLIDQTEY	55
CC5-1strainN315HsdSB	1	-MSNTQKKNVPELRF	PPGFE---GEWEEKL	GDLDLDRVIRKKNLES----	KKPLTISGQLGLIDQTEY	60
CC93-2 b*	1	-MSNTQKKNVPELRF	PEFE---GEWEEKL	EDTLEFIKDGTHGTH-----	ENVNNGPWLLSAKNI	56
Consensus_ss:			eeeeeeeeeeee		eeeeeee e	

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Conservation:	5	5
CC80-3 f*	52 DEKSRKLN-YQNLTP---DELNNKYKLRNDILFARTGAST-----GKS-YIHKEEKDIYNYFAGFL	110
CC45-1L	53 GYRPAIY-KVSKFVSE-NTYNNWFREHLKENDILFSTVGNT-----GIV-SLMDN----YKAVIAQNI	109
CC97 c*	53 EKANINRV-----LKLGATNYKRFEGQFIYKQNFN-----GAF-DIVPKK--FDGLYSSSDV	104
CC22-1I	57 KKGYIDFNVEAKYGNLD-LYTRMWRGNELYKQVLFTEAPM-----GNV-AQVPD---NKGYILSQRT	115
CC873D	53 DRKDN-----SSKNKSNYKVVVRKNDIAYNSMRMQ-----GAS-GKSNY-----NGIVSPAY	98
CC5-1D	53 DRKDN-----SSKDKSNYKVVVRKNDIAYNSMRMQ-----GAS-GKSNY-----NGIVSPAY	98
CC30-1D	53 DRKDN-----SSKDKSNYKVVVRKNDIAYNSMRMQ-----GAS-GRSNY-----NGIVSPAY	98
CC5-2H	47 GLILQSD---YYKDRKT-FAESNIGYFILPKNHITYRSRSD---GIFKFNLNLN---IDVGIISKYY	104
CC133-2fromED133 d*	60 IYDISNF---RYIINE-NKYKEMQSFVQPNDIIMSCSGTI-----GRL-ALIPHN---YTKGIINQAL	115
CC72-2S	55 ENGL-----INP-RIYTRVTKLIQKDEIILTVRAPV-----GKL-AMAQI-----NACIGRGV	101
CC93-3 a*	55 NQKYLK---GNKGIS---KDAAKNYMKVKNLTLIMSFKLT---GKL-AIVKA-----PLYTNEAI	106
CC93-2K	55 KVNDLILQ-QCNKFIK-NSIELSSAKLIPANSIAIVTRVGV-----GKL-CLVEF-----DYATSQDF	110
CC30-2K	55 KVNDLILR-QCNKFIK-NSIELSSAKLIPANSIAIVTRVGV-----GKL-CLVEF-----DYATSQDF	110
CC80-2W	57 TNQNGKIY-HLEQKLS---IEGQEKTRVLVTHLLLSIAASI-----GKP-VMNFV-----KTGVHDGF	110
CC75-2W	57 TNQNGKIY-HLEQKLS---IEGQEKTRVLVTHLLLSIAASI-----GKP-VMNFV-----KTGVHDGF	110
CC59Q	54 IDYV-----DDFIFDGNLYLLIGEDGA-NIITRSAPLVYLVNG-----KFWVNNHA	97
CC72-1Q	54 IDYV-----DDFIFDGNLYLLIGEDGA-NIITRSAPLVYLVNG-----KFWVNNHA	97
CC1-2G	54 IDYVK-----DYLFNNEERLLIGEDGA-KWGQFETSS-FIANG-----QYVNNHA	97
ST425-1E	52 VYKI-----NTFSYEGEAILTVGDGVGV-----GKV-FHYVN---GKFDYHQRV	91
CC15TRD2E	52 VYKI-----NTFSYEGEAILTVGDGVGV-----GKV-FHYVN---GKFDYHQRV	91
CC133_771E	52 VYKI-----NTFSYEGEAILTVGDGVGV-----GKV-FHYVN---GKFDYHQRV	91
CC398-1E	52 VYKI-----NTFSYEGEAILTVGDGVGV-----GKV-FHYVN---GKFDYHQRV	91
CC80-1Y	55 NNSKNY---ESENKLT---QEGYKARQLPENTLLVTCIASI-----GKN-ALLRK---QCSNQQI	107
CC75-1U	55 IKG-----DSNYDIEESYISILKDGAGV-----GRL-NLRPG---KSSVIGTM	93
CC1-1F	62 SINSTYN--DQNIQV---KNKTEKYILSKGDLAMVLNDKTKDGKIIGRS-IFIDK---DNQYIYNQRT	122
CC873 e*	57 PKKETP-----KGSNDTQYKRRAGQLMYKGLDFLN---CAF-GIVPD---SLNNYESTID	105
CC80-2Z	59 RLKIYIN-----SKEFSSQKNKFNPNQVLYGKLRPYL-----NKY-YFTKK---SGVCSSEI	106
CC80-3XS.Sau11819ORF2227P	60 LRPNS-----QTKY-FIENPPQSVIANKEDILMTRTGTN-----GKV-VTNVF---GAFHNFF	108
CC80-1X	60 LRPNS-----QTKY-FIENPPQSVIANKEDILMTRTGTN-----GKV-VTNVF---GAFHNFF	108
CC75-1T	60 NNSKVTH---SKEKITE-YAMKSLKLVKNSVLIAMYGGFNQI---GRT-GLLKI---DATINQAI	116
ST130-1T	60 NNSKVTH---SKEKITE-YAMNSLKLKLVKNSVLIAMYGGFNQI---GRT-GLLKI---DATINQAI	116
CC93-3M	57 HSKFKTV--LDS DGNIP-NIIEKAVFELIQKGDIVFADASEDYSDL--GKA-VMIDFE--PNSLISGLHT	118
CC133_771-1strain32320Hsd	57 HSKFKTV--LDS DGNIP-NIIEKAVFELIQKGDIVFADASEDYSDL--GKA-VMIDFK--PNSLISGLHT	118
CC133-2fromED133J	55 MTS-----VSEPLSEIDAVGIGRKGTI-----NKP-YLLEA---PFWTVDTL	92
CC72-2J	55 MTS-----VSEPLSEIDAVGIGRKGTI-----NKP-YLLEA---PFWTVDTL	92
CC51TRD1J	55 MTS-----VSEPLSEIDAVGIGRKGTI-----NKP-YLLEA---PFWTVDTL	92
CC30-2strainMRSA252HsdSJ	55 MTS-----VSEPLSEIDAVGIGRKGTI-----NKP-YLLEA---PFWTVDTL	92
CC59-1J	55 MTS-----VSEPLSEIDAVGIGRKGTI-----NKP-YLLEA---PFWTVDTL	92
CC72-1R	56 KTLNSS----KYISE-EAFEKEFKIRPEFGDILMTRIGDI-----GTP-NIVSS---NEKFAYYVSL	108
CC15TRD1O	59 HNIKREN---ITNFITE-KGLNNESSAKLITNEALIAMYGQKTR---GMS-ALNLF---EATTNQAC	115
CC398-1strain398HsdSN	61 FDKEIS-----NIDNIKKYVVVEENDFVYNPRMSNYAPF--GPV-NRNLK---GKKGVMSPLY	112
ST425-1C	56 FNNRSINT--NNLTGKVN-VNSKELKNYSVEKGDVFFTRTSEVIGEI--GYP-SVILND--PENTVFSGFV	118
CC30-1strainMRSA252HsdSC	56 FNNRSINT--NNLTGKVN-VNSKELKNYSVEKGDVFFTRTSEVIGEI--GYP-SVILND--PENTVFSGFV	118
CC45-1strain3067HsdSC	56 FNNRSINT--NNLTGKVN-VNSKELKNYSVEKGDVFFTRTSEVIGEI--GYP-SVILND--PENTVFSGFV	118
CC97A	60 RNGKLN--NDLVYISK-DIDDEMKNSTRYYGDVLLNITGASI-----GRT-AINSIV--ETHANLNQHV	118
CC1-2strainMW2HsdSA	60 RNGKLN--NDLVYISK-DIDDEMKNSTRYYGDVLLNITGASI-----GRT-AINSIV--ETHANLNQHV	118
CC1-1strainMW2HsdSA	60 RNGKLN--NDLVYISK-DIDDEMKNSTRYYGDVLLNITGASI-----GRT-AINSIV--ETHANLNQHV	118
CC5-2strainN315HsdSA	60 RNGKLN--NDLVYISK-DIDDEMKNSTRYYGDVLLNITGASI-----GRT-AINSIV--ETHANLNQHV	118
CC75-2V	66 GDGYFYN--SNKVFTSN-EKAEVLKSCNVFPGDIVIAKMDAPI-----ARA-AIVPDN-NIGKYLMSADG	125
CC22-1strain5096HsdSB	61 FSKSVS-----SKNLENYTLIKNGEFAYNKSYNGYPL--GAI-KRLTR---YDSGVLSLly	111
CC51TRD2P	56 FSKSVS-----SKNLENYTLIKNGEFAYNKSYNGYPL--GAI-KRLTR---YDSGVLSLly	106
CC5-1strainN315HsdSB	61 FSKSVS-----SKNLENYTLIKNGEFAYNKSYNGYPL--GAI-KRLTR---YDSGVLSLly	111
CC93-2 b*	57 KNNKIIIS-SDDRKISESDYKIKYKYLEKGLDILLTIVGTI-----GRA-AIVKN---PNNIAFORSV	115

Consensus_ss: e e ee eeeee eee eee eeee e

	Conservation:	7	6	98 775
1				
2	CC80-3 f*	111	IKFKINE---QNSPLFIYQFTLTSKFNKWKVMS---VRSQQPGINSEYAKLPLVLPN-KLEQQKIAK	172
3	CC45-1L	110	VGLRVNN---NNLPSFIYMLSYKGNQKKIKRIQ---MGAVQPSVKVVSQFKFKIYLVPI-KDEQEKVAK	171
4	CC97 c*	105	PAFEINT--EKIEPNYFISYISRPSFYKSKEKYS---TGTGSKRIHENTVNLNPSLHLPC-LNEQLKIAS	167
5	CC22-1I	116	IAFNSNE---KITDNFLASLLSENVDLLKLC---SGATAKGVSQKLNLRLYVTI PHSISEQEBIAE	178
6	CC873D	99	TVLYPTQ---NTSSLFIYGFKFKTHRMHFKFKINSQ--GLTSDTWNLKYKQLKNINIDIPV-LLEEQEKIGD	162
7	CC5-1D	99	TVLYPTQ---NTSSLFIYGFKFKTHRMHFKFKINSQ--GLTSDTWNLKYKQLKNINIDIPV-LLEEQEKIGD	162
8	CC30-1D	99	TVLYPTQ---NTSSLFIYGFKFKTHRMHFKFKINSQ--GLTSDTWNLKYKQLKNINIDIPV-LLEEQEKIGD	162
9	CC5-2H	105	PVFKGI---DANQYLLTLHLNYQ-LKKEYIKYA---TGTSQVLVLSQKDLQNIKTKLPS-YEEQQKIGD	164
10	CC133-2fromED133 d*	116	IRFRTNH---KIRSEFFLIFMRSNQMQQRKILEAN---PGSAITNLVVPVKELKLI PFPPLPV-KFEQDKISQ	178
11	CC72-2S	102	CSIKGD-----KFLYYFLEWFATQNKWIRFS---QGSTFESISGNDIRNIHIIKIPV-EDERTKIIK	158
12	CC93-3 a*	107	CHFHWK---NKINTEFIYYLNS---LNISTFGV---QAVKGVTLNNDINSINSIIVKLPN-EEEQNI IAK	166
13	CC93-2K	111	LSLSSL---KYDKLYSLYSLLY--TMKKISANL---QGTSIKGITKKELLDSIIKIPHNLEEQQKIGD	170
14	CC30-2K	111	LSLSSL---KYDKLYSLYSLLY--TMKKISANL---QGTSIKGITKKELLDSIIKIPHNLEEQQKIGD	170
15	CC80-2W	111	LIFLNP---KFNLFMYWLEY--FKDKWSKYG---QPGSQVNLNTEIVKSTQLNMPN-NHEQEKVQG	169
16	CC75-2W	111	LIFLKP---KFNLFMYWLEY--FKDKWSKYG---QPGSQVNLNTEIVKSTQLNMPN-NHEQEKVQG	169
17	CC59Q	98	HILSPL---NGNIQYLYQVAEL---VNYEKYN---TGTAQPKLNIQNLKI INVVISNLEEQQKIGS	155
18	CC72-1Q	98	HILSPL---NGNIQYLYQVAEL---VNYEKYN---TGTAQPKLNIQNLKI INVVISNLEEQQKIGS	155
19	CC1-2G	98	HVVKSN---DHNLFMYWLEY--FKDKWSKYG---QPGSQVNLNTEIVKSTQLNMPN-NHEQEKVQG	169
20	ST425-1E	92	YKISDFK---NYYGLLLFFYFYSQ-NFLKETKKYS---AKTSVDSVRKDMVANMKVPRPI-YIEQEKIGQ	152
21	CC15TRD2E	92	YKISDFK---NYYGLLLFFYFYSQ-NFLKETKKYS---AKTSVDSVRKDMVANMKVPRPI-YIEQEKIGQ	152
22	CC133_771E	92	YKISDFK---NYYGLLLFFYFYSQ-NFLKETKKYS---AKTSVDSVRKDMVANMKVPRPI-YIEQEKIGQ	152
23	CC398-1E	92	YKISDFK---NYYGLLLFFYFYSQ-NFLKETKKYS---AKTSVDSVRKDMVANMKVPRPI-YIEQEKIGQ	152
24	CC80-1Y	108	NAVVPFE---NINIDYLYYISDS--LSTFMKSI A---GKTATQIVNKNTFENLEIY LAP-FEEQKRIAG	167
25	CC75-1U	94	GYIQSN---NVDIEFLYYRMKV---VDFPKYI---IGSTIPHLVFKDYKSTETLPPS IQETQAKIGM	151
26	CC1-1F	123	ERLIPFA---ENDNKFLWFLMNTDLIRNKIKGMM---QGATQVYINYSIKLISIQIQLPL-LEEQQKIRG	184
27	CC873 e*	106	SPSDFPI---NGDSKFLLEIRIKLKS FYKFGDIA---NGSRKAKRINQDTPFLSIPVPAPK-YDEQRLRIG	168
28	CC80-2Z	107	WVLKSTKE-DKLNLFYFIQTKRYS-DVASKS---AGSKMPRADWGLIENIRVYFPE-LCEQDKVIGQ	169
29	CC80-3XS_Sau11819ORF2227P	109	KIKFDKN---LYDRFLVEVLNSSKIQNKILSLA---GSSTIPDLNHSDFYSISSYPL-LREQQKIGK	170
30	CC80-1X	109	KIKFDKN---LYDRFLVEVLNSSKIQNKILSLA---GSSTIPDLNHSDFYSISSYPL-LREQQKIGK	170
31	CC75-1T	117	SALLMNH---ETNPEFIQAFNLNYQV-KGWKRYAA---SSRKDPNITKKDIEQFKVPYVS-INEQQKIGE	177
32	ST130-1T	117	SALLMNH---ETNPEFIQAFNLNYQV-KGWKRYAA---SSRKDPNITKKDIEQFKVPYVS-INEQQKIGE	177
33	CC93-3M	119	HLFRPLN---NAISNFIYTKTLSYKFFIRQQG---TGISVLGISKKSLNINLVLI PRSELEQQKIGQ	181
34	CC133_771-1strain32320Hsd	119	HLFRPLN---NAISNFIYTKTLSYKFFIRQQG---TGISVLGISKKSLNINLVLI PRSELEQQKIGQ	181
35	CC133-2fromED133J	93	FYCTPKK---ETDILFILSLFRKIN---WKVYD---ESTGVPVSLSKQTINKINRFVPT-NKEQQKIGK	150
36	CC72-2J	93	FYCTPKK---ETDILFILSLFRKIN---WKVYD---ESTGVPVSLSKQTINKINRFVPT-NKEQQKIGK	150
37	CC51TRD1J	93	FYCTPKK---ETDILFILSLFRKIN---WKVYD---ESTGVPVSLSKQTINKINRFVPT-NKEQQKIGK	150
38	CC30-2strainMRSA252HsdSJ	93	FYCTPKK---ETDILFILSLFRKIN---WKVYD---ESTGVPVSLSKQTINKINRFVPT-NKEQQKIGK	150
39	CC59-1J	93	FYCTPEK---EADILFILSLFRKIN---WKLYD---ESTGVPVSLSKQTINKINRNVPT-NKEQQKIGE	150
40	CC72-1R	109	ALLKTK---NLNSYFLKNLILSSSIQNELWRKT---LHVAFPKKINKNEIGKIKINYPK-KQEQQKIGQ	170
41	CC15TRD1O	116	AIYQTN---QININVFQYFQK--LYEFLRSL S---NEGSQKNLSLSLKEITLNYPN-EQEQQKIGD	173
42	CC398-1strain398HsdSN	113	TVFKIQ---NIDLNFIEFYFKSSKWYRFMALNGD-SGARADRFSIKDRTFMEMPLHIPC-MDEQIKIGQ	176
43	ST425-1C	119	LRGRPKSGIDLINNNFKRYVFFTNSFRKEMITKS---SMTTRALTSGTAINMKVIVPVSAKEQKIGD	184
44	CC30-1strainMRSA252HsdSC	119	LRGRPKSGIDLINNNFKRYVFFTNSFRKEMITKS---SMTTRALTSGTAINMKVIVPVSAKEQKIGD	184
45	CC45-1strain3067HsdSC	119	LRGRPKSGIDLINNNFKRYVFFTNSFRKEMITKS---SMTTRALTSGTAINMKVIVPVSAKEQKIGD	184
46	CC97A	119	CIIRLKK---EYNYFFGQYLLSRKGRKIFLAQ---SGGSREGLNFKEIANLKI FTPTIFEEQQKIGK	181
47	CC1-2strainMW2HsdSA	119	CIIRLKK---EYNYFFGQYLLSRKGRKIFLAQ---SGGSREGLNFKEIANLKI FTPTIFEEQQKIGK	181
48	CC1-1strainMW2HsdSA	119	CIIRLKK---EYNYFFGQYLLSRKGRKIFLAQ---SGGSREGLNFKEIANLKI FTPTIFEEQQKIGK	181
49	CC5-2strainN315HsdSA	119	CIIRLKK---EYNYFFGQYLLSRKGRKIFLAQ---SGGSREGLNFKEIANLKI FTPTIFEEQQKIGK	181
50	CC75-2V	126	IRLSVDT---VHFNTKFVLECI NRKSFRRKVEDNS---SGSTRMIRGLS TLGSLTLTKTT-LKEQQKIGQ	188
51	CC22-1strain5096HsdSB	112	ICFSIKS---EMSKDFMEAYFDSTHWYREVSGIAVEGARNHGLLNVS VNDFFTLILIKYPS-LLEEQQKIGK	177
52	CC51TRD2P	107	ICFSIKS---EMSKDFMEAYFDSTHWYREVSGIAVEGARNHGLLNVS VNDFFTLILIKYPS-LLEEQQKIGK	172
53	CC5-1strainN315HsdSB	112	ICFSIKS---EMSKDFMEAYFDSTHWYREVSGIAVEGARNHGLLNVS VNDFFTLILIKYPS-LLEEQQKIGK	177
54	CC93-2 b*	116	AILKTKA---TYDVGFIFQLFQTKYFKNLLLRKQ---VVSAQPGLYLGDIRKIKISITNIEEQRKIGI	178
55	Consensus_ss:		eeeeee hhhhhhhh hhhhhhhh hhhhh ee hhhhhhhh	

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Conservation:	75	6	5	76	66	597	568568	
CC80-3 f*	173	FLDRFDRQIELEKQKIEILQQQKKGLLQSMFI						204
CC45-1L	172	LLIEIDKLVNKQLIKIEILLQQRKKALLKSMFI						203
CC97 c*	168	FVCFLNKRIEELLERKIYLIKKQKQALLQQMFI						199
CC22-1I	179	FFRKINQLVELQKYKIEHTKSQKQVFLQKMFI						210
CC873D	163	FFKKMDILISKQKIKIEILEKEKQSFLOKMFL						194
CC5-1D	163	FFKKMDILISKQKMKIEILEKEKQSFLOKMFL						194
CC30-1D	163	FFKKMDILISKQKIKIEILEKEKQSFLOKMFL						194
CC5-2H	165	FFSEIDRLVEKQSSKVGRLKVRKELLQKMFI						196
CC133-2fromED133 d*	179	FIHIINRRIEQSEKKIESLKNRKGFLQKLFV						210
CC72-2S	159	LLNSLDVLSKTDLKIQLNLRKQKQSLLOKIFV						190
CC93-3 a*	167	FLEVDKTVNQLVTKLLKQKGLLQRMFI						198
CC93-2K	171	LFYKIDKYISFNKCKIEMLKSLKQGLLKKMFI						202
CC30-2K	171	LFYKIDKYISFNKCKIEILKSLKQGLLQKIFI						202
CC80-2W	170	FFNRNEKLIELQEQEKIMYLRKRKQVLLQKMFI						201
CC75-2W	170	FFNRNEKLIELQEQEKIMYLRKRKQVLLQKMFI						201
CC59Q	156	FLSKLDRQIDLEEQKLELLQQRKKALLKSMFI						187
CC72-1Q		-----						
CC1-2G	155	LLKSIDNKMNQMNRIELLKERKELLQKMFI						186
ST425-1E	153	FIKKVDNKIKIQKQVIELLQQRKKALLQKMFI						184
CC15TRD2E	153	FIKRVDNKTKIQKQVIELLQQRKKALLQKMFI						184
CC133_771E	153	FIKKVDNKIKIQKQVIELLQQRKKALLQKMFI						184
CC398-1E	153	FIKRVDNKTKIQKQVIELLQQRKSLLOKMF						184
CC80-1Y	168	LISSELEELIEKQASKLIKMKSRKQGLQIMFI						199
CC75-1U	152	FISNLDKLIENKLNKLNKQLKQGLLQSMFI						183
CC1-1F	185	FLEVLSGITTKQLHKIDQLKERKKAFLQKMFI						216
CC873 e*	169	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						200
CC80-2Z	170	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						201
CC80-3XS.Sau11819ORF2227P	171	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						202
CC80-1X	171	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						202
CC75-1T	178	FFSKIDHQIELEEQKLELLQQQKKGYMOKIFS						209
ST130-1T	178	FFSKLDRQIELEEQKLELLQQQKK-----						201
CC93-3M	182	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						213
CC133_771-1strain32320Hsd	182	FFSKLDRQIELEEQKLELLQQQKKGYIQKIFS						213
CC133-2fromED133J	151	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						182
CC72-2J	151	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						182
CC51TRD1J	151	FFSKLDRQIELEEQKLELLFQQQKKGYMOKIFS						182
CC30-2strainMRSA252HsdSJ	151	FFIKLDRQIELEEQKLELLQQQKKGYMOKIFS						182
CC59-1J	151	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						182
CC72-1R	171	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						202
CC15TRD1O	174	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						205
CC398-1strain398HsdSN	177	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						208
ST425-1C	185	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFT						216
CC30-1strainMRSA252HsdSC	185	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						216
CC45-1strain3067HsdSC	185	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						216
CC97A	182	FFSKLDRQIELEEQKLELLQQQKKGYLQKIFS						213
CC1-2strainMW2HsdSA	182	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFT						213
CC1-1strainMW2HsdSA	182	FISKLDRQIELEEQKLELLQQQKKGYMOKIFS						213
CC5-2strainN315HsdSA	182	FFSKLDQIELEEQKLELLQQQKKCYIQKIFS						213
CC75-2V	189	FFSKLDRQIVLEEQKLELLQQQKKGYMOKIFS						220
CC22-1strain5096HsdSB	178	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						209
CC51TRD2P	173	FFIKLDRQIELEEQKLELLQQRKKALLKSMFI						204
CC5-1strainN315HsdSB	178	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						209
CC93-2 b*	179	FFSKLDRQIELEEQKLELLQQQKKGYMOKIFS						210
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