



Review Article

Endocrinol Metab 2015;30:456-466 http://dx.doi.org/10.3803/EnM.2015.30.4.456 pISSN 2093-596X · eISSN 2093-5978

Congenital Hypogonadotropic Hypogonadism and Kallmann Syndrome: Past, Present, and Future

Soo-Hyun Kim

Molecular Cell Sciences Research Centre, St. George's Medical School, University of London, London, United Kingdom

The proper development and coordination of the hypothalamic-pituitary-gonadal (HPG) axis are essential for normal reproductive competence. The key factor that regulates the function of the HPG axis is gonadotrophin-releasing hormone (GnRH). Timely release of GnRH is critical for the onset of puberty and subsequent sexual maturation. Misregulation in this system can result in delayed or absent puberty and infertility. Congenital hypogonadotropic hypogonadism (CHH) and Kallmann syndrome (KS) are genetic disorders that are rooted in a GnRH deficiency but often accompanied by a variety of non-reproductive phenotypes such as the loss of the sense of smell and defects of the skeleton, eye, ear, kidney, and heart. Recent progress in DNA sequencing technology has produced a wealth of information regarding the genetic makeup of CHH and KS patients and revealed the resilient yet complex nature of the human reproductive neuroendocrine system. Further research on the molecular basis of the disease and the diverse signal pathways involved will aid in improving the diagnosis, treatment, and management of CHH and KS patients as well as in developing more precise genetic screening and counseling regime.

Keywords: Hypogonadism; Kallmann syndrome; Puberty; Olfaction disorders; Infertility; Gonadotropins

INTRODUCTION

Idiopathic congenital hypogonadotropic hypogonadism (CHH) is a rare reproductive disorder that is primarily caused by a gonadotrophin-releasing hormone (GnRH) deficiency but with significant genetic heterogeneity. Clinically, this disorder is characterized by abnormally low plasma levels of luteinizing hormone (LH) and follicle-stimulating hormone (FSH) in conjunction with low or undetectable concentrations of circulating sex steroids. In approximately 50% of cases, CHH patients also suffer from a reduced or deficient sense of smell (hyposmia or anosmia, respectively), which is then termed as Kallmann syndrome (KS) [1,2].

KS was first recognized in 1856 by Maestre de San Juan [1] who observed patients with defective olfactory structures and a microphallus. A few years later, Kallmann et al. [2] identified the hereditary nature of this condition. In the 1950s, De Morsier and Gauthier [3] further described the partial or complete absence of the olfactory bulb (OB) and its axons in multiple hypogonadal males. Since then, a variety of non-reproductive dysfunctions and developmental anomalies have been observed in association with hypogonadism. This review aimed to provide a brief overview of the genetics, molecular pathogenesis, diagnosis, and treatment of CHH and KS, with a particular focus on recent progress in the field.

Received: 29 September 2015, Revised: 8 October 2015,

Accepted: 15 October 2015

Corresponding author: Soo-Hyun Kim

Molecular Cell Sciences Research Centre, St. George's Medical School, University of London, Cranmer Terrace, London SW17 0RE, United Kingdom Tel: +44-208-266-6198, Fax: +44-208-725-2993, E-mail: skim@sgul.ac.uk

Copyright © 2015 Korean Endocrine Society

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Sertoli & Leydig cells

Sex hormone production

gametogenesis

THE HYPOTHALAMIC-PITUITARY-**GONADAL AXIS**

Multiple developmental and neuroendocrine signaling pathways regulate the ontogeny and homeostasis of the GnRH neurons including the production, secretion, and action of GnRH [4]. The development of the olfactory system and GnRH neurons are intimately connected with each other and are often modulated by common cell surface receptors and chemoattractant or chemorepellent molecules. During early embryogenesis, GnRH neurons originate from the neural crest and ectodermal progenitors within the olfactory placode outside of the brain and then migrate in close association with the growing axons of olfactory receptor neurons and terminal nerves. After penetrating through the cribriform plate, the GnRH neurons arrive in the hypothalamus, where they detach from the olfactory axonal guides, become non-motile, and then disperse further into the brain basal lamina before undergoing terminal differentiation [5]. In mice, this migratory process begins at embryonic day E10.5 and is completed by E17.5 [6]. The timed and coordinated expressions of various cell adhesion molecules, axonal guidance cues, and extracellular matrix proteins, along with different neurotransmitters, transcription factors, and growth factors that regulate the migration of GnRH neurons, have been documented [5].

Within the hypothalamus, functional GnRH neurons extend their axonal processes to the medial eminence through which pulsatile GnRH is secreted into circulation via the hypophyseal portal system. GnRH binds to GnRH receptor 1 (GnRHR1) on gonadotroph cells in the anterior pituitary and stimulates the synthesis and secretion of LH and FSH. Subsequently, LH acts on the testes to stimulate testosterone production, and LH and FSH act on the ovaries to induce estrogen production which, in turn, leads to steroidogenesis and germ cell production [4]. GnRH is temporarily secreted at 3 to 6 months postnatally, which is more evident in boys than girls and is sometimes called "mini-puberty." GnRH secretion then remains dormant until the onset of puberty, when its reactivation initiates secondary sexual maturation [4]. Therefore, the normal development and properly coordinated actions of the hypothalamic-pituitary-gonadal (HPG) axis are essential for GnRH pulse generation and normal reproductive function (Fig. 1).

The examination of a 19-week-old human aborted fetus with X-linked KS revealed that the GnRH neurons were arrested in a tangle above the cribriform plate [7]. Because the initial differentiation and migration of the GnRH precursor cells ap-

Hypothalamus Preoptic area ARC GnRH neurons GnRH secretion Hypophyseal Median eminence portal vein network Gonadotrophs with GnRH receptors Anterior pituitary Posterior pituitary Hypophyseal veins **FSH** LH Systemic blood circulation Testis Ovary

Cumulus, mural &

Thecal cells

Fig. 1. The hypothalamic-pituitary-gonadal (HPG) axis. During early brain development, gonadotrophin-releasing hormone (GnRH)-releasing neurons (green) migrate from the nasal region to the hypothalamus, where they permanently reside and differentiate. Hypothalamic GnRH neurons secrete GnRH at the median eminence into the hypophyseal portal system and release pulsatile GnRH to the anterior pituitary. GnRH then binds to GnRH receptor 1 on the gonadotrophs to stimulate these cells to produce luteinizing hormone (LH) and follicle-stimulating hormone (FSH), which enter the systemic blood stream through the hypophyseal veins. LH and FSH act on the gonads (Sertoli and Leydig cells in testes and cumulus, mural and thecal cells in ovaries) to induce steroidogenesis and germ cell production which, in turn, maintains sexual competence. The release of kisspeptin from the hypothalamic neurons located in the arcuate (ARC) and anteroventral periventricular nuclei within the preoptic area is critically important for the re-initiation of pulsatile GnRH secretion at puberty. The developmental failure or misregulation of any one or combination of the genes involved in GnRH migration, secretion, and activity at any stage of development may result in congenital hypogonadotropic hypogonadism and Kallmann syndrome. AVPV, anteroventral periventricular nucleus.

peared to be normal, it was speculated that the subsequent axonal elongation, path-finding, and/or terminal differentiation

EnM

processes might have been disrupted and prevented the GnRH neurons from reaching the forebrain [7]. It was also proposed that the defective targeting, innervation, and synaptogenesis of the axons of olfactory sensory neurons to the OB anlage might have caused OB dysgenesis in KS patients. Following this study, CHH and KS were defined as the partial or complete failure of sexual development secondary to the failed embryonic establishment of hypothalamic GnRH neurons, which causes deficits in the secretion of sex hormones from the anterior pituitary and gonads [4-7].

GENETICS AND GENETIC TESTING

Recently, significant progress in genetic research regarding CHH has led to the identification of more than 31 different putative loci for this disorder, 17 of which are also associated with KS [8]. The current list of genes and information on their presumed biological activities and inheritance patterns are provided in Tables 1-3. Although these genes may be implicated in the etiology of approximately 50% of CHH/KS cases, the mutations in each of the genes account for less than 10% of such cases; furthermore, the majority of the underlying mechanisms have yet to be fully characterized [9,10].

CHH/KS may present as either a sporadic or a familial case following autosomal dominant, autosomal recessive, or X-linked recessive modes of inheritance. The variable phenotypes

among individuals carrying the same mutation within a pedigree, even between monozygotic twins [11], have led to proposals of digenicity or oligogenicity, in which mutations of multiple genes synergize to produce a more severe phenotype but contribute to the phenotype with a variable penetrance [12]. For instance, loss-of-function mutations of semaphorin 7A (SEMA3A) have been reported in KS patients, but monoallelic mutations of this gene are not sufficient to cause the disease phenotype [13]. Thus, mutations of SEMA3A likely contribute to the pathogenesis through synergistic effects with mutations in other genes. Heparan sulphate 6-O-sulfotransferase 1 (HS6ST1) mutations account for approximately 2% of CHH cases and have been identified in combination with fibroblast growth factor receptor 1 (FGFR1) mutations in KS patients [14]. Additionally, mutations of prokineticin receptor-2 (PROKR2) in combination with mutations of anosmin-1 (ANOSI) [15] and FGFRI [16] have been identified as well as potentially digenic patterns of nasal embryonic luteinizing hormone-releasing hormone factor (NELF)/ANOS1 and NELF/ tachykinin receptor 3 (TACR3) mutations [17]. Patients with complex genetic makeups such as these are estimated to comprise at least 20% of all cases [10].

Establishing a phenotype-genotype association can aid phenotype-driven priority screening and better inform patient management and counseling [9]. Recent advancements in DNA sequencing technology have led to a wealth of genetic informa-

Gene and protein (alternative names)	OMIM	Known biological activity	Reversible	Oligogenicity	Inheritance
ANOSI (KALI) (anosmin-1)	300836	Extracellular matrix protein modulating FGFR1 and integrin signaling. Guidance molecule for GnRH neuronal migration and survival.	Yes	Yes	X-linked recessive
FEZF1 (FEZ family zinc finger 1)	613301	Zinc finger-containing transcriptional repressor regulating the development of forebrain and neocortex. GnRH neuronal migration and survival.	ND	ND	Autosomal recessive
HESX1 (homeobox gene expressed in ES cells 1)	182230	Transcription repressor for homeodomain-containing genes. Also involved in combined pituitary hormone deficiency and septo-optic dysplasia.	ND	ND	Autosomal recessive/ dominant
IL17RD (SEF) (interleukin 17 receptor D)	606807	Negative regulator and interactant of FGFR1.	ND	Yes	Autosomal dominant
SEMA3A (semaphorin-3A)	614897	Guidance molecule for GnRH neuronal migration and axonal pathfinding.	ND	Yes	Autosomal dominant
SOX10 (SRY-related HMG-box gene 10)	602229	Related to testis-determining transcription factor SRY. Regulate neural crest development. Also involved in Waardenburg-Shah syndrome.	ND	ND	Autosomal dominant

mined; SRY, sex-determining region Y.



tion regarding CHH/KS patients. In particular, the recognition of oligogenic traits [10,12] has influenced perspectives on the genetic testing, diagnosis, and counseling for CHH/KS. For example, if an X-linked mode of inheritance is apparent, a mutation of KAL1 (note: following a nomenclature change by the Human Genome Organisation, this gene is now designated as

Gene and protein (alternative names)	OMIM	Known biological activity	Reversible	Oligogenicity	Inheritance
DMXL2 (Rabconnectin-3α)	616113	Synaptic protein involved in stimulation and homeostasis of GnRH neurons and gonadotrophs. Also mutated in polyendocrine-polyneuropathy syndrome.	ND	ND	Autosomal recessive
GNRH1 (gonadotropin-releasing hormone 1)	614841	Exclusively expressed by GnRH-releasing neurons. Binds to its receptor GnRHR to stimulate HPG axis.	ND	ND	Autosomal recessiv
GNRHR (gonadotropin-releasing hormone receptor)	146110	G-protein-coupled 7-transmembrane-domain receptor for GnRH, expressed in the gonadotrophs in the pituitary.	Yes	Yes	Autosomal recessiv
KISS1 (kisspeptin; metastin)	614842	Secreted by the hypothalamic neurons of arcuate and anteroventral periventricular nucleus. Binds to its receptor GPR54 to regulate GnRH neurons.	ND	ND	Autosomal recessiv
KISS1R (G-protein-coupled receptor 54)	614837	G-protein-coupled receptor for kisspeptin, expressed in hypothalamic GnRH neurons.	ND	Yes	A utosomal recessive dominant
LEP (leptin)	614962	Adipocyte-specific hormone regulating food intake, energy balance and fat metabolism. Associated with obesity.	ND	ND	Autosomal recessiv
LEPR (leptin receptor)	614963	Single-transmembrane-domain receptor for leptin. Preferentially expressed in hypothalamus.	ND	ND	ND
NR0B1 (DAX1) (nuclear receptor subfamily 0, group B)	300200	Negative regulator of retinoic acid receptor. Mutated in X-linked congenital adrenal hypoplasia with HH.	ND	ND	X-linked recessive
OTUD4 (OTU domain-containing protein 4)	611744	De-ubiquitinase found to be mutated in Gordon Holmes syndrome, a hypogonadism associated with cerebellar ataxia.	ND	ND	Autosomal recessive
PCSK1 (proprotein convertase-1)	162150	Required for processing of various pre-hormones including proopiomelanocortin, proinsulin, and proglucagon.	ND	ND	ND
PNPLA6 (patatin-like phospholipase domain-containing protein 6)	603197	Catalyzes the de-esterification of membrane phosphatidylcholine. Also mutated in Gordon Holmes and Boucher-Neuhauser syndrome.	ND	ND	Autosomal recessiv
RNF216 (ring finger protein 216)	609948	Zinc finger protein that binds and inhibits TNF and NF-κB. Also mutated in Gordon Holmes syndrome.	ND	ND	Autosomal recessiv
TAC3 (tachykinin 3; neurokinin B, neuromedin-K)	614839	Secreted in the hypothalamic neurons of arcuate nucleus. Binds to its receptor TACR3 to regulate the secretion and homeostasis of GnRH neurons.	Yes	Yes	Autosomal recessiv
TACR3 (tachykinin receptor 3; neurokinin B receptor, neuromedin-K receptor)	614840	G-protein-coupled receptor for TAC3. Expressed in hypothalamic GnRH neurons to regulate secretion and homeostasis of GnRH.	Yes	Yes	Autosomal recessiv

OMIM, Online Mendelian Inheritance in Man; ND, not determined; HPG, hypothalamic-pituitary-gonadal; GPR54, G-protein-coupled receptor 54; DAX1, dosage-sensitive sex reversal, adrenal hypoplasia critical region, on chromosome X, gene 1; HH, hypogonadotropic hypogonadism; TNF, tumor necrosis factor; NF-κB, nuclear factor-κB.

Gene and protein (alternative names)	OMIM	Known biological activity	Reversible	Oligogenicity	Inheritance
AXL (AXL receptor tyrosine kinase)	109135	Receptor tyrosine kinase containing fibronectin type III domain with oncogenic activity. Required for GnRH neuron migration.	ND	ND	ND
CHD7 (chromodomain helicase DNA- binding protein 7)	612370	Transcriptional regulator essential for the formation of neural crest and the development of forebrain, craniofacial bones and heart.	Yes	ND	Autosomal dominan
FGF8 (fibroblast growth factor 8)	612702	Ligand for FGFR1. Essential morphogen for development of forebrain, olfactory GnRH system, skeletal structure and heart.	ND	Yes	Autosomal dominan
FGF17 (fibroblast growth factor 17)	603725	Similar to FGF8 as a ligand for FGFR1, but more in patterning the dorsal frontal cortex.	ND	Yes	ND
FGFR1 (fibroblast growth factor receptor 1)	147950	Receptor tyrosine kinase essential for development of forebrain, craniofacial niche, and stimulation and secretion of GnRH neurons and gonadotrophs.	Yes	Yes	Autosomal dominar
HS6ST1 (heparan-sulphate 6O-sulphotransferase 1)	614880	Catalyzes the transfer of sulphate to position 6 of the N-sulfoglucosamine residue of heparan sulphate, essential for FGFR1 signaling activity.	Yes	Yes	Autosomal dominar
NSMF (nasal embryonic luteinizing hormone- releasing hormone factor; NELF)	614838	Guidance molecule for olfactory axon projections required for the axonophilic migration of GnRH neurons.	Yes	Yes	ND
PROK2 (prokineticin 2)	610628	Secreted by the hypothalamic neurons of suprachiasmatic nucleus that regulate circadian clock. Chemoattractant for subventricular zone neuronal progenitors. Involved in olfactory bulb morphogenesis and the migration and stimulation of GnRH neurons.	ND	Yes	A utosomal recessive dominant
PROKR2 (prokineticin receptor-2; GPR73L 1)	607123	G-protein-coupled receptor for PROK2. Regulate the formation of olfactory bulb, GnRH neuron and reproductive organs.	Yes	Yes	A utosomal recessive dominant
SEMA7A (semaphorin 7A)	607961	Membrane-anchored guidance molecule of the semaphorin family. Enhances axon outgrowth and interacts with integrin receptors.	ND	Yes	ND
WDR11 (WD repeat protein 11)	614858	Member of the WD repeat-containing protein family. Expressed in the forebrain and HPG axis.	ND	Yes	Autosomal dominar

ANOS1) is highly likely, especially when unilateral renal agenesis (30% to 40% of cases) and bimanual synkinesis (~75% of cases) are present [18,19]. Kidney anomalies have not been associated with FGFR1 mutations, which is consistent with findings showing that the conditional knockout of FGFR1 in the mouse ureteric bud did not result in any kidney defects [20]. On the other hand, dental agenesis, cleft palate, and/or skeletal anomalies accompanied by varying degrees of hypogonadism with or without anosmia are indicative of a defective fibroblast

growth factor (FGF) signaling pathway; thus, screening for *FGFR1*, *FGF8*, and *HS6ST1* is recommended [14,21].

If autosomal recessive inheritance is observed, alternations in *GNRHR* [22], kisspeptin receptor (*KISS1R*) [23], *TACR3* [24], *PROKR2* [25], or FEZ family zinc finger 1 (*FEZF1*) [26] can be suspected. If there are signs of CHARGE syndrome (Coloboma, Heart anomalies, Choanal atresia, Retardation of growth and/or development, Genital and/or urinary defects, and Ear anomalies and/or deafness), the chromodomain heli-

case DNA-binding protein 7 (*CHD7*) gene mutation should be considered a priority [27]. Mutations in dosage-sensitive sex reversal, adrenal hypoplasia critical region, on chromosome X, gene 1 (*DAXI*) cause CHH associated with adrenal insufficiency [28] and, if congenital deafness is present, then *CHD7*, SRY-related HMG-box gene 10 (*SOX10*), and/or interleukin 17 receptor D (*IL17RD*) can be suspected [21,29]. CHH with obesity may indicate a mutation of leptin (*LEP*), leptin receptor (*LEPR*), or proprotein convertase-1 (*PCSKI*) [30,31], whereas signs of other syndromic conditions, such as congenital ichthyosis [32] or spherocytosis [33], require that a comparative hybridization array or karyotype analysis be performed to detect any aberrant chromosomes.

THE EMERGING PICTURE OF MOLECULAR PATHOGENESIS

The developmental failure or misregulation of any one or combination of the genes and the signal pathways that are involved in GnRH migration, secretion, and/or activity at any stage will result in CHH and KS. For example, FGFR1 plays pleiotropic roles in human organogenesis including forebrain, skeleton, and neuroendocrine systems. The activity of this receptor is regulated not only by the binding of specific FGF ligands but also by alternative isoform expressions and other regulatory interactants. There is strong evidence to suggest that defects within the FGF signaling pathway underlie the pathogenesis of CHH/KS. Mutations in FGF8 (the ligand) and FGFR1 (the receptor) account for approximately 12% of CHH/KS cases [34]. Anosmin-1, a secreted extracellular matrix protein encoded by ANOS1, is the first mutated gene to be identified in KS patients [35]. Later it was shown that anosmin-1 binds to FGFR1 as a co-ligand and acts to fine-tune receptor signaling activity [36].

Heparan sulphate proteoglycans (HSPGs) are cell surface co-receptors that are essential for the formation of functional FGF/FGFR signaling complexes. The sequence specificity, length, and sulphation patterns of heparan sulphate are important regulators of receptor activity. Anosmin-1 may play a role in the recruitment of specific types of heparan sulphate to the FGF8/FGFR1 signaling complex [37]. In addition, mutations of *HS6ST1*, which is an enzyme that regulates the sugar modification of HSPG, have been identified in CHH patients [14]. More recently, mutations in other modulators of the FGFR1 signaling pathway, including FGF17, IL17RD, dual specificity phosphatase 6 (DUSP6), sprout homolog 4 (SPRY4), and fibronectin leucine rich transmembrane protein 3 (FLRT3), have

been identified [21]; all of these were initially identified as part of the so-called FGF8 synexpression group.

The kisspeptin and tachykinin signaling systems also play key roles in the expression and release of GnRH at the time of puberty. Subpopulations of neurons that co-express kisspeptin and neurokinin-B are located in the arcuate and infundibular hypothalamic regions [38], and kisspeptin-producing neurons comprise the major afferents to GnRH neurons, which act to regulate the tonic feedback control of GnRH and/or gonadotropin secretion as well as the pre-ovulatory surge [39]. However, during development, signaling via kisspeptin and its receptor GPR54 are not necessarily required for GnRH neuronal migration *per se* [40]. Signaling mediated by tachykinin-3 (TAC3), which is the precursor of neurokinin-B, and TACR3, which is also known as neuromedin-K receptor (NKR), are also important for the metabolic regulation of GnRH neurons [38].

Because reproduction requires an adequate energy supply, the metabolic status is important for the regulation, stimulation, and homeostasis of GnRH neurons and gonadotrophs. In animal models, the insulin and leptin signaling pathways stimulate the reproductive endocrine system and regulate GnRH neuronal function [41]. It has also been shown that kisspeptin-expressing neurons are sensitive to changes in leptin concentrations and mutations in *LEP* or *LEPR*, result in CHH [30]. However, GnRH and kisspeptin neurons do not express the leptin receptor [41]; therefore, the mechanism by which leptin signaling can be detected and transmitted to kisspeptin/GnRH neurons remains unclear.

Although the molecular mechanisms of many of these putative genes remain under investigation, they can be divided into four categories according to their presumed biological activities: (1) development of craniofacial and forebrain structures; (2) embryonic migration of GnRH neurons; (3) expression and secretion of GnRH; and (4) maintenance and homeostasis of GnRH neurons and gonadotrophs. These divisions are not mutually exclusive because the associated events are often interdependent and multiple signal pathways cross-talk with one another. Ironically, genes that affect human reproduction should not be conserved in evolution. Nonetheless, a recent study revealed that founder mutations in four known KS/CHH genes (GNRH1, TAC3, PROKR2, and GNRHR) are shared by multiple unrelated individuals and that one founder allele of PROKR2 originated 9,000 years ago, implying there might be an advantage associated with this ancient mutation [42].

DIAGNOSIS AND DIFFERENTIAL DIAGNOSIS

Although it is a relatively rare disorder, CHH/KS is the most common form of gonadotrophin deficiency, and it is about five times more prevalent in males. The reported frequencies vary across studies, ranging from 1 in 8,000 men and 40,000 women to 1 in 29,000 men and 130,000 women [43,44]; ORPHA suggests that an average figure of 3.75 in 100,000 people is appropriate (www.orpha.net). A diagnosis of KS is often difficult to make due to genetic heterogeneities and the broad spectrum of phenotypic presentations. Patients with CHH are commonly diagnosed in late adolescence or early adulthood. Delayed puberty is classically defined as the absence of virilization and testicular enlargement (testicular volume <4 mL) in conjunction with the lack of or low sperm production by age 14 years in males and as primary amenorrhea and the absence of breast development (Tanner stage I) by age 13 years in females [45]. Any signs of eunuchoidal properties along with either anosmia or hyposmia should confirm the diagnosis of KS. When either a micropenis (5% to 10% of cases) or cryptorchidism (30% of cases) is present in infant males in conjunction with a lack of neonatal activation of the HPG axis at 3 to 6 months (mini-puberty), an early diagnosis of CHH can be made and then confirmed by the hormonal profiling of serum gonadotropins, sex steroids, anti-Müllerian hormone (AMH), insulin-like 3 (INSL3), and inhibin B [46,47]. However, the occurrence of severe genital anomalies, such as hypospadias, likely suggests the presence of a human chorionic gonadotropin deficiency. Patients may also exhibit a variety of non-reproductive anomalies including craniofacial defects, such as cleft lip and/or palate and hypodontia, and eye defects, such as iris coloboma, ptosis, hypertelorism, and oculomotor palsy. Additionally, neurological symptoms, such as bimanual synkinesis, corpus callosum agenesis, and hearing impairments, and/or medical conditions, such as unilateral renal aplasia, may also be present [48].

Hypoplasia or aplasia of the OB and its tracts can be confirmed using high-resolution magnetic resonance imaging (MRI) but this does not always correlate with the clinical status of olfactory function; thus, a diagnosis of anosmia should be confirmed by formal smell testing and detailed questioning [49]. MRI scans are also useful during the CHH/KS examination to investigate the possible presence of any tumors or lesions in the hypothalamic and pituitary regions and to detect any problems in inner ear structures. To assess bone age and epiphyseal closure, bone densitometry can be used, whereas

abdominal and/or pelvic ultrasonography aid in the detection of dysmorphisms in the kidney, ovaries, uterus, or testes [43,48]. Low serum concentrations of FSH and LH, along with low testosterone (<3.7 nmol/L) and estradiol (<0.18 pg/mL) levels, are typically a consequence of decreased gonadotropin production [47]. Thus, a detailed assessment of anterior pituitary function must be undertaken to rule out other endocrinological conditions involving growth hormone and thyroid or adrenal functioning. Hyperprolactinemia may result in acquired HH (AHH) and, therefore, normal prolactin levels should also be confirmed [50].

Other complex syndromes present with clinical features that are similar to those of CHH or KS; these include combined pituitary hormone deficiency, septo-optic dysplasia, CHARGE syndrome, adrenal hypoplasia congenita with HH, Waardenburg syndrome, Bardet-Biedl syndrome, Gordon Holmes syndrome, Morning Glory syndrome, Hartsfield syndrome, and Dandy-Walker syndrome [8]. However, the most challenging differential diagnosis is that of constitutional delay of puberty and growth (CDPG). In the absence of anosmia, hormone profiling is not always fully informative regarding the confirmation of a diagnosis, although the combination of GnRH and human chorionic gonadotropin stimulation tests might help [51]. Signs that support a diagnosis of CHH include mild pubic and axillary hair growth due to the presence of adrenal androgens and the lack of a pubertal growth spurt while exhibiting steady linear growth. Thus, normal growth but longer arms and legs due to prolonged epiphyseal development may be suggestive of CHH, whereas individuals who are shorter than normal but have a proportional stature and delayed skeletal maturation while showing poor growth velocity are more likely to have CDPG [50]. Unlike CHH/KS, puberty is spontaneously initiated and eventually completed in patients with CDGP. Moreover, the presence of a micropenis or cryptorchidism, a cleft palate, sensorineural deafness, and anosmia are not observed in patients with CDGP. Nonetheless, no conclusive differential diagnostic tests are currently available for the differentiation of CHH/KS from CDPG.

Patients with CHARGE syndrome have both hypogonadism and OB aplasia/hypoplasia and can also share additional traits with KS patients, including a cleft lip or palate [52], which makes distinguishing this condition from KS extremely difficult. Potential systematic causes should also be eliminated prior to the confirmation of the diagnosis. Celiac disease, eating disorders, excessive exercise, and chronic diseases of the liver and/ or kidney can all lead to AHH. The possibility of hemochroma-

tosis, pituitary adenomas, and/or brain tumors should also be considered [8]. For further details regarding the diagnostic clues of CHH/KS, refer to a previously reported case series [48].

TREATMENT AND MANAGEMENT

CHH/KS can be managed more effectively if it is recognized during the early stages. The initial assessment of this disease typically relies on the physical examination and data obtained in primary clinics. Because these patients are often admitted by a variety of different clinics (endocrinology, pediatrics, neurosurgery, urology, obstetrics, and gynecology), a correct initial diagnosis and subsequent referrals are critical for appropriate follow-up visits and the systematic management of the disease on a long-term basis. Timely treatment to induce puberty can be also crucial for sexual, bone, and metabolic health and for the minimization of the psychological effects that can be associated with CHH/KS. The treatment regimen for CHH is principally determined by whether the goal is to develop secondary sexual characteristics (virilization or estrogenization) or to induce fertility as well. In most cases, fertility can be induced under specific regimens of testosterone treatment in males and estrogen/progesterone treatment in females, but only a few predictors have been identified [8]. In cases where gonadotrophs retain their functional secretory capability, the administration of exogenous pulsatile GnRH invariably induces the phasic release of LH and FSH and can restore puberty and fertility [50]. Patients typically require lifelong treatment to maintain normal sexual function, yet some patients sustain function of the gonads and remain fertile after testosterone withdrawal, which is called as "reversible" case [53]. Reversal of CHH may be more prevalent than appreciated. According to a retrospective study, the estimated lifetime incidence of the spontaneous recovery of reproductive function might be as high as 22% [54]. Interestingly, in these reversible cases, there is an enrichment of neurokinin B mutations, and the absence of the OB does not hinder the reversal. On the other hand, a subsequent relapse can also occur and, therefore, these patients require ongoing monitoring for reversal and/or relapse [54]. However, no predictors are currently available.

Recently, inhibin B, INSL3, AMH, and kisspeptin have emerged as potential biomarkers for the diagnosis and treatment of CHH [45-47]. Kisspeptin may also be a potentially useful therapeutic target since a male patient reversed his condition after receiving exogenous kisspeptin infusions [45]. Although earlier studies indicate that the neonatal administration of gonadotropins could be beneficial [44,50], further investigation is necessary to confirm its value for fertility outcomes and long-term effects. A comparative assessment of gonadotropin treatment during adolescence vs. adulthood and the benefits of prior treatment with androgens or FSH before pubertal induction are still under consideration [50].

Many studies have focused on testicular growth and gametogenesis in males [46,47], but the rate of pregnancy in females with CHH/KS has yet to be sufficiently investigated, although this may be partly due to rarity of the disease and the long-term follow-up assessments required for gathering such information. Patients with cleft lip and/or palate, hearing loss, skeletal defects, or cryptorchidism require surgical and specialist interventions early in life. CHH/KS patients also have an increased risk of metabolic problems such as type 2 diabetes [55]. In females, low estrogen levels can result in osteopenia/osteoporosis [50] and, thus, bone density should be monitored. Due to the congenital nature of the disease, patients with KS who exhibit anosmia or hyposmia do not complain of altered flavor perception [49]. However, due to the potential detrimental effects of KS on quality of life and mental health issues, patients with KS should be carefully advised of this disability.

It is also important to recognize the implications of fertility treatment in CHH/KS patients because it leads to the inevitable risk of passing on the mutated gene to the next generation; this is especially true for genes with an autosomal dominant inheritance that have a 50% chance of transmission. Thus, genetic counseling should be provided to enable informed medical and personal decisions that rest on a clear understanding of the risks involved in conception, which can be even more critical in the case of suspected oligogenicity [9,10,50]. Potential complications that may arise during pregnancy in female patients with certain anomalies should also be carefully assessed.

The major challenges associated with the treatment and management of CHH/KS include the appropriate timing of the treatment and how to promote adherence to treatment, especially during the transitioning of young adults from pediatric to adult care because any gaps can have considerable consequences. According to the consensus of the European Consortium on GnRH biology (COST Action BM1105, http://www.gnrhnetwork.eu/), a structured transition is preferable. Nonetheless, medical treatment provides only some aspects of the overall solution as many of the patients are emotionally traumatized, particularly by psychosexual issues and low self-esteem [9]. Thus, targeted psychosocial interventions, counseling, and peer support should aid in the amelioration of some of these issues.

CONCLUSIONS

Advancements in next-generation DNA sequencing technology hold much hope for more accurate detection and diagnostic procedures, personalized genetic counseling and screening, and tailored treatments for CHH/KS patients. Furthermore, future international multicenter trials that include large patient populations should aid in the development of an organized consensus for optimal management plans and fertility treatment options for these patients. Predictions based on genotype-phenotype correlations that consider oligogenicity, the identification of novel biomarkers, and the development of targeted therapies might also become realistic possibilities. Additionally, recent findings on the reversibility and oligogenicity of CHH/KS with variable phenotypes have challenged the conventional beliefs and current dogma associated with congenital defects. CHH/ KS was previously believed to be secondary hypogonadism due to the absence of GnRH function [4-7], but many of the genes associated with CHH/KS are broadly expressed in the HPG axis. Thus, the impact of the mutations may not be limited to hypothalamic GnRH neurons but may also contribute to primary defects in the pituitary or gonads downstream of GnRH receptor. Although several genes that ensure proper gonadal development have been identified, their links with CHH/ KS has not been established. Additionally, the reversibility of this condition underscores the plasticity of the GnRH neuroendocrine system and suggests that further research is necessary.

Over the years, the distinction between CHH and KS has become even more blurry, in part because the mutations of known CHH/KS genes have been identified in other congenital syndromes that include CHH features but are accompanied by different symptoms [8]. The assumption is that it may not be a simple "all-or-none" story but rather a complex scenario that involves multiple feedback mechanisms. A broad range of associated phenotypes found in multiple disorders with HH potentially indicate that these issues are caused by overlapping signal pathways with different tissue specificities. Therefore, the need to redefine these congenital disorders and revise the existing perceptions of them is becoming more apparent, and it will be necessary to implement stratified medical strategies for these patients in the future.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

This work has been supported by the Medical Research Council (MRC Grant Reference MR/L020378/1) and St. George's Medical School, University of London.

REFERENCES

- Maestre de San Juan A. Falta total de los nerviosolfactorios con anosmia en un individuo en quien existia una atrofia congenita de los testiculos y miembro viril. Siglo Med 1856;131:211.
- 2. Kallmann FJ, Schoenfeld WA, Barrera SE. The genetic aspects of primary eunuchoidism. Am J Ment Defic 1944; 158:203-36.
- 3. De Morsier G, Gauthier G. Olfacto-genital dysplasia. Pathol Biol (Paris) 1963;11:1267-72.
- 4. Wierman ME, Kiseljak-Vassiliades K, Tobet S. Gonadotropin-releasing hormone (GnRH) neuron migration: initiation, maintenance and cessation as critical steps to ensure normal reproductive function. Front Neuroendocrinol 2011; 32:43-52.
- 5. Forni PE, Wray S. GnRH, anosmia and hypogonadotropic hypogonadism: where are we? Front Neuroendocrinol 2015; 36:165-77.
- Schwarting GA, Wierman ME, Tobet SA. Gonadotropinreleasing hormone neuronal migration. Semin Reprod Med 2007;25:305-12.
- Schwanzel-Fukuda M, Pfaff DW. Origin of luteinizing hormone-releasing hormone neurons. Nature 1989;338:161-4.
- Boehm U, Bouloux PM, Dattani MT, de Roux N, Dode C, Dunkel L, et al. Expert consensus document: European Consensus Statement on congenital hypogonadotropic hypogonadism: pathogenesis, diagnosis and treatment. Nat Rev Endocrinol 2015;11:547-64.
- Au MG, Crowley WF Jr, Buck CL. Genetic counseling for isolated GnRH deficiency. Mol Cell Endocrinol 2011;346: 102-9.
- 10. Sykiotis GP, Plummer L, Hughes VA, Au M, Durrani S, Nayak-Young S, et al. Oligogenic basis of isolated gonadotropinreleasing hormone deficiency. Proc Natl Acad Sci U S A 2010;107:15140-4.
- 11. Matsuo T, Okamoto S, Izumi Y, Hosokawa A, Takegawa T, Fukui H, et al. A novel mutation of the KAL1 gene in monozygotic twins with Kallmann syndrome. Eur J Endocrinol 2000;143:783-7.

- 12. Pitteloud N, Quinton R, Pearce S, Raivio T, Acierno J, Dwyer A, et al. Digenic mutations account for variable phenotypes in idiopathic hypogonadotropic hypogonadism. J Clin Invest 2007:117:457-63.
- 13. Hanchate NK, Giacobini P, Lhuillier P, Parkash J, Espy C, Fouveaut C, et al. SEMA3A, a gene involved in axonal pathfinding, is mutated in patients with Kallmann syndrome. PLoS Genet 2012;8:e1002896.
- 14. Tornberg J, Sykiotis GP, Keefe K, Plummer L, Hoang X, Hall JE, et al. Heparan sulfate 6-O-sulfotransferase 1, a gene involved in extracellular sugar modifications, is mutated in patients with idiopathic hypogonadotrophic hypogonadism. Proc Natl Acad Sci U S A 2011;108:11524-9.
- 15. Canto P, Munguia P, Soderlund D, Castro JJ, Mendez JP. Genetic analysis in patients with Kallmann syndrome: coexistence of mutations in prokinetic receptor 2 and KAL1. J Androl 2009;30:41-5.
- 16. Leroy C, Fouveaut C, Leclercq S, Jacquemont S, Boullay HD, Lespinasse J, et al. Biallelic mutations in the prokineticin-2 gene in two sporadic cases of Kallmann syndrome. Eur J Hum Genet 2008;16:865-8.
- 17. Xu N, Kim HG, Bhagavath B, Cho SG, Lee JH, Ha K, et al. Nasal embryonic LHRH factor (NELF) mutations in patients with normosmic hypogonadotropic hypogonadism and Kallmann syndrome. Fertil Steril 2011;95:1613-20.e1-7.
- 18. Costa-Barbosa FA, Balasubramanian R, Keefe KW, Shaw ND, Al-Tassan N, Plummer L, et al. Prioritizing genetic testing in patients with Kallmann syndrome using clinical phenotypes. J Clin Endocrinol Metab 2013;98:E943-53.
- 19. Quinton R, Duke VM, Robertson A, Kirk JM, Matfin G, de Zoysa PA, et al. Idiopathic gonadotrophin deficiency: genetic questions addressed through phenotypic characterization. Clin Endocrinol (Oxf) 2001;55:163-74.
- 20. Zhao H, Kegg H, Grady S, Truong HT, Robinson ML, Baum M, et al. Role of fibroblast growth factor receptors 1 and 2 in the ureteric bud. Dev Biol 2004;276:403-15.
- 21. Miraoui H, Dwyer AA, Sykiotis GP, Plummer L, Chung W, Feng B, et al. Mutations in FGF17, IL17RD, DUSP6, SPRY4, and FLRT3 are identified in individuals with congenital hypogonadotropic hypogonadism. Am J Hum Genet 2013;92:725-43.
- 22. de Roux N, Young J, Misrahi M, Schaison G, Milgrom E. Loss of function mutations of the GnRH receptor: a new cause of hypogonadotropic hypogonadism. J Pediatr Endocrinol Metab 1999;12 Suppl 1:267-75.
- 23. de Roux N, Genin E, Carel JC, Matsuda F, Chaussain JL,

- Milgrom E. Hypogonadotropic hypogonadism due to loss of function of the KiSS1-derived peptide receptor GPR54. Proc Natl Acad Sci U S A 2003;100:10972-6.
- 24. Topaloglu AK, Reimann F, Guclu M, Yalin AS, Kotan LD, Porter KM, et al. TAC3 and TACR3 mutations in familial hypogonadotropic hypogonadism reveal a key role for Neurokinin B in the central control of reproduction. Nat Genet 2009;41:354-8.
- 25. Dode C, Teixeira L, Levilliers J, Fouveaut C, Bouchard P, Kottler ML, et al. Kallmann syndrome: mutations in the genes encoding prokineticin-2 and prokineticin receptor-2. PLoS Genet 2006;2:e175.
- 26. Watanabe Y, Inoue K, Okuyama-Yamamoto A, Nakai N, Nakatani J, Nibu K, et al. Fezfl is required for penetration of the basal lamina by olfactory axons to promote olfactory development. J Comp Neurol 2009;515:565-84.
- 27. Kim HG, Kurth I, Lan F, Meliciani I, Wenzel W, Eom SH, et al. Mutations in CHD7, encoding a chromatin-remodeling protein, cause idiopathic hypogonadotropic hypogonadism and Kallmann syndrome. Am J Hum Genet 2008:83: 511-9.
- 28. Achermann JC, Gu WX, Kotlar TJ, Meeks JJ, Sabacan LP, Seminara SB, et al. Mutational analysis of DAX1 in patients with hypogonadotropic hypogonadism or pubertal delay. J Clin Endocrinol Metab 1999;84:4497-500.
- 29. Pingault V, Bodereau V, Baral V, Marcos S, Watanabe Y, Chaoui A, et al. Loss-of-function mutations in SOX10 cause Kallmann syndrome with deafness. Am J Hum Genet 2013;92:707-24.
- 30. Farooqi IS, O'Rahilly S. Mutations in ligands and receptors of the leptin-melanocortin pathway that lead to obesity. Nat Clin Pract Endocrinol Metab 2008;4:569-77.
- 31. Jackson RS, Creemers JW, Ohagi S, Raffin-Sanson ML, Sanders L, Montague CT, et al. Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene. Nat Genet 1997;16:303-6.
- 32. Ballabio A, Parenti G, Tippett P, Mondello C, Di Maio S, Tenore A, et al. X-linked ichthyosis, due to steroid sulphatase deficiency, associated with Kallmann syndrome (hypogonadotropic hypogonadism and anosmia): linkage relationships with Xg and cloned DNA sequences from the distal short arm of the X chromosome. Hum Genet 1986;72:237-
- 33. Vermeulen S, Messiaen L, Scheir P, De Bie S, Speleman F, De Paepe A. Kallmann syndrome in a patient with congenital spherocytosis and an interstitial 8p11.2 deletion. Am J

EnM

- Med Genet 2002;108:315-8.
- 34. Falardeau J, Chung WC, Beenken A, Raivio T, Plummer L, Sidis Y, et al. Decreased FGF8 signaling causes deficiency of gonadotropin-releasing hormone in humans and mice. J Clin Invest 2008;118:2822-31.
- 35. Franco B, Guioli S, Pragliola A, Incerti B, Bardoni B, Tonlorenzi R, et al. A gene deleted in Kallmann's syndrome shares homology with neural cell adhesion and axonal pathfinding molecules. Nature 1991;353:529-36.
- 36. Hu Y, Guimond SE, Travers P, Cadman S, Hohenester E, Turnbull JE, et al. Novel mechanisms of fibroblast growth factor receptor 1 regulation by extracellular matrix protein anosmin-1. J Biol Chem 2009;284:29905-20.
- 37. Bulow HE, Berry KL, Topper LH, Peles E, Hobert O. Heparan sulfate proteoglycan-dependent induction of axon branching and axon misrouting by the Kallmann syndrome gene kal-1. Proc Natl Acad Sci U S A 2002;99:6346-51.
- Navarro VM, Tena-Sempere M. Neuroendocrine control by kisspeptins: role in metabolic regulation of fertility. Nat Rev Endocrinol 2012;8:40-53.
- 39. Mayer C, Acosta-Martinez M, Dubois SL, Wolfe A, Radovick S, Boehm U, et al. Timing and completion of puberty in female mice depend on estrogen receptor alpha-signaling in kisspeptin neurons. Proc Natl Acad Sci U S A 2010;107:22693-8.
- Pinilla L, Aguilar E, Dieguez C, Millar RP, Tena-Sempere M. Kisspeptins and reproduction: physiological roles and regulatory mechanisms. Physiol Rev 2012;92:1235-316.
- 41. Quennell JH, Mulligan AC, Tups A, Liu X, Phipps SJ, Kemp CJ, et al. Leptin indirectly regulates gonadotropin-releasing hormone neuronal function. Endocrinology 2009;150:2805-12.
- 42. Avbelj Stefanija M, Jeanpierre M, Sykiotis GP, Young J, Quinton R, Abreu AP, et al. An ancient founder mutation in PROKR2 impairs human reproduction. Hum Mol Genet 2012;21:4314-24.
- 43. Laitinen EM, Vaaralahti K, Tommiska J, Eklund E, Tervaniemi M, Valanne L, et al. Incidence, phenotypic features and molecular genetics of Kallmann syndrome in Finland. Orphanet J Rare Dis 2011;6:41.
- 44. Meczekalski B, Podfigurna-Stopa A, Smolarczyk R, Katulski K, Genazzani AR. Kallmann syndrome in women: from genes to diagnosis and treatment. Gynecol Endocrinol 2013;29:296-300.
- 45. Chan YM. Effects of kisspeptin on hormone secretion in humans. Adv Exp Med Biol 2013;784:89-112.

- 46. Kuiri-Hanninen T, Seuri R, Tyrvainen E, Turpeinen U, Hamalainen E, Stenman UH, et al. Increased activity of the hypothalamic-pituitary-testicular axis in infancy results in increased androgen action in premature boys. J Clin Endocrinol Metab 2011;96:98-105.
- 47. Andersson AM, Juul A, Petersen JH, Muller J, Groome NP, Skakkebaek NE. Serum inhibin B in healthy pubertal and adolescent boys: relation to age, stage of puberty, and follicle-stimulating hormone, luteinizing hormone, testosterone, and estradiol levels. J Clin Endocrinol Metab 1997;82:3976-81.
- 48. Kaplan JD, Bernstein JA, Kwan A, Hudgins L. Clues to an early diagnosis of Kallmann syndrome. Am J Med Genet A 2010:152A:2796-801.
- 49. Maione L, Cantone E, Nettore IC, Cerbone G, De Brasi D, Maione N, et al. Flavor perception test: evaluation in patients with Kallmann syndrome. Endocrine 2015 Jul 25 [Epub]. http://dx.doi.org/10.1007/s12020-015-0690-y.
- Fechner A, Fong S, McGovern P. A review of Kallmann syndrome: genetics, pathophysiology, and clinical management. Obstet Gynecol Surv 2008;63:189-94.
- 51. Segal TY, Mehta A, Anazodo A, Hindmarsh PC, Dattani MT. Role of gonadotropin-releasing hormone and human chorionic gonadotropin stimulation tests in differentiating patients with hypogonadotropic hypogonadism from those with constitutional delay of growth and puberty. J Clin Endocrinol Metab 2009:94:780-5.
- 52. Pinto G, Abadie V, Mesnage R, Blustajn J, Cabrol S, Amiel J, et al. CHARGE syndrome includes hypogonadotropic hypogonadism and abnormal olfactory bulb development. J Clin Endocrinol Metab 2005;90:5621-6.
- 53. Pitteloud N, Acierno JS Jr, Meysing AU, Dwyer AA, Hayes FJ, Crowley WF Jr. Reversible kallmann syndrome, delayed puberty, and isolated anosmia occurring in a single family with a mutation in the fibroblast growth factor receptor 1 gene. J Clin Endocrinol Metab 2005;90:1317-22.
- 54. Sidhoum VF, Chan YM, Lippincott MF, Balasubramanian R, Quinton R, Plummer L, et al. Reversal and relapse of hypogonadotropic hypogonadism: resilience and fragility of the reproductive neuroendocrine system. J Clin Endocrinol Metab 2014;99:861-70.
- Ding EL, Song Y, Malik VS, Liu S. Sex differences of endogenous sex hormones and risk of type 2 diabetes: a systematic review and meta-analysis. JAMA 2006;295:1288-99.